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Transcriptomic Profile of *Arabidopsis* Rosette Leaves during the Reproductive Stage after Exposure to Ionizing Radiation

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We attempted to obtain a transcriptomic profile of ionizing radiation-responsive genes in *Arabidopsis* plants using Affymetrix ATH1 whole-genome microarrays. The *Arabidopsis* plants were irradiated with 200 Gy γ rays at the early reproduction stage, 33 days after sowing. Rosette leaves were harvested during the postirradiation period from 36 to 49 days after sowing and used for the microarray analysis. The most remarkable changes in the genome-wide expression were observed at 42 days after sowing (9 days after the irradiation). We identified 2165 genes as γ -ray inducible and 1735 genes as γ -ray repressible. These numbers of affected genes were almost two to seven times higher than those at other times. In a comparison of the control and irradiated groups, we also identified 354 differentially expressed genes as significant by applying Welch's *t* test and fold change analysis. The gene ontology analysis showed that radiation up-regulated defense/stress responses but down-regulated rhythm/growth responses. Specific expression patterns of 10 genes for antioxidant enzymes, photosynthesis or chlorophyll synthesis after irradiation were also obtained using real-time quantitative PCR analysis. We discuss physiological and genetic alterations in the antioxidative defense system, photosynthesis and chlorophyll metabolism after irradiation at the reproductive stage.

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INTRODUCTION

The development of techniques for genetic evaluation of biochemical and physiological responses of plant cells to environmental stress factors has allowed enormous advances in our understanding of stress-inducible signaling mechanisms as well as offering promising genetic targets to generate transgenic plants that can tolerate multiple stresses (1–3). However, few advances have been reported for plant research in the field of radiation biology because of both

experimental limitations and political/social restrictions. The lack of large-capacity facilities for irradiation and/or low-dose radiation fields (or greenhouses) for long-term research makes plants unattractive for radiation biology studies. Therefore, it has been unusual for ionizing radiation to be used as a mutagen to broaden the genetic spectrum of major crop plants (4). The mechanism of action by which radiation induces genetic alterations in cells has been elucidated by studies using *in vitro* systems (5–8). A recent report on radiation-induced DNA damages inducing mutations reinforced the concept that the sensitivity of genes to radiation depends on the base composition and/or the spatial localization of the gene on the chromosome (8). The scientific bases of this theory are universal enough to be applicable to both plant and animal systems. Genome-wide transcript profiling was performed using human cell lines exposed to radiation to elucidate changes in intracellular signaling (9). In that study, 19 genes were identified to be differentially expressed in cell cycle regulation, three in apoptosis, and four in nucleotide excision repair. This information increases our understanding of radiation responses at the gene expression level. In spite of some functional similarities of genes in plant and animal systems, however, differences in the genetic backgrounds have limited the use of expression profiles to identify cell signaling in both plants and animals (10). Accordingly, the need for genome-wide analyses of plant transcripts is increasing in the field of radiation biology. Transcriptional and post-transcriptional regulation to determine radiation-mediated signaling in plant cells has rarely been investigated on a genome-wide scale. In contrast, the physiological/ecological responses in irradiated plants have been investigated extensively (11–15).

It has generally been presumed that radiation-induced damages to DNA occur through both direct energy deposition into the sugar-phosphate moiety and interactions with reactive oxygen species (ROS), generally hydroxyl radical (OH) produced by water radiolysis (8, 16, 17). In addition, cellular responses to radiation can occur through direct participation of ROS in cell signaling (18, 19) and/or indirect involvement of low-molecular-weight signaling factors released from the reactions of ROS and neighboring cellular

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TABLE 1
Primer Sequences Used in Quantitative RT-PCR Experiment

| Locus name | Gene name | Forward/reverse primer |
|------------|----------------|---|
| At1g49240 | <i>actin2</i> | 5'GCCCAGAAGTCTGTTCCAG3'/ 5'CTTGGTGCAAGTGCTGTGAT3' |
| At1g58290 | <i>hemA1</i> | 5'AGGAAAGCAATGGAAGCTCA3'/ 5'AAGTCATCAACCGCTCTCGT3' |
| At4g27440 | <i>porB</i> | 5'CACTTTCGCTTCGCTTTACC3'/ 5'CAGCTCCAATAAACCCCTGA3' |
| At2g40100 | <i>lhcb4.3</i> | 5'AAAACCTTGCCAAGAACGTG3'/ 5'TTGCCATGCAATTCTGTAA3' |
| At1g08830 | <i>csd1</i> | 5'ACATTCAACCCCGATGGTA3'/ 5'GAGGTCATCAGGGTCTGCAT3' |
| At2g28190 | <i>csd2</i> | 5'AAGAAGGCTGTTGCAGTGCT3'/ 5'AGGGTTGAAATGTGGTCTCG3' |
| At4g25100 | <i>fsd1</i> | 5'TCACTGGGGAAAACATCACA3'/ 5'CTCCACCACCTGGTTTCATT3' |
| At4g08390 | <i>sapx</i> | 5'GGTCCTGAGGATTGTCCAGA3'/ 5'CTCCAGGTCTCTTTCTCGTG3' |
| At4g35090 | <i>cat2</i> | 5'TTCAAACCCGTGTCTTCTCC3'/ 5'TTCTCAGCATGACGAACCTG3' |
| At1g 19570 | <i>dhar</i> | 5'TCCAATGACGGATCTGAACA3'/ 5'TATGGACATGGGGAAAGCTC3' |
| At4g31870 | <i>gp</i> | 5'TCAAACCTGAGATCAAGCA3'/ 5'TCTCGACAACCTTGCCCTTT3' |

components (20, 21). Therefore, ROS are critical mediators of intracellular signaling of radiation responses, and ROS-scavenging (antioxidant) enzyme activities are also crucial factors in propagation of radiation responses. Accordingly, studies on the physiology of stress using radiation may provide information about its ability to confer tolerance to multiple stresses to plants (11, 13, 15). Such studies may help

improve the survival of plants under deteriorating environmental conditions, e.g., frequent droughts and floods, and increased exposure to UV radiation and may have applications in the detection and evaluation of radioactive materials near nuclear accident sites (15, 22). Profiling of radiation-induced/repressed transcripts can be used to increase the efficiencies of mutation and screening for desirable phenotypes in plants. To date, genome-wide changes in the expressions of genes in *Arabidopsis* have been documented after treatments with auxin and brassinosteroid (1), chitin (23), hypoxia (24), and oxidative stress-causing agents such as methyl viologen, *Alternaria alternata* toxin, 3-aminotriazole and ozone (2). Profiling of candidate genes involved in cell cycle regulation (25, 26) or wax biosynthesis (27) has been also reported using *Arabidopsis* microarrays. *Arabidopsis* is an attractive plant model for radiation biology because of its short generation time, genetic simplicity, and availability of whole genome information. Recently, the responses of hundreds of plant genes to radiation were characterized through a short-term global transcription analysis in *Arabidopsis*, and *cis*-acting elements in the promoter regions of the radiation-responsive genes were also analyzed (28–35). Most altered genes were involved in cell growth, cell division, development, translation, general metabolism, signal transduction, and stress/defense response. However, a long-term genome-scale profiling of transcripts responding to radiation in plants at the reproductive stage has not been reported for any plant species, including *Arabidopsis*.

Massive doses of radiation have been shown to induce physical changes in plants, including enhancement of respiration, increase in ethylene production, induction of en-

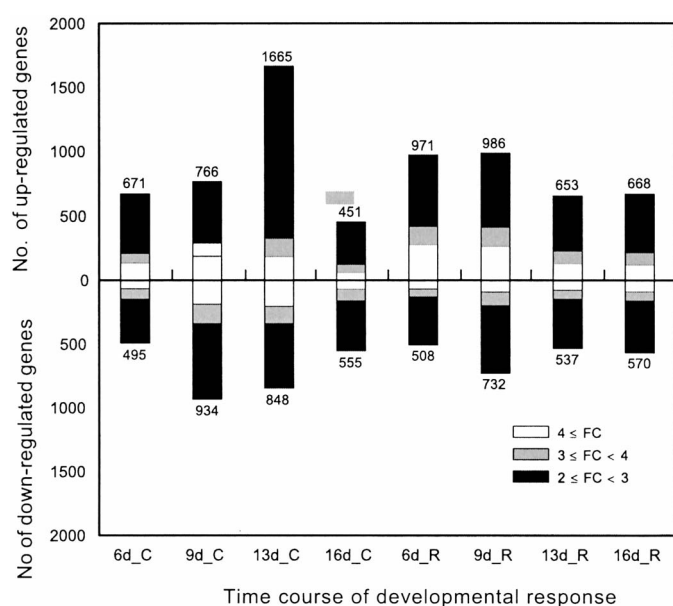


FIG. 1. Transcript level changes in leaves during the reproduction stage. The numbers of genes with a two-, three-, or fourfold increase or decrease in expression in the 6-, 9-, 13- or 16-day plants compared to the 3-day plants after γ irradiation are represented by black, gray and white bars, respectively. Data were analyzed as described in the Materials and Methods. FC, fold change; C, control; R, γ -irradiated.

