

## A review of prevalence and phylogeny of the bacterial symbiont *Cardinium* in mites (subclass: Acari)

YAN-KAI ZHANG<sup>1,2</sup>, YA-TING CHEN<sup>1</sup>, KUN YANG<sup>1</sup> & XIAO-YUE HONG<sup>1\*</sup>

<sup>1</sup>Department of Entomology, Nanjing Agricultural University, Nanjing, Jiangsu 210095, China

<sup>2</sup>Institute of Zoology, Chinese Academy of Sciences, Beijing, 100101, China

\* Corresponding author. E-mail: xyhong@njau.edu.cn

### Abstract

Many species of mites are of agricultural and veterinary importance to human. The bacterial symbiont *Cardinium* is assumed to be widespread and involved in manipulating its mite host's reproduction. We investigated *Cardinium* infections in phylogenetically diverse mite species based on published data. *Cardinium* was reported in 45 mite species, representing 3 orders and 14 families. Some *Cardinium* strains can influence their mite hosts' reproduction via two strategies: cytoplasmic incompatibility and feminization. The phylogeny of the examined mites was well resolved based on the *18S rRNA* gene, whereas *Cardinium* phylogeny was not completely concordant with its associated host phylogeny, raising the possibility that horizontal transmission of *Cardinium* has occurred among species. In addition, *Cardinium* phylogeny was associated with the ecology of the hosts. *Cardinium* cooperating with other endosymbionts influences the ecology and evolution of its host.

**Keywords:** Mites, bacterial symbionts, reproduction manipulation, horizontal transmission

### Introduction

Endosymbiotic bacteria are widespread in arthropods. Traditionally, they can be grouped into primary and secondary symbionts. In 2001, the secondary symbiont *Cardinium hertigii* was first reported from the parasitoid wasps *Encarsia* (Weeks *et al.* 2001; Zchori-Fein *et al.* 2001). Several screening studies indicated that about 5–10% of arthropods species are infected with *Cardinium* symbionts (Weeks *et al.* 2003; Zchori-Fein & Perlman 2004; Duron *et al.* 2008). *Cardinium* can manipulate the reproduction of their hosts through various strategies, including parthenogenesis (Zchori-Fein *et al.* 2001), feminization (Weeks *et al.* 2001; Zchori-Fein *et al.* 2004; Giorgini *et al.* 2009), cytoplasmic incompatibility (CI) (Hunter *et al.* 2003; Gotoh *et al.* 2007; Ros & Breeuwer 2009) and altered fecundity (Weeks & Stouthamer 2004). These manipulations play significant roles in the host ecology and evolution (Breeuwer *et al.* 2012; Kageyama *et al.* 2012). Thus, *Cardinium* and other symbionts have received intensive attention.

Mites are the largest group within the arthropod class Arachnida, with over 48,200 described species before 2000 (Halliday *et al.* 2000) and 55,214 described species by 2013 (Zhang 2013). They live in diverse habitats and act as predators, parasites, herbivores or detritivores (Krantz & Walter 2009). Mites of the family Tetranychidae and Tenuipalpidae are known as serious pests of agricultural crops (Helle & Sabelis 1985), those of the family Sarcoptidae and Dermanyssidae are parasites to animals and humans, acting as direct agents of diseases (Moro *et al.* 2009a; Mounsey *et al.* 2013) and those of the family Acaridae are pests of stored products (Palyvos *et al.* 2008). On the other hand, some mites (belonging to families Phytoseiidae, Laelapidae, Macrochelidae) are known for their beneficial effects, acting as biological control agents against agricultural pests (Gerson *et al.* 2003).

