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Morphology and histology of Lyonet's gland of the tropical tasar silkworm, *Antheraea mylitta*

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Abstract

The morphology and histology of Lyonet's gland in the second to fifth instar larvae of *Antheraea mylitta* Drury (Lepidoptera: Saturniidae) are described. Each of the paired silk glands of this silk worm were associated with a Lyonet's gland. The paired Lyonet's glands were located on the ventrolateral sides of the esophagus, close to the subesophageal ganglion. Whole mount and SEM observations revealed that each Lyonet's gland consisted of a rosette of glandular mass, and a short narrow tubular duct opening into the anterior part of the silk gland (ASG), close to the common excretory duct. In each instar, these glands were unequal in size. The glandular mass was innervated by fine nerves from the subesophageal ganglion, suggesting a neural control for the glandular activity. The glandular mass was made up of clustered long cells wrapped by a thin basal lamina, which was continuous over the non-secretory low columnar cells of the Lyonet's gland duct and ASG. The narrow bases of long cells of each glandular mass led into the lumen of the duct of the gland. Histochemical analysis of fully developed Lyonet's gland showed clustered lipid granules in the gland cells.

Keywords: Daba TV ecorace, silk gland

Abbreviations: ASG, anterior part of the silk gland; Cd, common silk gland duct; Hyp, hypopharynx; Lly, left Lyonet's gland; Lu, lumen; Lyd, Lyonet's gland duct; Msp, muscles of silk press; Rly, right Lyonet's gland; Sp, silk press; Spn, spinneret; Ti, tunica intima; Tm, tunica media; Tp, tunica propria; Tra, tracheoles

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Introduction

The Lyonet's gland, first described in 1760 by Lyonet in lepidopteran larvae (Machida 1965), is often referred to as "Filippi's gland" in the silkworm *Bombyx mori* L. (Lepidoptera: Bombycidae) (Waku and Sumimoto 1974; Akai 1984). This gland usually occurs close to the excretory duct of the silk gland, and communicates with it (Waku and Sumimoto 1974). It has been considered as an accessory gland of the silk gland (Waku and Sumimoto 1974; Sehnaal and Akai 1990).

The function of Lyonet's gland is still uncertain (Victoriano and Gregorio 2004). Its role in the exchange of small molecules, such as water and ions (Waku and Sumimoto 1974), in the secretory process of cementing substance for the silk elements (Day and Waterhouse 1953; Wigglesworth 1972), and secretion of some lubricating substance that helps in the extrusion of silk from the silk glands (Glasgow 1936; Day and Waterhouse 1953) have been suggested.

Antheraea mylitta D. (Lepidoptera: Saturniidae) is the producer of commercial tasar silk in tropical India. A survey of literature reveals that no information is available on the Lyonet's glands in the larvae of this silk moth. The present work is an attempt to describe the morphology, histology, and histochemical properties of these glands.

Materials and Methods

Second to fifth instar larvae of *A. mylitta* (Daba TV ecorace) were procured from the field during rearing periods from Tasar Pilot Project Centre, Salboni, Purulia (West Bengal). The Lyonet's glands were removed, and fixed in appropriate fixatives for whole

mounts, histology and histochemical studies. The glands of five larvae of each second to fifth instars were measured using the micrometer. 6 μ thick sections of the gland were stained with Hematoxylin and Eosin/Triple Mallory, Mercuric bromophenol blue, PAS reagents, and Sudan black-B. For scanning electron microscopy (SEM), the Lyonet's glands of fifth instar larvae were fixed in 2.5% glutaraldehyde in 0.1 M phosphate buffer (P^H 7.2 to 7.4) at 4° C for 2-3 hours, and then post-fixed in 1% osmium tetroxide in a similar buffer for 2 hours. The post-fixed specimens were dehydrated through graded series of alcohol and acetone, critical point dried with liquid CO₂, and gold coated in a sputter. Scanning of specimens was performed by field emission scanning electron microscope.

