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Control of acetolactate synthase inhibitor/glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*) in isoxaflutole/glufosinate/glyphosate-resistant soybean

Jasmine Mausbach[1], Suat Irmak[2], Debalin Sarangi[3], John Lindquist[4] and Amit J. Jhala[5]

[1]Graduate Research Assistant, Department of Agronomy and Horticulture, University of Nebraska–Lincoln, Lincoln, NE, USA; [2]Harold W. Eberhard Distinguished Professor, Department of Biological Systems Engineering, University of Nebraska–Lincoln, Lincoln, NE, USA; [3]Assistant Professor, Department of Agronomy and Plant Genetics, 411 Borlaug Hall, University of Minnesota, St. Paul, MN, USA; [4]Professor, Department of Agronomy and Horticulture, University of Nebraska–Lincoln, Lincoln, NE, USA and [5]Associate Professor, Department of Agronomy and Horticulture, University of Nebraska–Lincoln, Lincoln, NE, USA

Research Article

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Author for correspondence:
Amit J. Jhala, Department of Agronomy and Horticulture, University of Nebraska–Lincoln, 279 Plant Science Hall, PO Box 830915, Lincoln, NE 68583. Email: Amit.Jhala@unl.edu

Abstract

Palmer amaranth is the most problematic and troublesome weed in agronomic cropping systems in the United States. Acetolactate synthase (ALS) inhibitor and glyphosate-resistant (GR) Palmer amaranth has been confirmed in Nebraska and it is widespread in several counties. Soybean resistant to isoxaflutole/glufosinate/glyphosate has been developed that provides additional herbicide site of action for control of herbicide-resistant weeds. The objectives of this study were to evaluate herbicide programs for control of ALS inhibitor/GR Palmer amaranth and their effect on Palmer amaranth density and biomass, as well as soybean injury and yield in isoxaflutole/glufosinate/glyphosate–resistant soybean. Field experiments were conducted in a grower's field infested with ALS inhibitor and GR Palmer amaranth near Carleton, Nebraska, in 2018 and 2019. Isoxaflutole applied alone or mixed with sulfentrazone/pyroxasulfone, flumioxazin/pyroxasulfone, or imazethapyr/saflufenacil/pyroxasulfone provided similar control (86%–99%) of Palmer amaranth 21 d after PRE (DAPRE). At 14 d after early-POST (DAEPOST), isoxaflutole applied PRE and PRE followed by (fb) POST controlled Palmer amaranth by 10% to 63% compared to 75% to 96% control with glufosinate applied EPOST in both years. A PRE herbicide fb glufosinate controlled Palmer amaranth 80% to 99% 21 d after late-POST (DALPOST) in 2018, and reduced density 89% to 100% in 2018 and 58% to 100% in 2019 at 14 DAEPOST. No soybean injury was observed from any of the herbicide programs tested in this study. Soybean yield in 2019 was relatively higher due to higher precipitation compared with 2018 with generally no differences between herbicide programs. This research indicates that herbicide programs are available for effective control of ALS inhibitor/GR Palmer amaranth in isoxaflutole/glufosinate/glyphosate-resistant soybean.

Introduction

Palmer amaranth is a summer annual broadleaf weed belonging to the Amaranthaceae family and is one of dioecious species among pigweeds (Steckel 2007). Human activities in the 20th century such as agricultural development, within- and between-field operations, and seed and equipment transportation have led Palmer amaranth to spread to the northern United States (Culpepper 2006). Since the first report of Palmer amaranth in Virginia in 1915 beyond its native habitat in the southwest United States, it has become one of the most problematic and troublesome weeds in agronomic cropping systems in the United States (Culpepper et al. 2010). Being dioecious, Palmer amaranth is an obligate outcrossing, wild pollinated species (Sosnoskie et al. 2012), resulting in wide genetic diversity (Jhala et al. 2021; Oliveira et al. 2018). High photosynthetic rate along with diaheliotropic movement (i.e., leaves orienting themselves perpendicular to incoming sunlight to intercept radiant energy and light) allow Palmer amaranth to fix carbon at higher rate, resulting in rapid growth (Ehleringer and Forseth 1980; Ehleringer 1985). In a 2-yr field study in Kansas, Horak and Loughin (2000) reported that Palmer amaranth had the highest plant dry weight, leaf area, water-use efficiency, and growth rate (0.10 to 0.21 cm per growing degree day) compared to redroot pigweed (*Amaranthus retroflexus* L.), tumble pigweed (*Amaranthus albus* L.), and waterhemp [*Amaranthus tuberculatus* (Moq.) Sauer].

Depending on environmental conditions, Palmer amaranth typically flowers during September and October, although decreasing day length can accelerate the flowering process

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(Bond and Oliver 2006). Female Palmer amaranth plants are prolific seed producers, even under competition with agronomic crops (Massinga et al. 2001). Seeds are usually dispersed by gravity forces; however, dispersal via irrigation, plowing, mowing, harvesting, birds, and mammals has been documented (Costea et al. 2004, 2005). Because Palmer amaranth's prolific seed production and aggressive growth habit make it difficult to control in agronomic cropping systems (Horak and Loughin 2000; Ward et al. 2013), it is vital to control Palmer amaranth early in the growing season by integrating mechanical, cultural, and chemical practices, including PRE herbicides with multiple sites of action (SOAs; de Sanctis et al. 2021; Norsworthy et al. 2012).

Globally, glyphosate is the most widely used agricultural pesticide and is used extensively in glyphosate-resistant (GR) canola (*Brassica napus* L.), corn (*Zea mays* L.), cotton (*Gossypium hirsutum* L.), sugarbeet (*Beta vulgaris var. saccharifera*), and soybean in the United States (Heap and Duke 2018). Since the commercialization of GR crops, particularly GR corn and soybean in the midwestern United States and GR cotton in the southern United States, continuous use of glyphosate multiple times in a year, along with a decline in the use of residual herbicides (Culpepper 2006; Young 2006), has resulted in the evolution of GR weeds (Beckie 2006). As of 2020, 50 weeds have been confirmed resistant to glyphosate worldwide (Heap 2021), including six broadleaf weeds such as common ragweed (*Ambrosia artemisiifolia* L.), giant ragweed (*Ambrosia trifida* L.), kochia [*Bassia scoparia* (L.) A. J. Scott], horseweed (*Erigeron canadensis* L.), waterhemp, and Palmer amaranth in Nebraska (Jhala 2017a).