Using the electron microscope, Hess & Hoy (1982) had observed two morphologically distinct forms of microorganisms in the tissues of the phytoseiid mite *Metaseiulus occidentalis*, one of which was later identified as *Cardinium* (Weeks & Breeuwer 2003). Subsequently, many mite species were reported to harbor this symbiont (Duron *et al.* 2008; Ros *et al.* 2012; Kopecky *et al.* 2013; Famah Sourassou *et al.* 2014; Weinert *et al.* 2015; Chaisiri *et al.* 2015). There are several studies reporting *Cardinium*-induced reproductive alterations in mites (Weeks & Stouthamer 2004; Groot & Breeuwer 2006; Gotoh *et al.* 2007; Ros & Breeuwer 2009; Liu *et al.* 2010; Xie *et al.* 2010; Wu & Hoy 2012; Zhu *et al.* 2012b; Zhao *et al.* 2013a; Zhao *et al.* 2013b; Famah Sourassou *et al.* 2014), and *Cardinium* seem to be essential for oviposition in the Ghana population of *Neoseiulus paepalivorus* (Famah Sourassou *et al.* 2014). The number of publications on the association mites-*Cardinium* has substantially increased, a comprehensive study is helpful for understanding the interactions of symbionts and hosts.

In the present paper, the prevalence of *Cardinium* symbionts and their reproductive effects in mites was reviewed based on published data. In addition, the phylogenies of mites and their associated *Cardinium* symbionts were presented and compared.

## Material and Methods

### *Available data on Cardinium in mites*

In this study, the reports of *Cardinium* found in mites were summarized by searching from 2 major scientific literature databases, PubMed ([www.ncbi.nlm.nih.gov/pubmed](http://www.ncbi.nlm.nih.gov/pubmed)) and Web of Science ([www.webofknowledge.com](http://www.webofknowledge.com)). The term 'mite' in combination with '*Cardinium*' or 'bacteria' were used as the keywords and applied to the title field or abstracts in those databases. In addition, we considered the unpublished studies on *Cardinium* in mites that have been described in the GenBank (<http://www.ncbi.nlm.nih.gov/genbank>).

### *Phylogenetic analyses of Cardinium and their hosts*

Theories suggested that *Cardinium* and other strict mutualistic maternally inherited symbionts might result in co-evolutionary (phylogenetic) patterns with their hosts (Moran & Baumann 1994; Lo *et al.* 2003). However, this phylogenetic pattern would be lost if horizontal transmission or recent invasion of endosymbionts occurs. Consequently, comparison of endosymbiont's and their hosts' phylogenies could infer their transmission patterns and evolutionary histories.

Previously, the 18S gene of the nuclear ribosomal DNA (*18S rRNA*) has been widely used for resolving the phylogeny of mites (Cruickshank 2002; Matsuda *et al.* 2014). Here, we constructed the phylogeny of *Cardinium*-infected mites based on the available *18S rRNA* sequences that were downloaded from GenBank. Similarly, the phylogeny of associated *Cardinium* was constructed based on the 16S gene of the nuclear ribosomal DNA (*16S rRNA*) that were obtained from our experiments and GenBank. Accession numbers of all available sequences were shown in Table 1. The analyzed sequences were aligned using the multiple alignment program ClustalW with default setting and edited manually in MEGA 6.0 (Tamura *et al.* 2013). In total of 33 sequences of mites *18S rRNA* were obtained and the final alignment was 1892 nucleotides. In *Cardinium 16S rRNA* phylogenetic analyses, there were 31 sequences that consisted of 351 nucleotides. Prior to phylogenetic analyses, jModelTest v2.1.6 (Darriba *et al.* 2012) was used to select the optimal evolution model by Akaike Information Criterion (AIC). Phylogenetic analyses were performed using Bayesian Inference (BI) and Maximum-Likelihood (ML) estimation for mites *18S rRNA* and *Cardinium 16S rRNA*, respectively. For Bayesian inference of mites and *Cardinium*, analyses were performed in MrBayes v3.1.2 (Ronquist & Huelsenbeck 2003) using the GTR+I+G model. Both

Markov chains and samplefreq were set to 1000000 generations and 100, respectively. At the same time, ML trees were constructed using MEGA 6.0, with gamma distributed with invariant sites (G+I), and the method of GTR as genetic distance model; bootstrap replications were set to 1000.

