

# TUBERCULOSIS IN EUROPEAN BADGERS (MELES MELES) AND THE CONTROL OF INFECTION WITH BACILLE CALMETTE-GUÉRIN VACCINATION

Authors: Corner, L. A. L., Murphy, D., Costello, E., and Gormley, E.

Source: Journal of Wildlife Diseases, 45(4): 1042-1047

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-45.4.1042

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <u>www.bioone.org/terms-of-use</u>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

# TUBERCULOSIS IN EUROPEAN BADGERS (MELES MELES) AND THE CONTROL OF INFECTION WITH BACILLE CALMETTE-GUÉRIN VACCINATION

#### L. A. L. Corner,<sup>1,3</sup> D. Murphy,<sup>1</sup> E. Costello,<sup>2</sup> and E. Gormley<sup>1</sup>

<sup>1</sup> School of Agriculture, Food Science and Veterinary Medicine, University College, Stillorgan Road, Belfield, Dublin 4, Ireland <sup>2</sup> Central Veterinary Research Laboratories, Department of Agriculture, Fisheries and Food, Backweston, Staccumy

Lane, Celbridge, Co. Kildare, Ireland <sup>3</sup> Corresponding author (email: leigh.corner@ucd.ie)

ABSTRACT: The eradication of tuberculosis (Mycobacterium bovis infection) from cattle herds may be compromised if infected wildlife species, such as European badgers (Meles meles), share the same environment and contribute to transfer of infection. Options for dealing with tuberculosis in this wild reservoir host are limited by conservation and social concerns, despite a clear implication that infected badgers are involved with the initiation of tuberculosis in cattle herds. Vaccination of badgers against  $\breve{M}$ . bovis, if successfully employed, would directly facilitate the completion of bovine tuberculosis eradication in affected areas. Vaccine trials in captive badgers have established that the M. bovis bacille Calmette-Guérin (BCG) vaccine can induce a protective response that limits the distribution and severity of tuberculosis disease following experimental challenge. The protective effect of the vaccine has been demonstrated when the vaccine was delivered by subcutaneous injection, deposited on mucous membranes, and given orally in a lipid formulation.  ${
m \dot{A}}$ large-scale field trial of oral BCG vaccine has been designed to measure the protection generated in wild badgers subjected to natural transmission of infection and to estimate vaccine efficacy. These parameters will be estimated by comparing the prevalence of M. bovis infection in vaccinated and nonvaccinated badgers. The results will provide a framework for the development and implementation of a national strategy to eliminate the disease in badger populations and if successful will remove this major impediment to bovine tuberculosis eradication.

Key words: Badger, Meles meles, tuberculosis, Mycobacterium bovis, BCG vaccine.

#### INTRODUCTION

Mycobacterium bovis is the causative agent of bovine tuberculosis in livestock and in a wide range of wild animals. Where wildlife species are infected with M. bovis they may act as a source of infection for domestic and wild animals (Corner, 2006). Tuberculosis in domestic livestock causes economic losses both directly from lost production and from the costs associated with eradication programs, and indirectly through the risk of zoonotic infection. Mycobacterium bovis infection is endemic in a number of wild animal species: brushtail possum (Trichosurus vulpecula) in New Zealand, white-tailed deer (Odocoileus virginianus) in North America, and European badger (Meles meles) in Ireland (Gormley and Collins, 2000) and Great Britain (Clifton-Hadley et al., 1993). Tuberculosis in the Irish cattle population has remained at a stable level over many years despite the application of testing programs and disease eradication procedures that have proven successful in the control of bovine tuberculosis in other countries.

That *M. bovis* infection in the badger population constitutes a significant reservoir of infection for cattle has been demonstrated in Ireland via the East Offaly (O'Mairtin et al., 1998a, b) and Four Area (Griffin et al., 2005) projects, and also in the UK (Woodroffe et al., 2005). Based on these studies, the Irish government has instigated a strategy of focused (reactive) culling of infected badgers to lower the density of infection in the badger population. This serves to limit the opportunity for transmission of infection to cattle and to decrease the economic impact of the disease in cattle. The decision to cull is made following an epidemiologic investigation of the possible causes of a breakdown in a cattle herd. If

badgers are identified as the probable source of infection, then the badgers in the immediate area (within 1 km of the farm) are culled. This is an interim strategy while research on alternative control strategies, including vaccination of badgers, is undertaken. Vaccination is an attractive control option for badgers because, if shown to be effective, it would reduce the burden of infection in this species and break the transmission cycle to cattle without provoking conservation and social concerns.