Glyphosate-resistant Palmer amaranth was first confirmed in Georgia in 2004 (Culpepper et al. 2006), and since then has been confirmed in 28 states in the United States (Heap 2021). Palmer amaranth has evolved resistance to herbicides from at least eight SOA groups: microtubule-, acetolactate synthase (ALS)–, 5-enolpyruvyl-shikimate-3-phosphate synthase (EPSPS)–, photosystem II (PS II)–, hydroxyphenylpyruvate dioxygenase (HPPD)–, very long chain fatty acid (VLCFA)–, protoporphyrinogen oxidase (PPO)–, and synthetic auxin inhibitors (Heap 2021). Palmer amaranth resistant to glufosinate was recently confirmed in Arkansas (Barber et al. 2021). Some populations of Palmer amaranth have also been found to have resistance to multiple herbicides in a few states, including ALS inhibitor/glyphosate resistance in Nebraska (Chahal et al. 2017), atrazine/HPPD inhibitor resistance in Nebraska (Jhala et al. 2014), and 2,4-D/ALS inhibitor/atrazine/glyphosate/HPPD inhibitor resistance in Kansas (Kumar et al. 2019).

Palmer amaranth resistance to ALS inhibitors was first confirmed in Kansas in 1994 and since then has been confirmed in 14 states (Heap 2021; Sprague et al. 1997). Isoxaflutole, an HPPD-inhibiting PRE corn herbicide, has been available commercially since 1998 (Spaunhorst and Johnson 2016). The recently available isoxaflutole/glufosinate/glyphosate-resistant soybean (LibertyLink GT27™) provides an opportunity to use isoxaflutole applied PRE alone or in mixture with other residual herbicides for early-season weed control. Glufosinate is a contact, POST herbicide for control of emerged broadleaf and grass weeds (Jhala et al. 2013). It is a nonselective herbicide traditionally used to control weeds in fruit and nut orchards and non-crop areas (Jhala et al. 2013). Norsworthy et al. (2008) reported 99% control of GR Palmer amaranth with glufosinate. Hoffner et al. (2012) found that glufosinate applied early-POST (EPOST) controlled Palmer amaranth by 73% compared to 76% control with glufosinate applied EPOST followed by a late-POST (LOST). Wiesbrook et al. (2001) found that glufosinate applied sequentially improved control of broadleaf weeds over a single application. Glufosinate applied EPOST resulted in 71% control, and a sequential LPOST application provided 76% control of GR waterhemp in glufosinate-resistant soybean in Nebraska (Jhala et al. 2017). An additional option for POST control of GR Palmer amaranth in glufosinate-resistant soybean is glufosinate mixed with residual herbicides such as acetochlor, pyroxasulfone, or *S*-metolachlor (Aulakh and Jhala 2015). This mixture provides foliar and residual control of Palmer amaranth through overlapping residual activity.

ALS inhibitor and/or GR Palmer amaranth has been observed in several corn/soybean production fields in south-central and west-central Nebraska, in addition to alfalfa (*Medicago sativa* L.), corn, and sugarbeet fields in western Nebraska (Vieira et al. 2018). To address the growing need to control GR weeds in cropping systems, multiple herbicide–resistant soybean traits have been developed. For example, isoxaflutole/glufosinate/glyphosate-resistant soybean has been developed to provide an additional herbicide SOA for control of herbicide-resistant weeds, primarily GR weeds; however, herbicide programs need to be developed and tested that provide season-long control of GR Palmer amaranth in this multiple herbicide-resistant soybean. The objectives of this research were to 1) evaluate isoxaflutole- and glufosinate-based herbicide programs for control of ALS inhibitor and GR Palmer amaranth in isoxaflutole/glufosinate/glyphosate-resistant soybean; and 2) evaluate the effect of herbicide programs on Palmer amaranth density and biomass, as well as soybean injury and grain yield.

Materials and Methods

Field Experiments

Field experiments were conducted in 2018 and 2019 in a grower's field near Carleton, NE (40.30°N, 97.67°W). The field had a GR corn-soybean rotation with reliance on glyphosate for weed control in a no-till production system for the last 10 yr and confirmed to have an ALS inhibitor and GR Palmer amaranth (Chahal et al. 2017). Hereafter, we refer to this as multiple herbicide-resistant (MHR) Palmer amaranth. The soil at the experimental site was silt loam (montmorillonitic, mesic, Pachic Argiustolls), pH 6.0; and 19% sand, 63% silt, 18% clay, and 2.6% organic matter content. Winter annual weeds were controlled with glyphosate at 900 g ae ha^{-1} + 2,4-D ester at 560 g ae ha^{-1} + liquid ammonium sulfate 3% vol/vol 2 wk prior to establishing an experiment. A soybean cultivar resistant to isoxaflutole/glufosinate/glyphosate was planted in a no-till seedbed at 345,800 seeds ha^{-1} in rows spaced 76 cm apart. Soybean was planted on May 10, 2018, and May 6, 2019. Individual experimental plot dimensions were 3 m wide by 9 m long. The experimental site was in a rainfed environment with no supplemental irrigation. The precipitation received during both years during crop growing season are listed (Table 1).

