

# Host Specificity Tests of Gratiana graminea (Coleoptera:Chrysomelidae), a Potential Biological Control Agent of Tropical Soda Apple, Solanum viarum (Solanaceae)

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Source: Florida Entomologist, 93(2): 231-242

Published By: Florida Entomological Society

URL: https://doi.org/10.1653/024.093.0213

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## HOST SPECIFICITY TESTS OF *GRATIANA GRAMINEA* (COLEOPTERA: CHRYSOMELIDAE), A POTENTIAL BIOLOGICAL CONTROL AGENT OF TROPICAL SODA APPLE, *SOLANUM VIARUM* (SOLANACEAE)

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## Abstract

Multiple-choice and no-choice tests were conducted at the Department of Agriculture, Division of Plant Industry Quarantine facility in Gainesville, FL to determine the specificity of the Brazilian leaf-beetle Gratiana graminea Klug, a candidate for biological control of Solanum viarum, tropical soda apple. One hundred fifteen plant species in 32 families were included in the feeding-oviposition multiple-choice tests including the target weed and the 5 major cultivated Solanaceae Capsicum annuum L., Lycopersicon sculentum Mill., Nicotiana tabacum L., Solanum melongena L., and Solanum tuberosum L. Eight to 12 plant species, including always the main target weed, growing in 1-gallon pots were simultaneously exposed to 20 G. graminea adults (10 males and 10 females that most of the time had recently emerged from pupae) in an aluminum cage  $(60 \times 60 \times 60 \text{ cm})$ . At the beginning of each test the insects were placed at the bottom center of each cage to allow them to orient by themselves to the tested plants. Plant species in each test were replicated 3-4 times (one replication of tested plants in each separate cage). Plants tested were exposed to G. graminea adults from 3-6 weeks. Observation of oviposition and feeding were made during almost all the weekdays. No-choice host specificity tests were conducted with G. graminea adults on potted plants in cages made of clear-plastic cylinders and with G. graminea larvae placed on cluster of leaves of each individual plant tested. Ten G. graminea adults were exposed to 29 plant species individually tested during 3 to 5 weeks, and 10 neonate larvae were exposed to 31 plant species. Plant species in each test were replicated 3-4 times. Results indicated that G. graminea fed and developed only on the target weed. The tests indicated that a host range expansion of G. graminea to any of the major cultivated Solanaceae species is highly unlikely. A petition for field release in Florida was submitted to the Technical Advisory Group for Biological Control Agents of Weeds (TAG) in Sep 2008.

Key Words: weed biological control, host-specificity tests, *Gratiana graminea*, *Solanum viarum*, tropical soda apple, Solanaceae

#### RESUMEN

Pruebas de ovoposición y alimentación (con y sin elección), se realizaron para evaluar la especificidad del escarabajo defoliador, de origen Brasileiro, Gratiana graminea Klug como agente potencial para el control biológico de tropical soda apple, Solanum viarum Dunal en los Estados Unidos. Las pruebas se efectuaron en la cuarentena del Departamento de Agricultura de la Florida, División de Industria de Plantas en Gainesville. Ciento quince especies de plantas, en 32 familias, fueron incluidas en las pruebas de especificidad de elección multiple, incluyendo la maleza objetivo y las cinco plantas cultivadas pertenecients a la familia Solanaceae más importantes: Capsicum annuum L., Lycopersicon sculentum Mill., Nicotiana tabacum L., Solanum melongena L., y Solanum tuberosum L. En cada prueba se utilizaron de ocho a doce plantas, icluyendo siempre la maleza objetivo, creciendo en macetas de un galón las cuales fueron expuestas a 20 adultos de G. graminea (10 machos y 10 hembras recién emergidos de pupa) durante 3 a 6 semanas. Registros de alimentación y ovoposición fueron realizados casi todos los días de la semana. Pruebas de alimentación/ovoposición sin elección fueron tambien realizadas usando plantas creciendo en macetas y jaulas cilíndricas hechas de plástico claro transparente. Diez adultos de G. graminea fueron expuestos a 29 especies de plantas en forma individual durante 3 a 5 semanas. Diez larvas recién nacidas fueron también expuestas a 31 especies de plantas en forma individual. Cada prueba tuvo 3-4 repeticiones. Los resultados indicaron que G. graminea se alimentó, colocó posturas y completó su desarrollo únicamente en la maleza objetivo. Las pruebas de especificidad indicaron que la posibilidad de G. graminea de llegar a ser una plaga de las Solanaceas cultivadas es muy remota. La solicitud a TAG para liberar el escarabajo fue presentada en septiembre 2008.

Translation provided by the authors.

Tropical soda apple, Solanum viarum Dunal (Solanaceae), is a perennial weed, originally from northeast Argentina, southern Brazil, Paraguay, and Uruguay, that has been spreading throughout Florida at an alarming rate during the last two decades. The pasture-land infested in 1992 was estimated to be approximately 60,000 hectares (Mullahey et al. 1993), and increased to more than 300,000 hectares in 1995-96 (Mullahey et al. 1997). Currently, the infested area is estimated at more than 400,000 hectares (Medal et al. 2008). Tropical soda apple, first reported in the United States in Glades County, Florida in 1988 (Coile 1993; Mullahey & Colvin 1993), is also present in Alabama, Georgia, Mississippi, North Carolina, South Carolina, Texas, and Puerto Rico (Bryson & Byrd, Jr. 1996; Dowler 1996; Mullahey et al. 1997; Medal et al. 2003). The potential range of tropical soda apple in the United States may be extended even further based on studies of the effects of temperatures and photoperiods conducted by Patterson (1996) in controlled environmental chambers. This invasive exotic weed was placed on the Florida and Federal Noxious Weed Lists in 1995.

In addition to its invasion of pasture lands and reduction of cattle carrying capacity (Mullahey et al. 1993; Bredow et al. 2007), tropical soda apple is known to harbor at least 6 viruses that affect cultivated solanaceous crops such as tomato, tobacco, and pepper (McGovern et al. 1994a, 1994b, 1996). Tropical soda apple is also an alternative host for key pests such as the Colorado potato beetle, Leptinotarsa decemlineata (Say) (Coleoptera: Chrysomelidae), major defoliating insect pest of potato in North America; the tomato hornworm, Manduca quinquemaculata (Haworth) and the tobacco hornworm, Manduca sexta (L.), (Lepidoptera: Sphingidae), major pests of tomato and tobacco plants; the silverleaf whitefly, Bemisia argentifolii Bellows and Perring (Homoptera: Aleyrodidae) one of the most troublesome insect pest worldwide of many field and vegetable crops; the tobacco budworm, Heliothis virescens (Fabr.) (Lepdoptera: Noctuidae) one of the most destructive pests of tobacco; the green peach aphid, Myzus persicae (Sulzer) an important pest of peach trees and vector of plant viruses to solanaceous plants and other food crops (Homoptera: Aphididae); the southern green stinkbug, Nezara viridula (L.) (Hemiptera: Pentatomidae) an important pest of soybean and vegetable crops; and the suckfly, *Tupiocoris notatus* (Distant) (Hemiptera: Miridae) a pest of several crops including tobacco (Habeck et al. 1996; Medal et al. 1999a; Sudbrink et al. 1999). Although it is very difficult to estimate the real (direct and indirect) economic losses due to this invasive weed, the production loss to Florida ranchers by tropical soda apple was estimated from \$6.5-16 million annually (Thomas 2007).

