

NATURAL DISTRIBUTION OF HYMENOPTERAN PARASITIDS OF SPODOPTERA FRUGIPERDA (LEPIDOPTERA: NOCTUIDAE) LARVAE IN MEXICO

Authors: Molina-Ochoa, Jaime, Carpenter, James E., Lezama-Gutiérrez, Roberto, Foster, John E., González-Ramírez, Martín, et al.

Source: Florida Entomologist, 87(4) : 461-472

Published By: Florida Entomological Society

URL: [https://doi.org/10.1653/0015-4040\(2004\)087\[0461:NDOHPO\]2.0.CO;2](https://doi.org/10.1653/0015-4040(2004)087[0461:NDOHPO]2.0.CO;2)

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

NATURAL DISTRIBUTION OF HYMENOPTERAN PARASITOIDS OF *SPODOPTERA FRUGIPERDA* (LEPIDOPTERA: NOCTUIDAE) LARVAE IN MEXICO

JAIME MOLINA-OCHOA¹, JAMES E. CARPENTER², ROBERTO LEZAMA-GUTIÉRREZ¹, JOHN E. FOSTER³, MARTÍN GONZÁLEZ-RAMÍREZ¹, CÉSAR ANDRES ANGEL-SAHAGÚN¹ AND JAVIER FARIAS-LARIOS¹

¹Universidad de Colima, Facultad de Ciencias Biológicas y Agropecuarias
Apartado postal 36, Tecomán, Colima 28100, México

²United States Department of Agriculture, Agricultural Research Service
Crop Protection & Management Research Laboratory, P.O. Box 748, Tifton, GA 31793-0748, USA

³University of Nebraska Lincoln, Department of Entomology
312F Plant Industry Building, Lincoln, NE 68583-0816, USA

ABSTRACT

A survey of parasitoids of fall armyworm (FAW), *Spodoptera frugiperda* (J. E. Smith), larvae was conducted in six Mexican states during August and September 2000. Thirteen genera of hymenopteran parasitoids were recovered representing the following 3 families, Braconidae: *Aleoidea*, *Chelonus*, *Cotesia*, *Glyptapanteles*, *Homolobus*, and *Meteorus*; Ichneumonidae: *Campoletis*, *Eiphosoma*, *Ophion*, and *Pristomerus*; and Eulophidae: *Aprostocetus*, *Euplectrus*, and *Horismenus*. Out of 5591 FAW larvae collected, 772 produced parasitoids, for a parasitism rate of 13.8%. The highest rate of parasitism from a single collection was 42.2%, representing three species of parasitoids in Michoacán. *Chelonus insularis* Cresson was the most widely distributed species occurring in 45.3% of the locations. *Pristomerus spinator* (F.), and *Meteorus laphygmae* (Viereck), exhibited the highest rates of parasitism for a single collection with 22.2% and 22.1%, in Sinaloa, and Michoacán, respectively. The results supported the hypothesis that natural distribution and rates of parasitism of FAW larvae may be related to more diverse habitats with more forests, orchards, and pastures near to cornfields.

Key Words: fall armyworm, *Chelonus*, *Pristomerus*, *Meteorus*, *Ophion*, *Campoletis*, corn, survey.

RESUMEN

Se llevó a cabo un inventario de parasitoides de larvas del gusano cogollero, *Spodoptera frugiperda* (J. E. Smith) (FAW) colectadas principalmente de maizales en estado de verticilio en seis estados mexicanos durante Agosto y Septiembre de 2000. Trece géneros de parasitoides himenópteros fueron recuperados, representando a tres familias, Braconidae: *Aleoidea*, *Chelonus*, *Cotesia*, *Glyptapanteles*, *Homolobus*, y *Meteorus*; Ichneumonidae: *Campoletis*, *Eiphosoma*, *Ophion*, y *Pristomerus*; y Eulophidae: *Aprostocetus*, *Euplectrus*, y *Horismenus*. De un total de 5591 larvas colectadas, 772 produjeron parasitoides, para una tasa de parasitismo de 13.8%. La tasa de parasitismo más alta para una colecta simple fué de 42.2%, representando a tres especies de parasitoides en Michoacán. La especie más ampliamente distribuida fué *Chelonus insularis* Cresson, presentándose en 45.3% de las localidades inventariadas. *Pristomerus spinator* (F.), y *Meteorus laphygmae* (Viereck), mostraron las tasa más altas de parasitismo para una colecta simple con 22.2% y 22.1%, en Sinaloa, y Michoacán, respectivamente. Los resultados apoyan la hipótesis de que la distribución natural y las tasas de parasitismo pueden estar relacionadas a lo diverso de los hábitat con la cercanía de más bosques, huertas y pastizales a los maizales.

Translation provided by the authors.

The therapeutic approach of killing pest organisms with toxic chemicals has prevailed as a pest control strategy for over 50 years (Lewis et al. 1997). In the 1950s environmental effects of persistent organochlorine insecticides such as DDT began to be observed. Currently, in agricultural pest control, the adverse effects of the use of insecticides are leading scientists to search for al-

ternatives to chemical control of insect pests based on health, environmental, wild life, and economic concerns (Johnson et al. 1998; Mattsson et al. 2000; Solomon & Schettler 2000).

Native insects and pathogens are normal parts of functioning agro-ecosystems and can profoundly influence the agricultural structure, species composition, and diversity. Agro-ecosystems

exhibit high biodiversity, mainly influenced by crops, weeds, microorganisms, and arthropods, but these factors are also influenced by geographical location, soil, and climatic characteristics, as well as human factors. Scientific evidence suggests that biodiversity can be used for improved pest management (Altieri 1991). The increased use of beneficial insects and interference with the colonization of fall armyworm in multiple cropping systems have prevented outbreaks in Latin America (Altieri 1994).

The fall armyworm (FAW), *Spodoptera frugiperda* (J. E. Smith), is a voracious pest inflicting damage to a multiplicity of annual crops in the Americas, and it is commonly controlled with synthetic insecticides, although insecticide resistance has been observed and is a concern (Yu 1991, 1992). Moreover, two strains of FAW have been identified according to their host preference, a corn-associated strain that feeds principally on corn, and a rice-associated strain that feeds primarily on forage grasses and rice (Pashley et al. 1987). Both FAW strains exhibited differences in resistance to chemical and biological insecticides (Adamczyk et al. 1997; López-Edwards et al. 1999), and have differences in their genetic population structure and population ecology (Pashley 1988; Lu & Adang 1996; Bossart & Prowell 1998; Levy et al. 2002; Meagher & Gallo-Meagher 2003; Nagoshi & Meagher 2003). These differences between FAW strains complicate the management of this pest.

Biological control is a highly desirable alternative to insecticides for controlling FAW infestations (Gross & Pair 1986). The value of parasitoids in reducing larval populations of this noctuid has long been recognized (Luginbill 1928; Vickery 1929). In order to develop a better understanding of the natural distribution of the FAW parasitoid complex and natural enemies, surveys have been carried out in different regions of Mexico (Carrillo 1980; Lezama-Gutiérrez et al. 2001; Molina-Ochoa et al. 2001, 2003a).

Here, we report the natural distribution of parasitoids of FAW larvae collected from whorl-stage corn, grain sorghum, forage sorghum, and Sudan grass fields from five Mexican states in the Pacific coast and one state in the Gulf of Mexico, during the summer of 2000.