## Results and Discussion

### *Cardinium prevalence in mites*

This paper includes a review of 34 papers reporting *Cardinium* infections in mites. *Cardinium* infections were reported in 45 mite species belonging to 3 orders and 14 families. Detailed information was present in Table 1. The associations of *Cardinium* and family Tetranychidae were relatively well documented. Additionally, *Cardinium* was found in more than one species of family Phytoseiidae, Acaridae, Glycyphagidae, Pyroglyphidae and Tenuipalpidae.

*Cardinium* is an intracellular symbiotic bacterium that cannot be cultured outside of host cells. Thus, the detection of its presence in mites is done mainly using PCR-based techniques, sometimes aided by bioassays (crossing breeding studies). Available molecular markers for its detection in mites include, but not limited to, the 16S ribosomal RNA (*16S rRNA*) and the Gyrase B (*gyrB*) (Hoy & Jeyaprakash 2005; Chigira & Miura 2005; Liu *et al.* 2006; Novelli *et al.* 2007; Enigl & Schausberger 2007; Novelli *et al.* 2008; Chen *et al.* 2009; Moro *et al.* 2009b; Jeong *et al.* 2009; Ros *et al.* 2012; Zhu *et al.* 2012b; Zhao *et al.* 2013a; Zhao *et al.* 2013b; Hubert *et al.* 2012; Famah Sourasson *et al.* 2014). The other techniques used to determine the presence and distribution of *Cardinium* in mites are transmission electron microscopy (Kitajima *et al.* 2007) and fluorescence in situ hybridization (Zhu *et al.* 2012a). Kitajima *et al.* (2007) presented a visual evidence for the presence of *Cardinium* in three *Brevipalpus* species, while Zhu *et al.* (2012a) demonstrated that *Cardinium* mainly enters the oocytes with trophic flow from nurse cells, midgut and oviduct in *Tetranychus piercei*.

*Cardinium* seems to be restricted to four arthropod lineages: Hymenoptera, Hemiptera, Diptera and Acari (Weeks *et al.* 2003; Zchori-Fein & Perlman 2004; Duron *et al.* 2008; Nakamura *et al.* 2009). Among these, mites probably represent *Cardinium* hotspots. For example, Weeks *et al.* (2003) had estimated that about 31.6% mite species should to be infected by *Cardinium*, which seems higher compared with infection rate in others (6.1% in Hemiptera, 10.4% in Hymenoptera). Nine out of 22 spider mite species and 13 astigmatid mite species were reported to harbor *Cardinium* (Nakamura *et al.* 2009; Kopecky *et al.* 2013). Zindel *et al.* (2012) screening for bacterial symbionts in 21 mite species, including phytophagous, predatory or parasitic species, had discussed their possible importance in biological control. To explain high prevalence of *Cardinium* in mites, Weinert *et al.* (2015) speculated that this might reflect evolutionary changes in arthropod immunity, as many of the groups with high *Cardinium* incidences lack components of immune deficiency (IMD) signal pathway, and IMD is activated by diaminopimelic acid-type peptidoglycan, which is produced by *Cardinium* (Penz *et al.* 2012).

Given high prevalence of *Cardinium* and mite species diversity, the number of infected mite species was obviously underestimated. In addition, the infection in mite species of economic importance appears to receive much attention, as indicated by well-documented data for these species. The screened mite species are only a small proportion, and the bacterial community of mites is influenced by environmental factors (Novelli *et al.* 2008; Wu & Hoy 2012; Kopecky *et al.* 2014). It is predicated that more infected species exist on the basis of extensive sampling and sensitive detection approaches. Meanwhile, extensive sampling of mites that have genetic divergence and live in various habitats would allow us to infer how host phylogeny and ecology affect their endosymbiotic bacterial infections.