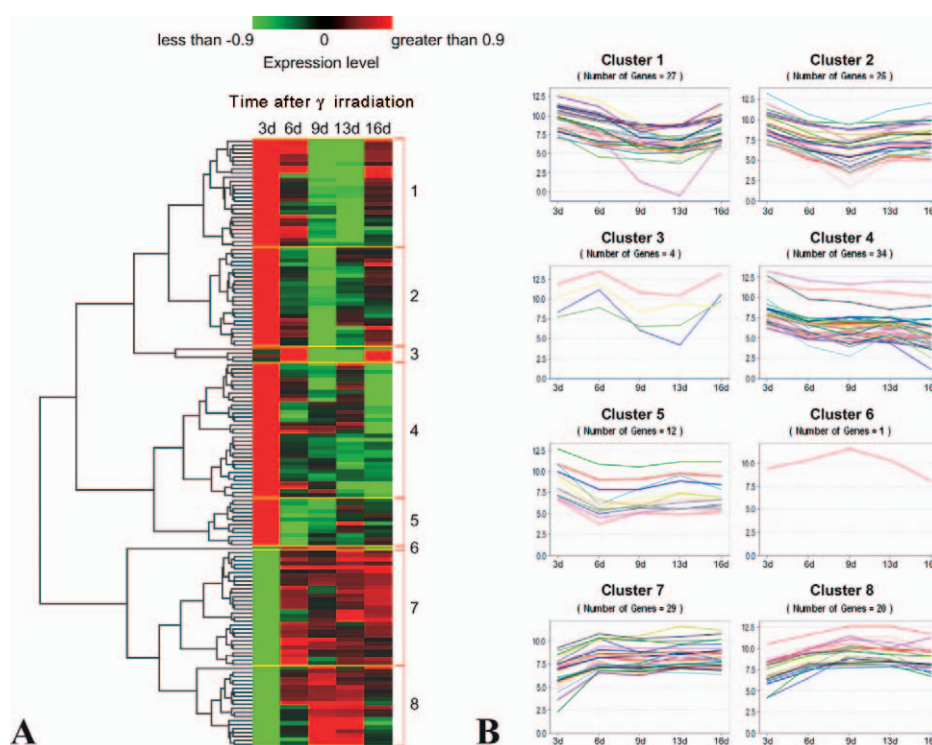


FIG. 2. Clustering analysis of gene expression in control leaves during the reproduction stage. Panel A: Hierarchical cluster analysis. Each horizontal line shows the expression data for one gene after normalization at the times indicated. Colors show the normalized expression level. Induction (or repression) ranges from black to red (or green) with a fold-change scale bar shown above the cluster. Numbers (1 to 8) indicate defined sub-branches or clusters. Panel B: The eight hypothetical profiles are displayed as Clusters 1 to 8 with the number of genes found in each cluster. Each line represents the expression of one gene.

zyme activities, and accumulation of sucrose and specific protein species (12). Photosynthesis and antioxidative defense systems have long been studied in relation to the radiation responses of various plant species (11, 13, 15, 30). These sensitive systems continue to be interesting subjects in plant radiation biology. However, the stress sensitivity of a plant can differ depending on the developmental stage (36, 37), and this is not limited to radiation. We first determined the developmental stage of *Arabidopsis* showing the most dramatic alterations in the phenotypic traits after exposure to γ rays at a dose of 200 Gy, which induced noticeable phenotypic changes in *Arabidopsis* seedlings in a preliminary experiment. Interestingly, the early reproduction stage within 4–5 weeks after seeds were sown was the most sensitive to radiation, with the irradiated plants showing inhibition of stem growth, retardation of leaf senescence, accumulation of chlorophyll and carotenoid, and elevation of photosynthetic saturation for light use. To gain more insight into how plants respond to radiation at the genome level, we carried out gene expression profiling using *Arabidopsis* leaves sampled at five times: 3, 6, 9, 13 and 16 days after the plants were irradiated with 200 Gy γ rays at the early reproduction stage. Affymetrix ATH1 oligonucleotide GeneChip arrays were used to cover the whole genome representing nearly all the nuclear, plastid and mitochondrial genes encoded in *Arabidopsis*. The goal was to generate an informative transcriptomic profile of

plant genes responding to radiation at the reproductive stage through a long-term genome-scale analysis of *Arabidopsis* genes. The results allowed for the genome-wide identification of genes with significant positive or negative changes in expression (induced and repressed, respectively) in response to radiation in plants at the reproductive stage. This paper presents characteristics of radiation responses during the reproduction stage, identifying the radiation-responsive genes related to photosynthesis and antioxidant enzymes.

MATERIALS AND METHODS

Plant Materials and γ Irradiation

Arabidopsis thaliana ecotype Columbia (Col-0) was used in this study. Seedlings were grown for 33 days after sowing at 22/18°C (day/night) with a 16-h photoperiod in a compound soil mixture (vermiculite:peat moss:perlite = 1:1:1). They were exposed to 200 Gy γ rays (dose rate 50 Gy/h) generated by a ^{60}Co γ irradiator (150 TBq of capacity; AECL, Canada) at the Korea Atomic Energy Research Institute. The irradiated seedlings were placed under the same growth conditions as described above. Then their rosette leaves were detached and immediately frozen in liquid nitrogen at 3, 6, 9, 13 and 16 days after irradiation, which corresponded to 36, 39, 42, 46 and 49 days after sowing, respectively. The leaves were stored at -80°C until RNA isolation.

RNA Preparation and Microarray Analysis

To minimize interplant variability, rosette leaves from a minimum of 10 plants were pooled for each RNA extraction. Total RNA was extracted

