

Phyllodes and bipinnate leaves of Acacia exhibit contemporary continental-scale environmental correlation and evolutionary transition-rate heterogeneity

Authors: Renner, Matt A. M., Foster, Charles S. P., Miller, Joseph T., and Murphy, Daniel J.

Source: Australian Systematic Botany, 34(6) : 595-608

Published By: CSIRO Publishing

URL: <https://doi.org/10.1071/SB21009>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Phyllodes and bipinnate leaves of *Acacia* exhibit contemporary continental-scale environmental correlation and evolutionary transition-rate heterogeneity

Matt A. M. Renner ^{A,E}, Charles S. P. Foster ^B, Joseph T. Miller ^C and Daniel J. Murphy ^D

^ANational Herbarium of New South Wales, Australian Institute of Botanical Science, Royal Botanic Gardens and Domain Trust, Mrs Macquaries Road, Sydney, NSW 2000, Australia.

^BSchool of Life and Environmental Sciences, University of Sydney, NSW 2006, Australia.

^CGlobal Biodiversity Information Facility, Universitetsparken 15, DK-2100 Copenhagen Ø, Denmark.

^DRoyal Botanic Gardens Victoria, Birdwood Avenue, Melbourne, Vic. 3004, Australia.

^ECorresponding author. Email: matt.renner@rbgsyd.nsw.gov.au

Abstract. In *Acacia*, 90% of species have drought-tolerant phyllodes as their adult foliage, the remaining species have bipinnate leaves. We conducted tests for relationships between foliage type and 35 bioclimatic variables at the continental scale and found significant correlations of both ‘moisture seasonality’ and ‘radiation in the coldest quarter’ with foliage type. Bipinnate species have lower species mean values of each variable, growing in stable soil moisture and generally darker environments (longer nights and lower incident radiation), on average. Evolutionary transformations between bipinnate and phyllodinous adult foliage exhibit asymmetry across the *Acacia* phylogeny, with transformations from bipinnate leaves to phyllodes occurring times faster than the reverse. At least three (and up to seven) transitions from phyllode to bipinnate adult foliage were inferred. Foliage type in the most recent common ancestor of extant *Acacia* is unresolved, some analyses favour a phyllodinous ancestor, others a bipinnate ancestor. Most ancestral nodes inferred as having bipinnate adult foliage had median age estimates of less than 5 million years (Ma), half having ages between 3 and 1.5 Ma. *Acacia* lineages with bipinnate adult foliage diversified during the Pliocene, perhaps in response to wetter climatic conditions experienced by the continental margin during this period.

Received 6 April 2021, accepted 28 October 2021, published online 3 December 2021

Introduction

Sclerophylly is hypothesised to be a response to nutrient-deficient soils, especially to those low in phosphorus (Loveless 1961; Beadle 1966; Eckstein *et al.* 1999). Sclerophyllous leaves are tougher and less prone to physical damage, both of which increase leaf longevity and so reduce nutrient loss by leaf-fall (Read and Sanson 2003). Sclerophyllous traits may be exaptations for drought tolerance (Boughton 1986; Hill and Brodribb 2001), and may have been further modified to enhance drought resistance and xeromorphy (Crisp and Cook 2013). Phyllodes are a sclerophyllous leaf type, involving the elaboration of petiolar tissue, and suppression of leaf blade expression (Gardner *et al.* 2005; Leroy and Heuret 2008).

Acacia Mill. (Leguminosae) is the only phyllode-bearing mimosoid legume genus in Australia, with more than 90% of the 1015 described *Acacia* species having phyllodes as their major adult foliage expression; the remaining species have

bipinnate adult leaves (Brown *et al.* 2011). With three known exceptions (*Acacia diaphylloidea* Maslin, *A. confusa* Merr. and *A. glutinosissima* Maiden & Blakely, Murphy *et al.* (2010)), all phyllodinous *Acacia* species are heteroblastic, that is, they experience a phase change from bipinnate juvenile to phyllodinous adult foliage as they grow (Wang *et al.* 2011). The ontogeny of seedling development in heteroblastic *Acacia* includes at least one bipinnate seedling leaf (Murphy *et al.* 2010; Brown *et al.* 2012), although the timing of the phase change can be flexible (Rose *et al.* 2019). In contrast, none of the *Acacia* species with bipinnate adult foliage experiences a heteroblastic phase change.

Acacia phyllode morphologies exhibit considerable variation (Fig. 1) and lability (Gardner *et al.* 2005), with the broadly circumscribed character states of uninerved versus multinerved phyllodes having multiple origins within *Acacia* (Ariati *et al.* 2006). Bipinnate leaves are thought to be the ancestral foliage type in the *Acacia* clade (Leroy and

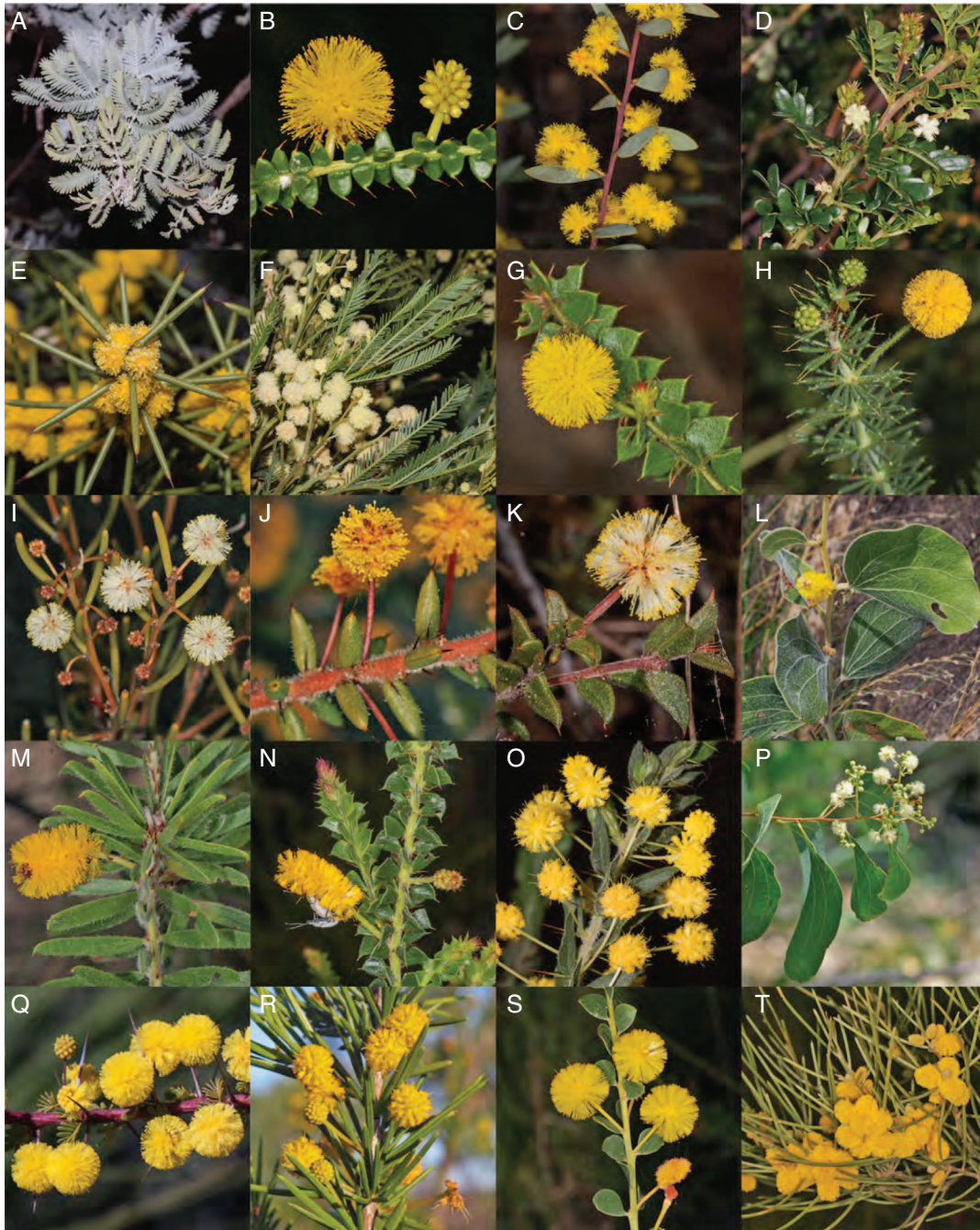


Fig. 1. A selection of *Acacia* species, showing variation in shoot, flower, phyllode and leaf morphology, not to scale. A. *A. baileyana*. B. *A. barretiorum*. C. *A. buxifolia*. D. *A. Cape le Grande*. E. *A. colletioides*. F. *A. dealbata*. G. *A. deltoidea*. H. *A. dimorpha*. I. *A. sp. indet.* J. *A. Grass Patch*. K. *A. gunnii*. L. *A. humifusa*. M. *A. kelleri*. N. *A. obtriangularis*. O. *A. paradoxa*. P. *A. platycarpa*. Q. *A. pulchella*. R. *A. tetragonophylla*. S. *A. translucens*. T. *A. Wathroo*. Bipinnate adult foliage illustrated in A, D, F and Q, others phyllodinous. All photos by Russell Barrett, with permission.

Heuret 2008), but this has not been tested empirically. In a broader context, bipinnate leaves are likely to be ancestral within the Caesalpinioideae, as all lineages, except *Acacia*, have species with bipinnate adult foliage. Within *Acacia*,

multiple lineages express bipinnate adult foliage, and at least two independent phylogenetic reversion events are recognised (Murphy *et al.* 2000, 2003). Certainly, phyllodes are an apomorphy within the Caesalpinioideae, but an

outstanding question is whether the most recent common ancestor of extant *Acacia* spp. possessed bipinnate leaves or phyllodes.