**TABLE 1.** Documented *Cardinium* infections in mite species.

Order Family Species <sup>a</sup>	Importance of mites	<i>Cardinium</i> -induced phenotypes	Collection sites	References	GenBank Accession numbers	
					<i>18S rRNA</i> of mites	<i>16S rRNA</i> of <i>Cardinium</i> <sup>b</sup>
Mesostigmata						
Dermanyssidae						
<i>Dermanyssus gallinae</i> <sup>w</sup>	Vertebrate parasite		UK, France	1	FJ911836	JX844812 <sup>*</sup>
Phytoseiidae						
<i>Metaseiulus occidentalis</i> <sup>w</sup>	Invertebrate predator	CI	USA	2, 3, 4, 5	-	AY635291 <sup>*</sup>
<i>Euseius finlandicus</i>			Austria	6	-	DQ910766
<i>Neoseiulus paspalivorus</i> <sup>w</sup>			Ghana	7	-	KF135427
Sarcoptiformes						
Acaridae						
<i>Acarus siro</i>	Stored products pest		The Czech Republic	8	AF022023	JQ406832 <sup>*</sup>
<i>Aleuroglyphus ovatus</i>			UK	8	JQ000109	JQ406781 <sup>*</sup>
<i>Sancassania berlessei</i>			UK	8	GQ864331	JQ406760 <sup>*</sup>
<i>Tyroborus lini</i>			The Czech Republic	8	EF203768	JQ406755 <sup>*</sup>
<i>Tyrophagus putrescentiae</i>			The Czech Republic	8	DQ025510	JQ406820 <sup>*</sup>
Aeroglyphidae						
<i>Aeroglyphus robustus</i>	Stored products pest		The Czech Republic	8	EU152490	JQ406701 <sup>*</sup>
Carpoglyphidae						
<i>Carpoglyphus lactis</i>	Stored products pest		Colombia	8	GQ864327	JQ406713 <sup>*</sup>
Chaetodactylidae						
<i>Chaetodactylus sp</i>	Invertebrate parasite		South Korea	9	-	EU930867 <sup>*</sup>
Chortoglyphidae						
<i>Chortoglyphus arcuatus</i>	Stored products pest		The Czech Republic	8	AF022028	JQ406809 <sup>*</sup>
Echimyopodidae						
<i>Blomia tropicalis</i>	Household pest		The Czech Republic	8	AY499556	JQ406805 <sup>*</sup>
Glycyphagidae						
<i>Glycyphagus domesticus</i>	Stored products pest		The Czech Republic	8	AY499555	JQ406725 <sup>*</sup>
<i>Lepidoglyphus destructor</i>			The Czech Republic	8	EF203771	JQ406843 <sup>*</sup>
Pyroglyphidae						
<i>Dermatophagoides pteronyssinus</i>	Household pest		The Czech Republic	8	DQ025511	JQ406770 <sup>*</sup>
<i>Dermatophagoides farina</i>			The Czech Republic	8	-	-
Oppiidae						
<i>Oppiella nova</i> <sup>w</sup>	Soil detritivore		USA	2	-	AY279414 <sup>*</sup>
Trombidiformes						
Erythraeidae						
<i>Balaustium sp</i>	Invertebrate predator		USA	2	EF203775	AY279411 <sup>*</sup>
Tenuipalpidae						
<i>Brevipalpus californicus</i>	Plant pest	Feminization	Japan	10	-	AB116514 <sup>*</sup>