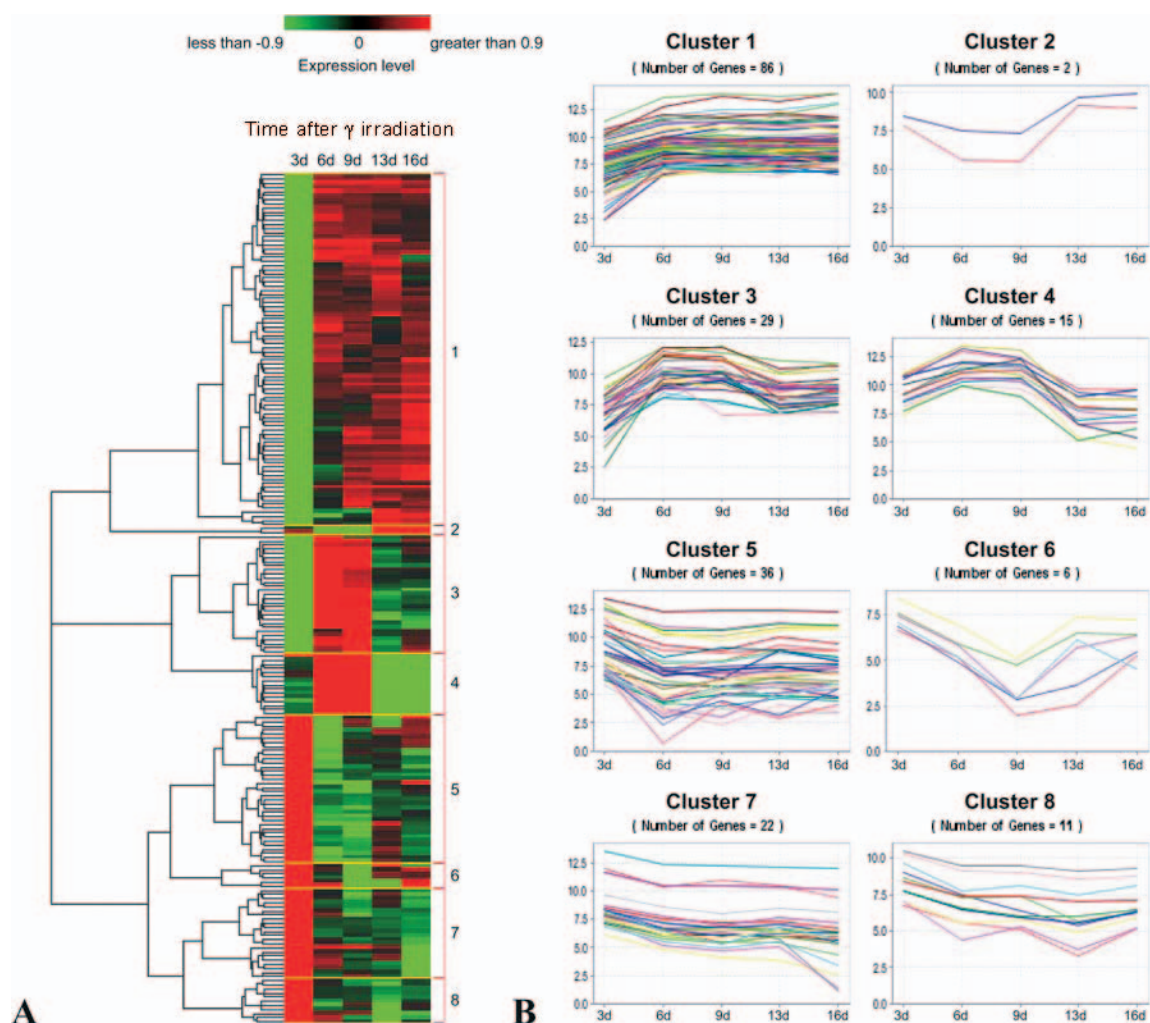


FIG. 3. Clustering analysis of gene expression in the irradiated leaves during the reproduction stage. Panel A: Hierarchical cluster analysis. Each horizontal line shows the expression data for one gene after normalization at the times indicated. Colors show the normalized expression level. Induction (or repression) ranges from black to red (or green) with a fold-change scale bar shown above the cluster. Numbers (1 to 8) indicate defined sub-branches or clusters. Panel B: The eight hypothetical profiles are displayed as Clusters 1 to 8 with the number of genes found in each cluster. Each line represents the expression of one gene.

from the leaves using Trizol (Invitrogen, Carlsbad, CA) and further purified using an RNeasy Mini Kit (Qiagen, Valencia, CA) following the manufacturer's specifications. The amount of RNA was determined by spectrophotometry at 260 nm, and its integrity was assessed by analyzing the ribosomal RNA bands after gel electrophoresis. Conversion of total RNA into double-stranded cDNA, generation of biotin-labeled cRNA from the double-stranded cDNA, and hybridization of the biotin-labeled cRNA with the GeneChip® Arabidopsis ATH1 Genome Array (Affymetrix, Santa Clara, CA) were performed and analyzed as recommended by Affymetrix.

Data Analysis

GenPlex® v1.8 software (ISTECH Inc., Goyang city, South Korea) was used for data analysis. The MAS5 algorithm was used for expression summary and signal calculation. Global scaling normalization using a GCOS algorithm was performed, and then the normalized data were log₂-transformed. Fold change and Welch's *t* test were applied to select the differentially expressed genes. The difference was more than twofold, and the significance level was 0.05. To better visualize and compare the two methods for differentially expressed genes, a volcano plot was used. The twofold differentially expressed genes were clustered using hierarchical

clustering with Pearson correlation as a similarity measure and complete linkage as a linkage method (38–40). The top 200 differentially expressed genes with the greatest fold change were classified into functional sub-groups by BINGO, which enables Gene Ontology significance analysis (41). Pathway analysis was performed using the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway database, which is available online (<http://www.genome.jp/kegg/pathway.html>).

Real-Time Quantitative PCR Analysis

One microgram of total RNA was reverse-transcribed into cDNA using AccuPower RT Premix (Bioneer, Daejeon, Korea) for 60 min at 42°C using 0.5 µg anchored oligo(dT)₁₈ V primers. Real-time PCR was carried out in a 25-µl reaction mixture containing 1 µl template cDNA, 12.5 µl SYBR®Premix Ex Taq™ (2×) (Takara Bio Inc., Otsu, Japan), 2 µl forward primer (10 µM), 2 µl reverse primer (10 µM), and 7.5 µl distilled water, using a Smart Cycler® II System (Cepheid, Sunnyvale, CA). Gene-specific primer sets used are listed in Table 1. The thermal profile consisted of 1 cycle at 95°C for 1 min followed by 45 cycles at 95°C for 5 s, 56°C for 20 s, and 72°C for 15 s. For each run, data acquisition and analysis were done using the Smart Cycler® software (version 2.0b, Cepheid). Then the relative transcript level was determined using the com-

TABLE 2
Gene Ontology Analysis of Differentially Expressed Genes in Control (A) and Irradiated (B) Samples during the Reproduction Stage

| Days after irradiation | GOC | GO_ID | GO term | C | P value | k |
|------------------------|-----|------------|--|--------------------|---------|----------------|
| A: Control | | | | | | |
| Up-regulation | | | | | | |
| 6 | BP | GO:0009628 | response to abiotic stimulus | 224/16475 (1.4) | 0.0168 | 7/150 (4.7) |
| | CC | GO:0005623 | cell | 11389/16475 (69.1) | 0.1123 | 112/150 (74.6) |
| | MF | GO:0016462 | pyrophosphatase activity | 235/16475 (1.4) | 0.0058 | 8/150 (5.3) |
| 9 | BP | GO:0009605 | response to external stimulus | 366/16475 (2.2) | 0.0002 | 13/158 (8.2) |
| | CC | GO:0005576 | extracellular region | 61/16475 (0.4) | 0.1161 | 3/158 (1.9) |
| | MF | GO:0008194 | UDP-glycosyltransferase activity | 109/16475 (0.7) | 0.0006 | 7/158 (4.4) |
| 13 | BP | GO:0006950 | response to stress | 376/16475 (2.3) | 0.0006 | 12/147 (8.2) |
| | CC | GO:0012505 | endomembrane system | 3426/16475 (20.8) | 0.0372 | 41/147 (27.9) |
| | MF | GO:0005386 | carrier activity | 336/16475 (2.0) | 0.0032 | 10/147 (6.8) |
| 16 | BP | GO:0006519 | amino acid and derivative metabolism | 254/16475 (1.5) | 0.0014 | 9/139 (6.4) |
| | CC | GO:0043231 | intracellular membrane-bound organelle | 6455/16475 (39.2) | 0.1893 | 61/139 (43.9) |
| | MF | GO:0004066 | asparagine synthase activity | 2/16475 (0.0) | 0.0168 | 2/139 (1.4) |
| Down-regulation | | | | | | |
| 6 | BP | GO:0009719 | response to endogenous stimulus | 369/16475 (2.2) | 0.0001 | 13/147 (8.8) |
| | CC | GO:0016020 | membrane | 4667/16475 (28.3) | 0.0039 | 58/147 (39.5) |
| | MF | GO:0003700 | transcription factor activity | 1342/16475 (8.1) | 0.0018 | 24/147 (16.3) |
| 9 | BP | GO:0042545 | cell wall modification | 40/16475 (0.2) | 0.0000 | 8/156 (5.1) |
| | CC | GO:0016020 | membrane | 4667/16475 (28.3) | 0.0000 | 81/156 (51.9) |
| | MF | GO:0016799 | hydrolase activity | 13/16475 (0.1) | 0.0002 | 4/156 (2.6) |
| 13 | BP | GO:0009828 | cell wall loosening | 32/16475 (0.2) | 0.0000 | 7/164 (4.3) |
| | CC | GO:0016020 | membrane | 4667/16475 (28.3) | 0.0000 | 94/164 (57.3) |
| | MF | GO:0016798 | hydrolase activity | 150/16475 (0.9) | 0.0001 | 9/164 (5.5) |
| 16 | BP | GO:0007275 | development | 382/16475 (2.3) | 0.0006 | 12/147 (8.2) |
| | CC | GO:0016020 | membrane | 4667/16475 (28.3) | 0.0000 | 66/147 (44.9) |
| | MF | GO:0015290 | transporter activity | 195/16475 (1.2) | 0.0018 | 8/147 (5.4) |
| B: Irradiated | | | | | | |
| Up-regulation | | | | | | |
| 6 | BP | GO:0006952 | defense response | 315/16475 (1.9) | 0.0000 | 13/155 (8.4) |
| | CC | GO:0031225 | anchored to membrane | 219/16475 (1.3) | 0.0011 | 9/155 (5.8) |
| | MF | GO:0030528 | transcription regulator activity | 1450/16475 (8.8) | 0.0022 | 26/155 (16.8) |
| 9 | BP | GO:0006952 | defense response | 315/16475 (1.9) | 0.0000 | 15/159 (9.4) |
| | CC | GO:0012505 | endomembrane system | 3426/16475 (20.8) | 0.0018 | 50/159 (31.4) |
| | MF | GO:0030528 | transcription regulator activity | 1450/16475 (8.8) | 0.0015 | 27/159 (17.0) |
| 13 | BP | GO:0006952 | defense response | 315/16475 (1.9) | 0.0000 | 18/143 (12.6) |
| | CC | GO:0012505 | endomembrane system | 3426/16475 (20.8) | 0.0002 | 49/143 (34.3) |
| | MF | GO:0016740 | transferase activity | 1908/16475 (11.6) | 0.0036 | 29/143 (20.3) |
| 16 | BP | GO:0009607 | response to biotic stimulus | 399/16475 (2.4) | 0.0000 | 20/154 (13.0) |
| | CC | GO:0016602 | CCAAT-binding factor complex | 11/16475 (0.1) | 0.0001 | 4/154 (2.6) |
| | MF | GO:0009055 | electron carrier activity | 49/16475 (0.3) | 0.0011 | 5/154 (3.2) |
| Down-regulation | | | | | | |
| 6 | BP | GO:0048511 | rhythmic process | 17/16475 (0.1) | 0.0004 | 4/142 (2.8) |
| | CC | GO:0005634 | nucleus | 697/16475 (4.2) | 0.3949 | 8/142 (5.6) |
| | MF | GO:0030528 | transcription regulator activity | 1450/16475 (8.8) | 0.0001 | 28/142 (19.7) |
| 9 | BP | GO:0007623 | circadian rhythm | 17/16475 (0.1) | 0.0003 | 4/131 (3.1) |
| | CC | GO:0031225 | anchored to membrane | 219/16475 (1.3) | 0.5219 | 3/131 (2.3) |
| | MF | GO:0030528 | transcription regulator activity | 1450/16475 (8.8) | 0.0051 | 22/131 (16.8) |
| 13 | BP | GO:0009733 | response to auxin stimulus | 117/16475 (0.7) | 0.0005 | 7/141 (5.0) |
| | CC | GO:0016020 | membrane | 4667/16475 (28.3) | 0.0001 | 62/141 (44.0) |
| | MF | GO:0005372 | water transporter activity | 28/16475 (0.2) | 0.0017 | 4/141 (2.8) |
| 16 | BP | GO:0009733 | response to auxin stimulus | 117/16475 (0.7) | 0.0000 | 11/148 (7.4) |
| | CC | GO:0016020 | membrane | 4667/16475 (28.3) | 0.0005 | 62/148 (41.9) |
| | MF | GO:0016837 | carbon-oxygen lyase activity | 26/16475 (0.2) | 0.0226 | 3/148 (2.0) |