Results

In *A. mylitta*, there was a pair of small, creamy white Lyonet's glands, each associated with the anterior parts of paired silk glands. These were located on the ventrolateral sides of the esophagus, close to the subesophageal ganglion (Figures 1, 2). Fine nerve fibers arising from the subesophageal ganglion innervated the Lyonet's gland, indicating a neural control for glandular function (Figure 3). Whole mount of the glands and SEM studies (Figure 4) revealed that each gland was made up of a rosette of glandular mass, and a narrow tubular duct that opens into the inner side of the anterior part of the silk gland (ASG), near the origin of common excretory ducts (Figures 4, 5, 6). These glands increased in size as the larval instar advanced. It was interesting to note that in the second to fourth instar larvae, the left Lyonet's gland was larger than the right gland, but in the fifth instar, the right

Table 1. Measurement of Lyonet’s glands and their ducts in five, second to fifth instar larvae of *Antheraea mylitta*.

Instar	No. of specimen	Left Lyonet's gland		Stalk of left Lyonet's gland		Right Lyonet's gland		Stalk of right Lyonet's gland	
		Length (μ)	Width (μ)	Length (μ)	Width (μ)	Length (μ)	Width (μ)	Length (μ)	Width (μ)
2nd	1	120	112	64	32	96	88	40	24
	2	120	80	56	32	120	64	24	32
	3	128	80	40	32	112	72	32	32
	4	104	72	48	24	88	88	40	32
	5	136	112	72	48	112	96	48	40
	Mean ± S.D	121.00 ± 10.61	91.20 ± 17.23	56.00 ± 11.31	33.60 ± 7.83	105.60 ± 11.75	81.60 ± 11.75	36.80 ± 8.15	32.00 ± 5.05
3rd	1	232	80	40	32	136	80	32	32
	2	160	136	120	40	144	88	24	24
	3	240	88	48	40	152	96	36	32
	4	208	120	128	48	184	100	48	40
	5	248	208	96	40	160	96	56	40
	Mean ± S.D	217.60 ± 14.25	126.40 ± 45.64	86.40 ± 36.27	40.00 ± 5.05	155.20 ± 16.47	92.00 ± 7.15	39.20 ± 11.42	33.60 ± 5.98
4th	1	304	216	196	80	280	208	176	44
	2	352	232	208	84	296	192	196	52
	3	280	200	200	64	240	160	180	48
	4	264	216	184	56	248	184	152	40
	5	288	224	192	64	232	176	184	48
	Mean ± S.D	297.60 ± 30.10	217.60 ± 10.57	196.00 ± 8.00	69.60 ± 10.61	259.20 ± 24.57	184.00 ± 16.00	177.60 ± 14.44	46.40 ± 4.07
5th	1	376	336	320	80	432	352	400	88
	2	320	240	232	80	336	264	256	80
	3	240	200	216	48	360	240	256	56
	4	280	224	216	72	408	360	272	80
	5	288	248	208	80	416	336	368	76
	Mean ± S.D	300.80 ± 45.47	249.60 ± 46.20	238.40 ± 41.53	72.00 ± 12.39	390.40 ± 36.27	310.40 ± 48.89	310.00 ± 61.21	76.00 ± 10.73

Lyonet’s gland was larger than the left one (Figures 5, 6). The morphometric records of the glands and their ducts from second to fifth instar larvae are presented in Table 1.

The basic histological features of Lyonet’s gland in the second to fifth instar larvae were similar (Figure 7). Each gland was composed of long cells of various lengths, arranged in whorls. The longest cell measured had a length of ~140μ. The whorls of long cells were wrapped by an extremely fine basal lamina for which the gland had a superficial rosette appearance. The bases of the glandular cells remained attached to the cuticular intimal layer of the Lyonet’s gland duct (Figures 8, 9). Each glandular cell contained a long polyploid nucleus. These cells were found to be richly supplied with tracheoles. Fine nerve fibers were also found, ending over the surfaces of the gland.