.....continued on the next page

Order Family Species <sup>a</sup>	Importance of mites	<i>Cardinium</i> -induced phenotypes	Collection sites	References	GenBank Accession numbers	
					18S rRNA of mites	16S rRNA of <i>Cardinium</i> <sup>b</sup>
<i>Brevipalpus obovatus</i>			USA	2	-	AY279401*
<i>Brevipalpus phoenicis</i>		Feminization	USA, Brazil	2, 11, 12	-	KJ675560*
<i>Breipalpus lewisi</i>			Japan	10	-	AB116515*
Tetranychidae						
<i>Bryobia rubrioculus</i> <sup>wo</sup>	Plant pest		Austria, France, Netherlands, Poland	6, 13	-	DQ910769
<i>Bryobia sarothamni</i> <sup>wo</sup>		CI	France	14, 15	-	JN572884
<i>Eotetranychus uncutus</i>			Austria	6	AB926275	DQ910768
<i>Eotetranychus suginamensis</i>		CI	Japan	10, 16	AB926271	AB241129*
<i>Tetranychus urticae</i> R ( <i>T. cinnabarinus</i> )		CI	Japan, China	10, 16, 17, 18	AB926313	KP828050*
<i>Tetranychus phaselus</i> <sup>wo</sup>		CI	China	19	AB926307	KP828051*
<i>Tetranychus kanzawai</i> <sup>wo</sup>			China	20	AB926298	KP828053*
<i>Tetranychus truncatus</i> <sup>wo</sup>		CI	China	21	AB926310	KP828054*
<i>Tetranychus pueraricola</i> <sup>wo</sup>			Japan	10, 16	AB926309	AB241135*
<i>Tetranychus piercei</i> <sup>wo</sup>		CI	China	22	AB926308	-
<i>Panonychus ulmi</i>			Switzerland	20	AB926242	JX844809*
<i>Panonychus mori</i> <sup>wo</sup>			Japan	16	AB926239	-
<i>Amphitetranychus viennensis</i> <sup>wo</sup>			China	20	AB926293	KP828052*
<i>Amphitetranychus quercivorus</i>			Japan	10, 16	AB926292	AB241131*
<i>Oligonychus ilicis</i>			Japan	10, 16	AB926284	AB241130*
<i>Oligonychus camelliae</i>			Japan	16	AB926278	-
<i>Oligonychus coffeae</i>			Japan	16	AB926281	-
<i>Oligonychus gotohi</i> <sup>wo</sup>			Japan	16	AB926282	-
<i>Petrobia harti</i> <sup>wo</sup>			USA	2, 14	AB926230	AY279410*
<i>Stigmaeopsis celarius</i>			Japan	16	AB926253	-
<i>Stigmaeopsis longus</i>			Japan	16	AB926254	-

<sup>a</sup>Superscripts on the mite species indicate *Wolbachia*-infected ones.

<sup>b</sup>Asterisks at the GenBank accession numbers indicate the 16S rRNA sequences used for *Cardinium* phylogenetic analysis.

1—De Luna *et al.* (2009); 2—Weeks *et al.* (2003); 3—Zchori-Fein & Perlman (2004); 4—Hoy & Jeyaparakash (2005); 5—Wu & Hoy (2012); 6—Enigl & Schausberger (2007); 7—Famah Sourassou *et al.* (2014); 8—Kopecky *et al.* (2013); 9—Jeong *et al.* (2009); 10—Gotoh *et al.* (2007); 11—Groot & Breeuwer (2006); 12—Weeks *et al.* (2001); 13—Ros *et al.* (2012); 14—Ros & Breeuwer (2009); 15—Ros *et al.* (2009); 16—Nakamura *et al.* (2009); 17—Liu *et al.* (2010); 18—Xie *et al.* (2010); 19—Zhao *et al.* (2013a); 20—unpublished; 21—Zhao *et al.* (2013b); 22—Zhu *et al.* (2012).

### *Cardinium*'s reproductive phenotypes in mites

*Cardinium* is known to affect their hosts in a variety of ways in order to favor their own transmission. These include CI and feminization. For example, *Cardinium* was found associated with nonreciprocal reproductive incompatibility in *M. occidentalis*, a phytoseiid mite that is an important predator of spider mite pests of agricultural crops (Wu & Hoy 2012). *Cardinium*-induced CI was also revealed in six phytophagous mite species, *Eotetranychus suginamensis* (Gotoh *et al.* 2007), *Bryobia sarothamni* (Ros & Breeuwer 2009), *Tetranychus cinnabarinus* (Xie *et al.* 2010), *T. piercei* (Zhu *et al.* 2012b), *Tetranychus phaselus* (Zhao *et al.* 2013a) and *Tetranychus truncatus* (Zhao *et al.* 2013b). All these CI types were expressed as female mortality, which is the only CI type found for *Cardinium* so far. Moreover, Groot & Breeuwer (2006) found *Cardinium* induce thelytoky by feminizing unfertilized haploid eggs in the related species *Brevipalpus californicus* and *B.*

*phoenicis*. Through reproductive manipulations, *Cardinium* could strongly impact mite population dynamics and demography as well as the mite genetic diversity (Groot 2007; Ros 2010).