Treatments were arranged in a randomized complete block design with four replications. Herbicide programs evaluated to control MHR Palmer amaranth consisted of PRE, EPOST, LPOST, and/or PRE fb POST herbicide programs (Table 2). A nontreated control was included for comparison. Herbicides were applied with a handheld CO_2-pressurized backpack sprayer equipped with AIXR 110015 flat-fan nozzles (TeeJet® Technologies, Wheaton, IL) calibrated to deliver a 140 L ha^{-1} flow rate at 276 kPa at a constant speed of 4.8 km h^{-1}. Glufosinate was mixed with liquid ammonium sulfate at 3% vol/vol (Anonymous 2017) and was applied with XR 11005

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Table 1. Monthly mean air temperature and total precipitation during the 2018 and 2019 growing seasons (March to October), along with the 30-yr average at the research site near Carleton, Nebraska.[a]

	Mean air temperature			Total precipitation		
Month	2018	2019	30-yr average	2018	2019	30-yr average
	C			mm		
March	4.5	1.1	4.6	23.6	85.6	45.2
April	5.9	11.8	10.6	26.4	16.0	66.3
May	20.6	14.6	16.4	78.0	172.7	135.4
June	25.0	21.8	22.3	96.0	153.2	115.1
July	24.7	25.1	24.9	95.5	137.2	105.2
August	23.3	23.1	23.7	92.2	154.9	94.0
September	20.6	22.6	19.1	153.4	120.4	66.0
October	10.6	9.6	12.1	99.8	118.1	58.4

[a]Data were obtained from National Oceanic and Atmospheric Administration (NOAA 2019).

flat-fan nozzles (TeeJet® Technologies). The PRE herbicides were applied after soybean planting on the same day (i.e., May 10) in 2018, and 4 d after soybean planting (i.e., May 10) in 2019. The EPOST herbicides were applied 31 d after PRE (DAPRE) herbicides were applied. Palmer amaranth was 1 to 8 cm tall depending on herbicide program. Soybean was at the first to second trifoliate (V1 to V2 growth stage). The LPOST herbicides were applied 20 to 22 DAEPOST herbicide application. Palmer amaranth was 8 to 25 cm tall depending on the herbicide program. Palmer amaranth plant height was variable because new plants had emerged and some plants had been partially controlled by the EPOST herbicide.

Data Collection

Palmer amaranth control was assessed visually at 21 DAPRE, 14 DAEPOST, and 14 and 28 DALPOST herbicide applications on a scale of 0% to 100% (0% indicating no control of Palmer amaranth and 100% indicating complete control). Palmer amaranth densities were recorded 21 DAPRE, 14 DAEPOST, 14 DALPOST, and 28 DALPOST by counting the number of Palmer amaranth plants in one 0.5-m^2 quadrat placed randomly between two center soybean rows in each plot. Soybean injury was assessed visually at 14 DAPRE, 14 DAEPOST, 14 and 28 DALPOST herbicide applications based on a scale of 0% to 100% (0% indicating no soybean injury and 100% indicating complete plant death). Palmer amaranth plants counted during density ratings were clipped at the soil surface, placed into paper bags, then placed in an oven at 65 C until they reached a constant weight. Aboveground biomass was converted into percent biomass reduction and was compared with the nontreated control using the following equation (Wortman 2014):

$$\%\,\text{Biomass reduction} = \left[\frac{\overline{C} - B}{\overline{C}}\right] * 100$$

where $\overline{C}$ is the biomass of the nontreated control and B is the biomass of an individual treatment plot. Soybean was harvested from the center two rows in each plot using a plot combine. Grain yield was adjusted to 13% moisture content and converted into kilograms per hectare.

Statistical Analysis

Data were subjected to ANOVA using the MIXED procedure in SAS version 9.3 (SAS Institute Inc, Cary, NC). Data were tested for normality with the use of UNIVARIATE procedure. Palmer amaranth control, density, and biomass data were arcsine square root–transformed before analysis; however, back-transformed data are presented with the mean separation based on the transformed data. Year and herbicide treatments were considered fixed effects, while replication was considered a random effect in the model. If year-by-treatment was nonsignificant, data from both years were combined. However, if the year-by-treatment interaction was significant, data were analyzed separately by year. Where the ANOVA indicated treatment effects were significant, means were separated at $P \leq 0.05$ using Tukey Kramer's pairwise comparison test.

Results and Discussion

Year-by-treatment interaction for MHR Palmer amaranth control 21 DAPRE was not significant ($P > 0.05$); therefore, data were combined for both years, while Palmer amaranth control estimates 14 DAEPOST and 28 DALPOST, Palmer amaranth density, and soybean yield were significant ($P < 0.05$); therefore, data were presented separately for both years. No soybean injury was observed from any herbicide program (data not shown), indicating that the herbicides evaluated in this study are safe to use in isoxaflutole/glufosinate/glyphosate–resistant soybean when applied according to label instructions. Schultz et al. (2015) also reported that isoxaflutole is safe to use in isoxaflutole-resistant soybean.

Temperature and Precipitation

The 2018 growing season started off warmer than average, with temperatures of 20.6 and 25.0 C for May and June, respectively, compared with 14.8 C and 21.8 C in 2019 (Table 1). Monthly precipitation varied from the 30-yr average in both years. Below-average precipitation occurred in 2018, with 78 and 96 mm in May and June, respectively, compared with the 30-yr average of 135 and 115 mm, whereas above-average precipitation was observed throughout the 2019 growing season (Table 1).

Palmer Amaranth Control

The PRE herbicides evaluated in this study controlled MHR Palmer amaranth by 86% to 99% 21 DAPRE (Table 3). Although statistically similar with other PRE herbicides, pyroxasulfone/sulfentrazone, flumioxazin/pyroxasulfone, and imazethapyr/pyroxasulfone/saflufenacil controlled Palmer amaranth by 97% to 99%. The contribution of the ALS-inhibiting herbicide (i.e., imazethapyr) was minimal; rather, the VLCFA inhibitor (i.e., pyroxasulfone) and PPO-inhibitor (i.e., saflufenacil) primarily contributed to the control. Shyam et al. (2021) reported similar findings 14 DAPRE with imazethapyr/pyroxasulfone/saflufenacil, when Palmer amaranth control ranged from 87% to 97% in a 2-yr study in 2,4-D choline/glufosinate/glyphosate-resistant soybean. Sarangi and Jhala (2019) reported at least 98% Palmer amaranth control 14 and 28 DAPRE with imazethapyr/dimethenamid-P/saflufenacil and flumioxazin/pyroxasulfone. Isoxaflutole applied PRE controlled Palmer amaranth by 86% to 89% 21 DAPRE (Table 3); however, variable control of Palmer amaranth has been reported with isoxaflutole in the literature. Meyer et al. (2016) and Johnson et al. (2012) reported at least 87% Palmer amaranth control with isoxaflutole 28 DAPRE. In contrast, Spaunhorst and Johnson (2016) reported GR Palmer amaranth control of 57% to 70% 21 DAPRE. The higher control occurred in a higher rainfall year, indicating the importance of moisture for herbicide activation (Spaunhorst and Johnson 2016). Isoxaflutole requires 12.7 to