Notes. Differentially expressed genes were selected to have more than a twofold change in the respective samples relative to the 3-day control. GOC, BP, CC and MF represent the GO category, biological process, cellular component, and molecular function. GO terms in the table were selected for their highest significance based on *P* values in each GO category. C, the total frequency of a given GO term in all annotations in the *Arabidopsis* genome; k, cluster frequency of a given GO term in the differentially expressed genes selected. Numbers in parentheses are percentages.

TABLE 3
Pathway Analysis of the Differentially Expressed Genes during the Reproduction Stage

| KEGG pathway | Gene counts |
|---|-------------|
| Control | |
| Stilbene, coumarine and lignin biosynthesis | 9 |
| gamma-Hexachlorocyclohexane degradation | 8 |
| Ribosome | 8 |
| Photosynthesis | 8 |
| Limonene and pinene degradation | 8 |
| Fluorene degradation | 8 |
| Ascorbate and aldarate metabolism | 8 |
| Oxidative phosphorylation | 5 |
| Phenylalanine metabolism | 4 |
| Tyrosine metabolism | 3 |
| Irradiated | |
| Stilbene, coumarine and lignin biosynthesis | 11 |
| Ribosome | 11 |
| gamma-Hexachlorocyclohexane degradation | 9 |
| Limonene and pinene degradation | 9 |
| Fluorene degradation | 9 |
| Ascorbate and aldarate metabolism | 9 |
| Oxidative phosphorylation | 5 |
| Purine metabolism | 4 |
| Photosynthesis | 3 |
| Glutathione metabolism | 3 |

Notes. List includes the top 10 pathways based on gene counts. Differentially expressed genes for the control and irradiated samples relative to the 3-day controls were 3347 and 3220 genes in combination, respectively. The pathway analysis was performed using 57 and 53 genes with pathway information among these, respectively.

parative ($2^{-\Delta\Delta Ct}$) method (42, 43). Relative quantification of gene expression using the $2^{-\Delta\Delta Ct}$ method was correlated with the absolute gene quantification obtained using calibration curves (44). *Actin2* was used as an endogenous control gene to normalize for differences in the amounts of total RNA.

RESULTS AND DISCUSSION

Despite many recent analyses of radiation-regulated gene expression in various animal cells by cDNA or oligonucleotide microarrays (9, 45–47), there are no genome-wide profiles for the long-term expression of genes in plants exposed to radiation. We attempted to analyze radiation-regulated cellular responses at the transcriptional level in *Arabidopsis* plants, specifically leaves, during the reproduction stage.

Gene Expression of the Control and Irradiated Groups during the Reproduction Stage

We first aimed to determine which genes are expressed and/or regulated during the reproduction stage in the control and irradiated plants. Leaves were harvested at 3, 6, 9, 13 and 16 days after irradiation, which corresponded to 36, 39, 42, 46 and 49 days after sowing, respectively. Transcript levels were calculated relative to the respective 3-day samples and differentially expressed genes were selected with a twofold change and a significance level of 0.05 after log₂

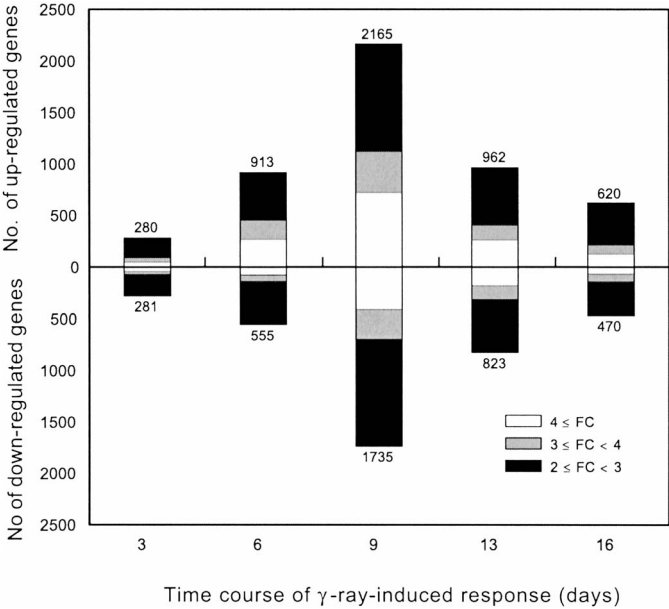


FIG. 4. Transcript level changes in leaves during the postirradiation period. The numbers of genes with a two-, three- or fourfold increase or decrease in expression in the γ -irradiated plants compared to the control plants at different times after γ irradiation are represented by black, gray and white bars, respectively. Data were processed as described in the Materials and Methods. FC, fold change.

transformation of the normalized data. The 3-day samples were used as a control instead of the 0-day samples to exclude the effects of short-term fluctuations in the expression of genes after γ irradiation. Gene expression levels in leaves decrease in terms of the amounts of transcripts during the reproduction stage (36). In the present study, however, the total number of genes affected increased in the control leaves up to 13 days after irradiation and then decreased at 16 days after the irradiation, while the irradiated leaves showed the maximum expression change at 9 days after the irradiation (Fig. 1). The proportion of genes showing an increased transcript level was generally higher in the irradiated leaves than in the control leaves except for the 13-day samples.