Phyllodes have been hypothesised to represent a key morphological innovation facilitating the diversification of *Acacia* in water-limited environments (Boughton 1986; Brodribb and Hill 1993). Supporting this hypothesis, phyllodes are more drought tolerant, and have higher water-use efficiency than do leaves in conditions of water deficit (Brodribb and Hill 1993; Wright and Westoby 2002). In contrast, bipinnate leaves have higher maximum rates of carbon dioxide uptake per unit mass, and higher photosynthetic rates than do phyllodes per unit mass and per unit area. They are also likely to be more efficient at light capture, given their horizontal orientation and spread of photosynthetic tissue (Rose *et al.* 2019). Both leaves and phyllodes of *Acacia* have high water-use efficiency compared with other C3 plants (Brodribb and Hill 1993), but being capable of high water-use efficiency in times of water stress does not necessarily imply high water-use efficiency at other times. For example, in South Africa, the bipinnate *Acacia mearnsii* has higher water use than does woody and grassland vegetation in riparian sites where it is invasive (Dye *et al.* 2001). It is possible that this higher water use supports higher relative growth rates.

The physiological differences between leaves and phyllodes are significant, given Australia's dry climate (Brodribb and Field 2010; Crisp and Cook 2012). Physiological performance studies have demonstrated that phyllodes are better capable of responding to, and recovering from, severe water stress than are bipinnate leaves (Brodribb and Hill 1993). *Acacia* species with bipinnate adult foliage would, therefore, be expected to occur in environments that are wetter, and not subject to severe water stress, than are species with phyllodes.

Acacia can be subdivided into two or three macroevolutionary cohorts with different diversification rates (Nge *et al.* 2020; Renner *et al.* 2020), one of which comprises a lineage occurring in mesic south-eastern Australia (Renner *et al.* 2020). This south-eastern mesic lineage has a higher mean diversification rate than the other extant *Acacia* lineages, contains both phyllodinous and bipinnate species, and may provide additional insight into the contribution that foliage type has made to the radiation of *Acacia*. The different diversification dynamics and high representation of bipinnate species may mean that transition rates among phyllode and bipinnate adult foliage are different in the south-eastern mesic lineage from the rest of the *Acacia* lineage.

The main aim of this paper is to examine why, given a progressively drying Australian climate, *Acacia* lineages have evidently reverted to bipinnate adult foliage. We address this question in two steps. We use geospatial and climatic data to ask whether bipinnate *Acacia* species occur within a continental climatic context predicted by the physiological performance of their leaves. We confirm that bipinnate adult foliage represents a phylogenetic reversion, and ask how many phylogenetic transitions to bipinnate foliage have occurred in *Acacia*, and at what rate. We then synthesise the results from

character reconstruction and analysis of climate variables to derive a testable model that may explain these observational data, wherein winter growth provides a competitive advantage to bipinnate *Acacia* in mesic habitats.

Materials and methods

Phylogeny reconstruction

Our methods followed those of Renner *et al.* (2020). We used the dataset published by Mishler *et al.* (2014), comprising two nuclear (nrITS and ETS) and four chloroplast markers (*psbA-trnH*, *trnL-trnF*, *rpl32-trnL*, *matK*) from 510 species, for phylogenetic reconstruction (see supplementary table 1 in Mishler *et al.* (2014) at <https://doi.org/10.1038/ncomms5473>, for voucher specimen details and GenBank accession numbers). Owing to the geographic location of the research groups working on this phylogenetic dataset for *Acacia*, the sampling of species for this dataset was skewed towards the eastern coast of Australia. This means that some groups of *Acacia* are over-represented in the data, for example, section *Botrycephalae* for which 82% of its constituent species were sampled, in contrast to an overall sampling rate of 47% for all species of *Acacia* (Renner *et al.* 2020). However, this is not as straightforward to assess as these numbers indicate, because section *Botrycephalae* is not monophyletic, similar to other sections of *Acacia*, so it is difficult to translate a geographic bias into an estimation of phylogenetic bias, especially as the relationships of unsampled species can be difficult to estimate on the basis of morphological data. We excluded three *Acacia* species with three or more missing markers that were non-overlapping within the data matrix, leaving 503 *Acacia* species and two outgroup taxa. The alignment was checked, and some sequences were manually re-aligned.

Incongruence between molecular markers is an important consideration in phylogenetic tree reconstruction. We concatenated chloroplast markers into a single alignment since the chloroplast is a predominantly non-recombining organelle, and concatenated the two nuclear markers into a single alignment because of their flanking positions in eukaryotic ribosomal cistrons. Incongruence among markers was assessed by comparing scores of Akaike information criterion for small sample sizes (AICc), similar to the approach of Walker *et al.* (2018). The optimal substitution model and the optimal partitioning scheme for each dataset were selected with PartitionFinder (ver. 2.1.1, see <http://www.robertlanfear.com/partitionfinder/>; Lanfear *et al.* 2014, 2017), with GTR+G being selected for all partitions (see supplementary table 2 in Renner *et al.* 2020). Maximum-likelihood trees were estimated for separate nuclear and chloroplast alignments, and for an overall concatenated alignment (chloroplast + nuclear) by using IQtree (ver. 1.6.12, see <http://www.iqtree.org>; Nguyen *et al.* 2015), with each partition having a separate GTR+G model. The log-likelihood scores for trees from IQtree were used in AICc calculations. AICc scores suggested congruence between chloroplast and nuclear markers, and supported our decision to concatenate genetic markers (see supplementary table 1 in Renner *et al.* 2020). We then estimated a maximum-likelihood

tree using RAxML (ver. 8, see <https://cme.h-its.org/exelixis/web/software/raxml/>; Stamatakis 2014), for use as a starting tree in divergence-time estimation.

To estimate a time-calibrated phylogeny, we employed BEAST (ver. 2.3.2, see <https://www.beast2.org>; Bouckaert *et al.* 2014). The substitution models for each marker were unlinked, and among site-rate variation modelled using a gamma distribution with four rate categories. Rate variation among lineages was modelled using a lognormal relaxed clock. The clock model was linked across molecular markers. The third codon position within *matK* was allowed to scale to a rate different from that of the first and second codon positions. A birth–death speciation model was used to generate a prior on the distribution of branch lengths and node depths, and the monophyly of the outgroup (*Paraserianthes*) was enforced. Markov-chain Monte Carlo (MCMC) chains were run for 100 million generations, logging trees sampled every 10 000 generations, and parameters sampled every 5000. A single time calibration point was used, based on fossil pollen with distinctive pseudocolpi on their surface (Macphail and Hill 2001), recorded in the late Oligocene (23 Ma), following Miller *et al.* (2013) and Renner *et al.* (2020). The use of a lognormal prior can be justified against the fossil pollen record itself; although *Acaciapollenites* pollen is present from the late Eocene onward (37.2–33.9 Ma; Macphail and Hill 2001), only younger records of this fossil genus can be confidently assigned on the basis of the possession of distinctive pseudocolpi on the pollen surface, characteristic of modern *Acacia* (Miller *et al.* 2013). Bounding the prior probability distribution of the *Acacia* origin against the younger ages of the *Acaciapollenites* record is appropriate because it reflects the origin of pseudocolporate pollen between these bounds, although it is likely closer to the younger limit. Therefore, to realistically model this prior, we applied a lognormal distribution with an offset of 23 Ma, median of 27.2 and 95% of the prior density between 23 and 34 Ma, following Miller *et al.* (2013), to the *Acacia* crown node. Maximum clade credibility trees were calculated from both runs after excluding the first 25% of each run as burnin, with TreeAnnotator (ver. 1.8.4, see <https://beast.community/treeannotator>).

Character scoring and trait analysis

Adult foliage type was scored from literature or herbarium specimens, with species having either bipinnate leaves or phyllodes. Species whose adult foliage comprises a mix of phyllodes and bipinnate leaves (heterophylly), such as *A. rubida*, were scored as phyllodinous (Forster and Bonser 2009). We compared four models of trait evolution using BayesTraits (ver. 3.0, see <http://www.evolution.rdg.ac.uk/BayesTraitsV3.0.5/BayesTraitsV3.0.5.html>; Pagel *et al.* 2004), which described whether transition rate parameters were linked or free, and whether transition rate parameters across the phylogeny were homogeneous or heterogeneous. Several phylogenetic transitions to phyllodes are localised within the bipinnate lineage that is part of the south-eastern mesic lineage, which was inferred to have different diversification dynamics from the rest of *Acacia* (see ‘event

subtree 2’ in Renner *et al.* 2020). Therefore, we estimated transition rate parameters for that lineage separately from the rest of *Acacia* to test for heterogeneity in morphological transformation rates across *Acacia*. Under the heterogeneous models, the south-eastern mesic lineage was allowed its own transition rate matrix, separate from the rest of *Acacia*. Transition rate matrices were estimated using Bayesian inference with a multistate continuous-time Markov model of character evolution for discrete data implemented by BayesTraits. For each Bayesian analysis, we seeded runs with an exponential prior with a mean of 10, because we expected rate parameters to be relatively small, and scaled the tree to have a mean branch length of 0.1, so the estimates would not be too small (for an explanation of what is too small, see the BayesTraits manual; Meade and Pagel 2016). MCMC chains were run for 10 million generations, sampling every 1000 generation. To accommodate phylogenetic uncertainty, we integrated estimates across a sample of 1000 trees from the posterior probability distribution sampled by BEAST. A burnin of 10 000 generations was discarded from each run. We estimated log Bayes factors (Kass and Raftery 1995) from the marginal likelihood estimated by stepping-stone analysis in BEAST. For all models, analyses used 100 stepping stones and 10 000 replicates. Each analysis was run independently three times to confirm convergence and consistency, and parameter traces and diagnostics were estimated and inspected with Tracer (ver. 1.6, see <https://beast.community/tracer>; Drummond *et al.*, 2012). We also used the variable-rates model implemented by BayesTraits to query the tree for regions having higher or lower rates of transformation. The variable-rates model uses reversible jump MCMC to identify parts of the tree in which the rate of character transformation differs. Because this model takes a single tree as input, we used the MCC tree from the BEAST analysis. The variable-rates model was run for 10 million generations, with a burnin of 1 million generations and scaled branch lengths, and the analysis was replicated three times to confirm run convergence.