Although tropical soda apple is able to spread vegetatively from the root system, the primary method of dispersal is by seed dissemination (Bryson et al. 1995; Medal et al. 1999b), which occurs mainly by livestock and wildlife that feed on the fruits and scarify the seeds (Akanda et al. 1996; Brown et al. 1996). A single plant of tropical soda apple can produces up to 150 fruits per year, with each fruit containing on average 400 seeds. The estimated seed production is 60,000 seeds/ plant/season with a viability of more than 75% (Mullahey & Colvin 1993; Pereira et al. 1997).

Currently recommended management practices for this invasive plant in southeastern United States include herbicide applications and mechanical techniques (mowing/tilling) (Mislevy et al. 1996; Mullahey et al. 1996; Sturgis & Colvin 1996; Akanda et al. 1997). These control tactics provide temporary weed suppression at an economic cost estimated at \$62 and \$47 per hectare in chemical and mechanical control methods, respectively (Thomas 2007). However, application of these control methods is often difficult to employ in remote and/or inaccessible areas.

A biological control project for tropical soda apple was started in Dec 1996 by the University of Florida in collaboration with the Universidade Estadual Paulista, Jabotical campus, Brazil, Universidade Federal do Parana in Curitiba, Brazil, Universidade Regional de Blumenau, Santa Catarina state, Brazil, Universidade Centro-Oeste in Irati, Parana state, Brazil, and the US-DAS-ARS, Biological Control Laboratory in Hurlingham, Buenos Aires Province, Argentina. The release of the Brazilian leaf-beetle Graminea graminea Klug (Coleoptera: Chrysomelidae) in Florida will complement the defoliation effects that Gratiana boliviana Spaeth has been making on tropical soda apple plants during the warm season in Florida since it was released in the summer 2003 (Medal et al. 2008; Overholt et al. 2008). Tropical soda apple defoliation by G. boliviana and G. graminea, in southern Brazil is causing a major suppressive effect on tropical soda apple density (Gandolfo et al. 2007; Medal et al. unpublished data). These two leaf-feeder beetles have a synergistic effect on tropical soda apple defoliation and occupy different niches in somewhat overlapped geographical regions in southern parts of Brazil.

In this paper we report the results of the hostspecificity tests conducted at the Florida Department of Agriculture-Division of Plant Industry quarantine facility in Gainesville with the leafbeetle *G. graminea* as a potential biological control agent of the non-native weed tropical soda apple.

### MATERIALS AND METHODS

Host-Feeding Specificity Tests

Plant-host specificity tests with *Gratiana* graminea adults and first instars were conducted

233

from Sep 2000 to Aug 2004 at the Florida Department of Agriculture and Consumer Services-Division of Plant Industry quarantine facility in Gainesville, Florida. Additional feeding/oviposition tests with *G. graminea* adults were conducted at the Gainesville quarantine facility from May to Sep 2008. *Gratiana graminea* (all developmental stages) were collected on tropical soda apple plants in Rio Grande do Sul, Brazil, introduced into Florida-quarantine, placed on caged plants of tropical soda apple growing in 1-gallon pots and eggs were removed twice a week to provide the insects required for testing.

## Multiple-Choice Feeding and Oviposition Tests

One hundred fifteen plant species in 32 families were included in the feeding and oviposition preference tests in quarantine (Table 1). The plants tested included 56 species in the family of the target weed (Solanaceae) of which 29 were from the genus Solanum and 27 from 15 other genera that include plants of agricultural or ecological importance. Ten species representing 5 families (Boraginaceae, Convolvulaceae, Ehretiaceae, Nolanaceae, Polemoniaceae) that are very closely related phyllogenetically to the Solanaceae and in the same order Polemoniales (Heywood 1993) were included. Forty-nine plant species representing 26 families, most of them with an economically and/or environmentally value in North America, were also tested. The major target weed (tropical soda apple), and 10 plant species in the Solanaceae were tested at least 3 times (Table 1). They included Solanum donianum Walpers that is in the list of Florida threatened plants (Coile 1998); 4 secondary target-weeds (Solanum tampicense Dunal, Solanum torvum Sw., Solanum capsicoides All., Solanum elaeagnifolium Cav.); and the 5 major cultivated Solanaceae (Capsicum annuum L., Lycopersicon esculentum Mill., Nicotiana tabacum L., Solanum melongena L., Solanum tuberosum L.). Eight to 12 plant species, including always the main target weed, growing in 1-gallon pots were simultaneously exposed to 20 G. graminea adults (10) males and 10 females which were newly emerged from pupae most of the time) in an aluminum cage  $(60 \times 60 \times 60 \text{ cm})$ . At the beginning of each test the insects were placed at the bottom center of each cage to allow them to orient by themselves to the tested plants. Plant species in each test were replicated 3-4 times (1 replication of tested plants in each separate cage). Plants were exposed to G. graminea adults from 3-6 weeks. Observations of oviposition and feeding were made during most of the weekdays. Plants consumed were replaced as needed. Plants were checked for oviposition sites and eggs were removed and counted weekly. On the last day of each experiment, plants were checked for feeding and eggs

laid on them. Leaf area consumed was measured with a Portable Area Meter Model LI-3000 (Lambda Instrument Corporation) and the leaffeeding area is reported on a scale from 0-5 (0 = no feeding, 1 = probing or <5% of leaf area consumed, 2 = light feeding or 5-20% of the area, 3 = moderate feeding or 21-40%, 4 = heavy feeding or 41-60%, and 5 = intense feeding or >60% of the leaf area consumed).