Many studies have shown that tuberculosis in badgers, as in most animal species, is primarily a respiratory disease involving the lung and associated thoracic lymph nodes (Gallagher et al., 1976; Gallagher and Clifton-Hadley, 2000; Gavier-Widen et al., 2001). Transmission between badgers is principally by inhalation of infectious aerosols (Nolan and Wilesmith, 1994). Following inhalation of M. bovis, infection becomes established in the lung and, by hematogenous dissemination, spreads to infect distal lymph nodes and visceral organs (Gallagher et al., 1998; Gallagher and Clifton-Hadley, 2000). In infected populations, less than 5% of infected badgers have generalized disease (Dolan, 1993; Gallagher, 1998; O'Boyle, 2003). In the majority of cases examined, infected badgers have no visible lesions (Clifton-Hadley et al., 1993; Corner, unpubl. data). This suggests that infection in the badger progresses slowly, or that it is contained and persists in a latent state. Tuberculosis does not appear to have a significant effect on the size or structure of badger populations (Wilkinson et al., 2000), and infected badgers may survive for several years (Little et al., 1982; Clifton-Hadley et al., 1993).

### VACCINATION AGAINST TUBERCULOSIS

The bacille Calmette-Guérin (BCG) vaccine (a live attenuated strain of M. *bovis*) is widely used in humans and studies have also shown its potential to limit the

spread of disease in wild animals. Two studies in wild brushtail possums in New Zealand showed that the BCG vaccine was protective. In the first study, vaccine efficacy was estimated to be 69% (Corner et al., 2002) and in the second, the estimated efficacy was 95–96% (Tompkins et al., 2009). These studies demonstrated that the BCG vaccine was more effective in wild possums than the results in captive studies had suggested. In the captive studies, as a consequence of the infection procedure used, all challenged possums developed lesions whereas in the wild possum studies vaccination prevented possums from becoming infected through natural transmission (Corner et al., 2002).

A cornerstone of developing a vaccine for tuberculosis in badgers is understanding the pathogenesis of the disease and how the infection progresses after an animal is infected. Experimental infections have been used to study pathogenesis in a variety of species and to test the protective efficacy of vaccines (Aldwell et al., 1995; Griffin et al., 1999; Hewinson et al., 2003; Buddle et al., 2005). In vaccine and challenge studies the objective of the experimental infections is to generate tuberculosis in the lungs in a uniform manner while maintaining a profile of lesion development and distribution that is consistent with natural *M. bovis* infection. We have shown that experimental endobronchial inoculation of badgers mimics natural disease (Corner et al., 2007, 2008a). Doses from <10 colony-forming units (cfu) to  $>10^3$  cfu lead to the establishment of infection and clearly demonstrated that badgers are very susceptible to M. bovis inoculated by this route, with infection consistently established with even with the lowest dose. The highest dose produced a uniform level of pathology when the response to inoculation was assessed over 24 wk.

In 1994, a joint consultative group of the World Health Organization, the Food and Agriculture Organization of the United Nations, and the Office International des Épizooties recommended that BCG Pasteur strain 1173P2 be used in animal studies (World Health Organization, 1994). This strain has been used successfully in studies in domestic cattle (Buddle et al., 1995), red deer (*Cervus elaphus*) (Griffin et al., 1999), ferrets (*Mustela furo*) (Qureshi et al., 1999), and brushtail possums (Aldwell et al., 1995). The BCG is an appropriate choice for vaccine studies in badgers because it has a long history of safe use in humans and animals (Murphy et al., 2008).

