BioOne COMPLETE

No Evidence of Persistent Yersina pestis Infection at Prairie Dog Colonies in North-Central Montana

Authors: Holmes, Brian E., Foresman, Kerry R., and Matchett, Marc R.

Source: Journal of Wildlife Diseases, 42(1) : 164-169

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-42.1.164

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

Journal of Wildlife Diseases, 42(1), 2006, pp. 164–169
© Wildlife Disease Association 2006

No Evidence of Persistent *Yersina pestis* Infection at Prairie Dog Colonies in North-Central Montana

Brian E. Holmes,[1,3] Kerry R. Foresman,[1] and Marc R. Matchett[2] [1] Division of Biological Sciences, University of Montana, Missoula, Montana 59812, USA; [2] Charles M. Russell National Wildlife Refuge, Lewistown, Montana 59457, USA; [3] Corresponding author (email: Brian_E_Holmes@blm.gov)

ABSTRACT: Sylvatic plague is a flea-borne zoonotic disease caused by the bacterium *Yersinia pestis*, which can cause extensive mortality among prairie dogs (*Cynomys*) in western North America. It is unclear whether the plague organism persists locally among resistant host species or elsewhere following epizootics. From June to August 2002 and 2003 we collected blood and flea samples from small mammals at prairie dog colonies with a history of plague, at prairie dog colonies with no history of plague, and from off-colony sites where plague history was unknown. Blood was screened for antibody to *Y. pestis* by means of enzyme-linked immunosorbent assay or passive hemagglutination assay and fleas were screened for *Y. pestis* DNA by polymerase chain reaction. All material was negative for *Y. pestis* including 156 blood samples and 553 fleas from colonies with a known history of plague. This and other studies provide evidence that *Y. pestis* may not persist at prairie dog colonies following an epizootic.

Key words: *Cynomys*, fleas, prairie dogs, sylvatic plague, *Yersinia pestis*.

Sylvatic plague is a flea-borne zoonotic disease of mammals caused by the bacterium *Yersinia pestis*. The disease primarily affects wild rodents, although many other groups of wild and commensal mammals can become infected (Gage et al., 1995). *Yersinia pestis* likely evolved in Asia with subsequent introduction onto all continents except Antarctica and Australia. In North America, the presence of *Y. pestis* was first identified in approximately 1900, having arrived in Pacific Coast ports via infected rats (*Rattus* sp.) on ships from Asia (Eskey and Haas, 1940). Today, the range of the disease in North America includes areas west of the 100th meridian (Cully et al., 2000).

The maintenance of *Y. pestis* in the wild depends on a complex set of interactions between host, vector, pathogen, and environmental factors that are poorly understood. In general, the sylvatic cycle of infection is characterized by relatively stable periods of enzootic activity where *Y. pestis* circulates at low levels within the "maintenance" host community, followed by explosive epizootics involving one or more species of "amplifying" host that often experience high mortality. In western North America, these epizootic hosts include species of prairie dogs (*Cynomys*) in which plague-associated die-offs can be particularly dramatic, with mortality often approaching 100% within colonies (Rayor, 1985; Menkens and Anderson, 1991).

Prairie dogs often repopulate colonies following epizootics (Menkens and Anderson, 1991; Cully et al., 1997) and these colonies may then persist for many years or experience a plague epizootic again. Barnes (1982) reported a recurrence of plague epizootics within 4 to 5 yr and Cully et al. (1997) reported an epizootic again after 3 yr. Whether these cases in which the same colonies experience plague again represent a continued presence of infection among hosts in that area or a reintroduction of *Y. pestis* from surrounding areas is not known. The objective of this study was to address two competing hypotheses regarding the maintenance of *Y. pestis* between epizootics: 1) following an epizootic, *Y. pestis* persists in an area at low levels within the host community or 2) following an epizootic, *Y. pestis* does not persist in a localized area and recurring plague epizootics result from reintroduction of the organism (i.e., *Y. pestis* is absent during the period of recovery). To test this we collected blood and flea samples from small mammals at prairie dog colonies with a history of