MATERIALS AND METHODS

During August and September of 2000, *S. frugiperda* larvae were collected from whorl-stage corn, grain and forage sorghum, and Sudan grass fields in 64 locations in the Mexican Pacific coast states of Sinaloa, Nayarit, Jalisco, Colima, and Michoacán, and in the Gulf of Mexico state of Veracruz. Egg masses and pupae were not collected.

FAW larvae were individually placed into 30-cc plastic cups with pinto bean diet (Burton & Perkins 1989), and held in the laboratory (Laboratory of Biological Control, Universidad de Colima, Facultad de Ciencias Biológicas y Agropecuarias, Tecmán, Colima, México) for emergence of parasitoids (Molina-Ochoa et al. 2001). Adult parasitoids were placed in 70% ethanol and then submitted to the USDA/ARS Systematic Entomology Laboratory, Beltsville, MD for identification. Collection size ranged from 33 to 119 FAW larvae. The number collected was corrected by subtracting the number that died from injury or unknown causes during the first few days after collection before calculating percent parasitism. Mortality due to pathogens and parasitic nematodes has been previously reported (Molina-Ochoa et al. 2003a).

Collection dates, geographic location, altitude, crop, sample size and total parasitism of FAW larvae in six Mexican states are presented in the Table 1. A Garmin GPS III Plus[®] was used for obtaining the coordinates and altitude data.

RESULTS AND DISCUSSION

Out of 5591 FAW larvae collected, 772 produced parasitoids, for a parasitism rate of 13.8%. These parasitoids represented 13 genera from three families of Hymenoptera: six Braconidae, four Ichneumonidae, and three Eulophidae. Nine of the 64 collections produced no parasitoids, six of 12 collections from whorl-stage corn in Michoacán, two of 13 in Jalisco, and only one of 11 in Colima. The highest rates of parasitism in each state were found in C4 (33.3%) in Colima, J12 (21.1%) in Jalisco, M12 (14.4%) in Michoacán, N9 (18.9%) in Nayarit, S5 (27.4%) in Sinaloa, and V4 (11.5%) in Veracruz (Table 1). The most diverse collections of parasitoids were found in the locations C5, J12, and N9 with 5, 4, and 4 species, respectively, (Tables 2 and 3). The collection from S5 produced the highest rate of parasitism for a single species with 22.1%; the braconid *Meteorus laphygmae* Viereck was the most common parasitoid collected from Sudangrass. Other parasitoids in that collection were the eulophid *Euplectrus plathypenae* Howard (2 individuals), and the ichneumonid *Ophion flavidus* Brulle (1 individual). The braconid *C. insularis* occurred in 29 of the 64 collections from the six states, and it was the most widely distributed parasitoid. Another important braconid was *M. laphygmae*, occurring in 21 of the 64 collections. The ichneumonid parasitoids, *O. flavidus*, and *Pristomerus spinator* F., occurred in 18, and 17 of the 64 collections, respectively. *E. plathypenae* was the most important and widely distributed eulophid, occurring in 16 of the 64 collections (Tables 2 and 3).

Chelonus insularis was the most widely distributed parasitoid of FAW larvae in this survey, occurring in all the six Mexican States, and it was the braconid species with the second highest parasitism rate per location with 16.7%. Thus, *C. in-*

TABLE 1. GEOGRAPHIC LOCATION, DATE, ALTITUDE, CROP (*), SAMPLE SIZE (N), AND TOTAL PERCENT *SPODOPTERA FRUGIPERDA* LARVAE PARASITIZED IN SIX MEXICAN STATES (**) DURING 2000.

| Code | Date | Location | Coordinates | Alt (m) | * | N | Percentage parasitized |
|------|-------|----------------------------|-----------------------------|---------|---|----|------------------------|
| C1 | 08/04 | El poblado, Coquimatlán | 19°3.698'N 103°47.722'W | 422 | C | 90 | 17.8 |
| C2 | 08/04 | Pueblo Juárez, Coquimatlán | 19°10.752'N 103°54.634'W | 279 | C | 90 | 4.4 |
| C3 | 08/04 | Amachico, Coquimatlán | 19°10.667'N 103°56.351'W | 328 | C | 90 | 12.2 |
| C4 | 08/06 | Los mezcales, Comala | 19°20.811'N 103°47.176'W | 608 | C | 90 | 33.3 |
| C5 | 08/06 | El remate, Comala | 19°24.825'N 103°47.639'W | 817 | C | 90 | 13.3 |
| C6 | 08/06 | Carrizalillo, Quesería | 19°25.389'N 103°41.000'W | 1550 | C | 90 | 1.1 |
| C7 | 08/06 | Quesería | 19°23.362'N 103°34.882'W | 1304 | c | 90 | 10.0 |
| C8 | 08/06 | Villa de Alvarez | 19°17.201'N 103°47.030'W | 515 | c | 90 | 4.4 |
| C9 | 08/06 | Juluapan, Villa de Alvarez | 19°18.890'N 103°49.611'W | 539 | c | 90 | 4.4 |
| C10 | 08/07 | Tepames, Colima | 19°08.231'N 103°37.996'W | 519 | c | 90 | 0.0 |
| C11 | 08/07 | Etapilla, Colima | 18°59.549'N 103°31.140'W | 304 | c | 90 | 21.1 |
| J1 | 08/08 | Ciudad Guzmán | 19°40.011'N 103°28.830'W | 1557 | c | 90 | 0.0 |
| J2 | 08/15 | Los pinitos, Tonila | 19°25.343'N 103°32.447'W | 1326 | c | 90 | 2.2 |
| J3 | 08/15 | Pialla, Tuxpan | 19°27.293'N 103°28.514'W | 1079 | c | 90 | 0.0 |
| J4 | 08/15 | Atenquique, Tuxpan | 19°31.778'N 103°27.851'W | 1338 | c | 90 | 1.1 |
| J5 | 08/17 | Canoas, Zapotiltic | 19°34.073'N 103°27.324'W | 1391 | c | 90 | 3.3 |
| J6 | 08/17 | Apastepe | 19°38.060'N 103°30.950'W | 1709 | c | 90 | 1.1 |
| J7 | 08/17 | Teocuitatlán | 20°07.035'N 103°32.704'W | 1369 | c | 90 | 10.0 |
| J8 | 08/17 | Zacoalco de Torres | 20°11.988'N 103°33.806'W | 1425 | c | 90 | 4.4 |
| J9 | 08/17 | Acatlán de Juárez | 20°25.362'N 103°33.406'W | 1575 | c | 96 | 2.1 |
| J10 | 08/17 | Tlajomulco de Zúñiga | 20°29.396'N 103°28.298'W | 1607 | c | 92 | 4.3 |
| J11 | 08/18 | Zapopan | 20°43.129'N 103°29.041'W | 1670 | c | 90 | 4.4 |
| J12 | 08/18 | Magdalena | 20°53.008'N 103°55.477'W | 1496 | c | 93 | 21.5 |
| J13 | 08/23 | Crucero de Magdalena | 20°56.300'N 104°02.509'W | 1386 | c | 92 | 2.2 |
| M1 | 08/09 | Totolán | 19°58.890'N 102°40.183'W | 1590 | c | 90 | 0.0 |
| M2 | 08/09 | Santa Inés Tocombo | 19°44.502'N 102°34.967'W | 1630 | c | 90 | 1.1 |
| M3 | 08/09 | Peribán | 19°33.106'N 102°26.586'W | 1475 | c | 90 | 1.1 |
| M4 | 08/10 | Cointzio | 19°41.609'N 101°16.398'W | 1932 | c | 90 | 0.0 |

*Corn (c), gran sorghum (gs), forage sorghum (fs), and Sudan grass (sg).
**Colima (C), Jalisco (J), Michoacan (M) Nayarit (N), Sinaloa (S), and Veracruz (V).