As mentioned previously, factors such as bacterial strains and densities, host genetic background or interactions between bacteria and host species can influence *Cardinium*-induced phenotypes. Gotoh *et al.* (2007) revealed that phylogenetic similarity of *Cardinium* was not related to their phenotypes, as CI-inducing *Cardinium* strain in *E. suginamensis* is closely related to the non-CI strains found in *T. pueraricola* and *Amphitetranychus quercivorus*, based on the *16S rRNA* tree. Furthermore, some studies revealed that *Cardinium* genotype, host genotype and *Cardinium* density may account for variations in CI expression (Xie *et al.* 2010; Zhu *et al.* 2012b; Zhao *et al.* 2013b). To date, studies on *Cardinium*-mediated reproductive alterations are still limited to a few phytophagous, predatory or parasitic species and more related studies especially in synanthropic mites infesting agricultural crops, stored products or acting as allergen are needed.

#### *Potential roles of Cardinium in mite biological control*

Predatory mites are widely used for biological control of phytophagous mites. *Cardinium*-induced CI in predatory mites *M. occidentalis* has been reported (Johanowicz & Hoy 1998; Wu & Hoy 2012). Weeks & Stouthamer (2004) also reported that three inbred lines of *M. occidentalis* showed a clear and significant increase in fecundity associated with infection by *Cardinium*. In these cases, *Cardinium*-mediated reproductive incompatibility may have significant implications for biological control programs using *M. occidentalis* because the release of *M. occidentalis* strains that are reproductively incompatible with the wild populations (caused by a different status of *Cardinium* infection) may reduce the effectiveness of this predatory mite as a biological control agent.

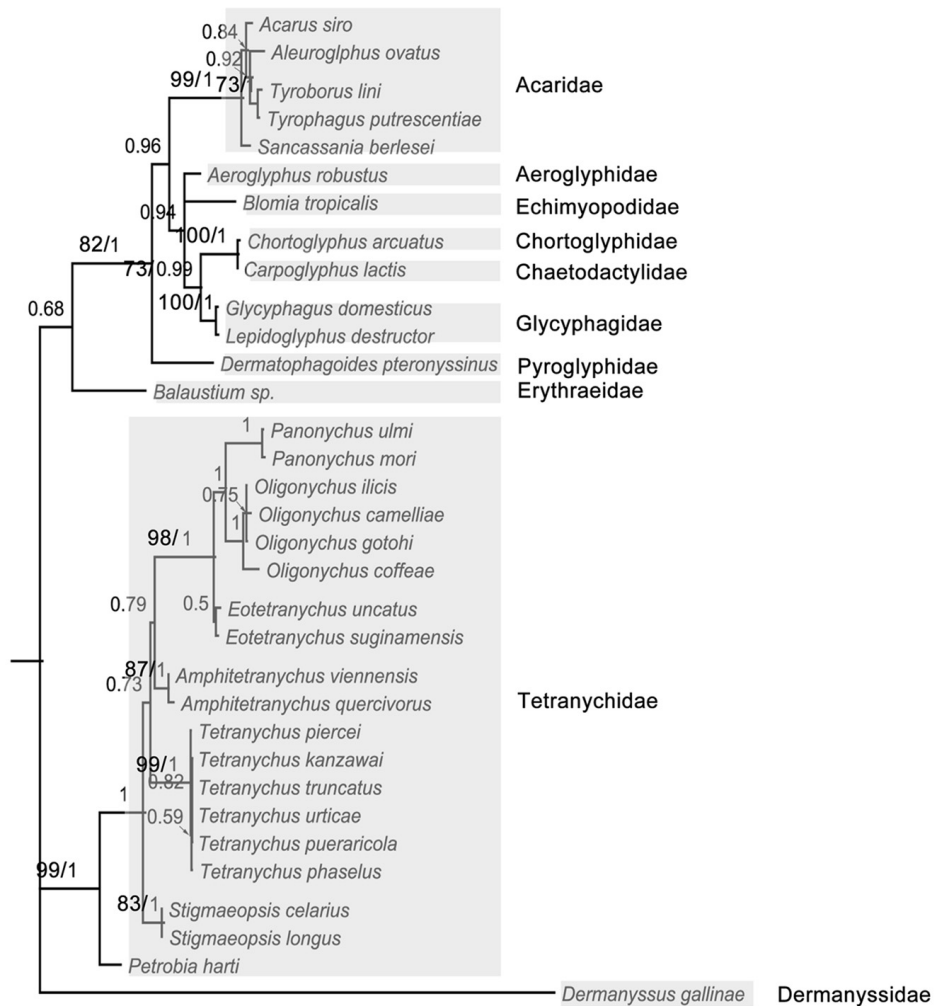
#### *Phylogenetic relationship of Cardinium and their mite hosts*