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

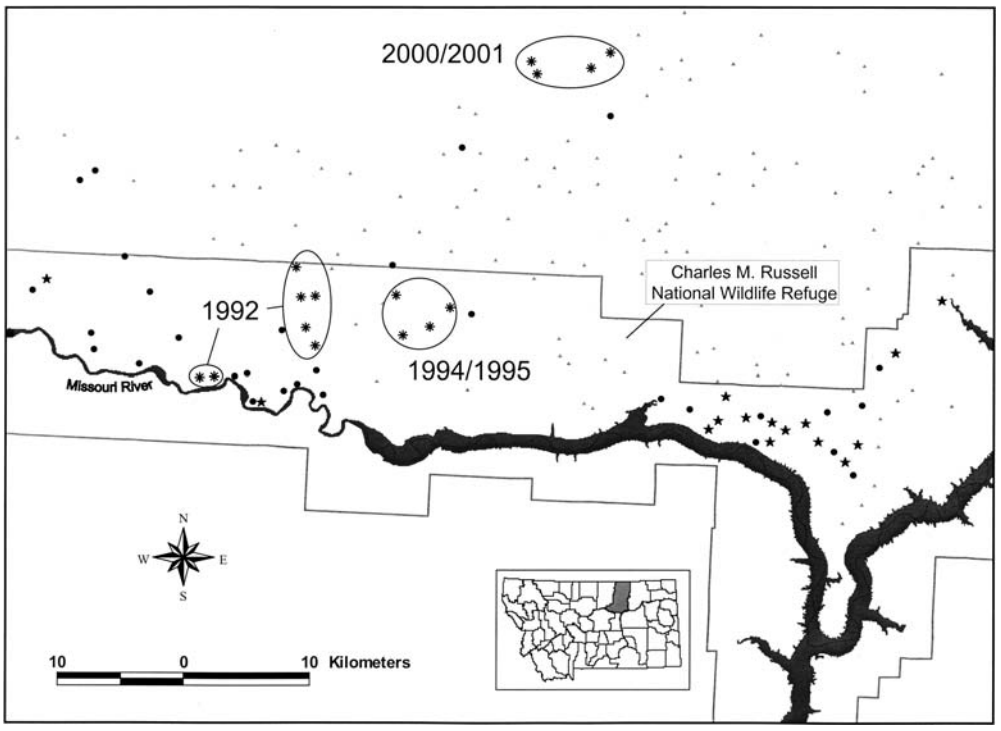


FIGURE 1. Map of southern Phillips County, Montana, showing the location of study sites including prairie dog colonies with a history of plague (asterisks), colonies with no history of plague (stars), and off-colony sites (circles). The year of plague epizootics also is indicated. Grey triangles are point locations of additional prairie dog colonies in the study area, some affected by plague, some not, and many where plague history is uncertain.

sylvatic plague epizootics and screened them for evidence of *Y. pestis*. In addition, we also collected samples from prairie dog colonies with no known history of plague and from off-colony sites where plague history was unknown.

The study took place in southern Phillips County, Montana, USA (47°35′ to 47°50′N, 107°45′ to 108°45′W) during June through August 2002 and 2003 (Fig. 1). The area is characterized by shrub and grassland habitats typical of the northern Great Plains, with big sagebrush (*Artemisia tridentata*), black greasewood (*Sarcobatus vermiculatus*), western wheatgrass (*Agropyron smithii*), and blue grama (*Bouteloua gracilis*) as common species. In addition, the southern margin of the county borders the Missouri River and consists of forested "breaks" topography with ponderosa pine (*Pinus ponderosa*), Douglas-fir (*Pseudotsuga menziesii*), and Rocky Mountain juniper (*Juniperus scopulorum*). Elevations of study sites are between 740 and 1,050 m. The area is a mosaic of federal, state, and private land ownership and has supported approximately 300 active black-tailed prairie dog (*C. ludovicianus*) colonies during the past 20 years. The majority of study sites were located on the Charles M. Russell National Wildlife Refuge (CMR) with the remainder located on adjacent Bureau of Land Management (BLM) lands north of the refuge.

We sampled 36 sites in 2002 and 60 sites (36 resampled from 2002 and 24 new) in 2003. In total, 15 sites were prairie dog colonies with a history of plague, 15 were prairie dog colonies with no history of plague, and 30 were off-colony sites. Sites with a history of plague were

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

identified through regular mapping efforts by CMR and BLM personnel such that the location and year of epizootics among prairie dogs were known. Plague epizootics occurred between 1992 and 2001 at sites included in this study (Fig. 1). We are confident that die-offs attributed to plague were in fact plague epizootics because no other disease has yet been identified that causes such high mortality in prairie dogs (Barnes, 1993) and antibody to *Y. pestis* has been consistently found in coyotes (*Canis latrans*) and badgers (*Taxidea taxus*) in the study area (Matchett, 1999). Off-colony sites occurred in a variety of habitats and were located >400 m from the nearest prairie dog colony. Plague history at these sites was unknown because prairie dogs were the sentinel species used to indicate the presence or absence of plague epizootics and off-colony sites had no prairie dogs present for at least the past 20 years.