TABLE 1. (CONTINUED) GEOGRAPHIC LOCATION, DATE, ALTITUDE, CROP (*), SAMPLE SIZE (N), AND TOTAL PERCENT *SPODOPTERA FRUGIPERDA* LARVAE PARASITIZED IN SIX MEXICAN STATES (**) DURING 2000.

| Code | Date | Location | Coordinates | Alt (m) | * | N | Percentage parasitized |
|------|-------|--------------------------|-----------------------------|---------|------|-----|------------------------|
| M5 | 08/10 | Cerro “La Esperanza” | 19°41.223’N 101°18.980’W | 1998 | c | 90 | 1.1 |
| M6 | 08/11 | Tejabán | 19°13.342’N 101°53.714’W | 587 | c | 90 | 0.0 |
| M7 | 08/11 | Carretera a Nueva Italia | 19°03.290’N 102°02.458’W | 442 | c | 90 | 0.0 |
| M8 | 08/11 | Presa de Zicuirán | 18°56.191’N 101°54.650’W | 292 | c | 63 | 0.0 |
| M9 | 08/11 | El ceñidor, Nueva Italia | 18°59.651’N 102°11.577’W | 350 | c | 57 | 1.8 |
| M10 | 08/12 | La Guadalupe Parácuaro | 19°07.472’N 102°12.519’W | 540 | fs | 90 | 1.1 |
| M11 | 08/12 | Las yeguas Parácuaro | 18°57.308’N 102°16.733’W | 359 | fs | 90 | 1.1 |
| M12 | 08/12 | El cirían, Nueva Italia | 18°53.661’N 102°07.483’W | 255 | c | 90 | 42.2 |
| N1 | 08/18 | Santa María del Oro | 21°20.121’N 104°40.174’W | 1160 | c | 90 | 3.3 |
| N2 | 08/18 | El rincón, Tepic | 21°32.472’N 104°56.123’W | 849 | c | 96 | 10.4 |
| N3 | 08/18 | El pichón, Tepic | 21°33.479’N 104°56.937’W | 774 | c | 95 | 4.2 |
| N4 | 08/19 | Xalisco | 21°19.601’N 104°55.060’W | 1042 | c | 107 | 2.8 |
| N5 | 08/19 | El refilión, Xalisco | 21°19.407’N 104°55.323’W | 964 | c | 90 | 8.9 |
| N6 | 08/19 | Compostela | 21°17.858’N 104°54.044’W | 920 | c | 93 | 1.1 |
| N7 | 08/19 | La presa, Compostela | 21°13.714’N 104°52.162’W | 928 | c | 90 | 1.1 |
| N8 | 08/20 | Las lumbres, Acaponeta | 22°20.795’N 105°18.141’W | 48 | C&gs | 60 | 5.0 |
| N9 | 08/23 | Seboruco | 21°20.850’N 104°40.749’W | 1134 | c | 90 | 18.9 |
| N10 | 08/23 | Ahuacatlán | 21°06.331’N 104°27.427’W | 1120 | c | 90 | 5.6 |
| S1 | 08/21 | Bacurimi, Culiacán | 24°51.668’N 107°29.478’W | 70 | gs | 97 | 4.1 |
| S2 | 08/21 | La campana, Culiacán | 24°58.415’N 107°33.517’W | 143 | gs | 100 | 5.0 |
| S3 | 08/21 | Pericos, Mocorito | 25°03.574’N 107°39.547’W | 80 | gs | 95 | 9.5 |
| S4 | 08/21 | Rancho viejo, Mocorito | 25°06.033’N 107°43.165’W | 89 | gs | 98 | 13.3 |
| S5 | 08/22 | Aguapepito, Mocorito | 25°03.861’N 107°39.547’W | 68 | sg | 95 | 27.4 |
| S6 | 08/22 | Comanito, Mocorito | 25°09.006’N 107°39.645’W | 91 | gs | 95 | 3.2 |
| S7 | 08/22 | La poma, Badiraguato | 25°15.749’N 107°40.739’W | 157 | c | 100 | 13.0 |
| S8 | 08/22 | La majada, Badiraguato | 25°14.076’N 107°39.781’W | 145 | c | 92 | 7.6 |
| V1 | 09/02 | Seis de Enero, Xalapa | 19°34.115’N 96°50.207’W | 950 | c | 91 | 6.6 |
| V2 | 09/02 | Altolucero, Almolonga | 19°35.063’N 96°47.384’W | 908 | c | 33 | 12.1 |

*Corn (c), gran sorghum (gs), forage sorghum (fs), and Sudan grass (sg).
**Colima (C), Jalisco (J), Michoacan (M) Nayarit (N), Sinaloa (S), and Veracruz (V).

TABLE 1. (CONTINUED) GEOGRAPHIC LOCATION, DATE, ALTITUDE, CROP (*), SAMPLE SIZE (N), AND TOTAL PERCENT *SPODOPTERA FRUGIPERDA* LARVAE PARASITIZED IN SIX MEXICAN STATES (**) DURING 2000.

| Code | Date | Location | Coordinates | Alt (m) | * | N | Percentage parasitized |
|------|-------|-----------------------|----------------------------|---------|---|-----|------------------------|
| V3 | 09/02 | Actopan | 19°34.623'N 96°48.589'W | 775 | c | 64 | 3.1 |
| V4 | 09/02 | Los González, Actopan | 19°31.894'N 96°41.294'W | 432 | c | 113 | 11.5 |
| V5 | 09/02 | Bocana, Actopan | 19°24.416'N 96°36.731'W | 311 | c | 119 | 4.2 |
| V6 | 09/03 | El volador, Coatepec | 19°21.594'N 96°51.037'W | 709 | c | 90 | 3.3 |
| V7 | 09/03 | Palmillas | 19°12.293'N 96°46.221'W | 702 | c | 59 | 6.8 |
| V8 | 09/03 | Tierra Colorada | 19°13.255'N 96°21.916'W | 46 | c | 45 | 4.4 |
| V9 | 09/04 | Cerro gordo | 19°25.252'N 96°39.566'W | 443 | c | 45 | 8.9 |
| V10 | 09/04 | La cumbre | 19°23.320'N 96°38.807'W | 366 | c | 66 | 6.1 |

*Corn (c), gran sorghum (gs), forage sorghum (fs), and Sudan grass (sg).
**Colima (C), Jalisco (J), Michoacan (M) Nayarit (N), Sinaloa (S), and Veracruz (V).

sularis is one of the most abundant natural enemies of fall armyworm larvae in the Western Coast and Gulf of Mexico. *Chelonus insularis* has been reported as an important parasitoid controlling FAW populations in the US (Luginill 1928; Vickery 1929). Ashley (1986) and Andrews (1988) listed *C. insularis* occurring in Central America and the US, highlighting its role as parasitoid of FAW in southern Florida where 63% of the FAW larvae were attacked. Recently, Molina-Ochoa et al., (2003b) reported *C. insularis* syn. *C. texanus* as the braconid with the broadest distribution in Latin America, including South America (Uruguay and Venezuela), the Caribbean Basin (Trinidad and Puerto Rico), and the US. In that inventory *Chelonus* sp. is also reported in Brazil, Mexico, and Peru. Lewis and Nordlund (1980) emphasized its role considering it as an excellent candidate for the following augmentative approaches: a) release throughout its overwintering zone; b) early-season colonization, and c) direct therapeutic release on target crops.