We reconstructed ancestral states using maximum likelihood and Bayesian inference methods. Likelihood-based ancestral-state reconstructions were completed using the *ace* function in the R package Ape (ver. 5.3, see <https://cran.r-project.org/web/packages/ape/index.html>; Paradis *et al.* 2004; Popescu *et al.* 2012; Paradis and Schliep 2019). These analyses also presented a visual summary of state evolution on the most credible clade tree. To address outgroup sampling, we manually created trees with 13 additional outgroup taxa following the topology (branching order and branch lengths) of the phylogeny presented by the Azani *et al.* (2017), which, although based only on *matK*, does give an indication that early diversification of the Caesalpinioideae was rapid, with lineages being separated by short branches. The ancestral-state reconstruction was repeated on this manually expanded outgroup tree. We also fixed the state at the *Acacia* crown node and compared marginal likelihoods and associated $2 \times \log$ Bayes factors ($2 \ln \text{BF}$) for models with phyllodes and leaves as the ancestral state under a homogeneous free model

of trait evolution, estimated by stepping-stone sampling of the prior, using 100 stepping stones and 10 000 replicates, with BayesTraits. Morphological data, the MCC tree, the sample of 1000 trees from the posterior probability distribution (PPD) and control file for the heterogeneous analysis are available in the Supplementary material (Tables S1–S4 of the Supplementary material) associated with this article.

Environmental variables

We used a set of 132 295 geo-referenced herbarium specimens of *Acacia* originally downloaded from the Australian Virtual Herbarium (see <https://www.ala.org.au>) and curated by González-Orozco *et al.* (2011), and later analysed by Mishler *et al.* (2014) and Renner *et al.* (2020). Thirty-five bioclimatic (BioClim) variables from the ANUCLIM (ver.6, β , see <https://fennerschool.anu.edu.au/research/products/anuclim>) layers, available through the Atlas of Living Australia's Spatial Portal (spatial.ala.gov.au), were downloaded for these ~132 000 voucher specimens. For each environmental variable, the species mean was calculated; the resulting means represent the realised climatic niche centroid for each species, and were used in the regression analyses. These species means are available in the supplementary material of Renner *et al.* (2020).

Correlation and covariation among environmental variables were assessed using the spectral decomposition method of principal-component analysis, accomplished using the *princomp* function of the stats package (ver. 4.0, R Foundation for Statistical Computing, see <https://stat.ethz.ch/R-manual/R-devel/library/stats/html/stats-package.html>) in R (ver. 3.5.6, R Foundation for Statistical Computing, see <http://www.R-project.org>). The variance–covariance matrix for Z-scaled data was calculated with the *cov* function. The kappa value suggested a high degree of multicollinearity within the environmental dataset, and variance inflation factors estimated by the VIF package (ver. 1.0, see <https://CRAN.R-project.org/package=VIF>; Lin *et al.* 2011) indicated the same result, and retained 28 variables. Because species with bipinnate adult foliage tend to be closely related to one another, we fitted a single multivariate phylogenetic binomial generalised linear model (Ives and Garland 2010) to examine the correlation between possession of bipinnate leaves or phyllodes and the 28 climate variables retained by VIF within a phylogenetic context, with the function *phyloglm* in package *phyloilm* (ver. 2.6, see <https://www.rdocumentation.org/packages/phyloilm/versions/2.6>, accessed 21 January 2020).

Results

Bipinnate leaf evolution

Reversions to bipinnate foliage have occurred at least three times, and in three different lineages within *Acacia* according to maximum-likelihood ancestral-state reconstruction (Fig. 2). Within the south-eastern mesothermic radiation (event Subtree 2), there have been four inferred transformations to and from bipinnate adult foliage. The $2 \times \log$ Bayes factors estimated by BayesTraits returned very strong support for the homogeneous free rates model as the best descriptor of state transformation in *Acacia* foliage (Table 1). Under the homogeneous free

model of trait evolution, the transition rates among phyllodinous and bipinnate adult foliage are asymmetrical (Table 2); estimated median transition rate of bipinnate leaves to phyllodes was two orders of magnitude higher than the transition rate for phyllodes to bipinnate leaves (Fig. 3). The variable-rate estimation using reversible jump MCMC recovered no evidence for significant rate heterogeneity across the phylogeny (results not shown), in accord with the inference that the homogeneous model provides a better fit with the databased on Bayes factor comparison.

Likelihood-based ancestral character reconstruction returned a high probability of the phyllodinous character state at the *Acacia* crown node, at all nodes along the *Acacia* backbone, and also at the MRCA of *Paraserianthes* plus *Pararchidendron* and *Acacia* (Fig. 2). Results from the phylogeny with an additional 13 outgroup taxa (and 13 nodes below the *Acacia* MRCA) had bipinnate adult foliage reconstructed along all nodes along the phylogeny backbone, and at the MRCA of *Paraserianthes* plus *Pararchidendron* and *Acacia*. The MRCA of *Acacia* was again reconstructed as phyllodinous, as in the original limited-outgroup dataset. The median age for the oldest node inferred to have been leafy is ~15 million years old, but this is an outlier; the median ages of most leafy nodes are less than 9 million years old and more than half are less than 3 million years old (Fig. 4).

Bayesian inference of ancestral states under a homogeneous free model returned positive evidence for bipinnate over phyllodinous adult leaves at the *Acacia* crown node, whereas the heterogeneous free model was indecisive (Table 3).

Environmental correlates

In all, 2 of the 28 bioclimatic variables were significantly correlated (after correction for phylogeny) with bipinnate or phyllodinous adult foliage, 'moisture seasonality' (Bio31) and 'radiation during the coldest quarter' (Bio27; Table 4), with bipinnate species means being clustered around lower values of both variables (Fig. 5). There was a trend towards 'moisture during the coldest quarter' (Bio35), 'moisture index during the highest period' (Bio29) and 'precipitation during the warmest period' (Bio18), correlating with the occurrence of phyllodinous and bipinnate foliage, albeit without passing our statistical significance threshold. In plots of the distribution of bipinnate and phyllodinous adult foliage along moisture variables, bipinnate species exhibited compression towards the lowest moisture seasonality values, representing those soil environments with the least difference between their wettest and driest moisture contents. This contrasts with the relatively wide distribution of bipinnate species on the axis of soil moisture values during the lowest quarter. Along the axis of radiation during the coldest quarter, bipinnate species clustered towards lower values (Fig. 5).

Discussion

Why phyllodes are so widespread across the Australian continent is a research question that has been addressed by

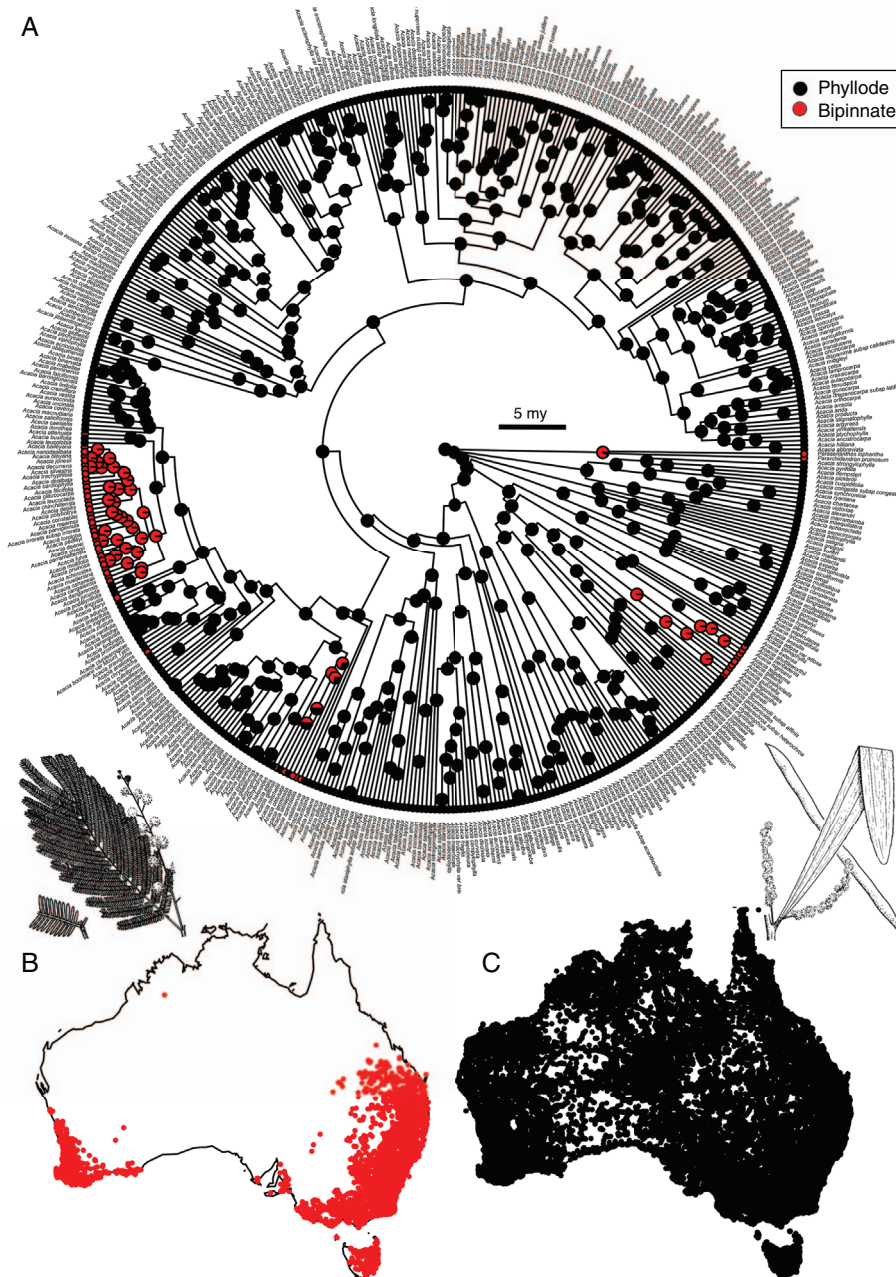


Fig. 2. A. Likelihood ancestral-state reconstruction for phyllodes and bipinnate adult foliage on the most credible clade time-tree from the time-calibration analysis; states show adult foliage. Red, bipinnate; black, phyllode. The time tree root-node median age is 26.82 Ma, the scale bar is 5 Ma. B. Distribution of bipinnate *Acacia* on the Australian continent. C. Distribution of phyllodinous *Acacia* in Australia. Records sourced from Australian Virtual Herbarium's Atlas of Living Australia.