If effective in badgers, vaccination could form an alternative strategy for controlling M. bovis infection in badger populations (Gormley and Collins, 2000). The purpose of vaccination would be to decrease the burden of infection in the population and thereby decrease the risk to cattle. It has been shown that vaccination of badgers with BCG is safe when administered by the intramuscular and subcutaneous route (Lesellier et al., 2006), and can induce a protective response when administered intradermally (Stuart et al., 1988).

# BACILLE CALMETTE-GUÉRIN VACCINATION IN CAPTIVE BADGERS

The badger vaccine research project in Ireland commenced in 2001. Studies were undertaken to determine if the BCG vaccine could induce a protective immune response in badgers against an endobronchial challenge with a virulent strain of M. bovis. The responses of vaccinated and control badgers to challenge were assessed by severity of disease and infection. In these studies, using BCG Pasteur, badgers were vaccinated by subcutaneous injection, by spraying the vaccine into the nasal cavity and instilling a drop of vaccine suspension into the conjunctival sac (mucosal vaccination) (Corner et al., 2008b), or by giving the BCG orally in a lipid formulation (Aldwell et al., 2003) to prevent degradation in the stomach. At 3 mo postchallenge, the badgers were examined postmortem to assess the pathologic and bacteriologic responses to challenge. Gross and histologic lesions of tuberculosis were observed and M. bovis was recovered from all challenged badgers. In both studies, infection in the vaccinated badgers was less severe than in the control group, demonstrating that BCG vaccine induced a significant protective effect. A follow-up study was conducted to compare BCG Pasteur and BCG Danish strains when delivered as lipid-formulated oral vaccines, because BCG Danish is the only vaccine strain currently registered for human use in the European Union. Following virulent challenge, both vaccines generated protective immunity that was significantly different from that in the nonvaccinated controls. However there was no difference in the levels of protection achieved by either strain.

# CONSIDERATIONS IN BCG VACCINE FIELD TRIAL DESIGN

Whereas captive badger studies are the most cost effective way of examining the protective response to vaccination, such studies cannot be used to predict whether BCG will be protective in free-ranging badgers or to estimate vaccine efficacy. Estimates of vaccine efficacy are extremely valuable in modelling potential vaccine strategies, but data from field trials are needed to reliably estimate protection and vaccine efficacy parameters. Any field trial will by necessity use an oral vaccine delivery system, because this is the likely method of choice for any broad-scale mass vaccination of free-ranging badger populations.

A BCG vaccine field trial likely will have two principal objectives. These are to validate the results of captive badger studies and show that BCG vaccine is protective in naturally exposed wild badgers, and to estimate vaccine efficacy under field conditions. These objectives will be met by comparing the prevalence of *M. bovis* infection in vaccinated badgers with that in nonvaccinated controls. A secondary outcome of field trials will be to measure the effect of BCG vaccine in badgers with pre-existing M. bovis infection. There is some evidence from laboratory animals that vaccination may exacerbate pre-existing infection (Moreira et al., 2002). However, the effects reported were observed in inbred mice and were marginal with a single dose of vaccine (but more pronounced after repeated vaccination). In addition to providing a measurement of protection and an estimate of vaccine efficacy, field trials will provide a practical basis for understanding the logistics of oral vaccine delivery to wild badger populations.

Three different trial designs have been considered. The first design was to compare disease prevalence in an area where 100% of the population was vaccinated with a matched control area. However, this design was regarded as difficult to implement because it would require replication with the inherent problems of matching study sites. The second candidate design was to compare disease prevalence in vaccinated and control badgers within one area. In this design, 50% of the badgers in the trial area would be vaccinated and the nonvaccinated animals would constitute the control group. Although this design adopts a more pragmatic approach and takes into account geographic clustering, it would only measure the vaccine efficacy in badgers as individuals and not at a population level. In the third candidate design, different proportions of a badger population would be vaccinated, for example, 0, 50, and 100%. The advantage of this latter design is that effects on vaccine efficacy arising from changes in the force of infection as a result of vaccination could be estimated. Consequently, this third design has been chosen for planned field studies. The required proportion of vaccinates will be achieved by systematically trapping the area. When first encountered, indivdual

badgers will be allocated to either the vaccination or control group as required for the particular area. To allow for continued exposure to infection, the trial will be conducted over a 3-yr period. It is estimated that an initial population of 300 badgers (100 in each of the treatment areas) will be required to accurately estimate vaccine efficacy, based on an assumed initial tuberculosis prevalence of 20–30% and vaccine efficacy of 50–70%. for an individual badger.