## No-Choice Larval Feeding Tests

No-choice host specificity tests were conducted with G. graminea neonate larvae in an environmental chamber at a temperature of  $22 \pm 2^{\circ}$ C, relative humidity of 55-65%, and a photoperiod of 12:12 (L:D). Recently hatched non-fed larvae were exposed to 31 plant species including 30 species in the family of the target weed (Solanaceae) and 1 species in the family Convolvulaceae. The species tested included 7 genera of plants very closely related phyllogenetically in the same family as the target weed, and with an economical and/or environmental value in North America (Table 2). Larvae were exposed to clusters of leaves of each individual plant tested by placing the clusters individually in a 30-mL plastic-cup containing water and fitted with a paper lid to avoid insect contact with the water. The leaf petiole was inserted through a hole (3-4 mm diameter) made in the middle of the paper-lid. The cup and plant cluster was placed inside a clear-plastic container covered with a plastic-lid having 6-7 small holes to allow air circulation. Moistened tissue paper was placed at the bottom of the plastic container and under the plastic-lid to provide moisture. The plants (treatments) were arranged in a completely randomized design. Three to 4 replications were used with 10 one-d- old larvae per replication. Each group of 10 G. graminea larvae was provided with only 1 plant species which they fed on or died. Daily observations of feeding were made and leaves were replaced as needed. Larval mortality counts were made 3 and 7 d after the experiment started.

## No-Choice Adult Feeding Tests

No-choice host specificity tests were conducted with *G. graminea* adults at the Gainesville quarantine facility with potted plants (20-60 cm height) in cages. *Gratiana graminea* adults were exposed to 29 plant species including *S. donianum* in the list of Florida threatened plants, all major cultivated Solanaceae, and 7 exotic (Table 3). Five to 6 plant species were individually tested each time due to limitation in availability of cages. Ten *G. graminea* adults (5 males, 5 females) per replication (3-4 replications) were exposed to plants during 21 to 35 d. Cages were made of clear plastic cylinders (15 cm diameter,

Plant Family Species   (*indicates native Solanum species)   No.   No.   Feeding Score <sup>1</sup> Eggs Laid per Femal     Category 1. Genetic types of the target weed species found in North America SOLANACEAE Tribe Solaneae Genus Solanum Subgenus Leptostemonum Section Acantophora Solanum viarum Dunal   Tropical soda apple   9   600   4-5   58-124     Category 2. Species in the same genus as the target weed, divided by subgenera (if applicable)   9   600   4-5   58-124
Category 1. Genetic types of the target weed species found in North America     SOLANACEAE     Tribe Solaneae     Genus Solanum     Subgenus Leptostemonum     Section Acantophora     Solanum viarum Dunal     Tropical soda apple   9     600   4-5     58-124
SOLANACEAETribe SolaneaeGenus SolanumSubgenus LeptostemonumSection AcantophoraSolanum viarum DunalTropical soda apple96004-558-124
Tribe Solaneae Genus SolanumSubgenus Leptostemonum Section Acantophora Solanum viarum DunalTropical soda apple96004-558-124
Genus SolanumSubgenus LeptostemonumSection AcantophoraSolanum viarum DunalTropical soda apple96004-558-124
Subgenus LeptostemonumSection AcantophoraSolanum viarum DunalTropical soda apple96004-558-124
Section AcantophoraSolanum viarum DunalTropical soda apple96004-558-124
Solanum viarum DunalTropical soda apple96004-558-124
Ualegory Z. Species in the same genus as the target weed invided by subgenera (i) applicable)
Tribe Solaneae
Genus Solanum
Subgenus Leptostemonum
Solanum capsicoides All. Red soda apple 3 220 1 0
Solanum mammosum L.Nipplefruit212010
Section Lasiocarpum
Solanum quitoense Lam. Naranjilla 2 120 0 0
Solanum pseudolulo Heise Falso Iulo 2 120 0 0
Solanum sessiliflorum Dunal Nightshade 2 120 0 0
Section Micracantha
Solanum tampicense Dunal Wetland nightshade 5 220 1 0
Solanum jamaicense Mill. Jamaican nightshade 2 120 0 0
Section Melongena
Subsection Lathyrocarpum
Solanum carolinense L. Horse nettle <sup>®</sup> 2 140 0 0
Solanum dimidiatum Raf. Western horsenettle <sup>*</sup> 2 140 0 0
Section Persicariae
Solanum bahamense Bahama nightshade 2 120 0 0
Solanum torvum Sw. Turkeyberry 6 420 1-3 0
Solanum verbascifolium L. Mullein nightshade <sup>*</sup> 2 140 0 0
Subgenus Solanum     Solanum americanum Mill.   American nightshade*   2   140   0   0
Solanum diphyllum L. Two-leaf nightshade <sup>*</sup> 2 140 0 0
Solanum erianthum Don. Potato tree <sup>®</sup> 2 140 0 0
Solanum jasminoides Paxt.Potato vine214000
Solanum mauritianum Scop.Earleaf nightshade214000
Solanum nigrescesns Mart. & Gal Divine nightshade <sup>*</sup> 2 140 0 0
Solanum nigrum L. Black nightshade <sup>*</sup> 2 160 0 0
Solanum parishii heller Parish nightshade <sup>*</sup> 2 140 0 0
Solanum ptycanthum Dunal Wonder berry <sup>*</sup> 2 140 0 0
Solanum seaforthianum Andr. Brazilian nightshade 2 140 0 0
Solanum tuberosum L. Potato 6 420 0 0
Category 3. Species in other genera in the same family as the target weed, divided by subfamily (if applicable) Genus <i>Acnistus</i>
Acnistus australe (Griseb.) Griseb. Acnistus 2 140 0 0 Genus Capsicum
Capsicum annuum L. Bell pepper 6 420 0 0
Capsicum frutescens L. Chile 2 140 0 0
Genus Iochroma
Iochroma sp. Iochroma 2 140 0 0
Genus Physalis
Physalis angulata L. Cutleaf Ground-Cherry 2 140 0 0
Physalis arenicola KearneyCypresshead214000

TABLE 1. HOST RANGE ADULT FEEDING AND OVIPOSITION TESTS WITH GRATIANA GRAMINEA IN FLORIDA.

Common Names

Each test included 3-4 replications with 20 adults (10 males, 10 females) per replication.

 $^{10}$  = no feeding, 1 = probing (<5% of leaf area), 2 = light feeding (5-20%), 3 = moderate feeding (21-40%), 4 = heavy feeding (41-60%), 5 = intense (60% of leaf-area).