The histology of the duct of Lyonet’s gland was quite similar to that of ASG, the walls comprised of three layers: the outer most thin tunica propria, or basal lamina; a middle epithelial layer, or tunica media, made up of single layered non secretory low columnar

cells with chromatin lumps; and the inner most thick tunica intima, or cuticular layer, surrounding the lumen of the duct. In fifth instar larvae, the thicknesses of tunica propria, tunica media, and tunica intima were 3μ, 16μ and 5μ respectively. The lumen of the Lyonet’s gland duct was 8μ wide.

The Lyonet’s glands became fully functional in the late fifth instar. This is evidenced by the presence of few secretory granules in the Lyonet’s gland cells of the fourth instar larvae, while in fifth instar larvae the glandular cells contained a much larger number of secretory granules.

The chemical nature of secretory materials of the Lyonet’s gland cells was studied using histochemical stains. It was found that the Lyonet’s gland cells were intensely mercuric bromophenol blue positive, indicating high protein content in the cells. Intense Periodic acid Schiff (PAS) reaction was confined to the basal and apical cytoplasm of the glandular cells, indicating high glycogen content in these regions. In the case of the Sudan Black-B reaction, the cells showed negative results, except for the clustered granules, which were

intensely positive, indicating high lipid content in them (Figure 10).

Discussion

The paired Lyonet's glands, characteristic of silk synthesizing Lepidoptera, have been studied in *Bombyx mori* (Waku and Sumimoto 1974), *Ostrinia nubilalis* (Drecktrah et.al. 1966), *Spodoptera frugiperda* (Chi et.al. 1975), and *Diatraea saccharalis* (Victoriano and Gregorio 2004). The results of the present study on the Lyonet's glands of *A. mylitta* revealed a similarity in the location and basic morphology of these accessory glands in all the species studied. However, as far as detailed morphology of these glands is concerned, in each case the arrangement of long glandular cells over the duct was different and characteristic for the species. It is also revealed that the basic histology of the gland and its duct were also similar.

The Lyonet's glands in *A. mylitta* appeared to be neuro-controlled, as they were supplied by fine nerves from the subesophageal ganglion. This was quite similar to that described in *D. saccharalis* (Victoriano and Gregorio 2004).

In *A. mylitta*, the histology of the Lyonet's gland duct was exactly the same as that of the ASG (Patra 2008). Furthermore, there is a clear continuity of the three layers (tunica propria, tunica media, and tunica intima), as well as the lumina of ASG, and the Lyonet's gland duct. This indicates that the Lyonet's gland duct was formed as a result of the out-pushing of the walls of the ASG during early development. The origin of the glandular cells of Lyonet's gland appeared to be due to the enormous elongation of tunica media cells of the duct at the growing end. These assumptions, however, require a detailed

study of the embryonic development of the Lyonet's gland and its duct.

Although the role played by the Lyonet's gland is still not clear, histochemical studies of the gland cells in *A. mylitta* have clearly revealed the presence of clustered lipid granules in their cytoplasm, which may be secreted as a lubricating substance, facilitating the extrusion of silk from the silk glands. A similar suggestion has been made by Glasgow (1936) and Day and Waterhouse (1953).

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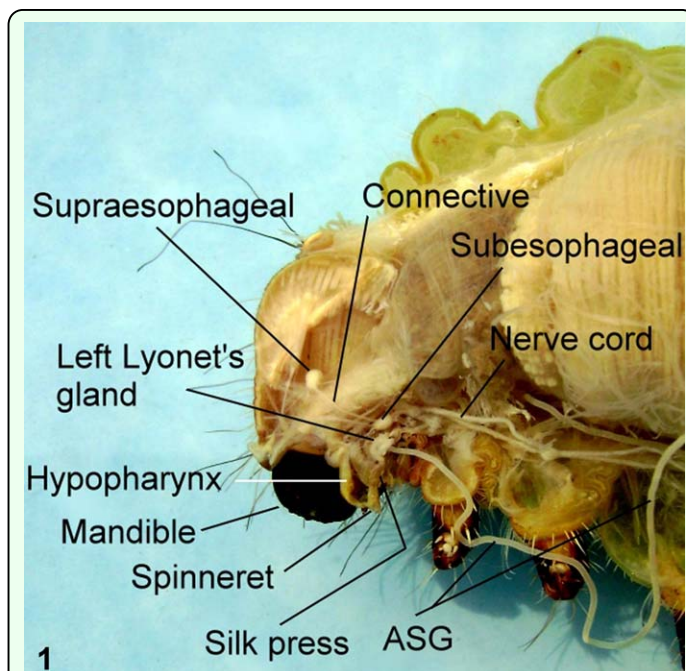


Figure 1. Dissected anterior region of fifth instar larva of *Antheraea mylitta* showing the location of Lyonet's gland close to the subesophageal ganglion. High quality figures are available online.