In a previous survey, Molina-Ochoa et al. (2001) commented on the importance and need of more study in Mexico on the taxonomy of the genus *Chelonus* (P. M. Marsh, pers. comm.).

Meteorus laphygmae occurred in 21 of the 64 collections. The highest rate of parasitism for a single location was obtained in S5 with 22.1%. This parasitoid occurred in all of the collections from Sinaloa, and the rate of parasitism ranged from 2.1 to 22.1%. *Meteorus laphygmae* was also collected in Colima, Nayarit, Michoacán, Jalisco, and Veracruz occurring in 45.5%, 30%, 25%, 10%, and 8.3% of the collections, respectively. This braconid was reported by Ashley (1986) occurring in

the Continental US, exhibiting its greatest impact on FAW collected from grass. Other reports were made by Alvarado-Rodríguez (1987) in Sinaloa, Mexico attacking *Spodoptera exigua* (Hübner) infesting tomatoes with a parasitism rate of 9.0%. A similar rate of parasitism was reported by Molina-Ochoa et al. (2001) in a single collection of FAW larvae made in El Mante, Tamaulipas with 10.3%. Molina-Ochoa et al. (2003b) listed several reports from countries of Central and South America, such as Honduras, Nicaragua, Mexico, Chile, Colombia, and Suriname, where *M. laphygmae* was collected from other crops such as maize, rice, cotton, sorghum, peanuts, and Bermudagrass, and was one of the most prevalent parasitoids in South America.

Low rates of occurrence and parasitization of *Cotesia* sp. probably *marginiventris* (Cresson), *Glyptapanteles* sp. probably *militaris* (Walsh), *Aleiodes* sp., and *Homolobus* sp. probably *mellea* (Cresson) were recorded. They were found in 5, 2, 1, and 1 of the 64 collections, respectively.

Cotesia sp. occurred in Colima, Jalisco, Nayarit with lower parasitization rates than 2.3%. Similar rates were reported by Molina-Ochoa et al. (2001) in a previous survey conducted in four Mexican States. This parasitoid is reported attacking FAW larvae in Argentina, Brazil, Chile, Honduras, Lesser Antilles, Mexico, Nicaragua, Puerto Rico, Suriname (Molina-Ochoa et al. 2003b), but it has been often reported as a parasitoid of FAW in the US (Ashley 1986) with parasitization rates of 6.3% on FAW larvae collected from maize (Riggin et al. 1993) and from less than 1% to 40% collected from maize and Bermuda-grass, respectively (Ashley et al. 1983).

TABLE 2. PERCENTAGE OF *SPODOPTERA FRUGIPERDA* LARVAE PARASITIZED BY EACH SPECIES OF BRACONIDAE AT EACH LOCATION.

| Code* | Braconidae | | | | | |
|-------|-----------------|-----------------|----------------|-----------------------|------------------|-----------------|
| | <i>Aleiodes</i> | <i>Chelonus</i> | <i>Cotesia</i> | <i>Glyptapanteles</i> | <i>Homolobus</i> | <i>Meteorus</i> |
| C1 | 0.0 | 3.3 | 0.0 | 0.0 | 0.0 | 2.2 |
| C2 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 1.1 |
| C3 | 0.0 | 7.8 | 0.0 | 0.0 | 0.0 | 0.0 |
| C4 | 0.0 | 16.7 | 1.1 | 0.0 | 0.0 | 0.0 |
| C5 | 0.0 | 1.1 | 1.1 | 0.0 | 0.0 | 4.4 |
| C6 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| C7 | 0.0 | 1.1 | 1.1 | 0.0 | 0.0 | 0.0 |
| C8 | 0.0 | 2.2 | 0.0 | 0.0 | 0.0 | 1.1 |
| C9 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 2.2 |
| C10 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| C11 | 0.0 | 14.4 | 0.0 | 0.0 | 0.0 | 0.0 |
| J1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J2 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J3 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J4 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J5 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J6 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| J7 | 0.0 | 3.3 | 0.0 | 0.0 | 0.0 | 1.1 |
| J8 | 0.0 | 4.4 | 0.0 | 0.0 | 0.0 | 0.0 |
| J9 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J10 | 0.0 | 3.3 | 0.0 | 0.0 | 0.0 | 0.0 |
| J11 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| J12 | 0.0 | 15.1 | 1.1 | 0.0 | 0.0 | 0.0 |
| J13 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| M1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| M2 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 1.1 |
| M3 | 0.0 | 0.0 | 0.0 | 0.0 | 1.1 | 0.0 |
| M4 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| M5 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| M6 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| M7 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| M8 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| M9 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 1.8 |
| M10 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| M11 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 1.1 |
| M12 | 0.0 | 14.4 | 0.0 | 0.0 | 0.0 | 0.0 |
| N1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| N2 | 1.0 | 1.0 | 0.0 | 8.3 | 0.0 | 0.0 |
| N3 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 2.1 |
| N4 | 0.0 | 0.0 | 0.0 | 1.9 | 0.0 | 0.9 |
| N5 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 2.2 |
| N6 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| N7 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| N8 | 0.0 | 5.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| N9 | 0.0 | 5.6 | 2.2 | 0.0 | 0.0 | 0.0 |
| N10 | 0.0 | 2.2 | 0.0 | 0.0 | 0.0 | 0.0 |
| S1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 2.1 |
| S2 | 0.0 | 1.0 | 0.0 | 0.0 | 0.0 | 4.0 |
| S3 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 8.4 |
| S4 | 0.0 | 1.0 | 0.0 | 0.0 | 0.0 | 12.2 |
| S5 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 22.1 |
| S6 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 3.2 |

Aleiodes sp., *Chelonus* sp. Probably *insularis* Cresson, *Cotesia* sp. probably *marginiventris* Cresson, *Glyptapanteles* sp. probably *militaris* Walsh, *Homolobus* sp. probably *mellea* Cresson, *Meteorus* sp. probably *laphygmae* Viereck.

TABLE 2. (CONTINUED) PERCENTAGE OF *SPODOPTERA FRUGIPERDA* LARVAE PARASITIZED BY EACH SPECIES OF BRACONIDAE AT EACH LOCATION.

| Code* | Braconidae | | | | | |
|-------|-----------------|-----------------|----------------|-----------------------|------------------|-----------------|
| | <i>Aleiodes</i> | <i>Chelonus</i> | <i>Cotesia</i> | <i>Glyptapanteles</i> | <i>Homolobus</i> | <i>Meteorus</i> |
| S7 | 0.0 | 2.0 | 0.0 | 0.0 | 0.0 | 10.0 |
| S8 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 6.5 |
| V1 | 0.0 | 3.3 | 0.0 | 0.0 | 0.0 | 1.1 |
| V2 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| V3 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| V4 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| V5 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| V6 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| V7 | 0.0 | 0.0 | 0.0 | 1.7 | 0.0 | 0.0 |
| V8 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| V9 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| V10 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |

Aleiodes sp., *Chelonus* sp. Probably *insularis* Cresson, *Cotesia* sp. probably *marginiventris* Cresson, *Glyptapanteles* sp. probably *militaris* Walsh, *Homolobus* sp. probably *mellea* Cresson, *Meteorus* sp. probably *laphygmae* Viereck.