The BCG Danish strain, encapsulated in a lipid formulation for oral administration and containing about  $10^8$  cfu/ml, will be used in the planned field trial. During the trial, badgers will be individually vaccinated by administration of the lipid vaccine or lipid placebo directly into the pharynx. Vaccine and placebo control samples will be "double-blind" coded. Badgers will be revaccinated annually and the population will be examined three times per year by trapping the entire study site in a continuous process. Throughout the trial, estimates of changing tuberculosis incidence will be made from the measurements of humoral and cellular immune responses.

At the end of the 3-yr study period, each site will be depopulated and all badgers will be examined for tuberculosis by detailed postmortem examination that will include an examination for visible lesions, histologic lesions, and mycobacteriology to demonstrate infection with *M. bovis*. The isolation of *M. bovis* from postmortem or clinical samples (wound exudates or tracheal swabs) will be used to define a case of tuberculosis.

### INCORPORATING BCG VACCINE INTO A NATIONAL CONTROL PROGRAM

Prior to implementing a wildlife vaccination strategy at a national level there would be many hurdles to overcome. Among these are issues related to licensing of the vaccine for use in badgers and development of efficient bait delivery systems to achieve high coverage in targeted populations. The environmental impact and effects of the vaccine on nontarget species also will need to be considered. In addition, the vaccination program will need to be carried out against the background of exhaustive investigation of tuberculosis in cattle, animal husbandry methods, and herd management-related factors that may affect cattle-to-cattle and badger-to-cattle transmission. Following vaccination, the risk to cattle posed by infected badgers would be expected to decrease as tuberculosis prevalence in badgers declines. By removing the influence of the reservoir host, an effective badger vaccination program could improve efficiency of the tuberculin testing program for controlling cattle-to-cattle spread and address a major impediment to the eradication of tuberculosis in Ireland and elsewhere.

#### ACKNOWLEDGMENTS

The badger vaccine project is supported through funds and staff of the Department of Agriculture, Fisheries and Food.

#### LITERATURE CITED

- ALDWELL, F. E., D. L. KEEN, V. C. STENT, A. THOMSON, G. F. YATES, G. W. DE LISLE, AND B. M. BUDDLE. 1995. Route of BCG administration in possums affects protection against bovine tuberculosis. New Zealand Veterinary Journal 43: 356–359.
  - —, —, N. A. PARLANE, M. A. SKINNER, G. W. DE LISLE, AND B. M. BUDDLE. 2003. Oral vaccination with *Mycobacterium bovis* BCG in a lipid formulation induces resistance to pulmonary tuberculosis in brushtail possums. Vaccine 22: 70–76.
- BUDDLE, B. M., G. W. DE LISLE, A. PFEFFER, AND F. E. ALDWELL. 1995. Immunological responses and protection against *Mycobacterium bovis* in calves vaccinated with a low dose of BCG. Vaccine 13: 1123–1130.
  - —, F. E. ALDWELL, M. A. SKINNER, G. W. DE LISLE, M. DENIS, H.-M. VORDERMEIER, R. G. HEWINSON, AND D. N. WEDLOCK. 2005. Effect of oral vaccination of cattle with lipid-formulated BCG on immune responses and protection against bovine tuberculosis. Vaccine 23: 3581–3589.
- CLIFTON-HADLEY, R. S., J. W. WILESMITH, AND F. A. STUART. 1993. Mycobacterium bovis in the

European badger (*Meles meles*): Epidemiological findings in tuberculous badgers from a naturally infected population. Epidemiology and Infection 111: 9–19.