The numbers of differentially expressed genes in the control and irradiated groups were 3347 and 3220 in combination during the postirradiation period, respectively, while only 152 and 207 differentially expressed genes existed in common throughout that period. These common differentially expressed genes were classified into two or three major groups using hierarchical clustering with Pearson correlation as a similarity measure and complete linkage as a linkage method (Figs. 2A and 3A). This result indicates that 6 days after the irradiation or 39 days after sowing can be a turning point in gene expression during the reproduction stage. Finally, the data set clusters suggest that although the majority of genes are either continuously up-regulated or down-regulated, there are also groups showing alternative patterns of expression (26). We identified two groups (Clus-

TABLE 4
Radiation Responsive Genes during the Postirradiation Period

| Days after irradiation | Average log ₂ (FC) | Transcript ID | Gene name |
|------------------------|-------------------------------|---------------|--|
| (A) | | | |
| Induced genes | | | |
| 3 | 2.4194489 | At5g40150 | peroxidase, putative |
| 6 | 2.4055262 | At4g33420 | peroxidase, putative |
| | 2.0559115 | At5g48450 | multi-copper oxidase type I family protein |
| | 1.765161 | At2g29170 | short-chain dehydrogenase/reductase (SDR) family protein |
| | 1.7520266 | At3g28200 | peroxidase, putative |
| | 1.7306409 | At3g50930 | AAA-type ATPase family protein |
| | 1.6923962 | At2g37540 | short-chain dehydrogenase/reductase (SDR) family protein |
| | 1.3828316 | At1g23020 | ferric-chelate reductase, putative |
| | 1.3707452 | At5g64120 | peroxidase, putative |
| 9 | 3.43536 | At2g28190 | superoxide dismutase (Cu-Zn), chloroplast (SODCP) (CSD2) |
| | 2.7038298 | At1g12520 | superoxide dismutase copper chaperone, putative |
| | 2.4422855 | At3g28200 | peroxidase, putative |
| | 2.279912 | At5g40150 | peroxidase, putative |
| | 2.144084 | At4g09010 | L-ascorbate peroxidase, chloroplast, putative |
| | 2.0108862 | At5g64120 | peroxidase, putative |
| | 1.9585624 | At5g51100 | superoxide dismutase (Fe), putative |
| | 1.8616467 | At1g71695 | peroxidase 12 (PER12) (PI2) (PRXR6) |
| | 1.6058083 | At1g08830 | superoxide dismutase (Cu-Zn) (SODCC) (CSD1) |
| | 1.5956535 | At5g23310 | superoxide dismutase (Fe) (FSD3) |
| | 1.5212107 | At1g77490 | L-ascorbate peroxidase, thylakoid-bound (tAPX) |
| | 1.4836998 | At5g51890 | peroxidase-related |
| | 1.383935 | At4g33420 | peroxidase, putative |
| 13 | 3.369529 | At5g51890 | peroxidase-related |
| | 2.350175 | At1g71695 | peroxidase 12 (PER12) (PI2) (PRXR6) |
| | 1.8598824 | At2g28190 | superoxide dismutase (Cu-Zn), chloroplast (SODCP) (CSD2) |
| | 1.7416306 | At4g03060 | 2-oxoglutarate-dependent dioxygenase, putative (AOP2) |
| | 1.6883192 | At1g19570 | dehydroascorbate reductase, putative |
| | 1.5280867 | At4g08390 | L-ascorbate peroxidase, stromal (sAPX) |
| | 1.3948259 | At4g09010 | L-ascorbate peroxidase, chloroplast, putative |
| | 1.3385196 | At2g22420 | peroxidase 17 (PER17) (PI7) |
| | 1.2742739 | At4g25100 | superoxide dismutase (Fe), chloroplast (SODB) (FSD1) |
| | 1.1855841 | At4g10500 | oxidoreductase, 2OG-Fe(II) oxygenase family protein |
| | 1.0841036 | At1g08830 | superoxide dismutase (Cu-Zn) (SODCC) (CSD1) |
| 16 | 3.0575595 | At4g08390 | L-ascorbate peroxidase, stromal (sAPX) |
| | 2.2580771 | At1g28030 | oxidoreductase, 2OG-Fe(II) oxygenase family protein |
| | 2.2224302 | At4g25100 | superoxide dismutase (Fe), chloroplast (SODB) (FSD1) |
| | 1.3870687 | At4g36430 | peroxidase, putative |
| | 1.3563209 | At2g29350 | tropinone reductase, putative |
| | 1.3140545 | At1g19570 | dehydroascorbate reductase, putative |
| Repressed genes | | | |
| 3 | -1.7209291 | At4g39830 | L-ascorbate oxidase, putative |
| | -1.3683543 | At1g29160 | Dof-type zinc finger domain-containing protein |
| | -1.1702714 | At5g19890 | peroxidase, putative |
| | -1.101944 | At1g17020 | oxidoreductase, 2OG-Fe(II) oxygenase family protein |
| 6 | -2.2576113 | At1g17020 | oxidoreductase, 2OG-Fe(II) oxygenase family protein |
| | -1.4576573 | At3g42570 | peroxidase, putative |
| | -1.3152676 | At4g08390 | L-ascorbate peroxidase, stromal (sAPX) |
| | -1.2995005 | At4g03060 | 2-oxoglutarate-dependent dioxygenase, putative (AOP2) |
| | -1.1865816 | At4g25100 | superoxide dismutase (Fe), chloroplast (SODB) (FSD1) |
| | -1.1730733 | At4g25300 | oxidoreductase, 2OG-Fe(II) oxygenase family protein |
| | -1.1638432 | At4g32320 | peroxidase family protein |
| | -1.039835 | At1g06620 | 2-oxoglutarate-dependent dioxygenase, putative |
| | -1.0072231 | At1g07890 | L-ascorbate peroxidase 1, cytosolic (APX1) |
| 9 | -2.6704993 | At4g10500 | oxidoreductase, 2OG-Fe(II) oxygenase family protein |
| | -2.6589546 | At3g42570 | peroxidase-related |
| | -2.4284115 | At4g25100 | superoxide dismutase (Fe), chloroplast (SODB) (FSD1) |
| | -2.3921642 | At1g17020 | oxidoreductase, 2OG-Fe(II) oxygenase family protein |

(continued on page 274)

TABLE 4
(continued)