Glyptapanteles sp. was found in Nayarit in two collections, N2 and N4, with parasitization rates of 8.3% and 1.9%, respectively, and in one location in Veracruz (V7) with 1.7% of parasitism rate. Rohlf's & Mack (1985), and Cave (1993) reported the occurrence of this parasitoid attacking FAW larvae in the US and Honduras, collected from sorghum and maize, respectively. Steffey (2001) reported *G. militaris* attacking armyworms and other caterpillars in Illinois. He speculated that this braconid and other natural enemies could suppress armyworm populations and keep them well below economic levels. Recently, Reis et al. (2003) suggested that the parasitoid may be well adapted to the Azorean agricultural systems in Portugal, characterized by prevalence of the grass, *Lolium perenne* L., throughout the year. The armyworm, *Pseudaletia unipuncta* (Haworth) when fed on fresh leaves of *L. perenne* is the most suitable host for the mass rearing of this braconid.

Aleiodes sp. occurred only in one collection in Nayarit (N2), and *Homolobus* sp. was found in Michoacán (M3), and their parasitism was lower than 1.2%. Ruíz-Cancino (1991) reported species of *Rogas* (Syn: *Aleiodes*) occurring in "La Reserva de la Biosfera El Cielo" in Tamaulipas, Mexico, and the family Braconidae is the second more abundant with 10% of the individuals, these braconids were attacking insect pest of annual, perennial and ornamental crops. *Aleiodes laphygmae* was reported by Molina-Ochoa et al. (2001) with a low parasitism rate (0.3%) on FAW larvae in Tamaulipas, Mexico. This braconid, *A. laphygmae* was the most abundant parasitoid attacking FAW larvae (12.8% parasitism) in South Georgia (Riggin et al. 1993).

Homolobus sp. probably *mellea* (Cresson), syn: *Zelee mellea* (Cresson) was previously found in

small numbers attacking FAW larvae in Honduras (Cave 1993), Nicaragua (Huis 1981) and the US (Vickery 1929; Wilson 1933; Riggin et al. 1992), but was not previously reported in Mexico. Parasitism by this species was low (1.1%), but finding it contributes to our knowledge on the occurrence and diversity of beneficial insects affecting FAW populations in Michoacán.

The ichneumonid parasitoids, *O. flavidus*, *P. spinator*, and *C. flavicincta* were the most frequently reared species in 18, 17, and 14 of the 64 collections, respectively. *Ophion flavidus* was recovered in more locations in Michoacán, and Colima (5 and 4 locations, respectively), but the highest parasitism rate for a single location was obtained in Colima (C7) with 6.7%. Similar results were reported by Molina et al. (2001), and Riggin et al. (1993). Recently, Molina-Ochoa et al., (2003b) listed the occurrence of *O. flavidus* in Argentina, Brazil, Honduras, Mexico, Nicaragua, and the US. Ashley et al. (1983) reported that *Ophion* sp. attacked FAW larvae developing on volunteer corn and Paragrass at Homestead, Florida. Gross & Pair (1991) emphasized that the tachinid *Archytas marmoratus* (Townsend) and *O. flavidus* provide opportunities for advancing biological strategies for managing FAW, with the development of economical methods for mass-propagation.

P. spinator was the second most widely distributed ichneumonid parasitoid. It was recovered in 17 of the 64 collections, 7 in Colima, 2 in Jalisco, 4 in Michoacán, and Nayarit, but this species was not recovered from Sinaloa, and Veracruz. The highest rate of parasitism for a single location was obtained in Michoacán (M12) with 22.2%. *Pristomerus spinator* has been reported in Mexico occurring in Quintana Roo, Tamaulipas (Carrillo 1980), and Michoacán, Colima, and Jalisco (Molina-Ochoa et al.

TABLE 3. PERCENTAGE OF *Spodoptera frugiperda* LARVAE PARASITIZED BY EACH SPECIES OF ICHNEUMONIDAE AND EU-
LOPHIDAE AT EACH LOCATION.

| Code* | Ichneumonidae | | | | Eulophidae | | |
|-------|---------------|-----|-----|------|------------|-----|------|
| | C.f | E.v | O.f | P.s | A.sp | E.p | H.sp |
| C1 | 0.0 | 0.0 | 0.0 | 12.2 | 0.0 | 0.0 | 0.0 |
| C2 | 0.0 | 2.2 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 |
| C3 | 0.0 | 0.0 | 0.0 | 4.4 | 0.0 | 0.0 | 0.0 |
| C4 | 0.0 | 1.1 | 0.0 | 14.4 | 0.0 | 0.0 | 0.0 |
| C5 | 0.0 | 0.0 | 5.6 | 1.1 | 0.0 | 0.0 | 0.0 |
| C6 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| C7 | 0.0 | 0.0 | 6.7 | 1.1 | 0.0 | 0.0 | 0.0 |
| C8 | 0.0 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| C9 | 0.0 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| C10 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| C11 | 0.0 | 0.0 | 0.0 | 6.7 | 0.0 | 0.0 | 0.0 |
| J1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J2 | 0.0 | 0.0 | 0.0 | 2.2 | 0.0 | 0.0 | 0.0 |
| J3 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J4 | 0.0 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| J5 | 0.0 | 0.0 | 0.0 | 3.3 | 0.0 | 0.0 | 0.0 |
| J6 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J7 | 1.1 | 0.0 | 4.4 | 0.0 | 0.0 | 0.0 | 0.0 |
| J8 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J9 | 2.1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J10 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J11 | 3.3 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| J12 | 3.2 | 0.0 | 2.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| J13 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 1.1 | 0.0 |
| M1 | 3.3 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| M2 | 2.2 | 0.0 | 0.0 | 3.3 | 0.0 | 0.0 | 0.0 |
| M3 | 0.0 | 0.0 | 3.3 | 1.1 | 0.0 | 0.0 | 0.0 |
| M4 | 0.0 | 0.0 | 1.1 | 1.1 | 0.0 | 0.0 | 0.0 |
| M5 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| M6 | 0.0 | 3.3 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| M7 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| M8 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 1.6 | 0.0 |
| M9 | 0.0 | 0.0 | 1.8 | 0.0 | 0.0 | 0.0 | 0.0 |
| M10 | 0.0 | 0.0 | 1.1 | 0.0 | 0.0 | 1.1 | 0.0 |
| M11 | 0.0 | 2.2 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| M12 | 0.0 | 5.6 | 0.0 | 22.2 | 0.0 | 0.0 | 0.0 |
| N1 | 2.2 | 0.0 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 |
| N2 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| N3 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| N4 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| N5 | 1.1 | 0.0 | 1.1 | 3.3 | 0.0 | 0.0 | 0.0 |
| N6 | 0.0 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| N7 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| N8 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| N9 | 3.3 | 0.0 | 0.0 | 7.8 | 0.0 | 0.0 | 0.0 |
| N10 | 2.2 | 0.0 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 |
| S1 | 0.0 | 0.0 | 2.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| S2 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| S3 | 0.0 | 0.0 | 1.1 | 0.0 | 0.0 | 0.0 | 0.0 |
| S4 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |
| S5 | 0.0 | 0.0 | 1.1 | 0.0 | 0.0 | 4.2 | 0.0 |
| S6 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 |

C.f = *Campoletis flavicincta* Ashmead, E.v = *Eiphosoma vitticolle* Cresson, O.f = *Ophion flavidus* Brulle, P.s = *Pristomerus spinator* Fabricius, A.sp. = *Aprostocetus* sp., E.p = *Euplectrus plathypenae* Howard, H.sp. = *Horismenus* sp.