- CORNER, L. A. L. 2006. The role of wild animal populations in the epidemiology of tuberculosis in domestic animals: How to assess risk. Veterinary Microbiology 112: 303–312.
  - —, S. NORTON, B. M. BUDDLE, AND R. S. MORRIS. 2002. The efficacy of bacille Calmette-Guérin vaccine in wild brushtail possums (*Trichosurus vulpecula*). Research in Veterinary Science 73: 145–152.
    - —, E. COSTELLO, S. LESELLIER, D. O'MEARA, D. P. SLEEMAN, AND E. GORMLEY. 2007. Experimental tuberculosis in the European badger (*Meles meles*) after endobronchial inoculation of Mycobacterium bovis: I. Pathology and bacteriology. Research in Veterinary Science 83: 53–62.
    - , \_\_\_\_, \_\_\_\_, \_\_\_\_, AND E. GORMLEY. 2008a. Experimental tuberculosis in the European badger (*Meles meles*) after endobronchial inoculation with *Mycobacterium bovis*: II. Progression of infection. Research in Veterinary Science (in press).
  - 2008b. Vaccination of European badgers (*Meles meles*) with BCG by the subcutaneous and mucosal routes induces protective immunity against endobronchial challenge with *Mycobacterium bovis*. Tuberculosis (in press).
- DOLAN, L. A. 1993. Badgers and bovine tuberculosis in Ireland: A review. In The badger, T. J. Hayden (ed.). Royal Irish Academy, Dublin, Ireland, pp. 108–116.
- GALLACHER, J. 1998. The natural history of spontaneous tuberculosis in wild badgers. PhD Thesis, University of London, London, UK, 286 pp.
- ——, AND R. S. CLIFTON-HADLEY. 2000. Tuberculosis in badgers; a review of the disease and its significance for other animals. Research in Veterinary Science 69: 203–217.
- —, R. H. MUIRHEAD, AND K. J. BURN. 1976. Tuberculosis in wild badgers (*Meles meles*) in Gloucestershire: Pathology. The Veterinary Record 98: 9–14.
- ——, R. MONIES, M. GAVIER-WIDEN, AND B. RULE. 1998. Role of infected, non-diseased badgers in the pathogenesis of tuberculosis in the badger. The Veterinary Record 142: 710–714.
- GAVIER-WIDEN, D., M. A. CHAMBERS, N. PALMER, D. G. NEWELL, AND R. G. HEWINSON. 2001. Pathology of natural *Mycobacterium bovis* infection in European badgers (*Meles meles*) and its relationship with bacterial excretion. Veterinary Record 148: 299–304.
- GORMLEY, E., AND J. D. COLLINS. 2000. The development of wildlife control strategies for eradication of tuberculosis in cattle in Ireland. Tubercle and Lung Disease 80: 229–236.