Plant Family Species	Common Names (*indicates native Solanum species)	No. of Tests	No. of Insects	$\begin{array}{c} \mathbf{Feeding} \\ \mathbf{Score}^1 \end{array}$	Eggs Laid per Female
Physalis crassifolia Benth	Ground-Cherry	2	140	0	0
Physalis gigantea L.	Ground-cherry	2	140	0	0
Physalis ixocarpa Brot.	Tomatillo	2	140	0	0
Physalis pubescens L.	Strawberry tomato	2	120	0	0
Physalis walteri Nutt.	Ground-cherry	2	140	0	0
Tribe Daturae Genus <i>Brugmansia</i>					
Brugmansia sanguinea (Ruiz & Pav.) Don	Red floripontio	2	140	0	0
Genus Datura					
Datura discolor Bernh	Angels'trumpet	2	140	0	0
Datura metel L.	Downy thorn apple	2	140	0	0
Datura meteloides D.	Datura	2	140	0	0
Datura stramonium L.	Jimson weed	2	140	0	0
Tribe Lycieae					
Genus Lycium					
Lycium carolinianum Walt.	Christmas berry	2	140	0	0
<i>Lycium fremontii</i> Gray. Genus Lycopersicon	Lycium	2	140	0	0
Lycopersicon esculentum Mill. Tribe: Nicandreae	Tomato	6	420	0	0
Genus: Nicandra Nicandra physaloides (L.) Gaertn.	Apple of Peru	2	140	0	0
Tribe Nicotianae					
Genus Nicotiana		0	100		0
Nicotiana tabacum L.	Tobacco	6	420	0	0
Nicotiana rustica L.	Wild tobacco	2	160	0	0
Nicotiana sylvestris Speg. & Comes Genus Nierembergia	Tobacco	2	140	0	0
Nierembergia scoparia Sendtri Genus Petunia	Cupflower	2	140	0	0
Petunia × hybrida Tribe Salpiglossidae	Garden-petunia	2	140	0	0
Genus Salpiglossis Salpiglossis sinuata Ruiz & Pav Genus Schizanthus	Painted tongue	2	140	0	0
Schizanthus spp. Tribe Solandeae	Butterfly flower	2	140	0	0
Genus Solandra Solandra glandiflora Swartz	Chalice vine	2	140	0	0
Category 4. Threatened and endanger and subfamily	ed species in the same fam	ily as the ta	rget weed di <sup>,</sup>	vided by su	bgenus, genus,
Section Torva Solanum donianum Walpers	${\bf Mullein\ nightshade}^{*}$	4	300	0	0
Category 5. Species in other families in similarities to the target weed	n the same order that have	e some phylo	genetic, mor	phological,	or biochemical
BORAGINACEAE Heliotrope sp.	Heliotrope	1	80	0	0
1 1	Heliotrope Forget Mo Not	1			
Myosotis alpestris Schmidt CONVOLVULACEAE	Forget-Me-Not	1	80	0	0
Convolvulus purpurea L.	Convolvulus	1	80	0	0

TABLE 1. (CONTINUED) HOST RANGE ADULT FEEDING AND OVIPOSITION TESTS WITH GRATIANA GRAMINEA IN FLORIDA.

Each test included 3-4 replications with 20 adults (10 males, 10 females) per replication.  $^{10}$  = no feeding, 1 = probing (<5% of leaf area), 2 = light feeding (5-20%), 3 = moderate feeding (21-40%), 4 = heavy feeding (41-40%), 4 = heavy feeding (41-60%), 5 = intense (60% of leaf-area).

Plant Family Species	Common Names (*indicates native <i>Solanum</i> species)	No. of Tests	No. of Insects	$\mathbf{Feeding}$ $\mathbf{Score}^{1}$	Eggs Laid per Female
Ipomoea batata (L.) Lam.	Sweet-potato	2	160	0	0
Evolvulus muttallianus EHRETIACEAE	Evolvulus	1	80	0	0
Cordia sebestena L. NOLANACEAE	Geiger tree	1	80	0	0
Nolana paradoxa Lindl. POLEMONIACEAE	Chilean bellflower	1	80	0	0
Cobaea scandens Cav.	Cobaea	1	80	0	0
Gilia tricolor Benth	Bird's-eyes	1	80	0	0
Phlox panuculata L.	Phlox	1	80	0	0
Category 6. Species in other orders that that share the same habitat ACERACEAE					-
Acer rubrum L. ACTINIDIACEAE	Red maple	1	80	0	0
Actinidia deliciosa Liang & Fergusson ANACARDIACEAE	Kiwi vine	1	80	0	0
Anacardium occidentale L.	Cashew	1	80	0	0
Mangifera indica L.	Mango	1	80	0	0
Pistacia vera L. APIACEAE	Cultivated pistacho	1	80	0	0
Daucus carota L. ASTERACEAE	Carrot	1	80	0	0
Helianthus annuus L.	Annual sunflower	1	80	0	0
Lactuca sativa L. CAMPANULACEAE	Lettuce	1	80	0	0
Campanula persicifolia L CARICACEAE	Bell flower	1	80	0	0
Carica papaya L. CHENOPODIACEAE	Papaya	1	80	0	0
Beta vulgaris L. CRUCIFERAE	Beet	1	80	0	0
Brassica oleracea L. var. botrytis	Broccoli/Cauliflower	2	160	0	0
Brassica oleracea L. var. capitata	Cabbage	1	80	0	0
Raphanus sativus L. CUCURBITACEAE	Radish	1	80	0	0
Citrullus lanatus (Thumb)	Watermelon	1	80	0	0
Cucumis milo L.	Cantaloupe	1	80	0	0
Cucurbita pepo (L.) Alef.	Pumpkin/Squash	1	80	0	0
Cucurbita sativus L. EBENACEAE	Cucumber	1	80	0	0
Diospyros virginiana ERICACEAE	Persimmon	1	80	0	0
<i>Vaccinium ashei</i> Rende. FABACEAE	Rabbit-eye blueberry	1	80	0	0
Arachis hypogaea L	Peanut	1	80	0	0
Glycine max (L.) Merrill	Soybean	1	80	0	0
Phaseolus vulgaris L.	Pinto bean	1	80	0	0
Phaseolus lunatus L.	Snowpea	1	80	0	0
Pisum sativum L.	Cowpea	1	80	0	0
Vigna unguiculata (L.) Walp.	Lima bean	1	80	0	0

TABLE 1. (CONTINUED) HOST RANGE ADULT FEEDING AND OVIPOSITION TESTS WITH GRATIANA GRAMINEA IN FLORIDA.

Each test included 3-4 replications with 20 adults (10 males, 10 females) per replication.

 $^{10}$  = no feeding, 1 = probing (<5% of leaf area), 2 = light feeding (5-20%), 3 = moderate feeding (21-40%), 4 = heavy feeding (41-60%), 5 = intense (60% of leaf-area).