TABLE 3. (CONTINUED) PERCENTAGE OF *Spodoptera frugiperda* LARVAE PARASITIZED BY EACH SPECIES OF ICHNEUMONIDAE AND EULOPHIDAE AT EACH LOCATION.

| Code* | Ichneumonidae | | | | Eulophidae | | |
|-------|---------------|-----|-----|-----|------------|------|------|
| | C.f | E.v | O.f | P.s | A.sp | E.p | H.sp |
| S7 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 1.0 | 0.0 |
| S8 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 1.1 | 0.0 |
| V1 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 2.2 | 0.0 |
| V2 | 0.0 | 0.0 | 0.0 | 0.0 | 3.0 | 6.1 | 3.0 |
| V3 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 3.1 | 0.0 |
| V4 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 11.5 | 0.0 |
| V5 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 4.2 | 0.0 |
| V6 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 2.2 | 0.0 |
| V7 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 5.1 | 0.0 |
| V8 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 4.4 | 0.0 |
| V9 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 8.9 | 0.0 |
| V10 | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 6.1 | 0.0 |

C.f = *Campoletis flavicincta* Ashmead, E.v = *Eiphosoma vitticole* Cresson, O.f = *Ophion flavidus* Brulle, P.s = *Pristomerus spinator* Fabricius, A.sp. = *Aprostocetus* sp., E.p = *Euplectrus plathypenae* Howard, H.sp. = *Horismenus* sp.

2001). Two collections from Michoacán during 1998 and 2000 exhibited the highest parasitism rates for a single location (El Hueso, and El Cirián, Nueva Italia) with 12.7%, and 22.2%, respectively. The ichneumonid was previously reported from Brazil, Honduras, Mexico, Nicaragua, and the US (Molina-Ochoa et al. 2003b).

Campoletis flavicincta was found in 14 of 64 collections, one in Colima, 5 in Jalisco, 3 in Michoacán, and 5 in Nayarit, but it was not recovered in Sinaloa, and Veracruz. *Campoletis flavicincta* had an overall parasitism range from 0 to 3.3%. The highest parasitism rate for a single location was obtained in N9. In a previous survey conducted by Molina-Ochoa et al. (2001), *C. flavicincta* accounted for 23% of parasitism in El Batillero, Michoacán, a location surrounded by avocado orchards and pine forest near to Apo, Michoacán; however, the FAW larvae from nearby locations in this survey (M1 and M2) showed low parasitism rates (3.3%, and 2.2%, respectively) by this parasitoid. It appears that, *C. flavicincta*, prefers or was associated with locations with high altitude; in this survey, it was found in locations with altitudes with an average of 1417 meters, as well as in locations near forests mainly constituted with pine and oak trees. Molina-Ochoa et al. (2003b) reported *C. flavicincta* occurring in Brazil, Honduras, Mexico, Nicaragua, and the US. This species was also reported attacking beet armyworm larvae fed on cotton in Georgia, USA (Ruberson et al. 1993, 1994).

Eiphosoma vitticole was the ichneumonid with the most limited distribution in this survey, found in 6 of the 64 collections. *E. vitticole* occurred in 2 locations in Colima, 3 locations in Michoacán, and 1 location in Nayarit. The highest rate of parasitism for a single location was re-

corded in M12 with 5.6%. This species showed low parasitism rates, and it was not found in Jalisco, Sinaloa, and Veracruz. It was collected from locations with an average altitude of 472m, with a range between 255 and 744m. Pair et al. (1986) reported the occurrence of *E. vitticole* in Texas, and Tamaulipas, Mexico. It also has been reported from Bolivia, Brazil, Colombia, Honduras, and Nicaragua (Molina-Ochoa et al. 2003b)

Three species of eulophid parasitoids were found in this survey, *Aprostocetus* sp., *Euplectrus plathypenae* Howard, and *Horismenus* sp. *Euplectrus plathypenae* was the most widely distributed eulophid, occurring in 16 of the 64 collections. It was found in Veracruz in all collections (10), Sinaloa in 3 collections, 2 in Michoacán, and one in Jalisco. Molina-Ochoa et al. (2001) reported a parasitism rate of 8.3% by *E. plathypenae* in a single collection in El Mante, Tamaulipas, similar rates in several locations in Veracruz, and low rate of about 1% in Michoacán. We also did not find levels higher than 1.6% in Michoacán; however, we found a range of parasitism in Sinaloa between 1% and 4.2%. The highest level of parasitism for a single location was obtained in the location V4 with 11.5%. Montoya-Burgos (1980) reported natural parasitism of about 15% by *Euplectrus* sp. against L2 FAW developing on corn in Veracruz. *Euplectrus plathypenae* is an important and well distributed parasitoid in the tropical Americas, and the US (Molina-Ochoa et al. (2003b).

The other eulophids, *Aprostocetus* sp. and *Horismenus* sp., occurred only in the location V2, with a parasitism rate of 3.0% for both species. This is the first report of *Aprostocetus* sp. and *Horismenus* sp. as parasitoids of FAW larvae. *Aprostocetus* sp. has been reported as a hyperparasitoid of *Gelechia senticetella* (Stgr.) (Lepidoptera:

Gelechiidae) fed on *Juniperus excelsa* in Bulgaria (Mirchev et al. 2001). *Aprostocetus* sp. also was reported as an egg parasitoid of mango leafhoppers (Fasih & Srivastava 1990). *Aprostocetus diplosis* Crawford is a parasitoid of *Stenodiplosis sorghicola*, a dipterous pest of sorghum in Brazil (Campos et al. 1998). *Horismenus* sp. has been reported to be a parasitoid of prepupae and pupae of the Citrus leafminer, *Phyllocnistis citrella* (Lepidoptera: Gracillariidae) in Mexico (Perales et al. 1996, Bautista-Martínez et al. 1998). Coffelt & Schultz (1993) mentioned that it is very common to find species of this genus acting as hyperparasitoids.

Our results demonstrate that hymenopteran parasitoids of FAW differentially occurred throughout the six Mexican states surveyed. However, this may have been influenced by the size of the FAW larvae collected. The hymenopteran parasitoids caused significant mortality of FAW larvae in most of the localities of this survey. It is important to highlight the occurrence and role on the FAW larval mortality caused by the braconids, *C. insularis*, and *M. laphygmae*, the ichneumonids, *O. flavidus*, *P. spinator*, and *C. flavicincta*, as well as the eulophid *E. plathypenae*. Our findings agree with Ashley (1986) in that no single parasitoid species exerted significant mortality throughout a major portion of the range of FAW. Another important aspect to note is the need for more taxonomic studies on two genera, *Chelonus* and *Meteorus*, which are important sources of mortality for FAW larvae.