Each study site consisted of a 10 by 10 grid of 100 Sherman live-traps (H. B. Sherman, Tallahassee, Florida, USA) with 10-m spacing and 20 Tomahawk live-traps (Tomahawk Live Trap Co., Tomahawk, Wisconsin, USA) placed at prairie dog burrows on colony sites and systematically throughout the grid at off-colony sites. We anesthetized captured animals with isoflurane ("IsoFlo" Abbot Laboratories, North Chicago, Illinois, USA or "IsoSol" Halocarbon Laboratories, River Edge, New Jersey, USA) before blood and flea sampling. Fleas were collected from animals by using a conventional flea comb as well as from prairie dog burrows by using a previously described swabbing technique (Holmes, 2003). Fleas were stored in vials containing 2% NaCl solution with a small amount (<0.01%) of Tween 80, identified to species, and then frozen. We collected blood samples of approximately 200 µl from the retro-orbital sinus of small rodents by using micro-hematocrit tubes (Chase Scientific, Rockwood, Tennessee, USA) and for larger animals such as prairie dogs and cottontails (*Sylvilagus audubonii*) we collected blood by clipping a hindfoot toenail to induce bleeding. In 2002, whole-blood samples were stored in a conventional (−20 C) freezer upon return from the field. In 2003, most blood samples were centrifuged the day of collection to separate serum, which was then stored as above. The remaining samples were collected onto individual Nobuto filter papers (Advantec MFS, Pleasanton, California, USA) which were air-dried, placed in paper envelopes, and stored at room temperature.

Laboratory diagnostics were performed at the Centers for Disease Control and Prevention, Division of Vector-Borne Infectious Diseases, Plague Section, Fort Collins, Colorado, USA. Serologic analyses followed protocols described by Chu (2000). Serum and blood samples were screened for the presence of antibody against *Y. pestis*-specific Fraction 1 antigen by using either a competitive enzyme-linked immunosorbent assay (cELISA) or a passive hemagglutination assay (PHA); all Nobuto strips were screened using PHA. Flea pools of one to 10 individuals (corresponding to the same species, host, date, and site of capture) were screened for the presence of *Y. pestis* with a multiplex polymerase chain reaction (PCR) assay described by Stevenson et al. (2003).

The number and source of samples screened for evidence of *Y. pestis* by serologic and PCR analysis is given in Tables 1 and 2, respectively. All 156 blood samples and 553 fleas from small mammals trapped at prairie dog colonies with a history of plague over two summers were negative for antibodies to and DNA from *Y. pestis*. Likewise, all materials from prairie dog colonies with no history of plague (369 blood, 1,894 fleas) and from off-colony sites (439 blood, 603 fleas) were negative for antibodies to and DNA from *Y. pestis*. Although we sampled an equal number of prairie dog colonies with and without a history of plague in each year of the study, the number of diagnostic samples collected at colonies with no

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

TABLE 1. Number and source of serum and blood samples collected in Phillips County, Montana, during June through August 2002 and 2003. Samples are from prairie dog colonies with a history of plague (Plague), prairie dog colonies with no history of plague (No Plague), and off-colony sites (Off).

Host	2002			2003		
	Plague	No Plague	Off	Plague	No Plague	Off
Cynomys ludovicianus	22	36	–	17	32	–
Microtus ochrogaster	–	–	–	1	–	20
Neotoma cinerea	–	–	–	–	–	4
Onychomys leucogaster	1	1	1	12	6	3
Peromyscus maniculatus	45	84	98	53	205	302
Reithrodontomys megalotis	–	–	–	–	–	4
Sylvilagus audubonii	3	1	–	2	4	4
Tamias minimus	–	–	–	–	–	3
Total	71	122	99	85	247	340

history of plague was greater because capture rates were consistently higher at those sites than at colonies with a history of plague.

Two previous studies (Lechleitner et al., 1968; Cully et al., 1997) followed the progression of plague epizootics among prairie dogs and demonstrated that, in general, *Y. pestis* positive fleas from prairie dogs, their burrows, and associated mammals are most likely to be collected during an epizootic. These studies also illustrated that one year following an epizootic, some *Y. pestis* positive fleas may still be present in prairie dog burrows, but after two years, there is little or no evidence of *Y. pestis* in the vector community. Serologic results from these

TABLE 2. Number and source of fleas collected for plague testing in Phillips County, Montana, during June through August 2002 and 2003. Samples are from prairie dog colonies with a history of plague (Plague), prairie dog colonies with no history of plague (No Plague), and off-colony sites (Off). The flea species and total number tested are given for each host.