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Table 2. Herbicides, application timings, and rates used for control of acetolactate synthase inhibitor and glyphosate-resistant Palmer amaranth in isoxaflutole/glufosinate/glyphosate–resistant soybean in field experiments conducted in 2018 and 2019.

Herbicide program[a]	Trade name	Application timing	Rate	Manufacturer
			g ae or ai ha^{-1}	
Isoxaflutole	Alite 27	PRE	105	BASF
Isoxaflutole fb isoxaflutole	Alite 27 fb Alite 27	PRE fb early POST	105, 105	BASF, BASF
Glufosinate	Liberty	Early POST	657	BASF
Glufosinate fb glufosinate	Liberty fb Liberty	Early POST fb late POST	657, 657	BASF, BASF
Isoxaflutole fb glufosinate	Alite 27 fb Liberty	PRE fb early POST	105, 657	BASF, BASF
Pyroxasulfone/sulfentrazone fb glufosinate	Authority Supreme fb Liberty	PRE fb early POST	292, 657	FMC, BASF
Pyroxasulfone/sulfentrazone + isoxaflutole fb glufosinate	Authority Supreme + Alite 27 fb Liberty	PRE fb early POST	292 + 105, 657	FMC + BASF, BASF
Flumioxazin/pyroxasulfone fb glufosinate	Fierce fb Liberty	PRE fb early POST	160, 657	Valent, BASF
Flumioxazin/pyroxasulfone + isoxaflutole fb glufosinate	Fierce + Alite 27 fb Liberty	PRE fb early POST	160 + 105, 657	Valent + BASF, BASF
Imazethapyr/pyroxasulfone/saflufenacil fb glufosinate	Zidua PRO fb Liberty	PRE fb early POST	215, 657	BASF, BASF
Imazethapyr/pyroxasulfone/saflufenacil + isoxaflutole fb glufosinate	Zidua PRO + Alite 27 fb Liberty	PRE fb early POST	215 + 105, 657	BASF, BASF
Isoxaflutole + glufosinate	Alite 27 + Liberty	Early POST	105 + 657	BASF
Isoxaflutole fb glufosinate fb glufosinate	Alite 27 fb Liberty fb Liberty	PRE fb early POST fb late POST	105, 657, 657	BASF, BASF, BASF
Isoxaflutole + glufosinate fb isoxaflutole + glufosinate	Alite 27 + Liberty fb Alite 27 + Liberty	Early POST fb late POST	105 + 657, 105 + 657	BASF, BASF
Glyphosate fb glyphosate	Roundup PowerMAX fb Roundup PowerMAX	Early POST fb late POST	1,260, 1,260	Bayer CropScience

[a]Glufosinate was mixed with ammonium sulfate (DSM Chemicals North America Inc., Augusta, GA) at 4.2 kg ha^{-1}.

25.4 mm of irrigation or rain to activate, although too much water can cause the herbicide to become diluted and leach, thus reducing its efficacy (Jhala 2017b). If moisture is adequate, isoxaflutole can provide 14 to 21 d of residual activity for Palmer amaranth control (Chahal et al. 2015).

Palmer amaranth control varied between years with a PRE fb EPOST herbicide programs (Table 3). Glufosinate applied alone controlled MHR Palmer amaranth 95% to 96% in 2018 and 75% in 2019. Glufosinate mixed with isoxaflutole controlled Palmer amaranth 92% to 95% in 2018 and 85% to 94% in 2019 (Table 3). Shyam et al. (2021) reported 88% Palmer amaranth control 14 DAEPOST with glufosinate. Conversely, Chahal and Jhala (2015) found that glufosinate in single and sequential applications provided 53% to 76% and 56% to 77% waterhemp control, respectively. Sequential glyphosate applications provided no control of MHR Palmer amaranth in this study, indicating that the population is highly resistant to glyphosate (Table 3). Chahal et al. (2017) reported 37-fold to 40-fold level of glyphosate resistance in MHR Palmer amaranth at this research site; therefore, no control with glyphosate was expected.

At 28 DALPOST, isoxaflutole applied PRE or in sequential applications (PRE fb EPOST) controlled MHR Palmer amaranth by 10% and 53% in 2018, respectively, while providing no control in 2019 (Table 3). This indicates that isoxaflutole applied alone at 105 g ai ha^{-1} will not provide effective control later in the growing season and that mixture with other herbicide(s) is needed to achieve economically acceptable control. In this study isoxaflutole was applied at 105 g ai ha^{-1}; however, it can be applied in a range of 140 to 210 g ai ha^{-1} in a single application with a season maximum of 210 g ai ha^{-1} (Anonymous 2020). Relatively lower use rate in this study is because the study was conducted before isoxaflutole label approved in 2020. In addition, isoxaflutole is primarily a residual herbicide with limited foliar activity; therefore, effective control of emerged Palmer amaranth at the time of application should not be expected. Janak and Grichar (2016) reported similar findings of 51% Palmer amaranth control with a single application of isoxaflutole 101 DAPRE. When mixed with metribuzin, isoxaflutole has been shown to provide 97% to 98% control of redroot pigweed and Powell amaranth (*Amaranthus powellii* S. Watson; Smith et al. 2019). With the exception of isoxaflutole, PRE fb POST herbicide programs provided 80% to 99% MHR Palmer amaranth control in 2018 and 78% to 99% control in 2019 at 28 DALPOST (Table 3). Whitaker et al. (2010) reported greater than 80% late-season control of GR Palmer amaranth with flumioxazin/*S*-metolachlor applied PRE fb fomesafen, although less than 30% late-season control was achieved with flumioxazin/*S*-metolachlor without fomesafen applied POST. A single herbicide application is less likely to provide a season-long control of Palmer amaranth and a PRE followed by a POST herbicide program is required for effective control and reducing Palmer amaranth seedbank (Norsworthy et al. 2012).