	Common Names (*indicates native	No.	No.	Feeding	Eggs Laid
Plant Family Species	Solanum species)	of Tests	of Insects	Score <sup>1</sup>	per Female
LAURACEAE					
Persea americana Mill.	Avocado	1	80	0	0
LOBELIACEAE					
Lobelia cardinalis L.	Cardinal flower	1	80	0	0
LOGANIACEAE		_			
Buddleia davidii Franch	Butterfly bush	1	80	0	0
MALVACEAE	Olma	1	90	0	0
Abelmoschus esculentus (L.) Gossypium hirsutum L.	Okra Cotton	1	80 80	0	0 0
Hibiscus spp.	Rose mallow	1	80 80	0	0
MORACEAE	Hose manow	T	80	0	0
Fucus aurea Nutt	Fig	1	80	0	0
MUSACEAE	8	-	00	0	Ū
Musa acuminata Colla.	Banana	1	80	0	0
MYRTACEAE					
Psidium guajaba L.	Tropical guava	1	80	0	0
PASSIFLORACEAE					
Passiflora edulis Sims	Passion fruit	1	80	0	0
POACEAE					
Oryza sativa L.	Rice	1	80	0	0
Saccharum officinarum L.	sugarcane	1	80	0	0
Zea mays L.	Sweet corn	1	80	0	0
PUNICACEAE			0.0	0	0
Punica gramatum L.	Pomegranate	1	80	0	0
ROSACEAE Malus pumilla Mill.	Apple	1	80	0	0
Prunus americana Marsh.	American plum	1	80 80	0	0
Rosa sp.	Miniature rose	1	80	0	0
Rubus betulifolius Small	Blackberry	1	80	0	0
RUTACEAE	Diaoingeriy	-	00	0	Ū
Citrus sinensis (L.) Osbeck	Sweet orange	1	80	0	0
Citrus limon (L.) Burm.	Lemon	1	80	0	0
Citrus paradise Mcfady	Grapefruit	1	80	0	0
SCROPHULARIACEAE	-				
Antirrhinum majus L.	Common snapdragon	1	80	0	0
Nemensia strumosa Benth.	Nemensia	1	80	0	0
Category 7. Any plant on which close found or recorded to feed/ or reproduc SOLANACEAE Section Melongena Subsection Lathyrocarpum		ontrol agen	t (within the	e same gent	ıs) have been
Solanum elaeagnifolium Cav.	Silverleaf nightshade*	4	300	0	0
Subsection Melongena			000	0.7	0.0
Solanum melongena L.	Eggplant	4	300	0-1	0-2
Subsection Cryptocarpum	Chiplers and all the line of the	0	100	0	0
Solanum sisymbriifolium Lam.	Sticky nightshade	2	120	0	0

TABLE 1. (CONTINUED) HOST RANGE ADULT FEEDING AND OVIPOSITION TESTS WITH GRATIANA GRAMINEA IN FLORIDA.

Each test included 3-4 replications with 20 adults (10 males, 10 females) per replication.

 $^{10}$  = no feeding, 1 = probing (<5% of leaf area), 2 = light feeding (5-20%), 3 = moderate feeding (21-40%), 4 = heavy feeding (41-60%), 5 = intense (60% of leaf-area).

50-60 cm height), with a mesh screen at the top and covering 6 circular holes (6 cm diameter) located in pairs at the bottom, middle, and upper part of the cylinder to allow for air circulation. Adults tested originated from F2- F3 generations reared in quarantine from larvae and adults collected on tropical soda apple plants in southern Brazil. The adults had either recently emerged

Plant family Species	Common names	No.of Tests	No. of Insects	$\begin{array}{c} \mathbf{Feeding} \\ \mathbf{Score}^1 \end{array}$	Mortality (%)
CONVOLVULACEAE					
Ipomoea batatas	Sweetpotato	1	30	0	100
SOLANACEAE					
Capsicum annuum	Bell pepper	2	70	0	100
Datura discolor	Angels' trumpet	1	30	0	100
Datura stramonium	Jimson weed	1	30	0	100
Lvcium carolinianum	Christamas berry	1	30	0	100
Lycopersicon esculentum	Tomato	2	70	0	100
Nierembergia scoparia	Cupflower	1	30	0	100
Nicotiana tabacum	Tobacco	2	60	0	100
Physalis angulata	Cutleaf Ground-Cherry	1	40	0	100
Physalis pubescens	Strawberry tomato	1	30	0	100
Solanum americanum	American nightshade	1	30	0	100
Solanum capsicoides	Red soda apple	2	70	$\overset{\circ}{2}$	100
Solanum carolinense	Horsenettle	1	30	0	100
Solanum citrullifolium	Watermelon nightshade	1	30	0	100
Solanum dimidiatum	Western horsnettle	1	30	ů 0	100
Solanum donianum	Mullein nightshade	2	70	0	100
Solanum elaeagnifolium	Silverleaf nightshade	1	30	0	100
Solanum heterodoxum	Melonleaf nightshade	1	30	ů 0	100
Solanum jamaicense	Jamaican nightshade	1	30	0	100
Solanum jasminoides	White potato vine	1	30	0	100
Solanum melongena	Eggplant	2	70	1	100
Solanum nigrescens	Divine nightshade	1	30	0	100
Solanum ptycanthum	Wonder berry	1	30	0	100
Solanum pumillum	Rock-outcrop Solanum	1	30	0	100
Solanum quitoense	Naranjilla	1	30	0	100
Solanum retroflexum	Sunberry	1	30	0	100
Solanum scabrum	Garden huckleberry	1	30	0	100
Solanum tampicense	Wetland nightshade	2	70	0	100
Solanum torvum	Turkeyberry	2	70	1	100
Solanum tuberosum	Potato	2	70	0	100
Solanum viarum	Tropical soda apple	2	70	5	26

## TABLE 2. HOST RANGE OF GRATIANA GRAMINEA FIRST INSTAR IN NO-CHOICE FEEDING TESTS IN FLORIDA QUARANTINE.

Each test included 3-4 replications with 10 neonate larvae per replication.

<sup>1</sup>= no feeding, = probing (<5% of leaf area), 2 = light feeding (5-20%), = moderate feeding (21-40%), 4 = heavy feeding (41-60%), 5 = intense feeding (>60% of leaf area).

from pupae or were still young (1-2 week old) and showing the intense green color that differentiate them from pale-yellow older adults. Eggs laid on plants, if any, were removed weekly and plants replaced as needed. At the end of the testing periods, feeding and adult mortality were recorded.

## RESULTS AND DISCUSSION

## Multiple-Choice Feeding-Oviposition Tests

In the quarantine multiple-choice tests (Table 1), *Gratiana graminea* adults fed heavily to intensively (41-100% of the leaf area offered) on the major target weed tropical soda apple. It fed lightly to moderately (5-40% of the leaf area offered) on turkeyberry, *Solanum torvum* Sw. (na-

tive to west Africa and on the list of Federal noxious weeds and on Florida's invasive species list of the Florida Exotic Pest Plant Council; webpage: http://www.fleppc.org/97list.htm). Minor or exploratory feeding (<5% of the leaf area offered) was observed on the non-native red soda apple, Solanum capsicoides All. (prickly weed of South American origin also present and spreading in Florida), on eggplant, Solanum melongena L. (crop of economic importance), on the non-native Solanum mamosum L. (native from Central America, not growing in USA), on the non-native Solanum tampicense Dunal (weed of Mexico, Central America, and Caribbean origin and now established and expanding in south Florida; also on the Florida's invasive species list of the Florida Exotic Pest Plant Council). No feeding was ob-

S IN FLORIDA QUARANTINE.	