many studies, including physiological studies demonstrating higher water use efficiency for phyllodes. Reversions to bipinnate adult foliage have occurred in three different *Acacia* lineages, which is surprising in the Australian context because Australia has become a dry continent (Martin 2006) subject to episodic severe water stress (Murphy and Timbal 2008), and phyllodes are more

resistant to drought than are bipinnate leaves. Why have some *Acacia* reverted to a more risky foliage type, given this continental climatic context? The answer *may* be found in the ecological setting wherein bipinnate species grow. With lower soil-moisture seasonality, and higher soil-moisture values generally, bipinnate *Acacia* grows in settings accessible to a range of other broadleaf and mesophyllous

Table 1. Bayes factor comparison of models of trait evolution, using 2ln Bayes factors

Positive values indicate support for the model in the column, negative values for the model in the row of the comparison. The model supported in all comparisons was the homogeneous free-parameter model, shown in bold

Model	log-Likelihood	Homogeneous linked	Homogeneous free	Heterogeneous linked	Heterogeneous free
log-Likelihood		-68.10	-64.38	-67.17	-73.61
Homogeneous linked	-68.10	0	7.45	1.85	-11.02
Homogeneous free	-64.38	-7.45	0	-5.59	-18.47
Heterogeneous linked	-67.17	-1.85	5.59	0	-12.88
Heterogeneous free	-73.61	11.02	18.47	12.88	0

Table 2. Parameters describing the rate of transformation among phyllodinous and bipinnate adult foliage in *Acacia*

ESS, effective sample size; s.e., standard error of the mean; s.d., standard deviation

Parameter	From bipinnate to phyllode	From phyllode to bipinnate
Mean	2.7571	0.0374
s.e.	0.00285	0.000143
s.d.	0.8312	0.0416
Variance	0.6909	0.00173
Median	2.8438	0.022
ESS	84763	84626

species, including rainforest pioneers. A ‘grow fast, die young’ strategy involving a winter growth period facilitated by high cold-quarter soil moisture may explain the relative restriction of bipinnate *Acacia* along that climatic axis. A seasonally dependent competitive growth advantage has not been posited as an explanation for the relative success of bipinnate *Acacia* in south-eastern Australia previously, but our results are consistent with this explanatory model, which we elaborate with reference to heteroblasty, character-state transition rates, physiological efficiencies, geographic occurrence, and environmental space occupancy in the following discussion.

Determinants and advantages of heteroblasty

Heteroblasty is a key innovation enabling sessile organisms to maximise performance in response to environmental change (Chitwood and Sinha 2016). Heteroblastic phase change can be developmentally programmed but environmentally induced (Ostria-Gallardo *et al.* 2016), and the timing of phase change can be plastic. Low light levels delay the phase change to phyllodinous leaves in *Acacia koa* and *A. implexa* (Forster *et al.* 2016; Rose *et al.* 2019). In *A. koa*, and other *Acacia*, bipinnately compound leaves contribute to rapid early seedling and sapling growth in forest gaps where light is, or will become, potentially limiting at low carbon cost (Brodrribb and Hill 1993; Craven *et al.* 2010; Pasquet-Kok *et al.* 2010). The existence of plasticity in the timing of phase changes means that different environments could act to hasten, delay or increase plasticity in the timing of phase transition (Pigliucci 1997). Differences in the timing of phase change are heritable, and both *A. koa* and *A. melanoxylon* exhibit population-level differences in the timing of phase

transition that are genetic (Farrell and Ashton 1978; Rose *et al.* 2019). In populations of *A. koa* exposed to regular drought, the phase change from bipinnate leaves to phyllodes occurs early in seedling growth, regardless of weather conditions experienced by seedlings, and so contributes to maximising survivorship in harsh, drought-prone sites (Rose *et al.* 2019).

MicroRNAs are evolutionarily conserved regulators of vegetative phase change, and play important roles in plant development (Lauter *et al.* 2005; Wu and Poethig 2006; Chuck *et al.* 2007; Wu *et al.* 2009; Cho *et al.* 2012). Wang *et al.* (2011) studied the transition from bipinnate to phyllodinous foliage during seedling ontogeny, and identified a likely causative signalling mechanism involving microRNA, wherein a reduction in microRNA expression correlates with a phase change to phyllode expression. The phase transition from bipinnate leaves to phyllodes in seedlings of *A. confusa* and *A. colei* is governed by opposing expression patterns of the microRNAs *miR156* and *miR172*, which regulate a conserved framework of phase changes in many, if not all, angiosperms (Wang *et al.* 2011). *miR156* represses the expression of squamosa promoter binding-like (*SPL*) transcription factors, shows high correlation with juvenile-like vegetative leaf traits, and its expression decreases during the juvenile to adult transformation (Wang *et al.* 2008, 2011). It follows, then, that overexpression of *miR156* in transgenic *Populus* was shown to prolong a juvenile phase (Wang *et al.* 2011).

A related observation in flowering plants, that of a prolonging of a juvenile phase in shade, is also consistent with a role for sugar-signalling in *miR156* expression, with high sugar levels signalling a down-regulation in the expression of *miR156* (Yang *et al.* 2013; Yu *et al.* 2013). Nutritional status, therefore, plays a demonstrable role in phase change in plants, as it does in animals (Chitwood and Sinha 2016). Wang *et al.* (2011) did not study microRNA expression in bipinnate *Acacia*; however, if *miR156* expression does govern the transition to phyllodes, then simply fixing the overexpression in *Acacia* seedlings could prolong indefinitely the bipinnate phase of ontogeny. This possibility is also consistent with fixed overexpression of *miR156* in transgenic *Populus × canadensis*, which results in a prolonged juvenile phase (Wang *et al.* 2011).

Fixed overexpression of *miR156* may alone explain leafy bipinnate adult foliage in bipinnate *Acacia*. However, whether patterns of *miR156* expression in other bipinnate species of Caesalpinioideae, such as, for example, *Paraserianthes* or

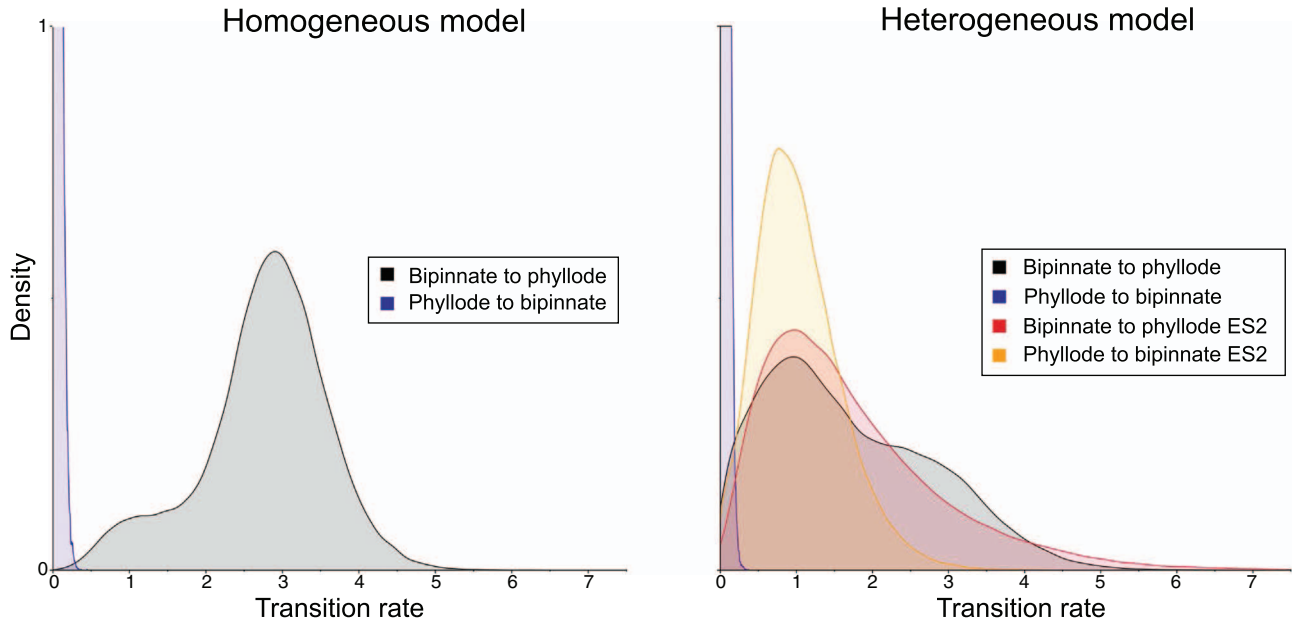


Fig. 3. Posterior probability distributions for transition rates among leaves and phyllodes, under homogeneous and heterogeneous models of trait evolution across the *Acacia* phylogeny. The homogeneous model was strongly favoured in a Bayes factor comparison.

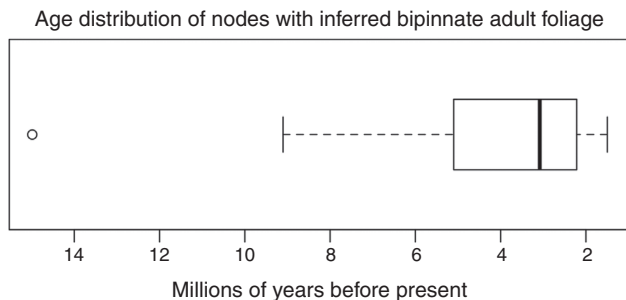


Fig. 4. Boxplot of age distribution for *Acacia* ancestral nodes inferred to have had bipinnate adult foliage.