Phylogenetic analyses indicated that the evolutionary relationship among *Cardinium* infecting mites was well resolved and the observed tree showed congruence with the classical taxonomy based on morphology, except for the family Chortoglyphidae and Carpoglyphidae (Fig. 1), suggesting *18S rRNA*'s better discriminating ability. Although the resolution of the phylogenetic tree of *Cardinium* using *16S rRNA* was low, it can provide information to identify two clusters of closely related *Cardinium* strains. As shown in Fig. 2, cluster I consisted of *Cardinium* from *T. pueraricola*, *T. phaselus*, *T. kanzawai*, *E. suginamensis*, *A. quercivorus*, *A. viennensis*, *Oligonychus illcis*, *B. obovatus*, *Petrobia harti* and *Balaustium sp.* And the remaining strains formed the cluster II.

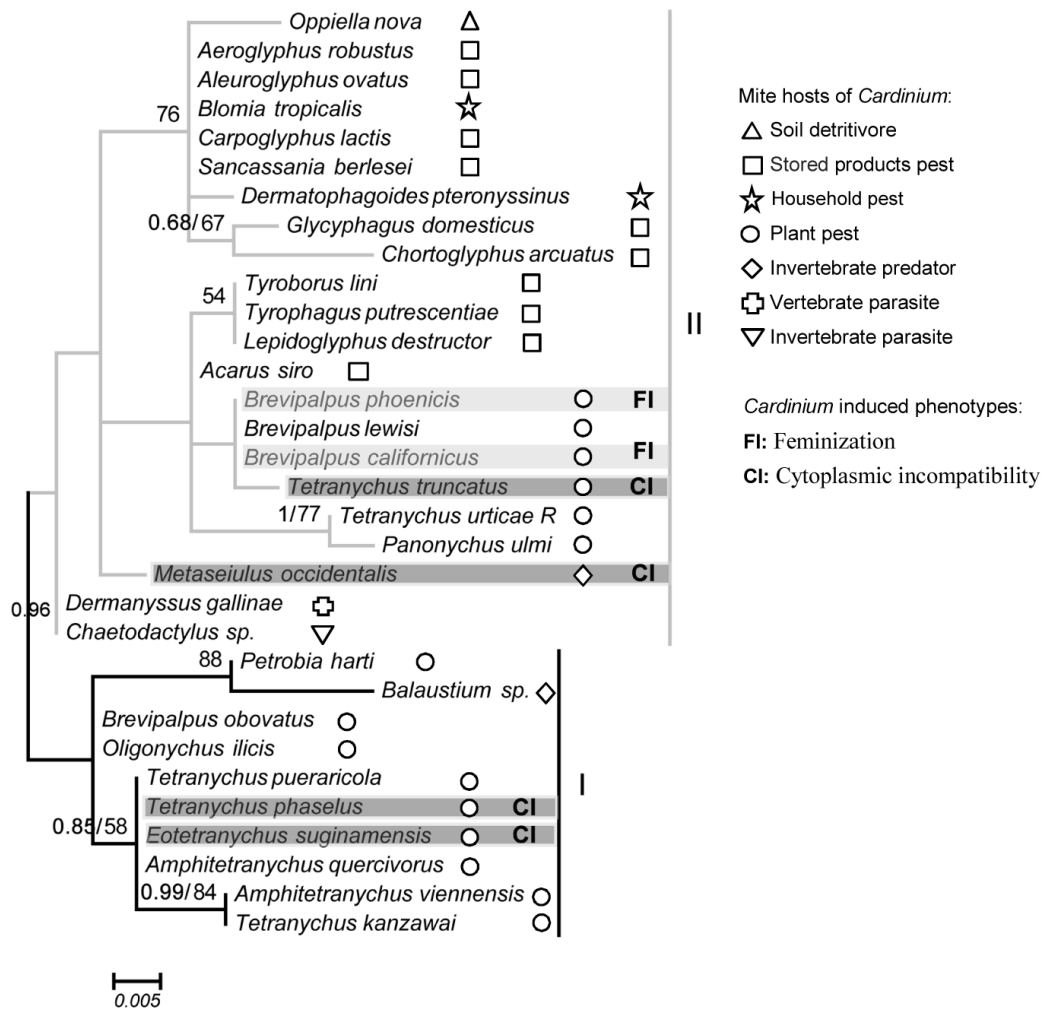
Comparisons between the evolutionary relationships among mites and associated *Cardinium* symbionts showed that there is no complete association between the two groups, as several *Cardinium* strains infecting different species were monophyletic. We cannot completely exclude the possibility that there has been repeated loss of infection post-speciation from an infected ancestor. This difference between the two phylogenies indicated that *Cardinium* was not solely acquired vertically, and points to the likelihood of horizontal transmission. Meanwhile, a degree of *Cardinium* specialization was observed, for example, closely related *Cardinium* strains clustered among three closely related *Brevipalpus* species and *Cardinium* infecting family Tetranychidae appeared to be closely related. These results were not surprising and similar trends have been observed in other *Cardinium*-arthropod associations (Weeks *et al.* 2003; Zchori-Fein & Perlman 2004; Duron *et al.* 2008; Kopecky *et al.* 2013) and in other endosymbionts (Weinert *et al.* 2009; Lachowska *et al.* 2010; Gerth *et al.* 2015). In addition, the single molecular marker will fail to assess the accurate phylogeny of *Cardinium* (Weeks *et al.* 2003; Duron *et al.* 2008). Thus more rapidly evolving and phylogenetically informative *Cardinium* genes are required in further study.