| Days after irradiation | Average log ₂ (FC) | Transcript ID | Gene name |
|------------------------|-------------------------------|---------------|--|
| 13 | -2.1933298 | At2g29370 | tropinone reductase, putative |
| | -1.8451958 | At2g29320 | tropinone reductase, putative |
| | -1.651741 | At2g29990 | pyridine nucleotide-disulphide oxidoreductase family protein |
| | -1.4436169 | At4g31870 | glutathione peroxidase, putative |
| | -1.4195185 | At4g32320 | peroxidase family protein |
| | -1.3083835 | At4g08390 | L-ascorbate peroxidase, stromal (sAPX) |
| | -1.2154627 | At4g35090 | catalase 2 |
| | -1.0904951 | At3g49120 | peroxidase 33 (PER33) (P33) (PRXCA) |
| | -2.2513075 | At4g31870 | glutathione peroxidase, putative |
| | -2.238213 | At2g29370 | tropinone reductase, putative |
| | -1.6643324 | At1g17020 | oxidoreductase, 2OG-Fe(II) oxygenase family protein |
| | -1.5291271 | At1g23020 | ferric-chelate reductase, putative |
| | -1.5120478 | At4g32320 | peroxidase family protein |
| 16 | -1.303483 | At4g35090 | catalase 2 |
| | -1.2666025 | At2g37130 | peroxidase 21 (PER21) (P21) (PRXR5) |
| | -1.2456388 | At2g31570 | glutathione peroxidase, putative |
| | -1.0318995 | At5g19890 | peroxidase, putative |
| | -1.0165234 | At1g29160 | Dof-type zinc finger domain-containing protein |
| | -1.0164618 | At4g33870 | peroxidase, putative |
| | -2.074337 | At1g23020 | ferric-chelate reductase, putative |
| | -1.218318 | At4g35090 | catalase 2 |
| | -1.2676253 | At4g33420 | peroxidase, putative |
| | -1.1945162 | At2g29300 | tropinone reductase, putative |
| (B) | | | |
| Induced genes | | | |
| 3 | 3.1086726 | At5g54190 | NADPH-protochlorophyllide oxidoreductase A (PORA) |
| 6 | 2.074326 | At5g54190 | NADPH-protochlorophyllide oxidoreductase A (PORA) |
| 9 | 1.8820562 | At3g27690 | chlorophyll A-B binding protein (LHCB2:4) |
| | 1.4740295 | At4g27440 | NADPH-protochlorophyllide oxidoreductase B (PORB) |
| | 2.663498 | At5g54190 | NADPH-protochlorophyllide oxidoreductase A (PORA) |
| | 2.5387745 | At3g27690 | chlorophyll A-B binding protein (LHCB2:4) |
| | 2.2573128 | At1g19150 | LHCI type II, putative |
| | 2.0492506 | At4g27440 | NADPH-protochlorophyllide oxidoreductase B (PORB) |
| | 2.0414686 | At5g45930 | Mg-protoporphyrin IX chelatase, putative |
| | 1.3696604 | At1g03630 | NADPH-protochlorophyllide oxidoreductase C (PORC) |
| | 1.3261137 | At1g19670 | coronatine-induced protein 1 (CORI1) |
| | 1.219584 | At1g64590 | short-chain dehydrogenase/reductase (SDR) family protein |
| | 4.059266 | At3g27690 | chlorophyll A-B binding protein (LHCB2:4) |
| | 2.08077 | At1g58290 | glutamyl-tRNA reductase 1 (HEMA1) |
| | 1.6156063 | At4g27440 | NADPH-protochlorophyllide oxidoreductase B (PORB) |
| 13 | 1.4812336 | At1g19670 | coronatine-induced protein 1 (CORI1) |
| | 1.1018019 | At1g19150 | LHCI type II, putative |
| | 1.0646944 | At1g15820 | chlorophyll A-B binding protein (LHCB6) |
| | 2.4675798 | At3g27690 | chlorophyll A-B binding protein (LHCB2:4) |
| | 1.7152658 | At1g58290 | glutamyl-tRNA reductase 1 (HEMA1) |
| | 1.5346851 | At4g27440 | NADPH-protochlorophyllide oxidoreductase B (PORB) |
| Repressed genes | | | |
| 6 | -1.3056841 | At2g40100 | chlorophyll A-B binding protein (LHCB4.3) |
| 9 | -1.8259163 | At2g40100 | chlorophyll A-B binding protein (LHCB4.3) |
| 13 | -1.11796 | At1g76570 | chlorophyll A-B binding family protein |
| | -1.0365758 | At3g14440 | 9-cis-epoxycarotenoid dioxygenase, putative |
| | -1.5352154 | At1g76570 | chlorophyll A-B binding family protein |
| | -1.5352154 | At2g40100 | chlorophyll A-B binding protein (LHCB4.3) |
| | -2.1161408 | At5g53090 | oxidoreductase, putative |
| | -1.4119272 | At1g76570 | chlorophyll A-B binding family protein |

Notes. (A) and (B) represent the genes for antioxidant enzymes and photosynthesis, which were selected by the authors by the gene name. Transcript levels were calculated as an average log₂(FC) of the irradiated samples relative to their respective controls. FC, fold change.

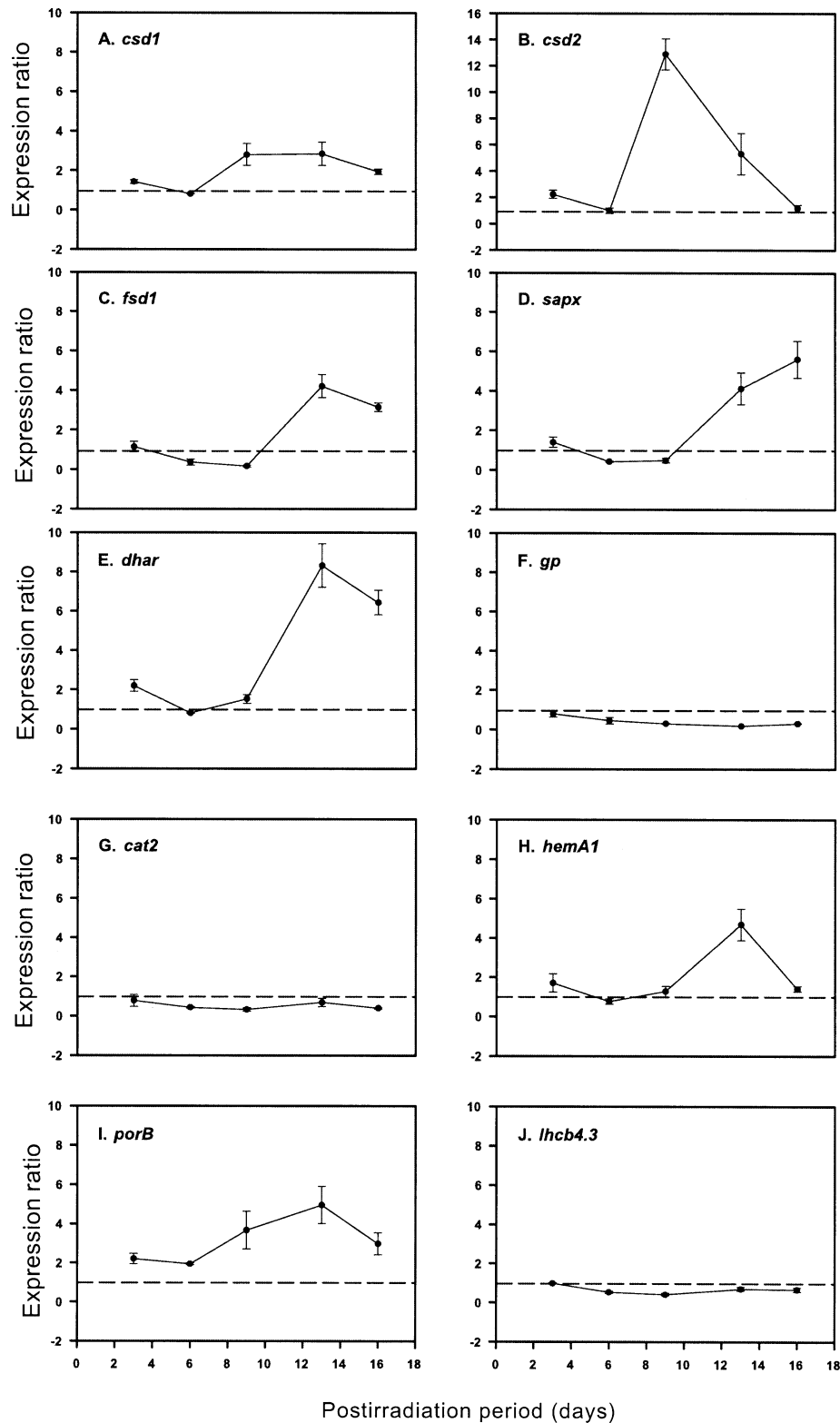


FIG. 5. Real-time quantitative PCR analysis of 10 genes selected from two radiation-responsive categories for validation of the microarray results. Panel A: Cytosolic Cu/Zn-superoxide dismutase (SOD) (*csd1*, At1g08830); panel B: chloroplastic Cu/Zn-SOD (*csd2*, At2g40100); panel C: chloroplastic Fe-SOD (*fsd1*, At4g25100); panel D: stromal ascorbate peroxidase (*sapx*, At4g08390); E, dehydroascorbate reductase (*dhar*, At1g19570); F glutathione peroxidase (*gp*, At4g31870); G, catalase 2 (*cat2*, At4g35090); H, glutamyl-tRNA reductase 1 (*hemA1*, At1g58290); I, NADPH-protochlorophyllide oxidoreductase B (*porB*, At4g27440); and J, chlorophyll A-B binding protein 4.3 (*lhcb4.3*, At2g40100). Expression ratios were calculated by the comparative ($2^{-\Delta\Delta Ct}$) method. *Actin2* was used as an endogenous control gene to normalize for differences in the amounts of total RNA. Error bars represent means \pm SD ($n = 3$).