239

Plant family Species	Common names	No. of tests	No. of Insects	$\begin{array}{c} \mathbf{Feeding} \\ \mathbf{Score}^1 \end{array}$	Eggs/ female
CONVOLVULACEAE					
Ipomoea batatas	Sweetpotato	1	40	0	0
SOLANACEAE					
Capsicum annuum	Bell pepper	3	110	0	0
Capsicum frutescens	Chile	1	30	0	0
Lycopersicon esculentum	Tomato	3	110	0	0
Nicotiana tabacum	Tobacco	$^{2}$	70	0	0
Nierembergia scoparia	Cupflower	1	30	0	0
Physalis crassifolia	Ground-cherry	1	30	0	0
Solanum americanum	American nightshade	1	40	0	0
Solanum bahamense	Bahama nightshade	1	30	0	0
Solanum capcicoides	Red soda apple	1	40	0	0
Solanum carolinense	Horse nettle	1	40	0	0
Solanum citrullifolium	Watermelon nightshade	1	30	0	0
Solanum dimidiatum	Western horsenettle	1	40	0	0
Solanum diphillum	Two-leaf nightshade	1	30	0	0
Solanum donianum	Mullein nightshade	2	70	0	0
Solanum elaeagnifolium	Silverleaf nightshade	2	70	1	0
Solanum heterodoxum	Melonleaf nightshade	1	30	0	0
Solanum jamaicense	Jamaican nightshade	1	40	0	0
Solanum jasminoides	White potato vine	1	40	0	0
Solanum mammosum	Nipplefruit	1	30	0-2	0
Solanum melongena	$\operatorname{Eggplant}$				
cv Black Beauty					
cv Classic		4	140	0-1	0
cv Ichiban		2	60	0	0
cv Italian-Nadia		<b>2</b>	60	0	0
cv Market		<b>2</b>	60	0	0
cv Neon		2	60	0-1	0
cv Orient charm		1	60	0	0
cv Orient express		<b>2</b>	60	0	0
cv Thai		<b>2</b>	60	0	0
Solanum nigrescens	Divine nightshade	1	30	0	0
Solanum pumilum	Rock-outcrop	1	30	0	0
Solanum ptycanthum	Wonder berry	1	30	0	0
Solanum retroflexum	Sunberry	1	30	0	0
Solanum scabrum	Garden huckleberry	1	30	0	0
Solanum seaforthianum	Brazilian nightshade	1	30	0	0
Solanum tampicense	Wetland nightshade	2	70	1	0
Solanum torvum	Turkeyberry	2	70	2	0
Solanum tuberosum	Potato	3	80	0	0
Solanum viarum	Tropical soda apple	4	140	5	47-61

Each test included 3-4 replications with 10 adults (5 females, 5 males) per replication.

 $^{10}$  = no feeding, 1 = probing (<5% of leaf area), 2 = light feeding (5-20%), 3 = moderate feeding (21-40%), 4 = heavy feeding (41-60%), 5 = intense feeding (>60% of leaf area).

served on any of an additional 109 plant species in 32 families that were tested. The adults laid from 58-104 eggs (average: 86) on tropical soda apple, and from 0-2 (average: 0.1 eggs) on eggplant (Table 1). No eggs were deposited on any of the other 113 plant species tested, including the threatened *S. donianum*. Although some minor feeding on eggplant has occurred in quarantine, this insect has never been recorded attacking eggplant in South America. Expanded host ranges of weed biocontrol candidates under confined quarantine laboratory conditions have been reported by South African researchers (Neser et al. 1989; Hill & Hulley 1995; Olckers et al. 1995; Hill & Hulley 1996; Olckers 1996). They indicated that almost all the agents that have been tested for biocontrol of *Solanum* weeds have shown feeding on closely related plant species, but they are never attacked under natural conditions. For example, *Gratiana spadicea* (Klug) (Coleoptera: Chrysomelidae) as a biocontrol agent against *Solanum sisymbrifolium* Lam. in South-Africa (Hill & Hulley 1995), fed and was successfully reared on eggplant in laboratory tests. This insect was field released in South Africa in 1994 based mainly on the lack of records as a pest of eggplant in South America. This insect has become established on S. *sisymbriifolium* with no reports of attacks of eggplant fields in South Africa.

### No-Choice Larval Feeding Tests

Larvae of *G. graminea* exposed to individual plants (31 species) in quarantine growth chambers (Table 2) completed development on the target weed tropical soda apple (74% reached the pupae stage, n = 70, 2 trials). Some feeding (5-20%) was observed on *Solanum capsicoides* (red soda apple), and also some probing or exploratory feeding (<5% of the leaf area offered) was observed on *Solanum melongena* (eggplant) and on *Solanum torvum* (turkeyberry), but larvae stopped feeding and died within a week after the experiment began. The rapid death of the *G. graminea* larvae with no feeding and no development on the 30 non-target plant species tested clearly indicated no risk of attack on these plants by this beetle.

### No-Choice Adult Feeding Tests

Starvation tests (no-choice) with G. graminea adults exposed to individual potted plants (29 species) in cages at the quarantine facility (Table 3) indicated that the insect fed and laid eggs (47-61 eggs per female; average: 57 per female) only on tropical soda apple. Feeding on tropical soda apple was intense (>60% of the leaf area offered) compared to a probing or exploratory feeding (<5%) observed in S. melongena (eggplant cultivars Black Beauty and Market), on S. tampicense, and on S. elaeagnifolium (secondary target weeds). Although there was some feeding (5-20% of the leaf area offered) on S. torvum (secondary target weed), and on S. mammosum (non-native), the females did not lay eggs on these plants. No eggs were laid on any of the 28 non-target plant species tested including the 9 eggplant cultivars (Black Beauty, Classic, Ichiban, Italian-Nadia, Market, Neon, Orient Charm, Orient Express, and Thai).

The high specificity shown by this beetle in the host range feeding tests and development only on the target weed, indicated no adverse impacts would be expected on the 6 solanaceous species that were not tested and are listed as threatened or endangered in Hawaii and Puerto Rico. Indirect beneficial effects on wildlife populations associated with release and establishment of *G. graminea* may be expected due to recolonization by native plants that have been displaced by the rapidly growing and highly competitive tropical soda apple plants.