Host	2002			2003		
	Plague	No Plague	Off	Plague	No Plague	Off
Cynomys ludovicianus[a]	67	683	–	83	310	–
Microtus ochrogaster[b]	–	–	–	4	–	44
Neotoma cinerea[c]	–	–	–	–	–	86
Onychomys leucogaster[d]	3	11	–	22	8	6
Peromyscus maniculatus[e]	125	233	119	129	402	328
Reithrodontomys megalotis[f]	–	–	–	–	–	3
Sylvilagus audubonii[f]	1	–	–	2	6	17
Prairie dog burrow[h]	48	145	–	69	96	–
Total	244	1,072	119	309	822	484

[a] *Oropsylla hirsuta* (*n*=208), *Oropsylla tuberculata* (*n*=68), *Pulex simulans* (*n*=867).

[b] *Aetheca wagneri* (*n*=4), *Malareus telchinus* (*n*=2), *Peromyscopsylla hesperomys* (*n*=2), *Orchopeas leucopus* (*n*=40).

[c] *Aetheca wagneri* (*n*=9), *Eumolpianus eumolpi* (*n*=1), *Orchopeas agilis* (*n*=76).

[d] *Aetheca wagneri* (*n*=22), *Foxella ignota* (*n*=5), *Malareus telchinus* (*n*=3), *Peromyscopsylla hesperomys* (*n*=20).

[e] *Aetheca wagneri* (*n*=1,145), *Callistopsyllus terinus* (*n*=4), *Epitedia wenmanni* (*n*=1), *Eumolpianus eumolpi* (*n*=1), *Foxella ignota* (*n*=1), *Malareus telchinus* (*n*=82), *Orchopeas leucopus* (*n*=11), *Peromyscopsylla hesperomys* (*n*=91).

[f] *Aetheca wagneri* (*n*=3).

[g] *Aetheca wagneri* (*n*=2), *Cediopsylla inaequalis* (*n*=24).

[h] *Aetheca wagneri* (*n*=11), *Oropsylla hirsuta* (*n*=147), *Oropsylla tuberculata* (*n*=40), *Peromyscopsylla hesperomys* (*n*=1), *Pulex simulans* (*n*=159).

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

two studies also failed to document evidence of persistent infection in the host community at affected colonies. Lechleitner et al. (1968) found antibodies to *Y. pestis* in only one of 108 deer mice (*Peromyscus maniculatus*) (the one seropositive animal was sampled during the active epizootic) and Cully et al. (1997) only found antibodies in prairie dogs. In both studies, the epizootic appeared to diminish over the course of about a year. Davis et al. (2004) also found evidence of a one to two year "fade-out" period following plague epizootics among populations of the great gerbil (*Rhombomys opimus*) in Asia when evidence of *Y. pestis* was still detectable but after which the populations were apparently plague-free.

We found no evidence that *Y. pestis* persists at black-tailed prairie dog colonies with a history of plague, at least in the host and vector species that we sampled. The most conservative interpretation of these negative data is that *Y. pestis* infection was not widespread among small mammals in southern Phillips County. However, if infection occurred in isolated pockets or was present in only a small proportion of the small mammal community we may not have been able to detect it with our sampling effort. What these data also suggest is that prairie dog colonies per se are not ideal focal areas for the long-term maintenance of *Y. pestis* in our study area.

The mechanism by which *Y. pestis* persists is still unclear. One potential scenario is that the disease continually moves across the landscape, driven by new infection of susceptible hosts. However, the patchy distribution of plague-affected prairie dog colonies in Phillips County does not appear to support this hypothesis, although several colonies in close proximity to one another were often affected at once. Another possibility is that *Y. pestis* persists in discrete enzootic foci that maintain appropriate conditions for long-term persistence, and these are a source of bacteria for epizootics among highly susceptible species such as prairie dogs. In Phillips County, this may occur off of prairie dog colonies where the diversity of potential enzootic hosts is higher (Holmes, 2003). Maintenance of *Y. pestis* is thought to be dependent on this continued circulation among competent hosts (Barnes, 1993) and several authors (Olsen, 1981; Gage et al., 1995; Biggins and Kosoy, 2001) have proposed that the factors most likely to support permanent plague foci include several host species co-occurring in areas of diverse or patchy habitats. If so, areas where such diverse habitats occur in proximity to prairie dog colonies—that is, where both the proposed enzootic and epizootic components of the plague system coexist—may prove important in supporting permanent plague foci and perpetuating epizootics.