TABLE 5
Pathway Analysis of the Differentially Expressed Genes during the Postirradiation Period

| Differentially expressed genes | No. of genes selected | No. of genes with pathway | KEGG pathway | Gene counts |
|--------------------------------|-----------------------|---------------------------|---|-------------|
| 3 | 561 | 6 | Pyrimidine metabolism | 2 |
| | | | Cysteine metabolism | 1 |
| | | | Flavonoid biosynthesis | 1 |
| | | | Glutathione metabolism | 1 |
| | | | Methionine metabolism | 1 |
| 6 | 1468 | 22 | Stilbene, coumarine and lignin biosynthesis | 6 |
| | | | Ascorbate and aldarate metabolism | 4 |
| | | | Fluorene degradation | 4 |
| | | | Limonene and pinene degradation | 4 |
| | | | Pyrimidine metabolism | 4 |
| 9 | 3900 | 72 | Ribosome | 24 |
| | | | Stilbene, coumarine and lignin biosynthesis | 12 |
| | | | Ascorbate and aldarate metabolism | 10 |
| | | | Fluorene degradation | 10 |
| | | | Limonene and pinene degradation | 10 |
| 13 | 1785 | 37 | Stilbene, coumarine and lignin biosynthesis | 7 |
| | | | gamma-Hexachlorocyclohexane degradation | 6 |
| | | | Limonene and pinene degradation | 6 |
| | | | Fluorene degradation | 6 |
| | | | Ascorbate and aldarate metabolism | 6 |
| 16 | 1090 | 17 | Glycine, serine and threonine metabolism | 4 |
| | | | Pyrimidine metabolism | 3 |
| | | | gamma-Hexachlorocyclohexane degradation | 2 |
| | | | Vitamin B6 metabolism | 2 |
| | | | Stilbene, coumarine and lignin biosynthesis | 2 |

Notes. List includes the top five pathways based on gene counts. Differentially expressed genes of the irradiated samples relative to their respective controls were selected, and then genes with pathway information were used for the analysis.

ters 3 and 6) in the control leaves and three (Clusters 2, 4, and 6) in the irradiated ones (Figs. 2B and 3B).

The 200 differentially expressed genes with the greatest changes from the data used for Fig. 1 were classified into functional subgroups by BINGO, an algorithm for Gene Ontology (GO) significance analysis (41) (Table 2). The GO category (GOC) consisted of biological process (BP), cellular component (CC), and molecular function (MF). Genes with stress/stimulus-responsive GO terms were up-regulated in the control leaves, but cell wall/membrane- and development-related genes were down-regulated (Table 2A). This is probably due to the progress of leaf senescence. In contrast, the irradiated leaves showed up-regulation in a defense response-related GO term (Table 2B), which is in good agreement with previous reports of increased gene expression and activities of antioxidant enzymes in irradiated plants (13, 15, 31). Together with these results, the reduced expressions of genes with rhythmic process-, circadian rhythm-, and auxin stimulus-related GO terms explain the inhibition in stem growth and retardation in leaf senescence we observed in the irradiated leaves (supplementary Fig. S1). Moreover, the pronounced change in expression of the transcription regulator-related genes may imply that a broad spectrum of genes are affected in irradiated leaves. However, the pathway analysis of the dif-

ferentially expressed genes demonstrated that the expressions of the photosynthesis-related genes are relatively constant in the irradiated leaves during the reproduction stage (Table 3).

Comparison of Gene Expression in Control and Irradiated Leaves during the Reproduction Stage

To investigate effects of γ radiation on genome-wide gene expression during the reproduction stage, transcript levels of genes in irradiated leaves were compared to those in the respective controls at 3, 6, 9, 13 and 16 days after irradiation. Differentially expressed genes with a twofold change were selected, and a significance level of 0.05 after \log_2 transformation of the normalized data was used. As discussed above for Table 2B, radiation caused a significant change in the expression of numerous genes (Fig. 4). The most striking change in gene expression was observed at 9 days after irradiation, when 2164 and 1735 genes were up- and down-regulated, respectively.

ROS are among the most decisive mediators in intracellular signaling of radiation responses. Cellular responses to radiation can occur through direct participation of ROS in cell signaling (18, 19) and/or indirect involvement of LMW signaling factors released from the reactions of ROS and

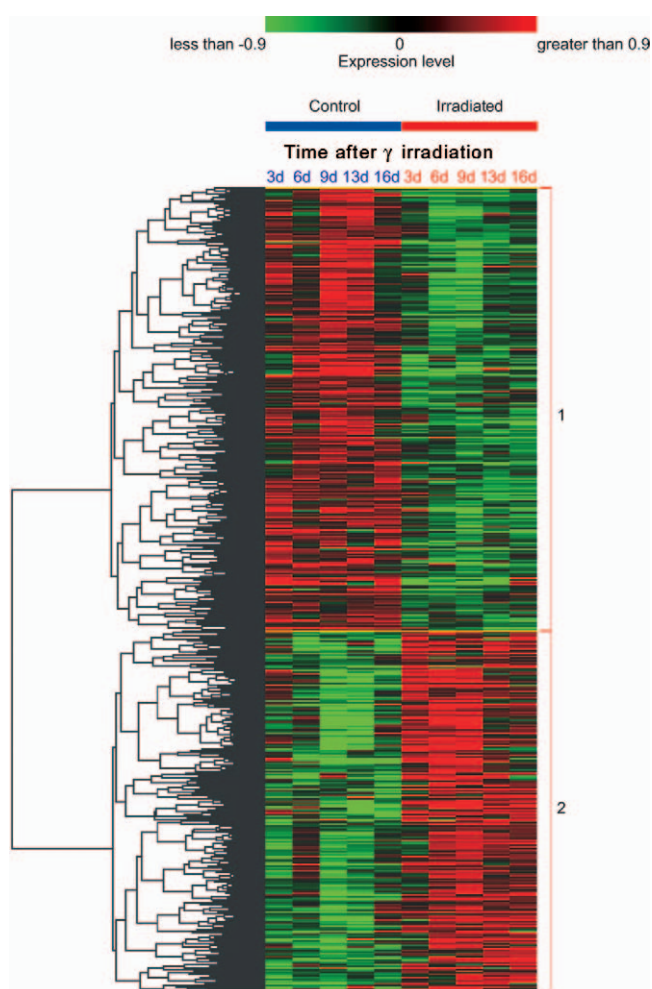


FIG. 6. Hierarchical cluster analysis of 2856 genes with ≤ 0.05 significance in the Welch's t test of the control and irradiated groups. Each horizontal line shows the expression data for one gene after normalization at the times indicated. Colors show the normalized expression levels. Induction (or repression) ranges from black to red (or green) with a fold-change scale bar shown above the cluster. Numbers (1 and 2) indicate defined sub-branches or clusters.

neighboring cellular components (20, 21). Therefore, we attempted to analyze the expression of genes for ROS-scavenging antioxidant enzymes as crucial factors in the propagation of radiation responses (Table 4A). Some of them were confirmed by real-time quantitative PCR using the same RNA samples.

Transcription levels of cytosolic Cu/Zn-superoxide dismutase (cytCu/Zn-SOD, *csd1*, accession no. At1g08830), chloroplastic Cu/Zn-SOD (chlCu/Zn-SOD, *csd2*, accession no. At2g40100), and dehydroascorbate reductase (DHAR, *dhar*, accession no. At1g19570) genes were higher overall in the irradiated leaves than in the control ones during the postirradiation period, reaching maximum levels at 9 or 13 days after irradiation (Fig. 5A, B and E). In contrast, the expressions of the glutathione peroxidase (GP, *gp*, accession no. At4g31870) and catalase 2 (CAT2, *cat2*, accession no. At4g35090) genes remained lower in the irradiated leaves

during the same period (Fig. 5F and G). Interestingly, the transcript levels of the chloroplastic Fe-SOD (chlFe-SOD, *fsd1*, accession no. At4g25100) and stromal ascorbate peroxidase (sAPX, *sapx*, accession no. At4g08390) genes fluctuated more dramatically, remaining lower in the irradiated leaves until 9 days after irradiation and becoming higher afterward (Fig. 5C and D). Although changes in the expression levels of the genes were not the same in the microarray and real-time PCR results due to the normalization for the former and the comparative expression for the latter using *actin2* as an internal control gene, the overall expression patterns of the genes were similar in the two systems. These data are in good agreement with high SOD and low CAT activities in plants after an acute exposure to relatively high doses of radiation (15, 48). However, the distinct expression patterns of antioxidant enzyme genes during the postirradiation period may indicate the existence of a complicated and controlled gene regulation system for radiation-induced gene responses in plants. Similarly, the differential expression patterns for the genes encoding the SOD, APX and CAT isoenzymes have been reported in rice plants after γ irradiation under salt stress (22). The results shown in Table 5 suggest that the differentially expressed genes for ascorbate/glutathione metabolism may contribute to the radiation-induced stress and defense signaling (49). The high gene count for ribosome at 9 days after irradiation can be correlated with the maximum change in gene expression, as shown in Fig. 4.