Albizia, are the same as in *Acacia* is not yet known. Given the flexibility in evolutionary transitions among leafy and phyllodinous adults in the *Acacia* phylogeny, the regulation of *miR156* expression may have fundamentally changed in bipinnate species. Measuring *miR156* expression in bipinnate *Acacia* species would be a first step in testing the role of this micro-RNA in governing adult leaf morphology. If it is overexpressed throughout the life cycle of bipinnate species, then silencing it may allow the expression of ‘hidden’ phyllodinous phenotypes in normally bipinnate species.

Presumably, the underlying capacity for phyllode expression is maintained by species with compound leaves. *Acacia* species with bipinnate adult foliage may represent a special case where selection has acted to delay phase transitions entirely; however, the genetic and regulatory mechanism for this is, as yet, incompletely understood. The transition to bipinnate adult foliage is predominantly a relatively recent phenomenon, with the majority of leafy ancestral nodes belonging to the most diverse and

Table 3. Bayes factor comparison of different ancestral-state reconstructions at the *Acacia* crown node, under homogeneous and heterogeneous free transition-rate models

Model	log L		Favoured	Bayes factor
	Bipinnate	Phyllode		
Homogeneous	-64.507	-66.087	Bipinnate	3.16
Heterogeneous	-66.413	-66.984	Neither	1.14

predominantly south-eastern Australian leafy lineage (comprising the bulk of section *Botrycephalae*), having median node ages less than 5 million years, and half having median ages between 3 and 1.5 Ma. This radiation was possibly promoted by wetter climatic conditions during a humid paleoclimate interval between 5.5 and 3.3 Ma that was identified to have occurred over Australia’s north-west (Karatsolis *et al.* 2020), and by the continental margin, more generally (Miller *et al.* 2012; Christensen *et al.* 2017).

Advantages of non-heteroblasty

Given that heteroblasty confers multiple benefits across different developmental stages in many plants (Jaya *et al.* 2010; Zotz *et al.* 2011), including *Acacia* (Pasquet-Kok *et al.* 2010; Wang *et al.* 2011), a key question is under what circumstances might heteroblasty be advantageously dispensed with? Occurrence of bipinnate species is significantly correlated with low moisture seasonality and low sunlight levels during winter (Table 4, Fig. 5), implying that most bipinnate species grow on soils that not only exhibit low moisture seasonality, but are also wetter on average than many Australian soils in absolute terms. This is as

Table 4. Regression statistics returned by phylogenetically corrected binary logistic regression assessing the correlation between phyllodinous and bipinnate *Acacia* species, and species mean environmental parameters

Significantly correlated variables are in bold. s.e., standard error; CI, confidence interval

Parameter	Estimate	s.e.	z-value	95% CI lower	95% CI upper	P-value
(Intercept)	5.981	2.153	2.778	4.762	6.336	0.005
Temp. annual mean (Bio01)	-73.265	38.565	-1.900	-73.560	-72.888	0.057.
Temp. diurnal range mean (Bio02)	4.320	3.015	1.433	3.656	4.985	0.152
Temp. isothermality (Bio03)	-2.125	1.955	-1.087	-2.667	-1.421	0.277
Temp. seasonality (Bio04)	40.686	27.030	1.505	40.222	41.094	0.132
Temp. warmest period max (Bio05)	-12.007	7.655	-1.568	-12.316	-11.661	0.117
Temp. wettest quarter mean (Bio08)	6.480	3.635	1.783	5.998	7.418	0.075.
Temp. driest quarter mean (Bio09)	2.524	2.104	1.200	1.764	3.186	0.230
Temp. warmest quarter (Bio10)	-28.190	36.790	-0.766	-28.503	-27.738	0.444
Temp. coldest quarter mean (Bio11)	111.395	58.724	1.897	111.110	111.799	0.059.
Precipitation annual (Bio12)	2.184	5.133	0.425	1.833	2.516	0.671
Precipitation wettest period (Bio13)	-4.291	4.872	-0.881	-4.705	-3.972	0.378
Precipitation seasonality (Bio15)	2.050	3.171	0.647	1.587	2.793	0.518
Precipitation wettest quarter (Bio16)	-6.530	6.070	-1.076	-6.926	-6.198	0.282
Precipitation driest quarter (Bio17)	0.252	1.531	0.165	-0.208	1.166	0.869
Precipitation warmest quarter (Bio18)	8.339	4.329	1.926	7.779	8.929	0.054.
Precipitation coldest quarter (Bio19)	3.662	2.291	1.598	3.225	4.613	0.110
Radiation highest period (Bio21)	-1.284	1.680	-0.764	-2.053	-0.634	0.445
Radiation lowest period (Bio22)	5.470	6.601	0.829	4.983	5.975	0.407
Radiation seasonality (Bio23)	-5.304	4.356	-1.218	-5.897	-4.585	0.223
Radiation wettest quarter (Bio24)	0.518	1.532	0.338	-0.023	1.202	0.735
Radiation warmest quarter (Bio26)	3.167	2.538	1.248	2.621	3.779	0.212
Radiation coldest quarter (Bio27)	-13.127	6.584	-1.994	-13.581	-12.612	0.046 *
Moisture index annual mean (Bio28)	4.794	4.099	1.170	4.447	5.256	0.242
Moisture index highest period (Bio29)	-10.466	5.583	-1.875	-10.967	-9.957	0.061.
Moisture index lowest period (Bio30)	0.341	0.952	0.358	-0.413	1.152	0.720
Moisture index seasonality (Bio31)	9.744	4.064	2.398	9.386	10.389	0.016 *
Moisture index highest quarter mean (Bio32)	8.144	6.853	1.188	7.673	8.531	0.235
Moisture index coldest quarter mean (Bio35)	-9.484	4.967	-1.909	-9.926	-8.794	0.056

expected from physiological studies of bipinnate leaves and phyllodes (Brodribb and Hill 1993), with bipinnate leaves performing poorly when subject to severe water limitation. These physiological performance parameters correlate with continental-scale occurrence of bipinnate species within the totality of Australia's climate space, across all *Acacia* and after correction for phylogeny. The significant correlation with winter light levels is also interesting, and what we might expect given the photosynthetic efficiency of leaves. Bipinnate leaves are more efficient photosynthetic organs than are phyllodes in terms of their maximum photosynthetic rate per unit mass (Brodribb and Hill 1993), which may confer a significant advantage to seedling and sapling growth rates (Morris *et al.* 2011). The higher mass of phyllodes is due in part to their investment in water-delivery tissues; primary nerve densities correlate with hydraulic conductance, and mass per area and bundle sheath extensions. Higher nerve density decreases the distance water travels within the phyllode, and phyllodes with high mass per area are more common in arid environments (Sommerville *et al.* 2012). *Acacia* species with bipinnate leaves have faster growth rates than do phyllodinous species, at least in part owing to the lower resource investment into water-delivery and support tissues in leaves (Atkin *et al.* 1998).

Bipinnate leaves should also be more efficient at capturing photosynthetically active radiation in low-light environments, given their horizontal orientation and spread of photosynthetic tissue (Brodribb and Hill 1993). This is not to say that phyllodinous species do not also occur in areas of low coldest-quarter radiation; they do, and, indeed, have the lowest mean values for this incident variable, below the bipinnate species clustered in the lower half of this variable. However, several observations from experimental studies of growth, development and ecology attest to the light-capturing efficiency of bipinnate leaves. Seedlings of *Acacia implexa* produce more leafy nodes under lower light levels (Forster *et al.* 2011), as do seedlings of *Acacia koa* (Rose *et al.* 2019), responses that reflect the nutritional status of the seedlings. Studies of invasive populations of the bipinnate species *Acacia dealbata* have shown that this species has high photosynthetic performance under low light, and considerable photosynthetic plasticity, both of which contribute to the shade tolerance of this species and, in turn, its invasiveness under established canopies outside its native range (Aguilera *et al.* 2015). *Acacia dealbata* invades forests in Europe (Rodríguez *et al.* 2017), Chile and Argentina (Spalazzi *et al.* 2019).

Phyllodinous *Acacia* species also occur at sites of low winter radiation, but they are not also restricted to sites of