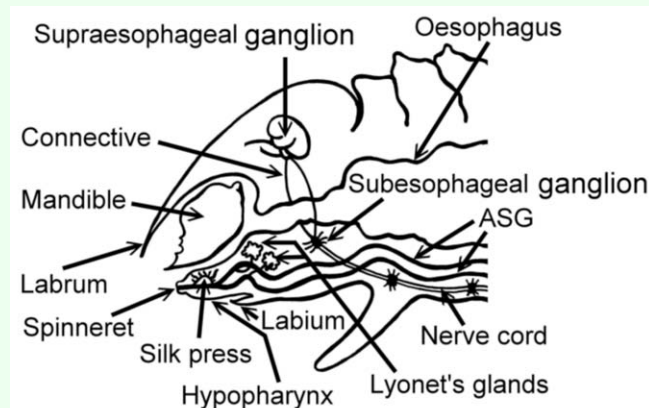


Figure 2. Location of Lyonet's glands in *Antheraea mylitta* larva (Diagrammatic). High quality figures are available online.

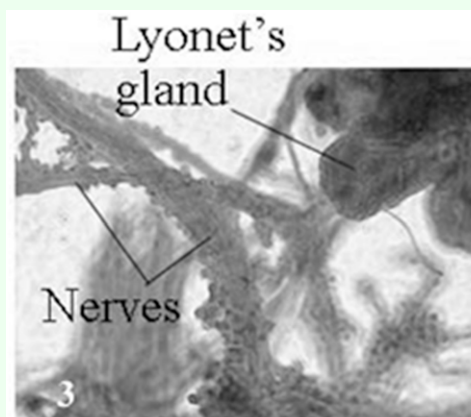


Figure 3. Lyonet's gland and associated nerves from the subesophageal ganglion in *Antheraea mylitta* larva (w.m, X 1000). High quality figures are available online.

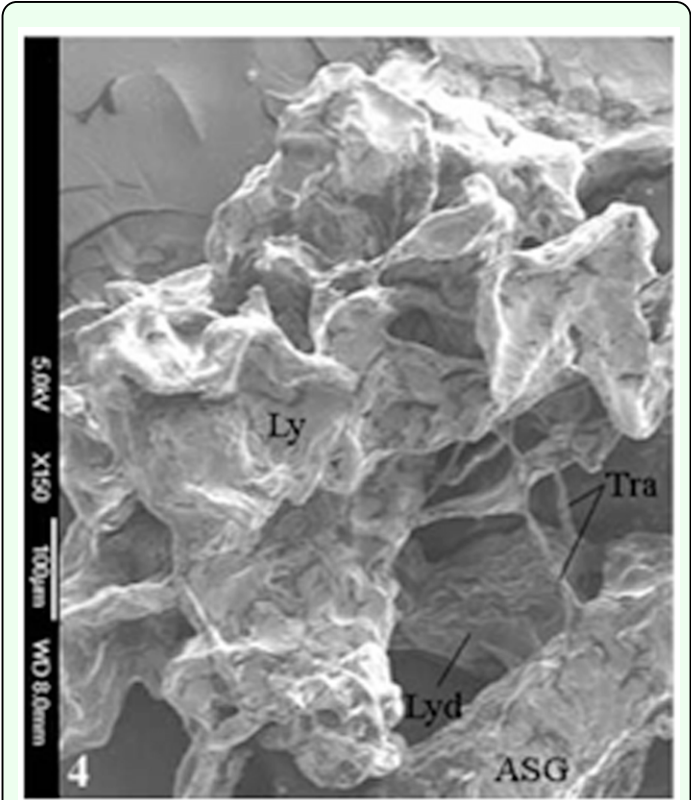


Figure 4. SEM image of Lyonet's gland surface in *Antheraea mylitta* larva. High quality figures are available online.