Palmer Amaranth Density and Biomass

Palmer amaranth density and biomass were affected by herbicide programs (Table 4). At 14 DAEPOST isoxaflutole reduced MHR Palmer amaranth density 0% and 48% in 2018 and 2019, respectively, whereas isoxaflutole applied PRE fb EPOST reduced density 49% and 53% in 2018 and 2019, respectively. Similarly, Meyer et al. (2016) reported 62% Palmer amaranth density reduction with isoxaflutole applied PRE. Meyer et al. (2015) reported 78% to 93% Palmer amaranth density reduction with flumioxazin/pyroxasulfone in soybean in a multiyear, multistate study, while Sarangi et al. (2017) reported 91% and 98% density reduction of GR waterhemp with flumioxazin/pyroxasulfone and imazethapyr/dimethenamid-P/saflufenacil, respectively.

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Table 3. Effect of herbicide programs on acetolactate synthase inhibitor and glyphosate-resistant Palmer amaranth control in isoxaflutole/glufosinate/glyphosate-resistant soybean 21 d after PRE, 14 d after early-POST, and 28 d after late-POST herbicide application in field experiments conducted in 2018 and 2019.[a,b,c]

Herbicide program	Application timing	Rate	Palmer amaranth control[d]				
			21 DAPRE[e]	14 DAEPOST		28 DALPOST	
		g ae or ai ha^{-1}	%				
			2018/2019	2018	2019	2018	2019
Isoxaflutole	PRE	105	89 a	41 c	38 c	10 d	0 e
Isoxaflutole fb isoxaflutole	PRE fb early POST	105, 105	86 a	63 b	10 d	53 c	0 e
Glufosinate	Early POST	657	-[e]	95 a	75 b	80 ab	36 cd
Glufosinate fb glufosinate	Early POST fb late POST	657, 657	-[e]	96 a	75 b	99 a	85 ab
Isoxaflutole fb glufosinate	PRE fb early POST	105, 657	88 a	97 a	95 ab	80 ab	34 cd
Pyroxasulfone/sulfentrazone fb glufosinate	PRE fb early POST	292, 657	98 a	98 a	99 a	91 ab	89 ab
Pyroxasulfone/sulfentrazone + isoxaflutole fb glufosinate	PRE fb early POST	292+105, 657	99 a	96 a	99 a	95 ab	88 ab
Flumioxazin/pyroxasulfone fb glufosinate	PRE fb early POST	160, 657	97 a	99 a	99 a	99 a	93 ab
Flumioxazin/pyroxasulfone + isoxaflutole fb glufosinate	PRE fb early POST	160+105, 657	95 a	99 a	99 a	97 ab	91 ab
Imazethapyr/pyroxasulfone/saflufenacil fb glufosinate	PRE fb early POST	215, 657	99 a	99 a	98 a	98 a	78 ab
Imazethapyr/pyroxasulfone/saflufenacil + isoxaflutole fb glufosinate	PRE fb early POST	215+105, 657	92 a	99 a	98 a	93 ab	61 bc
Isoxaflutole + glufosinate	Early POST	105+657	-[e]	95 a	85 ab	76 b	29 de
Isoxaflutole fb glufosinate fb glufosinate	PRE fb early POST fb late POST	105, 657, 657	88 a	94 a	92 ab	97 ab	93 a
Isoxaflutole + glufosinate fb isoxaflutole + glufosinate	Early POST fb late POST	105+657, 105+657	-[e]	92 a	94 ab	95 ab	89 ab
Glyphosate fb glyphosate	Early POST fb late POST	1,260, 1,260	-[e]	0 d	0 d	0 d	0 d

[a]Year-by-treatment interaction for Palmer amaranth control 14 DAPRE was not significant; therefore, data were combined across years.
[b]Year-by-treatment interaction for Palmer amaranth control 14 DAEPOST and 28 DALPOST was significant; therefore, data are presented separately for both years.
[c]Abbreviations: DAEPRE, days after PRE herbicide application, DAEPOST, days after early-POST herbicide application; DALPOST, days after late-POST herbicide application; fb, followed by.
[d]Means presented within each column with no common letter(s) are significantly different according to Fisher's protected LSD test at $P \leq 0.05$.
[e]POST herbicides were not applied at the time of evaluation 21 DAPRE.

Herbicides applied PRE fb glufosinate reduced MHR Palmer amaranth density by at least 87% in 2018 and 2019. Similar findings were reported by Shyam et al. (2021) and Norsworthy et al. (2016). Glufosinate applied alone reduced MHR Palmer amaranth density 89% and 58% in 2018 and 2019, respectively, whereas glufosinate mixed with isoxaflutole reduced density by 63% to 100% in 2018, and 85% to 94% in 2019 (Table 4). Chahal and Jhala (2015) reported 50% waterhemp density reduction with glufosinate applied EPOST; and 83% density reduction with glufosinate applied EPOST fb LPOST 45 DALPOST in glufosinate-resistant soybean in Nebraska.