We thank K. Gage and staff at the CDC, DVBID, Plague Section for providing laboratory space and expertise. Comments by J. Cully and two anonymous reviewers were helpful in improving the manuscript. Field methods were approved by the Institutional Animal Care and Use Committee at the University of Montana. Funding for portions of this work was provided by the United States Fish and Wildlife Service, The Nature Conservancy, and the American Museum of Natural History (Theodore Roosevelt Memorial Grants).

LITERATURE CITED

BARNES, A. M. 1982. Surveillance and control of bubonic plague in the United States. *In* Animal disease in relation to animal conservation, M. A. Edwards and U. McDonnell (eds.). Academic Press, London, UK, pp. 237–270.

———. 1993. A review of plague and its relevance to prairie dog populations and the black-footed ferret. *In* Proceedings of the symposium on the management of prairie dog complexes for the reintroduction of the black-footed ferret, J. L. Oldemeyer, D. E. Biggins, B. J. Miller and R. Crete. (eds.). US Fish and Wildlife Service, Washington, D.C., pp. 28–37.

BIGGINS, D. E., AND M. Y. KOSOY. 2001. Influences of introduced plague on North American mammals: implications from ecology of plague in Asia. Journal of Mammalogy 82: 906–916.

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

CHU, M. C. 2000. Laboratory manual of plague diagnostic tests. Centers for Disease Control and Prevention, Division of Vector-Borne Infectious Diseases, Fort Collins, Colorado, 129 pp.

CULLY, J. F. JR., A. M. BARNES, T. J. QUAN, AND G. MAUPIN. 1997. Dynamics of plague in a Gunnison's prairie dog colony complex from New Mexico. Journal of Wildlife Diseases 33: 706–719.

———, L. G. CARTER, AND K. L. GAGE. 2000. New records of sylvatic plague in Kansas. Journal of Wildlife Diseases 36: 389–392.

DAVIS, S., M. BEGON, L. DE BRUYN, V. S. AGEYEV, N. L. KLASSOVSKIY, S. B. POLE, H. VILJUGREIN, N. C. STENSETH, AND H. LEIRS. 2004. Predictive thresholds for plague in Kazakhstan. Science 304: 736–738.

ESKEY, C. R., AND V. H. HAAS. 1940. Plague in the western part of the United States. US Public Health Service. Public Health Bulletin 254: 1–83.

GAGE, K. L., R. S. OSTFELD, AND J. G. OLSON. 1995. Nonviral vector-borne zoonoses associated with mammals in the United States. Journal of Mammalogy 76: 695–715.

HOLMES, B. E. 2003. Ecology and persistence of sylvatic plague in Phillips County, Montana. MS Thesis, University of Montana, Missoula, Montana, 72 pp.

LECHLEITNER, R. R., L. KARTMAN, M. I. GOLDENBERG, AND B. W. HUDSON. 1968. An epizootic of plague in Gunnison's prairie dogs (*Cynomys gunnisoni*) in south-central Colorado. Ecology 49: 734–743.

MATCHETT, M. R. 1999. Black-footed ferret recovery activities on the U. L. Bend and Charles M. Russell National Wildlife Refuges, Phillips County, Montana: 1998 annual report and 5-year summary. US Fish and Wildlife Service, Lewistown, Montana, 38 pp.

MENKENS, G. E. JR., AND S. H. ANDERSON. 1991. Population dynamics of white-tailed prairie dogs during an epizootic of sylvatic plague. Journal of Mammalogy 72: 328–331.

OLSEN, P. F. 1981. Sylvatic plague. *In* Infectious diseases of wild mammals, J. W. Davis, L. H. Karstad and D. O. Trainer (eds.). Iowa State University Press, Ames, Iowa, pp. 232–243.

RAYOR, L. S. 1985. Dynamics of a plague outbreak in Gunnison's prairie dog. Journal of Mammalogy 66: 194–196.

STEVENSON, H. L., Y. BAI, M. Y. KOSOY, J. A. MONTENIERI, J. L. LOWELL, M. C. CHU, AND K. L. GAGE. 2003. Detection of novel *Bartonella* strains and *Yersinia pestis* in prairie dogs and their fleas (Siphonaptera: Ceratophyllidae and Pulcidae) using multiplex polymerase chain reaction. Journal of Medical Entomology 40: 329–337.

Received for publication 26 January 2005.

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use