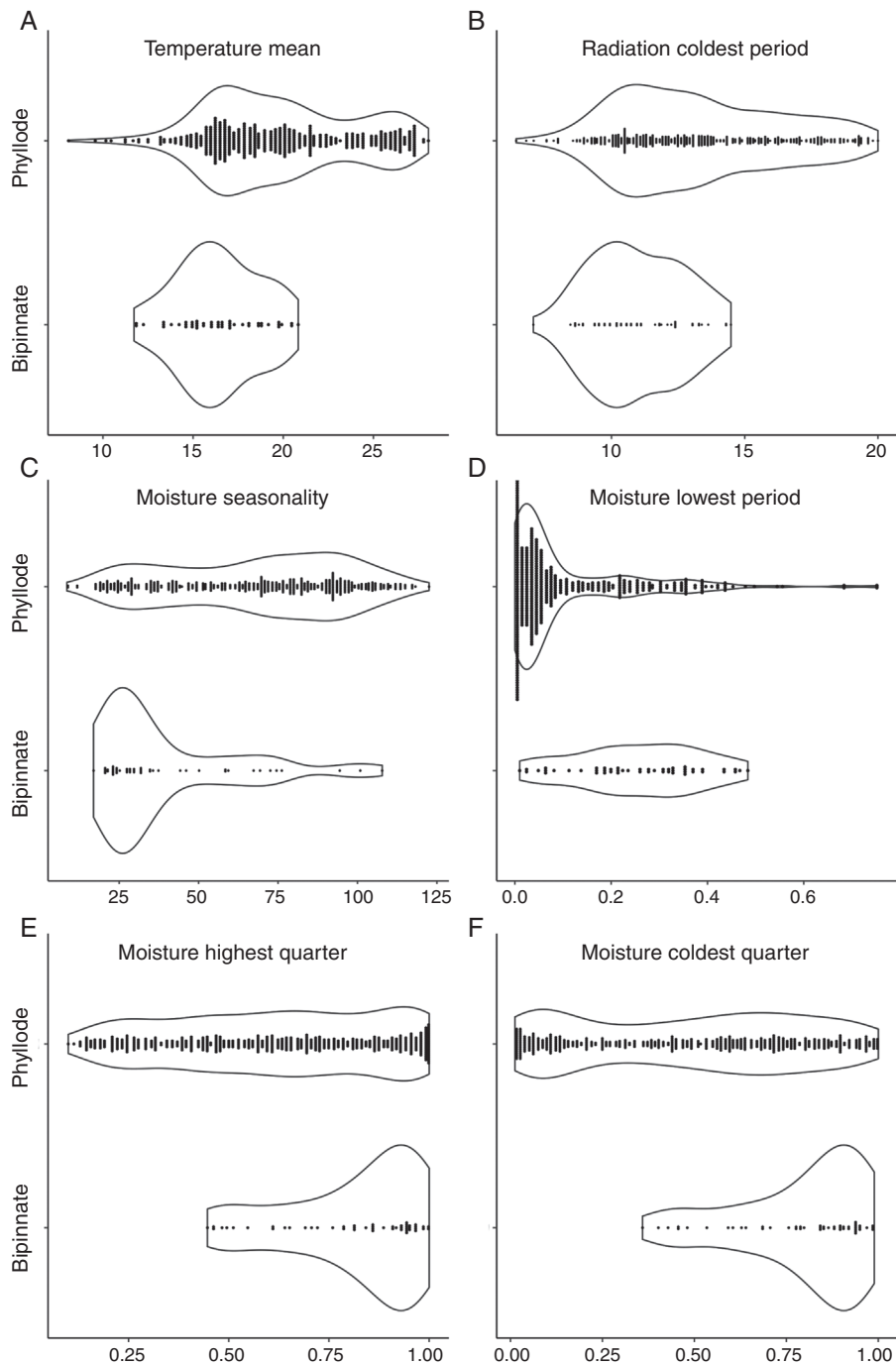


Fig. 5. Violin plots showing the distribution of phyllodinous (1) and bipinnate (0) species along a selection of climate variables. Significant correlations were returned for radiation in the coldest quarter and moisture seasonality. Note the contrasting distribution of bipinnate species along the moisture coldest-quarter, and moisture lowest- and warmest-quarter axes.

high coldest-quarter soil moisture. The occurrence of both phyllodes and bipinnate adult foliage in similar environmental space may represent a solution to simultaneous trade-offs along many environmental variables. For example, light and water availability impose challenges that interact within the Australian climate context.

Within the climatic envelope in which bipinnate species are found, they are compressed towards the high coldest-quarter soil-moisture values (Fig. 5, Bio35). This contrasts with their scattered distribution along the driest quarter soil-moisture axis, for which the correlation between bipinnate species mean values and lowest-quarter moisture index was *not* significant

(Fig. 5). Perhaps, surprisingly, this suggests that many bipinnate species have an innate capacity to tolerate drought, possibly by mechanisms of avoidance, as suggested by resource investment in root biomass under water stress (Forster *et al.* 2016; Rose *et al.* 2019). An innate drought tolerance ought not preclude the occupancy of lower coldest-quarter moisture environments by bipinnate *Acacia* species. Yet, on average, they do not occur in these lower coldest-quarter moisture environments. This restriction may have an explanation in biotic factors that affect how competitive bipinnate *Acacia* species are with other woody species. For example, a period of active winter growth during times of relatively low evapotranspiration stress may provide a competitive advantage for bipinnate *Acacia* over other woody species. Conversely, there are three bipinnate species among those sampled that do occur in environments with high seasonal variation in soil moisture. *Acacia epacantha*, *A. fagonioides* and *A. guinetti* are all spinescent shrubs from south-western Western Australia, with bipinnate leaves that are smaller in size than most other species of *Acacia*, comprising a single pair of pinnae, each with 2–5 pairs of pinnules. These three species are nested within a group of bipinnate species that are sister to most of the *Acacia* phylogeny, and were traditionally recognised in section *Pulchellae* (Benth.) Taub. Each of these three species deploys bipinnate foliage in quite different ecological contexts from others in the current study, and a comparative investigation of their performance response to water deficit and stress could be rewarding.

Macroevolution of foliage type in *Acacia*

Phyllodes and bipinnate adult foliage are ecologically important traits that are optimised to different climatic contexts (Renner *et al.* 2020). Physiological and life-history studies have previously demonstrated their optimisation within different life-history stages at the individual level, and these studies, along with our current results, suggest that the different performances of leaves and phyllodes have ecological consequences from the individual (as demonstrated by previous physiological studies) through to the macroecological and continental scale (as found here).

Within *Acacia*, bipinnate adult leaves have at least three separate origins within the genus. Multiple origins for bipinnate foliage were previously identified when the leafy section *Botrycephalae* (Benth.) Taub. was resolved as polyphyletic (Brown *et al.* 2006). Multiple reversals to bipinnate foliage have been inferred (Murphy *et al.* 2003), with neoteny being the likely developmental pathway for bipinnate adult leaves (Murphy *et al.* 2010). However, in the present study, it was newly discovered that transformations from phyllode to leafy adult foliage may have occurred in as few as three lineages within *Acacia*, and that each transformation to leaves is linked with many subsequent transformations back to phyllodes. This is consistent with Bayesian estimation of transition rates under the optimal model of trait evolution, in which the rate of transitions from phyllodinous to bipinnate adult foliage across the *Acacia* phylogeny was low.

Given the regulation of phyllode expression and transition from juvenile to adult foliage by miRNA156, it is likely that paedomorphosis describes each evolutionary transition to bipinnate adult foliage in *Acacia*; however, whether post-displacement or neoteny (*sensu* Alberch *et al.* 1979) is involved will not be resolved until the underlying gene-regulation pathway is better characterised. The existence of plasticity in the timing of phase-change initiation, pre- and post-displacement at the level of individual (see Pryer and Hearn 2009 for an explanation of terminology), suggests that evolutionary lability in phase change is possible as a result of fixation of plasticity in timing that exists within individuals. However, whether the same gene regulatory mechanism(s) are involved in each case of paedomorphosis in *Acacia* is currently unknown. We may hypothesise that it is not necessarily the same pathway for all bipinnate *Acacia*. The diversity of adult bipinnate leaf morphologies, phylogenetic distance and evolutionary time separating bipinnate lineages, and, in particular, the reduced leaves of species in section *Pulchellae* compared to those species in south-eastern Australia, suggest that more than one paedomorphic mode may well be involved.

Bipinnate adult foliage represents a derived character in extant *Acacia*, even though it is a phylogenetic transition to a symplesiomorphic state within the broader Caesalpinoideae. Bayesian estimation of ancestral states suggests that the most recent common ancestor of *Acacia* had phyllodinous adult foliage, and the transition to phyllodes preceded the radiation of *Acacia*. However, this reconstruction is based on the assumption that all phyllodes are homologous, and there is some evidence that suggests that they are not (Gardner *et al.* 2005). Studies of ontogeny, such as those on *A. mangium*, inform homology relations between phyllodes and bipinnate leaves (Leroy and Heuret 2008); however, as yet, no decisive arbitration among the three competing hypotheses of the *Acacia* phyllode origin has been achieved (Gardner *et al.* 2008). Character coding that better reflects the morphological diversity, and possible homology relations among phyllodes, may result in different ancestral-state reconstruction at the *Acacia* most recent common ancestor, as hinted at by Bayesian comparisons of the alternative states fixed at the *Acacia* crown node. This could be worth pursuing on the basis of a detailed appraisal of phyllode diversity.

Data availability

No new data were generated for this study; however, the molecular sequence data used in this study are available from GenBank <https://www.ncbi.nlm.nih.gov/genbank/> and see the voucher information provided at <https://doi.org/10.1038/ncomms5473>; the bioclimatic data are available at the Atlas of Living Australia <http://www.ala.org.au>, the Supplementary material associated with this publication, or from the authors by request, the morphological data are as reported in regional flora treatments and worldwide wattle, see <http://worldwidewattle.com>, the character matrix for which is also available in the Supplementary material associated with this publication, or from the authors by request.

Declaration of funding

This study received no specific funding.

Conflicts of interest

Dr Daniel Murphy is an editor for *Australian Systematic Botany* but did not at any stage have editor-level access to this manuscript while in peer review, as is the standard practice when handling manuscripts submitted by an editor to this journal. *Australian Systematic Botany* encourages its editors to publish in the journal and they are kept totally separate from the decision-making processes for their manuscripts. The authors have no further conflicts of interest to declare.

Acknowledgements

We thank Russell Barrett for his permission to include his photos in Fig. 1, and four reviewers for constructive criticism and comments that improved the study. The authors acknowledge the facilities and the technical assistance of the Sydney Informatics Hub and access to the high-performance computing facility, Artemis, at the University of Sydney.