ACKNOWLEDGMENTS

The authors thank Dr. R. W. Carlson, Dr. E. E. Grissell, Dr. P. M. Marsh, Dr. N. E. Woodley, and Dr. N. W. Gates (USDA-ARS, Systematic Entomology Laboratory, Beltsville, MD) for the insect identifications, and J. J. Molina-Cárdenas, M. A. Rodríguez-Vega, and F. Arceo-Palacios (Universidad de Colima, Tecmán, Colima) for assistance during the collections and preparation of the specimens. The authors express gratitude to Dr. Carlos Salazar-Silva, Rector of the Universidad de Colima, and CONACYT-Mexico for supporting this research, and for a grant to the senior author, respectively. This paper is a contribution of the Universidad de Colima-Facultad de Ciencias Biológicas y Agropecuarias, Tecmán, Colima, México, the USDA-ARS Crop Protection & Management Research Laboratory, Tifton, GA 31793, and the University of Nebraska Agricultural Research Division, Lincoln, NE 68583, Journal Series No.14482, Department of Entomology, University of Nebraska Lincoln. The authors also thank Dr. John J. Hamm USDA-ARS, Crop Protection & Management Research Laboratory, P.O. Box 748, Tifton, GA 31793-0748, USA, and Dr. John R. Ruberson, Department of Entomology, University of Georgia Tifton, Tifton, GA 31793, USA, for critical review of the manuscript.

REFERENCES CITED

ADAMCZYK, J. J., JR., J. W. HOLLOWAY, B. R. LEONARD, AND J. B. GRAVES. 1997. Susceptibility of fall army-

worm collected from different plant hosts to selected insecticides and transgenic Bt cotton. J. Cotton Sci. 1: 21-28.

- ALTIERI, M. A. 1991. Increasing biodiversity to improve insect pest management in agro-ecosystems. Chapter XIV, pp. 165-182 In D. L. Hawksworth [ed.] The Biodiversity of Microorganisms and Invertebrates: Its Role in Sustainable Agriculture. CAB International, Wallingford, UK.
- ALTIERI, M. A. 1994. Biodiversity and pest management in agroecosystems. Haworth Press, New York.
- ALVARADO-RODRÍGUEZ, B. 1987. Parasites and disease associated with larvae of beet armyworm *Spodoptera exigua* (Lepidoptera: Noctuidae), infesting processing tomatoes in Sinaloa, Mexico. Florida Entomol. 70: 444-449.
- ANDREWS, K. L. 1988. Latin American Research on *Spodoptera frugiperda* (Lepidoptera: Noctuidae). Florida Entomol. 71: 630-653.
- ASHLEY, T. R., C. S. BARFIELD, V. H. WADDILL, AND E. R. MITCHELL. 1983. Parasitization of fall armyworm larvae on volunteer corn, Bermudagrass, and paragrass. Florida Entomol. 66: 267-271.
- ASHLEY, T. R. 1986. Geographical distributions and parasitization levels for parasitoids of the fall armyworm, *Spodoptera frugiperda*. Florida Entomol. 69: 516-524.
- BAUTISTA-MARTÍNEZ, N., J. L. CARRILLO-SÁNCHEZ, H. BRAVO-MOJICA, AND S. D. KOCH. 1998. Natural parasitism of *Phyllocnistis citrella* (Lepidoptera: Gracillariidae) at Cuiclahuac, Veracruz, México. Florida Entomol. 81: 30-37.
- BOSSART, J. L., AND D. P. PROWELL. 1998. Genetic estimates of population structure and gene flow: limitations, lessons and new directions. Tree 13:202-206.
- BURTON, R. L., AND W. D. PERKINS. 1989. Rearing the corn earworm and fall armyworm for maize resistance studies, pp. 37-45 In CIMMYT, 1989. Toward Insect Resistant Maize for Third World: Proceedings of the International Symposium on Methodologies for Developing Host Plant Resistance to Maize Insects, México, D. F., CIMMYT.
- CAMPOS, A. R., F. M. LARA, AND O. R. CAMPOS. 1998. Influencia de genótipos de sorgo sobre a mosca *Stenodiplosis sorghicola* (Diptera: Cecidomyiidae) e seus parasitoides *Aprostocetus diplosis* Crawford, 1907 (Hymenoptera: Eulophidae). Cult. Agron. 7: 91-100.
- CARRILLO, H. 1980. Determinación del parasitismo natural de gusano cogollero, *Spodoptera frugiperda* (J. E. Smith) en Quintana Roo. Folia Entomol. Méx. 45: 111-112.
- CAVE, R. D. 1993. Parasitoides larvales y pupales de *Spodoptera frugiperda* (Smith) (Lepidoptera: Noctuidae) en Centro América con una clave para las especies encontradas en Honduras. Ceiba 34: 33-56.
- COFFELT, M. A., AND P. B. SCHULTZ. 1993. Larval parasitism of orangestriped oakworm (Lepidoptera: Saturniidae) in the urban shade tree environment. Dept. Entomology. University of Virginia. Biological Control 3: 127-134.
- FASIH, M., AND R. P. SRIVASTAVA. 1990. Parasites and predators of insect pest of mango. Int. Pest. Control 32: 39-41.
- GROSS, H. R., JR., AND S. D. PAIR. 1986. The fall armyworm: Status and expectations of biological control with parasitoids and predators. Florida Entomol. 69: 502-515.
- GROSS, H. R., AND S. D. PAIR. 1991. Seasonal distribution, response to host developmental stage, and