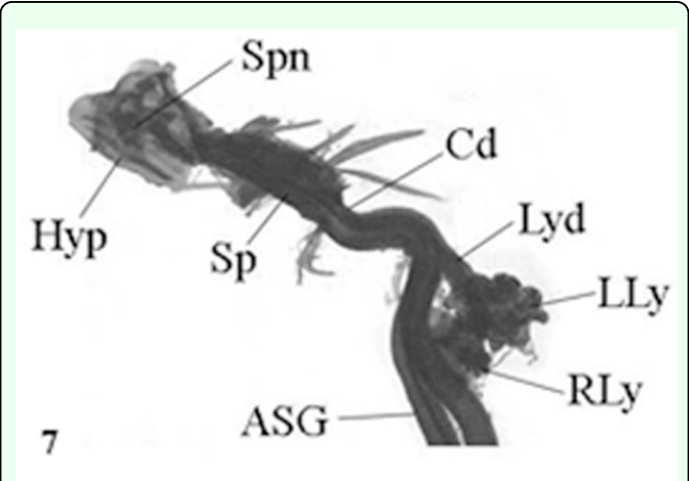


Figure 5. Lyonet's glands of fourth instar *Antheraea mylitta* larva (w.m. X 50). High quality figures are available online.

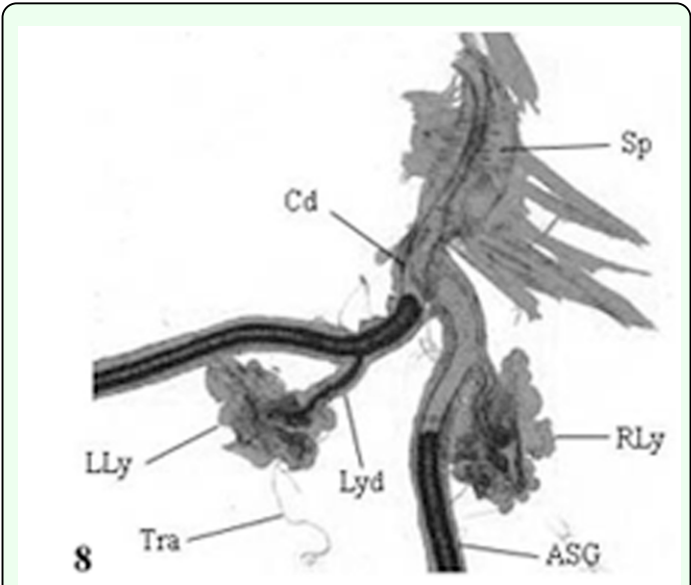


Figure 6. Lyonet's glands of fifth instar *Antheraea mylitta* larva (w.m. X 50). High quality figures are available online.

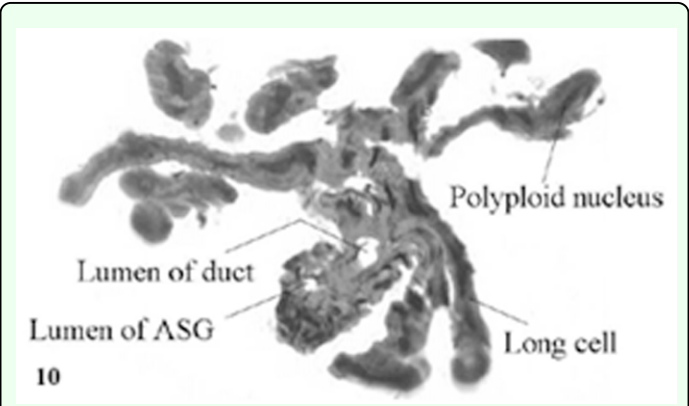


Figure 7. L. S. Lyonet's gland of third instar *Antheraea mylitta* larva showing cells and polyloid nuclei (X 450). High quality figures are available online.