References

- Aguilera N, Sanhueza C, Guedes LM, Becerra J, Carrasco S, Hernández V (2015) Does *Acacia dealbata* express shade tolerance in Mediterranean forest ecosystems of South America? *Ecology and Evolution* **5**, 3338–3351. doi:10.1002/ece3.1606
- Alberch P, Gould SJ, Oster GF, Wake DB (1979) Size and shape in ontogeny and phylogeny. *Paleobiology* **5**, 296–317. doi:10.1017/S0094837300006588
- Ariati SR, Murphy DJ, Udovicic F, Ladiges PY (2006) Molecular phylogeny of three groups of acacias (*Acacia* subgenus *Phyllodineae*) in arid Australia based on the internal and external transcribed spacer regions of nrDNA. *Systematics and Biodiversity* **4**, 417–426. doi:10.1017/S1477200006001952
- Atkin OK, Schortemeyer M, McFarlane N, Evans JR (1998) Variation in the components of relative growth rate in ten *Acacia* species from contrasting environments. *Plant, Cell & Environment* **21**, 1007–1017. doi:10.1046/j.1365-3040.1998.00356.x
- Azani N, Babineau M, Bailey CD, Banks H, Barbosa AR, Pinto RB, Boatwright JS, Borges LM, Brown GK, Bruneau A, Candido E, Cardoso D, Chung K, Clark RP, Conceição AS, Crisp M, Cubas P, Delgado-Salinas A, Dexter KG, Doyle JJ, Duminil J, Egan AN, de la Estrella M, Falcão MJ, Filatov DA, Fortuna-Perez AP, Fortunato RH, Gagnon E, Gasson P, Rando JG, de Azevedo Tozzi AMG, Gunn B, Harris D, Haston E, Hawkins JA, Herendeen PS, Hughes CE, Iganci JR, Javadi F, Kanu SA, Kazempour-Osaloo S, Kite GC, Klitgaard BB, Kochanovski FJ, Koenen EJ, Kovar L, Lavin M, le Roux M, Lewis GP, de Lima HC, López-Roberts MC, Mackinder B, Maia VH, Malécot V, Mansano VF, Marazzi B, Mattapha S, Miller JT, Mitsuyuki C, Moura T, Murphy DJ, Nageswara-Rao M, Nevado B, Neves D, Ojeda DI, Pennington RT, Prado DE, Prenner G, de Queiroz LP, Ramos G, Filardi FLR, Ribeiro PG, de Lourdes Rico-Arce M, Sanderson MJ, Santos-Silva J, São-Mateus WM, Silva MJ, Simon MF, Sinou C, Snak C, de Souza ÉR, Sprent J, Steele KP, Steier JE, Steeves R, Stirton CH, Tagane S, Torke BM, Toyama H, da Cruz DT, Vatanparast M, Wieringa JJ, Wink M, Wojciechowski MF, Yahara T, Yi T, Zimmerman E (2017) A new subfamily classification of the Leguminosae based on a taxonomically comprehensive phylogeny: the Legume Phylogeny Working Group (LPWG). *Taxon* **66**, 44–77. doi:10.12705/661.3
- Beadle NCW (1966) Soil phosphate and its role in molding segments of the Australian flora and vegetation, with special reference to xeromorphy and sclerophylly. *Ecology* **47**, 992–1007. doi:10.2307/1935647
- Bouckaert R, Heled J, Kühnert D, Vaughan T, Wu C-H, Xie D, Suchard MA, Rambaut A, Drummond AJ (2014) BEAST 2: a software platform for Bayesian evolutionary analysis. *PLoS Computational Biology* **10**, e1003537. doi:10.1371/journal.pcbi.1003537
- Boughton VH (1986) Phyllode structure, taxonomy and distribution in some Australian acacias. *Australian Journal of Botany* **34**, 663–674. doi:10.1071/BT9860663
- Brodrribb TJ, Field TS (2010) Leaf hydraulic evolution led a surge in leaf photosynthetic capacity during early angiosperm diversification. *Ecology Letters* **13**, 175–183. doi:10.1111/j.1461-0248.2009.01410.x
- Brodrribb T, Hill RS (1993) A physiological comparison of leaves and phyllodes in *Acacia melanoxylon*. *Australian Journal of Botany* **41**, 293–305. doi:10.1071/BT9930293
- Brown GK, Ariati SR, Murphy DJ, Miller JT, Ladiges PY (2006) Bipinnate acacias (*Acacia* subg. *Phyllodineae* sect. *Botrycephalae*) of eastern Australia are polyphyletic based on DNA sequence data. *Australian Systematic Botany* **19**, 315–326. doi:10.1071/SB05039
- Brown GK, Murphy DJ, Ladiges PY (2011) Relationships of the Australo-Malesian genus *Paraserianthes* (Mimosoideae: Leguminosae) identifies the sister group of *Acacia sensu stricto* and two biogeographical tracks. *Cladistics* **27**(4), 380–390. doi:10.1111/j.1096-0031.2011.00349.x
- Brown GK, Murphy DJ, Kidman J, Ladiges PY (2012) Phylogenetic connections of phyllodinous species of *Acacia* outside Australia are explained by geological history and human-mediated dispersal. *Australian Systematic Botany* **25**, 390–403. doi:10.1071/SB12027
- Chitwood DH, Sinha NR (2016) Evolutionary and environmental forces sculpting leaf development. *Current Biology* **26**, R297–R306. doi:10.1016/j.cub.2016.02.033
- Cho SH, Coruh C, Axtell M (2012) *miR156* and *mi390* regulate tasiRNA accumulation and developmental timing in *Physcomitrella patens*. *The Plant Cell* **24**, 4837–4849. doi:10.1105/tpc.112.103176
- Christensen BA, Renema W, Henderiks J, De Vleeschouwer D, Groeneveld J, Castaneda IS, Ruening L, Bogus K, Auer G, Ishiwa T, McHugh CM, Gallagher SJ, Fulthorpe CS IODP Expedition 356 Scientists (2017) Indonesian throughflow drove Australian climate from humid Pliocene to arid Pleistocene. *Geophysical Research Letters* **44**, 6914–6925. doi:10.1002/2017GL072977
- Chuck G, Cigan AM, Saetern K, Hake S (2007) The heterochronic maize mutant *Corngrass1* results from overexpression of a tandem microRNA. *Nature Genetics* **39**, 544–549. doi:10.1038/ng2001
- Craven D, Gulamhussein S, Berlyn GP (2010) Physiological and anatomical responses of *Acacia koa* (Gray) seedlings to varying light and drought conditions. *Environmental and Experimental Botany* **69**, 205–213. doi:10.1016/j.envexpbot.2010.04.002
- Crisp MD, Cook LG (2012) Phylogenetic niche conservatism: what are the underlying evolutionary and ecological causes? *New Phytologist* **196**, 681–694. doi:10.1111/j.1469-8137.2012.04298.x
- Crisp MD, Cook LG (2013) How was the Australian flora assembled over the last 65 million years? A molecular phylogenetic perspective. *Annual Review of Ecology, Evolution, and Systematics* **44**, 303–324. doi:10.1146/annurev-ecolsys-110512-135910
- Drummond AJ, Suchard MA, Xie D, Rambaut A (2012) Bayesian phylogenetics with BEAUti and the BEAST 1.7. *Molecular Biology and Evolution* **29**, 1969–1973. doi:10.1093/molbev/mss075
- Dye P, Moses G, Vilakazi P, Ndlela R, Royappen M (2001) Comparative water use of wattle thickets and indigenous plant communities at riparian sites in the Western Cape and KwaZulu-Natal. *Water S.A.* **27**, 529–538. doi:10.4314/wsa.v27i4.4967
- Eckstein RL, Karlsson PS, Weih M (1999) Leaf life span and nutrient resorption as determinants of plant nutrient conservation in