The host specificity tests in quarantine indicated that *G. graminea* is safe to release. Occasional temporary feeding might occur on the nonnative weeds *S. torvum* and *S. tampicense* (in the Federal Noxious Weed list), and *S. capsicoides*, a prickly weed introduced from South America (Kissman & Groth 1995). Noticeable damage to eggplant is unlikely to occur based on our host tests. The lack of a record as a crop pest in the native range of the beetle support our findings on the specificity and safety of *G. graminea* as a biocontrol agent of tropical soda apple.

Based on the specificity of *G. graminea* feeding and developing only on the target weed, we consider this beetle safe for field release against tropical soda apple. Therefore, a petition to release the Brazilian leaf-beetle *G. graminea* for the control of tropical soda apple in the southeastern United States was submitted to the USDA-APHIS-PPQ Technical Advisory Group (TAG) members on Sep 2008.

### ACKNOWLEDGMENTS

We thank Howard Frank (University of Florida, Entomology and Nematology Department), and Julieta Brambila (United States Department of Agriculture, Animal and Plant Health Inspection Service) for reviewing the manuscript. We thank Zundir Buzzi (Universidade Federal do Paraná, Curitiba, Brazil) for identification of *Gratiana graminea*. This research was funded by USDA-APHIS, and by the Florida Department of Agriculture and Consumer Services, Division of Plant Industry.

### References Cited

- AKANDA, R. A., MULLAHEY, J. J., AND SHILLING, D. G. 1996. Growth and reproduction of tropical soda apple (Solanum viarum Dunal) in Florida, pp. 15-22 In Proc. Tropical Soda Apple Symp. Bartow, Florida. University of Florida, IFAS.
- AKANDA, R. A., MULLAHEY, J. J., SHILLING, D. G. 1997. Tropical soda apple (Solanum viarum) and bahiagrass (Paspalum notatum) response to selected PPI, PRE, and POST herbicides, p. 35 In Abstracts of the Weed Science Society of America meeting, Orlando, Florida. WSSA Abstracts Vol. 37.
- BREDOW, E., PEDROSA, J. H., MEDAL, J. C., CUDA, J. P. 2007. Open field host specificity tests in Brazil for risk assessment of *Metriona elatior* (Coleoptera: Chrysomelidae), a potential biological control agent of *Solanum viarum* (Solanaceae) in Florida. Florida Entomol. 90: 559-564.
- BROWN, W. F., MULLAHEY, J. J., AND AKANDA, R. A. 1996. Survivability of tropical soda apple seed in the gastro-intestinal tract of cattle, pp. 35-39 *In* Proc. Tropical Soda Apple Symp. Bartow, Florida. University of Florida, IFAS.

- BRYSON, C. T., BYRD, JR., J. D., AND WESTBROOKS, R. G. 1995. Tropical SodaAapple (Solanum viarum Dunal) in the United States. Mississippi Dept. Agric. and Commerce-Bureau of Plant Industry Circular. 2 pp.
- BRYSON, C. T., AND BYRD, JR., J. D. 1996. Tropical soda apple in Mississippi, pp. 55-60 *In* Proc. Tropical Soda Apple Symp. Bartow, Florida. University of Florida, IFAS.
- COILE, N. C. 1998. Notes on Florida's Endangered and Threatened Plants. Florida Dept. Agric. and Consumer Ser., Bureau of Entomol., Nematol., and Plant Pathol. Botany Section Contribution No. 38, 2<sup>nd</sup> edition. 119 pp.
- COILE, N. C. 1993. Tropical Soda Apple, Solanum viarum Dunal: The Plant from Hell. Botany Circular No. 27. Florida Dept. Agric., and Consumer Serv., Division of Plant Industry.
- DOWLER, C. C. 1996. Some potential management approaches to tropical soda apple in Georgia, pp. 41-54 In Proc. Tropical Soda Apple Symp. Bartow, Florida. University of Florida, IFAS.
- GANDOLFO, D., MCKAY, F., MEDAL, J. C., AND CUDA, J. P. 2007. Open-field host specificity test of *Gratiana boliviana* (Chrysomelidae), a biocontrol agent of tropical soda apple in the USA. Florida Entomol. 90: 223-228.
- HABECK, D. H., MEDAL, J. C., AND CUDA, J. P. 1996. Biological control of tropical soda apple, pp. 73-78 *In* Proc. Tropical Soda Apple Symp. Bartow, Florida. University of Florida, IFAS.
- HILL, M. P., AND HULLEY, P. E. 1995. Biology and host range of *Gratiana spadicea* (Klug, 1829) (Coleoptera: Chrysomelidae: Cassidinae), a potential biological control agent for the weed *Solanum sisymbrifolium* Lamarck (Solanaceae) in South Africa. Biol. Control 5: 345-352.
- HILL, M. P., AND HULLEY, P. E. 1996. Suitability of Metriona elatior (Klug) (Coleoptera: Chrysomelidae: Cassidinae) as a biological control agent for Solanum sisymbrifolium Lam. (Solanaceae). African Entomol. 4: 117-123.
- MCGOVERN, R. J., POLSTON, J. E., DANYLUK, G. M., HEIBERT, E., ABOUZID, A. M., AND STANSLY, P. A. 1994a. Identification of a natural weed host of tomato mottle geminivirus in Florida. Plant Dis. 78: 1102-1106.
- MCGOVERN, R. J., POLSTON, J. E., AND MULLAHEY, J. J. 1994b. Solanum viarum: weed reservoir of plant viruses in Florida. Intl. J. Pest Management 40: 270-273.
- MCGOVERN, R. J., POLSTON, J. E., AND MULLAHEY, J. J. 1996. Tropical soda apple (*Solanum viarum* Dunal): Host of tomato, pepper, and tobacco viruses in Florida, pp. 31-34 *In* Proc. Tropical Soda Apple Symp. Bartow, Florida. University of Florida, IFAS.
- MEDAL, J. C., CHARUDATTAN, R., MULLAHEY, J. J., AND PITELLI, R. A. 1996. An exploratory insect survey of tropical soda apple in Brazil and Paraguay. Florida Entomol. 79: 70-73.
- MEDAL, J. C., AND CUDA, J. P. 2000. Biological control of some exotic weed by means of insects, pp. 75-82 In Proc. Caribbean Basin Administrative Group Workshop on Approaches to Mitigating the Effects of Exotic Pests on Trade and Agriculture in the Caribbean Region, 16-18 June 1999, Homestead, FL. University of Florida, Tropical Research Education Center, Homestead.