At 14 DALPOST in 2019, PRE herbicide fb glufosinate applied EPOST reduced MHR Palmer amaranth biomass by 49% to 97% compared to 95% biomass reduction with glufosinate applied LPOST (Table 4). Aulakh and Jhala (2015) reported 79% to 88% weed biomass reduction with dimethenamid-P/saflufenacil, or imazethapyr/sulfentrazone fb glufosinate. Shyam et al. (2021) reported 100% Palmer amaranth biomass reduction with imazethapyr/pyroxasulfone/saflufenacil fb glufosinate and 99% biomass with glufosinate applied EPOST followed by LPOST in 2,4-D choline/glufosinate/glyphosate–resistant soybean. Single or sequential applications of isoxaflutole resulted in no biomass reduction due to poor Palmer amaranth control (Table 4). Chahal and Jhala (2015) reported 80% to 91% and 92% to 95% biomass reduction with glufosinate applied in single and sequential applications, respectively, in glufosinate-resistant soybean. Thus, a PRE herbicide with multiple SOAs fb glufosinate has consistently provided >90% control of Palmer amaranth and >90% density and biomass reduction in most studies.

Soybean Yield

Year-by-treatment interaction was significant ($P < 0.05$); therefore, yield data are presented separately for both years (Table 4). Soybean yield in 2019 was higher compared to 2018 due to higher precipitation in 2019 that provided sufficient moisture for soybean growth and development (Table 1). Isoxaflutole mixed with pyroxasulfone/sulfentrazone applied PRE fb glufosinate had soybean grain yield of 2,290 kg ha^{-1} in 2018, and it was comparable with several herbicide programs (Table 4). In 2019, several herbicide programs resulted in similar soybean yield in the range of 3,140 to 4,282 kg ha^{-1} (Table 4). Shyam et al. (2021) reported soybean yields with similar PRE herbicides used in combination with glufosinate.

Practical Implications

A new soybean trait resistant to isoxaflutole/glufosinate/glyphosate has been available commercially since the 2019 growing season in the United States. Results of this study suggest that herbicide programs are available for effective control of MHR Palmer amaranth in isoxaflutole/glufosinate/glyphosate–resistant soybean. No soybean injury was observed with any of the herbicide programs evaluated in this study, including isoxaflutole applied in sequential applications. Isoxaflutole (Alite™ 27) was registered in 2020 for application in isoxaflutole-resistant soybean; however, use of this herbicide is limited to certain counties in a few states. For example, isoxaflutole (Alite™ 27) is labeled for application in only four southwest counties (Chase, Dundy, Hitchcock, and Red Willow) in Nebraska (Anonymous 2020). In addition, isoxaflutole cannot be applied on coarse-textured soils (e.g., sandy, sandy loam, loamy sand) with less than 1.5% organic matter content, limiting the use of this herbicide. The majority of soybean in Nebraska is grown in eastern Nebraska, so although growers can plant isoxaflutole/glufosinate/glyphosate–resistant soybean in this region, they cannot use isoxaflutole (Alite™ 27) due to label restriction (Anonymous 2020). Therefore, adoption of soybean resistant to isoxaflutole/glufosinate/glyphosate in Nebraska will likely be very limited.

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Table 4. Effect of herbicide programs on glyphosate-resistant Palmer amaranth density reduction and biomass reduction and isoxaflutole/glufosinate/glyphosate-resistant soybean yield in field experiments conducted in 2018 and 2019.[a]

Herbicide program	Application timing	Rate	Palmer amaranth density reduction[b,d]		Biomass reduction[c,d]	Soybean yield[b,d]	
			14 DAEPOST	14 DAEPOST	14 DALPOST		
		g ae or ai ha^{-1}	%		%	kg ha^{-1}	
			2018	2019	2019	2018	2019
Nontreated control	–	–	0	0	0	960 b	2,130 c
Isoxaflutole	PRE	105	0 c	48 d	0 d	990 b	2,710 bc
Isoxaflutole fb isoxaflutole	PRE fb early POST	105, 105	49 b	53 cd	0 d	1,110 ab	2,160 c
Glufosinate	Early POST	657	89 a	58 bcd	41 c	1,480 ab	2,040 c
Glufosinate fb glufosinate	Early POST fb late POST	657, 657	92 a	78 abcd	90 a	1,700 ab	4,230 a
Isoxaflutole fb glufosinate	PRE fb early POST	105, 657	95 a	92 abc	49 bc	1,040 b	2,670 bc
Pyroxasulfone/sulfentrazone fb glufosinate	PRE fb early POST	292, 657	100 a	100 a	87 ab	1,750 ab	3,970 ab
Pyroxasulfone/sulfentrazone + isoxaflutole fb glufosinate	PRE fb early POST	292 + 105, 657	100 a	100 a	95 a	2,290 a	4,020 ab
Flumioxazin/pyroxasulfone fb glufosinate	PRE fb early POST	160, 657	100 a	100 a	92 a	1,940 ab	4,050 ab
Flumioxazin/pyroxasulfone + isoxaflutole fb glufosinate	PRE fb early POST	160 + 105, 657	100 a	100 a	97 a	2,120 ab	3,630 ab
Imazethapyr/pyroxasulfone/saflufenacil fb glufosinate	PRE fb early POST	215, 657	100 a	99 ab	84 ab	2,030 ab	3,660 ab
Imazethapyr/pyroxasulfone/saflufenacil + isoxaflutole fb glufosinate	PRE fb early POST	215 + 105, 657	100 a	99 ab	83 ab	1,460 ab	3,370 abc
Isoxaflutole + glufosinate	Early POST	105 + 657	100 a	85 abcd	59 abc	1,570 ab	2,030 c
Isoxaflutole fb glufosinate fb glufosinate	PRE fb early POST fb late POST	105, 657; 657	95 a	87 abcd	95 a	1,980 ab	4,280 a
Isoxaflutole + glufosinate fb isoxaflutole + glufosinate	Early POST fb late POST	105 + 657, 105 + 657	63 ab	94 abc	96 a	1,020 b	3,140 abc
Glyphosate fb glyphosate	Early POST fb late POST	1,260, 1,260	0 c	0 e	0 d	950 b	2,130 c

[a]Abbreviations: DAEPOST, days after early-POST herbicide application; DALPOST, days after late-POST herbicide application; fb, followed by.
[b]Year-by-treatment interaction for glyphosate-resistant Palmer amaranth density and soybean yield were significant; therefore, data were not combined across the two years.
[c]Biomass reduction data is only available for 2019.
[d]Means presented within each column with no common letter(s) are significantly different according to Fisher's protected least significant difference test at $P \leq 0.05$.