- temperate–arctic regions. *New Phytologist* **143**, 177–189. doi:10.1046/j.1469-8137.1999.00429.x
- Farrell TP, Ashton DH (1978) Population studies on *Acacia melanoxylon* R.Br. I. Variation in seed and vegetative characters. *Australian Journal of Botany* **26**, 365–379. doi:10.1071/BT9780365
- Forster MA, Bonser SP (2009) Heteroblastic development and the optimal partitioning of traits among contrasting environments in *Acacia implexa*. *Annals of Botany* **103**, 95–105. doi:10.1093/aob/mcn210
- Forster MA, Ladd B, Bonser SP (2011) Optimal allocation of resources in response to shading and neighbours in the heteroblastic species, *Acacia implexa*. *Annals of Botany* **107**, 219–228. doi:10.1093/aob/mcq228
- Forster MA, Dalrymple RL, Bonser SP (2016) A low watering treatment alters biomass allocation and growth rate but not heteroplastic development in an *Acacia* species. *Trees* **30**, 2051–2059. doi:10.1007/s00468-016-1432-3
- Gardner SK, Murphy DJ, Newbigin E, Drinnan AN, Ladiges PY (2005) An investigation of phyllode variation in *Acacia verniciflua* and *A. leprosa* (Mimosaceae), and implications for taxonomy. *Australian Systematic Botany* **18**, 383–398. doi:10.1071/SB04052
- Gardner SK, Drinnan A, Newbigin E, Ladiges PY (2008) Leaf ontogeny and morphology in *Acacia* Mill. (Mimosaceae). *Muelleria* **26**, 43–51.
- González-Orozco CE, Laffan SW, Miller JT (2011) Spatial distribution of species richness and endemism of the genus *Acacia* in Australia. *Australian Journal of Botany* **59**, 601–608. doi:10.1071/BT11112
- Hill RS, Brodribb TJ (2001) Macrofossil evidence for the onset of xeromorphy in Australian Casuarinaceae and tribe Banksieae (Proteaceae). *Journal of Mediterranean Ecology* **2**, 127–136.
- Ives AR, Garland T Jr (2010) Phylogenetic logistic regression for binary dependent variables. *Systematic Biology* **59**, 9–26. doi:10.1093/sysbio/syp074
- Jaya E, Kubien DS, Jameson PE, Clemens J (2010) Vegetative phase change and photosynthesis in *Eucalyptus occidentalis*: architectural simplification prolongs juvenile traits. *Tree Physiology* **30**, 393–403. doi:10.1093/treephys/tp128
- Karatsolis BT, De Vleeschouwer D, Groeneveld J, Christensen B, Hendericks J (2020) The late Miocene to early Pliocene ‘Humid Interval’ on the NW Australian Shelf: disentangling climate forcing from regional basin evolution. *Paleoceanography and Paleoclimatology* **35**, e2019PA003780. doi:10.1029/2019PA003780
- Kass RE, Raftery AE (1995) Bayes Factors. *Journal of the American Statistical Association* **90**(430), 773–795. doi:10.1080/01621459.1995.10476572
- Lanfear R, Calcott B, Kainer D, Mayer C, Stamatakis A (2014) Selecting optimal partitioning schemes for phylogenomic datasets. *BMC Evolutionary Biology* **14**, 82. doi:10.1186/1471-2148-14-82
- Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B (2017) PartitionFinder 2: new methods for selecting partitioned models of evolution for molecular and morphological phylogenetic analyses. *Molecular Biology and Evolution* **34**, 772–773.
- Lauter N, Kampani A, Carlson S, Goebel M, Moose SP (2005) *microRNA172* down-regulates *glossy15* to promote vegetative phase change in maize. *Proceedings of the National Academy of Sciences of the United States of America* **102**, 9412–9417. doi:10.1073/pnas.0503927102
- Leroy C, Heuret P (2008) Modelling changes in leaf shape prior to phyllode acquisition in *Acacia mangium* Willd. seedlings. *Comptes Rendus Biologies* **331**, 127–136. doi:10.1016/j.crvi.2007.11.006
- Lin D, Foster DP, Ungar LH (2011) VIF-regression: a fast regression algorithm for large data. *Journal of the American Statistical Association* **106**(493), 232–247. doi:10.1198/jasa.2011.tm10113
- Loveless AR (1961) A nutritional interpretation of sclerophylly based on differences in the chemical composition of sclerophyllous and mesophytic leaves. *Annals of Botany* **25**, 168–184. doi:10.1093/oxfordjournals.aob.a083740
- Macphail MK, Hill RS (2001) Fossil record of *Acacia* in Australia: Eocene to recent. In ‘Flora of Australia. Vol. 11A: Mimosaceae, *Acacia* Part 1’. (Eds AE Orchard, AJG Wilson) pp. 13–29. (CSIRO Publishing: Melbourne, Vic., Australia; and Australian Biological Resources Study: Canberra, ACT, Australia)
- Martin HA (2006) Cenozoic climatic change and the development of the arid vegetation in Australia. *Journal of Arid Environments* **66**, 533–563. doi:10.1016/j.jaridenv.2006.01.009
- Meade A, Pagel M (2016) BayesTraitsV3.0.5 Manual. August 2021 version. Available at <http://www.evolution.rdg.ac.uk/BayesTraitsV3.0.5/Files/BayesTraitsV3.0.5-Manual.pdf>
- Miller CR, James NP, Bone Y (2012) Prolonged carbonate diagenesis under an evolving late Cenozoic climate; Nullarbor Plain, southern Australia. *Sedimentary Geology* **261–262**, 33–49. doi:10.1016/j.sedgeo.2012.03.002
- Miller JT, Murphy DJ, Ho SYW, Cantrill DJ, Seigler D (2013) Comparative dating of *Acacia*: combining fossils and multiple phylogenies to infer ages of clades with poor fossil records. *Australian Journal of Botany* **61**, 436–445. doi:10.1071/BT13149
- Mishler BD, Knerr N, González-Orozco CE, Thornhill AH, Laffan SW, Miller JT (2014) Phylogenetic measures of biodiversity and neo- and paleo-endemism in Australian *Acacia*. *Nature Communications* **5**, 4473. doi:10.1038/ncomms5473
- Morris TL, Esler KJ, Barger JJ, Jacobs SM, Cramer MD (2011) Ecophysiological traits associated with the competitive ability of invasive Australian acacias. *Diversity & Distributions* **17**, 898–910. doi:10.1111/j.1472-4642.2011.00802.x
- Murphy BF, Timbal B (2008) A review of recent climate variability and climate change in southeastern Australia. *International Journal of Climatology* **28**, 859–879. doi:10.1002/joc.1627
- Murphy DJ, Udovicic F, Ladiges PY (2000) Phylogenetic analysis of Australian *Acacia* (Leguminosae: Mimosoideae) by using sequence variations of an intron and two intergenic spacers of chloroplast DNA. *Australian Systematic Botany* **13**, 745–754. doi:10.1071/SB99027
- Murphy DJ, Miller JT, Bayer RJ, Ladiges PY (2003) Molecular phylogeny of *Acacia* subgenus *Phyllodineae* (Mimosoideae: Leguminosae) based on DNA sequences of the internal transcribed spacer region. *Australian Systematic Botany* **16**, 19–26. doi:10.1071/SB01042
- Murphy DJ, Brown GK, Miller JT, Ladiges PY (2010) Molecular phylogeny of *Acacia* Mill. (Mimosoideae: Leguminosae): Evidence for major clades and informal classification. *Taxon* **59**, 7–19. doi:10.1002/tax.591002
- Nge FJ, Biffin E, Thiele KR, Waycott M (2020) Extinction pulse at Eocene–Oligocene boundary drives diversification dynamics of two Australian temperate floras. *Proceedings. Biological Sciences* **287**, 20192546. doi:10.1098/rspb.2019.2546
- Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ (2015) IQ-TREE: a fast and effective stochastic algorithm for estimating maximum likelihood phylogenies. *Molecular Biology and Evolution* **32**, 268–274. doi:10.1093/molbev/msu300
- Ostria-Gallardo E, Ranjan A, Zumstein K, Chitwood DH, Kumar R, Townsley BT, Ichihashi Y, Corcuera LJ, Sinha NR (2016) Transcriptomic analysis suggests a key role for SQUAMOSA PROMOTOR BINDING PROTEIN LIKE, NAC and YUCCA genes in the heteroblastic development of the temperate rainforest tree *Gevuina avellana* (Proteaceae). *New Phytologist* **210**, 694–708. doi:10.1111/nph.13776
- Pagel M, Meade A, Barker D (2004) Bayesian estimation of ancestral character states on phylogenies. *Systematic Biology* **53**, 673–684. doi:10.1080/10635150490522232

- Paradis E, Schliep K (2019) ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics* **35**, 526–528. doi:10.1093/bioinformatics/bty633
- Paradis E, Claude J, Strimmer K (2004) APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290. doi:10.1093/bioinformatics/btg412
- Pasquet-Kok J, Creese C, Sack L (2010) Turning over a new 'leaf': multiple functional significances of leaves versus phyllodes in Hawaiian *Acacia koa*. *Plant, Cell & Environment* **33**, 2084–2100. doi:10.1111/j.1365-3040.2010.02207.x
- Pigliucci M (1997) Ontogenetic phenotypic plasticity during the reproductive phase in *Arabidopsis thaliana* (Brassicaceae). *American Journal of Botany* **84**, 887–895. doi:10.2307/2446278
- Popescu A-A, Huber KT, Paradis E (2012) ape 3.0: new tools for distance based phylogenetics and evolutionary analysis in R. *Bioinformatics* **28**, 1536–1537. doi:10.1093/bioinformatics/bts184
- Pryer K, Hearn D (2009) Evolution of leaf form in marsileaceous ferns: evidence for heterochrony. *Evolution* **63**, 498–513. doi:10.1111/j.1558-5646.2008.00562.x
- Read J, Sanson GD (2003) Characterising sclerophylly: the mechanical properties of a diverse range of leaf types. *New Phytologist* **160**, 81–99. doi:10.1046/j.1469-8137.2003.00855.x
- Renner MAM, Foster CS, Miller JT, Murphy DM (2020) Increased diversification rates are coupled with increased rates of climate space exploration in *Acacia* (Caesalpinioideae). *New Phytologist* **226**, 609–622. doi:10.1111/nph.16349
- Rodríguez J, Lorenzo P, González L (2017) Different growth strategies to invade undisturbed plant communities by *Acacia dealbata* Link. *Forest Ecology and Management* **399**, 47–53. doi:10.1016/j.foreco.2017.05.007
- Rose KME, Mickelbart MV, Jacobs DF (2019) Plasticity of phenotype and heteroblasty in contrasting populations of *Acacia koa*. *Annals of Botany* **124**, 399–409. doi:10.1093/aob/mcz083
- Sommerville KE, Sack L, Ball MC (2012) Hydraulic conductance of *Acacia* phyllodes (foliage) is driven by primary nerve (vein) conductance and density. *Plant, Cell & Environment* **35**, 158–168. doi:10.1111/j.1365-3040.2011.02425.x
- Spalazzi F, Tecco PA, Funes G (2019) Overcoming lag phase: do regenerative attributes onset *Acacia dealbata* spread in a newly invaded system? *Australian Journal of Botany* **67**, 46–54. doi:10.1071/BT18097
- Stamatakis A (2014) RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**, 1312–1313. doi:10.1093/bioinformatics/btu033
- Walker JF, Brown JW, Smith SA (2018) Analysing contentious relationships and outlier genes in phylogenomics. *Systematic Biology* **67**, 916–924. doi:10.1093/sysbio/syy043
- Wang JW, Schwab R, Czech B, Mica E, Weigel D (2008) Dual effects of *miR156*-targeted SPL genes and CYP78A5/KLUH on plastochron length and organ size in *Arabidopsis thaliana*. *The Plant Cell* **20**, 1231–1243. doi:10.1105/tpc.108.058180
- Wang JW, Park MY, Wang LJ, Koo Y, Chen XY, Weigel D, Poethig RS (2011) miRNA control of vegetative phase change in trees. *PLoS Genetics* **7**(2), e1002012. doi:10.1371/journal.pgen.1002012
- Wright IJ, Westoby M (2002) Leaves at low versus high rainfall: coordination of structure, lifespan and physiology. *New Phytologist* **155**, 403–416. doi:10.1046/j.1469-8137.2002.00479.x
- Wu G, Poethig RS (2006) Temporal regulation of shoot development in *Arabidopsis thaliana* by *miR156* and its target SPL3. *Development* **133**, 3539–3547. doi:10.1242/dev.02521
- Wu G, Park MY, Conway SR, Wang JW, Weigel D, Poethig RS (2009) The sequential action of *miR156* and *miR172* regulates developmental timing in *Arabidopsis*. *Cell* **138**, 750–759. doi:10.1016/j.cell.2009.06.031
- Yang L, Xu M, Koo Y, He J, Poethig RS (2013) Sugar promotes vegetative phase change in *Arabidopsis thaliana* by repressing the expression of *miR156C*. *eLife* **2**, e00260. doi:10.7554/eLife.00260
- Yu S, Cao L, Zhou CM, Zhang TQ, Lian H, Sun Y, Wu J, Huang J, Wang G, Wang JW (2013) Sugar is an endogenous cue for juvenile to adult phase transition in plants. *eLife* **2**, e00269. doi:10.7554/eLife.00269
- Zotz G, Wilhelm K, Becker A (2011) Heteroblasty – a review. *Botanical Review* **77**, 109–151. doi:10.1007/s12229-010-9062-8

Handling editor: Jennifer Tate