- GRIFFIN, J. F. T., C. G. MACKINTOSH, L. SLOBBE, A. J. THOMSON, AND G. S. BUCHAN. 1999. Vaccine protocols to optimise the protective efficacy of BCG. Tubercle and Lung Disease 79: 135– 143.
- GRIFFIN, J. M., D. H. WILLIAMS, G. E. KELLY, T. A. CLEGG, I. O'BOYLE, J. D. COLLINS, AND S. J. MORE. 2005. The impact of badger removal on the control of tuberculosis in cattle herds in Ireland. Preventive Veterinary Medicine 67: 237–266.
- HEWINSON, R. G., H. M. VORDERMEIER, AND B. M. BUDDLE. 2003. Use of the bovine model of tuberculosis for the development of improved vaccines and diagnostics. Tuberculosis 83: 119– 130.
- LESELLIER, S., S. PALMER, D. J. DALLEY, D. DAVE, L. JOHNSON, R. G. HEWINSON, AND M. A. CHAMBERS. 2006. The safety and immunogenicity of Bacillus Calmette-Guérin (BCG) vaccine in European badgers (*Meles meles*). Veterinary Immunology and Immunopathology 112: 24–37.
- LITTLE, T. W. A., P. F. NAYLOR, AND J. W. WILESMITH. 1982. Laboratory study of *Mycobacterium bovis* infection in badgers and calves. The Veterinary Record 111: 550–557.
- MOREIRA, A. L., L. TSENOVA, M. H. AMAN, L. G. BEKKER, S. FREEMAN, B. MANGALISO, U. SCHRO-DER, J. JAGIRDAR, W. N. ROM, M. G. TOVEY, V. H. FREEMAN, AND G. KAPLAN. 2002. Mycobacterial antigens exacerbate disease manifestations in *Mycobacterium tuberculosis*—infected mice. Infection and Immunity 70: 2100–2107.
- MURPHY, D., L. A. L. CORNER, AND E. GORMLEY. 2008. Adverse reactions to *Mycobacterium bovis* bacille Calmette-Guérin (BCG) vaccination against tuberculosis in humans, veterinary animals and wildlife species. Tuberculosis 88: 344–357.
- NOLAN, A., AND J. W. WILESMITH. 1994. Tuberculosis in badgers (*Meles meles*). Veterinary Microbiology 40: 179–191.
- O'BOYLE, I., E. COSTELLO, E. P. POWER, P. F. KELLEHER, J. BRADLEY, E. REDAHAN, F. QUIGLEY, U. FOGARTY, AND I. HIGGINS. 2003. Review of badger (*Meles meles*) research licenses in 2002. *In Selected papers 2002–2003. Veterinary Epidemiology and Tuberculosis Investigation Unit, University College Dublin, Dublin, Ireland, pp. 13–18.*

- O'MAIRTIN, D., D. H. WILLIAMS, L. DOLAN, J. A. EVES, AND J. D. COLLINS. 1998a. The influence of selected herd factors and a badger-intervention tuberculosis-control programme on the risk of a herd-level trade restriction to a bovine population in Ireland. Preventive Veterinary Medicine 34: 79–90.
  - —, —, J. M. GRIFFIN, L. A. DOLAN, AND J. A. EVES. 1998b. The effect of a badger removal programme on the incidence of tuberculosis in an Irish cattle population. Preventive Veterinary Medicine 34: 47–56.
- QURESHI, T., R. E. LABES, M. L. CROSS, J. F. T. GRIFFIN, AND C. G. MACKINTOSH. 1999. Partial protection against oral challenge with *Mycobacterium bovis* in ferrets (*Mustela furo*) following oral vaccination with BCG. International Journal of Tuberculosis and Lung Disease 3: 1025–1033.
- STUART, F. A., K. H. MAHMOOD, J. L. STANFORD, AND D. G. PRITCHARD. 1988. Development of diagnostic tests, and vaccination against, tuberculosis in badgers. Mammal Review 18: 74–75.
- TOMPKINS, D. M., D. S. L. RAMSEY, M. L. CROSS, F. E. ALDWELL, G. W. DE LISLE, AND B. M. BUDDLE. 2009. Oral vaccination reduces the incidence of tuberculosis in free-living brushtail possums. Proceedings of the Royal Society Series B 276: 2987–2995.
- WILKINSON, D., G. C. SMITH, R. J. DELAHAY, L. M. ROGERS, C. L. CHEESEMAN, AND R. S. CLIFTON-HADLEY. 2000. The effects of bovine tuberculosis (*Mycobacterium bovis*) on the mortality in a badger (*Meles meles*) population in England. Journal of Zoology, London 250: 389–395.
- WOODROFFE, R., C. A. DONNELLY, W. T. JOHNSTON, F. J. BOURNE, C. L. CHEESEMAN, R. S. CLIFTON-HADLEY, D. R. COX, G. GETTINBY, R. G. HEWINSON, A. M. LE FEVRE, J. P. MCINERNEY, AND W. I. MORRISON. 2005. Spatial association of *Mycobacterium bovis* infection in cattle and badgers *Meles meles*. Journal of Applied Ecology 42: 1–10.
- WORLD HEALTH ORGANIZATION. 1994. Report of WHO/FAO/OIE consultation on animal tuberculosis. World Health Organization, Veterinary Public Health Unit, Geneva, Switzerland WHO/ CDS/VPH/94.138.

Received for publication 6 June 2008.