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References

Anonymous (2017) Liberty herbicide product label. EPA Reg. No. 264-829. Research Triangle Park, NC: Bayer Crop Science 29 p

Anonymous (2020) Alite™ 27 herbicide product label. EPA Reg. No. 7969-433. Research Triangle Park, NC: BASF. 18 p

Aulakh JS, Jhala AJ (2015) Comparison of glufosinate-based herbicide programs for broad-spectrum weed control in glufosinate-resistant soybean. Weed Technol 29:419–430

Barber T, Norsworthy J, Butts T (2021) Arkansas Palmer amaranth found resistant to field rates of glufosinate. https://arkansascrops.uaex.edu/posts/weeds/palmer-amaranth.aspx Accessed: June 7, 2021

Beckie HJ (2006) Herbicide-resistant weeds: management tactics and practices. Weed Technol 20:793–814

Bond JA, Oliver LR (2006) Comparative growth of Palmer amaranth (*Amaranthus palmeri*) accessions. Weed Sci 54:121–126

Chahal PS, Jhala AJ (2015) Herbicide programs for control of glyphosate-resistant volunteer corn in glufosinate-resistant soybean. Weed Technol 29:431–443

Chahal PS, Aulakh JS, Jugulam M, Jhala AJ (2015) Herbicide-resistant Palmer amaranth (*Amaranthus palmeri* S. Wats.) in the United States — mechanisms of resistance, impact, and management. Pages 1–40 *in* Price AJ, JA Kelton, L Sarunaite (eds), Herbicides, Agronomic Crops and Weed Biology. London: InTechOpen

Chahal PS, Varanasi VK, Jugulam M, Jhala AJ (2017) Glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*) in Nebraska: confirmation, EPSPS gene amplification, and response to POST corn and soybean herbicides. Weed Technol 31:80–93

Costea M, Weaver SE, Tardif FJ (2004) The biology of Canadian weeds. 130. *Amaranthus retroflexus* L., A. *powellii* S. Watson and A. *hybridus* L. Can J Plant Sci 84:631–668

Costea M, Weaver SE, Tardif FJ (2005) The biology of invasive alien plants in Canada. 3. *Amaranthus tuberculatus* (Moq.) Sauer var. *rudis* (Sauer). Can J Plant Sci 85:507–522

Culpepper AS (2006) Glyphosate-induced weed shifts. Weed Technol 20:277–281

Culpepper AS, Grey TL, Vencill WK, Kichler JM, Webster TM, Brown SM, York AC, Davis JW, Hanna WW (2006) Glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*) confirmed in Georgia. Weed Sci 54:620–626

Culpepper AS, Webster TM, Sosnoskie LM, York A (2010) Glyphosate-resistant Palmer amaranth in the United States. Pages 195–212 *in* Nandula VK, ed. Glyphosate Resistance: Evolution, Mechanisms, and Management. Hoboken, NJ: John Wiley

de Sanctis JH, Barnes ER, Knezevic SZ, Kumar V, Jhala AJ (2021) Residual herbicides affect critical time of Palmer amaranth (*Amaranthus palmeri*) removal in soybean. Agron J 113:1920–1933

Ehleringer J (1985) Annuals and perennials of warm deserts. Page 162–180 *in* Chabot BF, Mooney HA, eds. Physiological Ecology of North American Plant Communities. Dordrecht: Springer Netherlands

Ehleringer J, Forseth I (1980) Solar tracking by plants. Science 210:1094–1098

Heap I (2021) The international herbicide-resistant weed database. http://www.weedscience.org/Pages/MOA.aspx?MOAID=12. Accessed: April 03, 2021