Distinctive changes in photosynthesis and pigment metabolism of plants after radiation exposure have been documented (11, 15, 50, 51). In particular, a high dose of radiation could cause specific physical alterations in *Arabidopsis* seedlings at certain developmental stages, such as accumulation of chlorophyll and carotenoid and elevation in photosynthetic saturation for light use as well as inhibition of stem growth and retardation of leaf senescence. Although the expressions of the photosynthesis-related genes are relatively constant in the irradiated leaves during the reproduction stage compared to those at 3 days after irradiation (Table 3), their relative expressions compared to the respective controls showed significant radiation-induced changes in photosynthesis and pigment metabolism (Table 4B, Fig. 5H, I and J). Expression of glutamyl-tRNA reductase 1 (HEMA1, *hemA1*, accession no. At1g58290), NADPH-protochlorophyllide oxidoreductase A (PORA, *porA*, accession no. At5g54190), and NADPH-protochlorophyllide oxidoreductase B (PORB, *porB*, accession no. At4g27440) was increased during the postirradiation period, with maximum levels reached at different times. Differential but increased transcript levels of these genes, which encode two major enzymes for chlorophyll synthesis (52, 53), may contribute to the increased chlorophyll content of the irradiated leaves at 13 and 16 days after irradiation (supplementary Fig. S1). In contrast, genes of chlorophyll A-B binding proteins 2:4 (LHCB2:4, *lhcb2:4*, accession no. At3g27690) and 4:3 (LHCB4:3, *lhcb4:3*, acces-

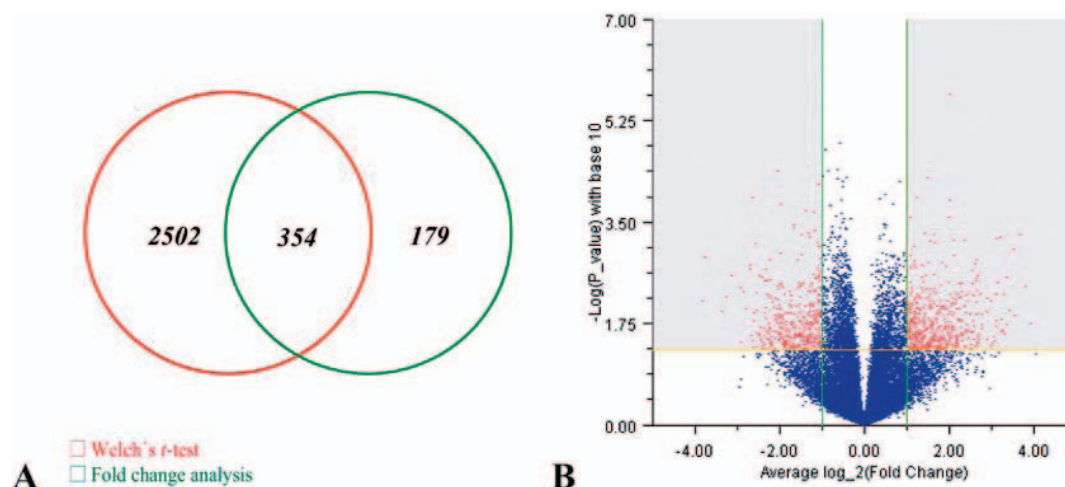


FIG. 7. Distribution of the genes from Welch's *t* test of the control and irradiated groups. Panel A: Filtered data for twofold-change genes (533 genes) and ≤ 0.05 significance in Welch's *t* test (2856 genes). Panel B: Unfiltered data for comparison of the control and irradiated groups. Red spots represent 354 genes in common of the fold-change analysis and the Welch's *t* test, while blue ones indicate the other 2681 of the total 3035 genes.

sion no. At2g40100) showed opposite expression patterns; the former showed higher transcript levels in the irradiated leaves than in the controls during the postirradiation period while the latter showed the reverse. The alternative expression of *lhcb2:4* can be correlated with retardation of leaf senescence in the irradiated leaves during the reproduction stage (supplementary Fig. S1).

Comparison of Gene Expression in the Control and Irradiated Groups throughout the Reproduction Stage

To compare transcript levels of genes in the control and irradiated groups during the postirradiation period, we carried out clustering analysis of 2856 genes below the 0.05 level of significance in Welch's *t* test (Fig. 6). The genes were divided into two subgroups (Clusters 1 and 2) based on the expression pattern. Expression of Cluster 1 genes (1570 genes) was decreased in the irradiated leaves, while that of Cluster 2 genes (1286 genes) was increased. Furthermore, filtered data for genes with more than a twofold change (533 genes) and changes with less than 0.05 significance in Welch's *t* test (2856 genes) included 354 differentially expressed genes, of which 225 were up-regulated and 129 down-regulated in the irradiated group (Fig. 7). However, only *porB*, *lhcb4.3*, *cat2* and *lhcb2:4* among the genes listed in Table 4 were selected in common with the 354 differentially expressed genes (supplementary Table S1). The other genes, especially the antioxidant genes confirmed in Fig. 5, were not selected as significant differentially expressed genes in the comparison of the control and irradiated groups. This can be attributed to the transience and fluctuation of their expressions, which changed throughout the postirradiation period. Instead, the involvement of two ribonucleotide reductase genes (accession nos. At2g21790 and At3g27060), which are critical for cell cycle progression, DNA damage repair, and plant development (54), implicated the most well-known genetic re-

sponse for recovery in the irradiated leaves. This may be involved in the sudden increase of affected genes at 9 days after irradiation (Fig. 4).

In the present study, we showed changes in transcript levels of *csd1*, *csd1*, *fsd1*, *sapx*, *dhar*, *cat2* and *gp* in leaves during the 16-day period after γ irradiation of plants at the early reproduction stage. Most of these enzymes belong to ascorbate-glutathione pathway, and they scavenge ROS or its derivatives (55). Interestingly, we found low transcript levels of *cat2* and *gp* in the irradiated leaves. Although these results are partly predictable from our previous studies (15, 22), there are also opposing results with chronic radiation-induced responses (13). Similarly, the expression of *cat2* was strongly induced by UV-light pulses (290 to 400 nm) in leaves of maize seedlings (56). Accordingly, the transcription of these genes appears to be inhibited after exposure to an acute dose of radiation. In contrast, the expression patterns of *hemA1*, *porA*, *porB*, *lhcb2:4* and *lhcb4.3* can be correlated with the physiological changes in the irradiated leaves, e.g., retardation in leaf senescence, accumulation of chlorophyll and carotenoid, and elevation of photosynthetic saturation for light use (Table 4 and supplementary Fig. S1). Together with *cat2*, however, *porB* and *lhcb4.3* belong to the most distinguishable genes in terms of the expression change throughout the postirradiation period (data not presented).

Recently, Nagata *et al.* (31) reported the results of microarray analysis of genes that respond to ionizing radiation in *Arabidopsis*. Their study focused on the short-term effect of ionizing radiation on the expression of genes within 24 h after γ irradiation with a dose of 3 kGy, which makes the subsequent growth and survival of plants impossible. In contrast, we attempted to determine the long-term effects of ionizing radiation on the genome-wide expression of genes in *Arabidopsis* rosette leaves during the reproductive stage, which is responsible for the phenotypic and physiological changes in the irradiated leaves.

In conclusion, we generated genome-wide transcriptomic profiles of radiation-responsive genes in plants at the reproductive stage using the *Arabidopsis* genome array and revealed the changes in expression of the genes involving in antioxidative defense, photosynthesis and chlorophyll synthesis. Considering the importance of the ROS scavenging system and pigment metabolism in the radiation-induced responses of plants, our results could allow deeper insight into the long-term changes in the functional genes in plants after radiation exposure. Next we are going to use the expression profile to elucidate transcription factors orchestrating the expressions of radiation-responsive genes, especially those related to antioxidative defense and pigment metabolism.

SUPPLEMENTARY INFORMATION

Supplemental document containing additional information about methodology: <http://dx.doi.org/10.1667/RR0963.1.s1>.

FIG. S1. Phenotypic and physiological differences between the control and irradiated seedlings after γ irradiation: <http://dx.doi.org/10.1667/RR0963.1.s2>.

TABLE S1. Expression profile of 225 induced and 129 repressed genes in the irradiation groups from the 354 differentially expressed genes selected in Fig. 6: <http://dx.doi.org/10.1667/RR0963.1.s3>.

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