- MEDAL, J. C., PITELLI, R. A., SANTANA, A., GANDOLFO, D., GRAVENA, R., AND HABECK, D. H. 1999a. Host specificity of *Metriona elatior* Klug (Coleoptera: Chrysomelidae) a potential biological control agent of tropical soda apple, *Solanum viarum* Dunal (Solanaceae), in the United States. BioControl 44: 421-436.
- MEDAL, J. C., GANDOLFO, D., PITELLI, R. A., SANTANA, A., CUDA, J. P., AND SUDBRINK, D. 1999b. Progress and prospects for biological control of *Solanum vi*arum in the USA, pp. 627-632 *In* Proc. X International Symp. Biol. Control of Weeds, 4-9. July 1999, Bozeman, MT. USDA-ARS/Montana State University, Bozeman.
- MEDAL, J. C., SUDBRINK, D., GANDOLFO, D., OHASHI, S., AND CUDA, J. P. 2002. Gratiana boliviana, a potential biocontrol agent of Solanum viarum: Quarantine host-specificity testing in Florida and field surveys in South America. BioControl 47: 445-461.
- MEDAL, J. C., GANDOLFO, D., AND CUDA, J. P. 2003. Biology of *Gratiana boliviana*, the First Biocontrol Agent Released to Control Tropical Soda Apple in the USA. University of Florida-IFAS Extension Circular ENY-826. 3pp.
- MEDAL, J., OHASHI, D., GANDOLFO, D., MCKAY, F., AND CUDA, J. 2004. Risk assessment of *Gratiana boliviana* (Chrysomelidae), a potential biocontrol agent of tropical soda apple, *Solanum viarum* (Solanaceae) in the USA, pp. 292-296 In J. M. Cullen et al. (eds.). Proc. XI International Symposium Biol. Control of Weeds, April 27-May 2, 2003. Canberra, Australia.
- MEDAL, J., ÖVERHOLT, W., STANSLY, P., RODA, A., OSBORNE, L., HIBBARD, K., GASKALLA, R., BURNS, E., CHONG, J., SELLERS, B., HIGHT, S., CUDA, J., VITORINO, M., BREDOW, E., PEDROSA-MACEDO, J., AND WIKLER, C. 2008. Establishment and initial impacts of *Gratiana boliviana* (Chrysomelidae) on *Solanum viarum* in Florida, pp. 591-596 *In* R. Sforza, M. C. Bon, H. C. Evans, P. E Hatcher, H. Z. Hinz, and B. G. Rector [eds.], Proc. XII Intl. Symp. Biol. Control of Weeds. La Grande Motte, France.
- MISLEVY, P., MULLAHEY, J. J., AND COLVIN, D. L. 1996. Management practices for tropical soda apple control: Update, pp. 61-67 *In* Proc. Tropical Soda Apple Symp. Bartow, Florida. University of Florida, IFAS.
- MISLEVY, P., MULLAHEY, J. J., AND MARTIN, F. G. 1997. Tropical soda apple (*Solanum viarum*) control as influenced by clipping frequency and herbicide rate, *In* Abstracts Weed Sci. Soc. of America Meeting, Orlando, Florida. WSSA Abstracts Vol. 37.
- MULLAHEY, J. J., AND COLVIN, D. L. 1993. Tropical Soda Apple: A New Noxious Weed in Florida. Univ. of Florida, Florida Cooperative Extension Service, Fact Sheet WRS-7.
- MULLAHEY, J. J., NEE, M., WUNDERLIN, R. P., AND DELANEY, K. R. 1993. Tropical soda apple (*Solanum viarum*): a new weed threat in subtropical regions. Weed Technol. 7: 783-786.
- MULLAHEY, J. J., MISLEVY, P., BROWN, W. F., AND KLINE, W. N. 1996. Tropical Soda Apple, an Exotic Weed Threatening Agriculture and Natural Systems. Dow Elanco. Down to Earth Vol. 51. No.1. 8 pp.
- MULLAHEY, J. J., AKANDA, R. A., AND SHERROD, B. 1997. Tropical soda apple (Solanum viarum) update from Florida, In Abstracts Weed Sci. Soc. of America Meeting, Orlando, Florida. WSSA Abstracts Vol. 37.
- NESER, S., ZIMMERMANN, H. G., ERB, H. E., AND HOFF-MANN, J. H. 1989. Progress and prospects for the bi-

ological control of two *Solanum* weeds in South Africa, pp. 371-381 *In* E. S. Delfose [ed.], Proc.VII Intl. Symp. Biol. Control of Weeds, Rome, Italy, Instituto Sperimentale per la Patologia Vegetale Ministerio dell Agriculture e delle Foreste, Rome.

- OLCKERS, T., ZIMMERMANN, H. G., AND HOFFMANN, J. H. 1995. Interpreting ambiguous results of hostspecificity tests in biological control of weeds: assessment of two *Leptinotarsa* species (Chrysomelidae) for the control of *Solanum elaeagnifolium* (Solanaceae) in South Africa. Biol. Control 5: 336-344.
- OLCKERS, T. 1996. Improved prospects for biological control of three *Solanum* weeds in South Africa, pp. 307-312 *In* V. C. Moran and J. H. Hoffmann [eds.], Proc. IX Intl. Symp. Biological Control of Weeds, Stellenbosch, South Africa. University of Cape Town, South Africa.
- OVERHOLT, W., MEDAL, J., HIBBARD, K., AND RODA, A. 2008. Biological Control of Tropical Soda Apple: A Success in the Making. The Florida Cattleman and Livestock Journal. July.
- PATTERSON, D. T. 1996. Effects of temperature and photoperiod on tropical soda apple (*Solanum viarum* Dunal) and its potential range in the United States,

pp. 29-30 *In* Proc. Tropical Soda Apple Symp. Bartow, Florida. University of Florida, IFAS.

- PEREIRA, A., PITELLI, R. A., NEMOTO, L. R., MULLAHEY, J., J., AND CHARUDATTAN, R. 1997. Seed production by tropical soda apple (*Solanum viarum* Dunal) in Brazil, p. 29 *In* Abstracts Weed Sci. Soc. of America Meeting, Orlando, Florida. 1997. WSSA Abstracts Vol. 37.
- STURGIS, A. K., AND COLVIN, D. L. 1996. Controlling tropical soda apple in pastures, p. 79 *In* Proc. Tropical Soda Apple Symp. Bartow, Florida. University of Florida, IFAS.
- SUDBRINK, JR. D. L., SNODGRASS, G. L., BRYSON, C. T., MEDAL, J. C., CUDA, J. P., AND GANDOLFO, D. 1999. Arthropods associated with tropical soda apple, Solanum viarum in the Southeastern USA, p.154 In Program Abstracts, X Intl. Symp. Biol. Control of Weeds, 4-9 July 1999. Bozeman, MT. USDA-ARS/ Montana State University, Bozeman.
- THOMAS, M., 2007. Impact of tropical soda apple on Florida's grazing land. The Florida Cattleman and Livestock Journal 71: 37-38.
- U.S. FISH AND WILDLIFE SERVICE. 1997. Endangered and threatened wildlife and plants. U.S. Government Printing Office, 52 pp.