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Heap I, Duke SO (2018) Overview of glyphosate-resistant weeds worldwide. Pest Manag Sci 74:1040–1049
Hoffner AE, Jordan DL, Chandi A, York AC, Dunphy EJ, Everman WJ (2012) Management of Palmer Amaranth (*Amaranthus palmeri*) in glufosinate-resistant soybean (*Glycine max*) with sequential applications of herbicides. ISRN Agron 2012:1–7
Horak MJ, Loughin TM (2000) Growth analysis of four *Amaranthus* species. Weed Sci 48:347–355
Janak TW, Grichar WJ (2016) Weed control in corn (*Zea mays* L.) as influenced by preemergence herbicides. Int J Agron 2016:1–9
Jhala AJ (2017a) Status of herbicide-resistant weeds in Nebraska. CropWatch, University of Nebraska–Lincoln. https://cropwatch.unl.edu/2017/status-herbicide-resistant-weeds-nebraska. Accessed: November 5, 2020
Jhala AJ (2017b) Effect of excessive rainfall on efficacy of residual herbicides applied in corn and soybean. CropWatch, University of Nebraska –Lincoln. https://cropwatch.unl.edu/2017/effect-excessive-rainfall-efficacy-residual-herbicides-applied-corn-and-soybean. Accessed: September 3, 2020
Jhala AJ, Norsworthy JK, Ganie ZA, Sosnoskie LM, Beckie HJ, Mallory-Smith CA, Liu J, Wei W, Wang J, Stoltenberg DE (2021) Pollen-mediated gene flow and transfer of resistance alleles from herbicide-resistant broadleaf weeds. Weed Technol 35:173–187
Jhala AJ, Ramirez AHM, and M Singh (2013) Tank mixing saflufenacil, glufosinate, and indaziflam improved burndown and residual weed control. Weed Technol 27:422–429
Jhala AJ, Sandell SD, Rana N, Kruger G, Knezevic SZ (2014) Confirmation and control of triazine and 4-hydroxyphenylpyruvate dioxygenase-inhibiting herbicide-resistant Palmer amaranth in Nebraska. Weed Technol 28:28–38
Jhala AJ, Sandell LD, Sarangi D, Kruger GR, Knezevic SZ (2017) Control of glyphosate-resistant common waterhemp (*Amaranthus rudis*) in glufosinate-resistant soybean. Weed Technol 31:32–45
Johnson WG, Chahal GS, Regehr DL (2012) Efficacy of various corn herbicides applied preplant incorporated and preemergence. Weed Technol 26:220–229
Kumar V, Liu R, Boyer G, Stahlman PW (2019) Confirmation of 2,4-D resistance and identification of multiple resistance in a Kansas Palmer amaranth population. Pest Manag Sci 75:2925–2933
Massinga RA, Currie RS, Horak MJ, Boyer J (2001) Interference of Palmer amaranth in corn. Weed Sci 49:202–208
Meyer CJ, Norsworthy JK, Young BG, Steckel LE, Bradley KW, Johnson WG, Loux MM, Davis VM, Kruger GR, Bararpour MT, Ikley JT, Spaunhorst DJ, Butts TR (2015) Herbicide program approaches for managing glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*) and waterhemp (*Amaranthus tuberculatus* and *Amaranthus rudis*) in future soybean-trait technologies. Weed Technol 29:716–729
Meyer CJ, Norsworthy JK, Young BG, Steckel LE, Bradley KW, Johnson WG, Loux MM, Davis VM, Kruger GR, Bararpour MT, Ikley JT, Spaunhorst DJ, Butts TR (2016) Early-season Palmer amaranth and waterhemp control from preemergence programs utilizing 4-hydroxyphenylpyruvate dioxygenase–inhibiting and auxinic herbicides in soybean. Weed Technol 30:67–75
[NOAA] National Ocean and Atmospheric Administration (2019) NOWData-NOAA Online Weather Data. https://w2.weather.gov/climate/xmacis.php?wfo=gid. Accessed January 2, 2020
Norsworthy JK, Griffith GM, Scott RC, Smith KL, Oliver LR (2008) Confirmation and control of glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*) in Arkansas. Weed Technol 22:108–113
Norsworthy JK, Korres NE, Walsh MJ, Powles SB (2016) Integrating herbicide programs with harvest weed seed control and other fall management practices for the control of glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*). Weed Sci 64:540–550
Norsworthy JK, Ward SM, Shaw DR, Llewellyn RS, Nichols RL, Webster TM, Bradley KW, Frisvold G, Powles SB, Burgos NR, Witt WW, Barrett M (2012) Reducing the risks of herbicide resistance: best management practices and recommendations. Weed Sci 60:31–62
Oliveira MC, Gaines TA, Patterson E, Jhala AJ, Irmak S, Keenan A, Knezevic SZ (2018) Interspecific and intraspecific metabolism-based herbicide resistance transfer in dioecious weedy *Amaranthus*. Plant J 96:1051–1063
Sarangi D, Jhala AJ (2019) Palmer amaranth (*Amaranthus palmeri*) and velvetleaf (*Abutilon theophrasti*) control in no-tillage conventional (non–genetically engineered) soybean using overlapping residual herbicide programs. Weed Technol 33:95–105
Sarangi D, Sandell LD, Kruger GR, Knezevic SZ, Irmak S, Jhala AJ (2017) Comparison of herbicide programs for season-long control of glyphosate-resistant common waterhemp (*Amaranthus rudis*) in soybean. Weed Technol 31:53–66
Schultz JL, Weber M, Allen J, Bradley KW (2015) Evaluation of weed management programs and response of FG72 soybean to HPPD-inhibiting herbicides. Weed Technol 29:653–664
Shyam C, Chahal PS, Jhala AJ, Jugulam M (2021) Management of glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*) in 2,4-D choline, glufosinate, and glyphosate-resistant soybean. Weed Technol 35:136–143
Smith A, Soltani N, Kaastra AJ, Hooker DC, Robinson DE, Sikkema PH (2019) Annual weed management in isoxaflutole-resistant soybean using a two-pass weed control strategy. Weed Technol 33:411–425
Sosnoskie LM, Webster TM, Kichler JM, MacRae AW, Grey TL, Culpepper AS (2012) Pollen-mediated dispersal of glyphosate-resistance in Palmer amaranth under field conditions. Weed Sci 60:366–373
Spaunhorst DJ, Johnson WG (2016) Palmer amaranth (*Amaranthus palmeri*) control with preplant herbicide programs containing dicamba, isoxaflutole, and 2,4-D. Crop Forage Turfgrass Manag 2:1–7
Sprague CL, Stoller EW, Wax LM, Horak MJ (1997) Palmer amaranth (*Amaranthus palmeri*) and common waterhemp (*Amaranthus rudis*) resistance to selected ALS-inhibiting herbicides. Weed Sci 45:192–197
Steckel LE (2007) The dioecious *Amaranthus* spp.: here to stay. Weed Technol 21:567–570
Vieira BC, Samuelson SL, Alves GS, Gaines TA, Werle R, Kruger GR (2018) Distribution of glyphosate-resistant *Amaranthus* spp. in Nebraska. Pest Manag Sci 74:2316–2324
Ward S, Webster TM, Steckel LE (2013) Palmer amaranth (*Amaranthus palmeri*): A review. Weed Technol 27:12–27
Whitaker JR, York AC, Jordan DL, Culpepper AS (2010) Palmer amaranth (*Amaranthus palmeri*) control in soybean with glyphosate and conventional herbicide systems. Weed Technol 24:403–410
Wiesbrook ML, Johnson WG, Hart SE, Bradley PR, Wax LM (2001) Comparison of weed management systems in narrow-row, glyphosate- and glufosinate-resistant soybean (*Glycine max*). Weed Technol 15:122–128
Wortman SE (2014) Integrating weed and vegetable crop management with multifunctional air-propelled abrasive grits. Weed Technol 28:243–252
Young BG (2006) Changes in herbicide use patterns and production practices resulting from glyphosate-resistant crops. Weed Technol 20:301–307

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