- screened-cage performance of *Archytas marmoratus* (Diptera: Tachinidae) and *Ophion flavidus* (Hymenoptera: Ichneumonidae) on *Spodoptera frugiperda* (Lepidoptera: Noctuidae) Florida Entomol. 74: 237-245.
- HUIS, A. V. 1981. Integrated pest management in the small farmer's maize crop in Nicaragua. Med. Landb. Wageningen 81: 221 pp.
- JOHNSON, D. E., F. J. SEIDLER, AND T. A. SLOTKIN. 1998. Early biochemical detection of delayed neurotoxicity resulting from developmental exposure to chlorpyrifos. Brain Research Bulletin 45: 143-147.
- LEWIS, W. J., AND D. A. NORDLUND. 1980. Employment of parasitoids and predators for fall armyworm control. Florida Entomol. 63: 433-438.
- LEWIS, W. J., J. C. VAN LENTEREN, S. C. PHATAK, AND J. H. TUMLINSON, III. 1997. A total system approach to sustainable pest management. Proc. Natl. Acad. Sci. 94: 12243-12248.
- LEZAMA-GUTIÉRREZ, R., J. J. HAMM, J. MOLINA-OCHOA, M. LÓPEZ-EDWARDS, A. PESCADOR-RUBIO, M. GONZÁLEZ-RAMÍREZ, AND E. STYER. 2001. Occurrence of entomopathogens of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in the Mexican States of Michoacán, Colima, Jalisco and Tamaulipas. Florida Entomol. 84: 23-30.
- LEVY, H. C., A. GARCÍA-MARUNIAK, AND J. E. MARUNIAK. 2002. Strain identification of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) insects and cell line: PCR-RFLP of cytochrome oxidase C subunit I gene. Florida Entomol. 85: 186-190.
- LÓPEZ-EDWARDS, M., J. L. HERNÁNDEZ-MENDOZA, A. PESCADOR-RUBIO, J. MOLINA-OCHOA, R. LEZAMA-GUTIÉRREZ, J. J. HAMM, AND B. R. WISEMAN. 1999. Biological differences between five populations of fall armyworm (Lepidoptera: Noctuidae) collected from corn in Mexico. Florida Entomol. 82: 254-262.
- LU, Y., AND M. J. ADANG. 1996. Distinguishing fall armyworm (Lepidoptera: Noctuidae) strains using a diagnostic mitochondrial DNA marker. Florida Entomol. 79: 48-55.
- LUGINBILL, P. 1928. The fall armyworm. U.S. Dep. Agric. Tech. Bull. No. 34.
- MATTSON, J. L., J. P. J. MAURISSEN, R. J. NOLAN, AND D. K. A. BRZAK. 2000. Lack of differential sensitivity to cholinesterase inhibition in fetuses and neonates compared to dams treated perinatally with chlorpyrifos. Toxicological Sciences 53: 438-446.
- MEAGHER, R. L. JR., AND M. GALLO-MEAGHER. 2003. Identifying host strains of fall armyworm (Lepidoptera: Noctuidae) in Florida using mitochondrial markers. Florida Entomol. 86: 450-455.
- MIRCHEV, P., T. GEORGIEV, AND G. TSANKOV. 2001. Studies on the parasitoids of *Gelechia senticetella* (Stgr)(Lepidoptera: Gelechiidae) in Bulgaria. Anzeiger für Schädlingkunde 74: 94-96.
- MOLINA-OCHOA, J. J., J. HAMM, R. LEZAMA-GUTIÉRREZ, M. LÓPEZ-EDWARDS, M. GONZÁLEZ-RAMÍREZ, AND A. PESCADOR-RUBIO. 2001. A survey of fall armyworm (Lepidoptera: Noctuidae) parasitoids in the Mexican States of Michoacán, Colima, Jalisco, and Tamaulipas. Florida Entomol. 84: 31-36.
- MOLINA-OCHOA, J., R. LEZAMA-GUTIÉRREZ, M. RAMÍREZ-GONZÁLEZ, M. LÓPEZ-EDWARDS, M. A. RODRÍGUEZ-VEGA, AND F. ARCEO-PALACIOS. 2003a. Pathogens and parasitic nematodes associated with populations of fall armyworm (Lepidoptera: Noctuidae) larvae in México. Florida Entomol. 86(3): 244-253.
- MOLINA-OCHOA, J., J. E. CARPENTER, E. A. HEINRICH, AND J. E. FOSTER. 2003b. Parasitoids and parasites of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in the Americas and Caribbean Basin: an inventory. Florida Entomol. 86: 254-289.
- MONTOYA-BURGOS, J. A. 1980. Observaciones sobre la importancia de *Euplectrus* sp., en la integración de métodos de control de plagas de maíz. VIII Reunión Nacional de Control Biológico. Secretaría de Agricultura y Recursos Hidráulicos, Sanidad vegetal, 22-25 de abril de 1980. Manzanillo, Colima, México, 7 pp.
- NAGOSHI, R. N., AND R. L. MEAGHER. 2003. FR Tandem—reapet sequence in fall armyworm (Lepidoptera: Noctuidae) host strains. Ann. Entomol. Soc. Am. 96: 329-335.
- PAIR, S. D., J. R. RAULSTON, A. N. SPARKS, AND P. B. MARTIN. 1986. Fall armyworm (Lepidoptera: Noctuidae) parasitoids: differential spring distribution and incidence on corn and sorghum in the Southern United States and Northeastern Mexico. Environ. Entomol. 15: 342-348.
- PASHLEY, D. P. 1988. Current status of fall armyworm host strains. Florida Entomol. 71: 227-234.
- PASHLEY, D. P., T. C. SPARKS, S. S. QUISENBERRY, T. JAMJANYA, AND P. F. DOWD. 1987. Two fall armyworm strains feed on corn, rice and bermudagrass. Louisiana Ag. 30: 8-9.
- PERALES, G. A., H. C. ARRENDONDO B., AND E. GARZA G. 1996. Native parasitoids of citrus leafminer *Phyllocnistis citrella* in Colima, Mexico. Southwestern Entomol. 21: 349-350.
- REIS, J., L. OLIVERA, AND P. GARCÍA. 2003. Effects of the larval diet of *Pseudaletia unipuncta* (Lepidoptera: Noctuidae) on the performance of the parasitoid *Glyptapanteles militaris* (Hymenoptera: Braconidae). Environ. Entomol. 32: 180-186.
- RIGGIN, T. M., B. R. WISEMAN, D. J. ISENHOUR, AND K. E. ESPELLE. 1992. Incidence of fall armyworm (Lepidoptera: Noctuidae) parasitoids on resistant and susceptible corn genotypes. Environ. Entomol. 21: 888-895.
- RIGGIN, T. M., K. E. ESPELIE, B. R. WISEMAN, AND D. J. ISENHOUR. 1993. Distribution of fall armyworm (Lepidoptera: Noctuidae) parasitoids on five corn genotypes in South Georgia. Florida Entomol. 76: 292-302.
- ROHLFS, W. M., AND T. P. MACK. 1985. Seasonal parasitism rates, host size, and adult emergence pattern of parasitoids of the fall armyworm, *Spodoptera frugiperda* (J. E. Smith), with emphasis on *Ophion flavidus* Brulle (Hymenoptera: Ichneumonidae). Ann. Entomol. Soc. Am. 78: 217-220.
- RUBERSON, J. R., G. A. HERZOG, AND W. J. LEWIS. 1993. Parasitism of the beet armyworm, *Spodoptera exigua*, in South Georgia cotton. Proc. 1993. Beltwide Cotton Prod. Conf. 3: 993-997.
- RUBERSON, J. R., G. A. HERZOG, W. R. LAMBERT, AND W. J. LEWIS. 1994. Management of the beet armyworm (Lepidoptera: Noctuidae) in cotton: role of natural enemies. Florida Entomol. 77: 440-453.
- RUÍZ-CANCINO, E. 1991. Hábitos parasíticos y alimenticios de las Familias de himenópteros de La Reserva de la Biosfera "El Cielo" de Tamaulipas. 8 pp. <http://ecologia.uat.mx/biotam/v2n3/art1.html>
- SOLOMON, G. M., AND T. SCHETTLER. 2000. Environment and health: 6. Endocrine disruption and potential human health implications. Canadian Medical Association or its Licensors (CMAL) 163: 1471-1476.

- STEFFEY, K. 2001. Parasitoids of armyworm have been common. <http://www.ag.uiuc.edu/cespubs/pest/articles/200114d.html>
- VICKERY, R. A. 1929. Studies of the fall armyworm in the Gulf Coast district of Texas. U.S. Dep. Agric. Tech. Bull. No. 138.
- WILSON, J. W. 1933. The biology of parasites and predators of *Laphygma exigua* Huebner reared during the season of 1932. Florida Entomol. 17: 1-15.
- YU, S. J. 1991. Insecticide resistance in the fall armyworm, *Spodoptera frugiperda* (J. E. Smith). Pestic. Biochem. Physiol. 39: 84-91.
- YU, S. J. 1992. Detection and biochemical characterization of insecticide resistance in fall armyworm (Lepidoptera: Noctuidae). J. Econ. Entomol. 85: 675-682.