*Correlation between Cardinium infections and mite hosts ecology*

Mites exhibit a breadth of ecological interactions unmatched in any other arthropod group (Krantz & Walter 2009). Diversity of habitats allowed us to test whether host ecology affects *Cardinium* strain infection patterns. Infected mite species can be spitted into seven types according to their habitats or economic importance. Species of the same type may share common environment or have ecological interactions. In general, mite ecology appeared to shape *Cardinium* strain similarity (Fig. 2). For example, the phylogenetic analysis revealed that *Cardinium* from phytophagous mites were more similar to one another. In addition, similar *Cardinium* strains were observed in mites that inhabiting house environment. These observations suggest that mite species sharing close ecological conditions easily transmit *Cardinium* among them. Ecologically mediated transmission of bacterial symbionts had also been reported in fruit feeding drosophilids (Haïne *et al.* 2005), mycophagous flies (Stahlhut *et al.* 2010) and *Megalomyrmex* social parasites and their fungus-growing ant hosts (Liberti *et al.* 2015).



**FIGURE 1. Bayesian phylogenetic tree of mite species infected with *Cardinium* symbionts based on 18S *rRNA* genes.** The topology resulting from Maximum Likelihood (ML) method was similar. ML bootstrap values (left numbers) and Bayesian probabilities (right numbers) were presented above the branches. Words on the tree indicated the family in which mite species is placed.



**FIGURE 2. Maximum Likelihood phylogenetic tree of *Cardinium* symbionts associated with several mite species based on *16S rRNA* genes.** Strains were characterized by the names of their host species. The topology resulting from Bayesian method was similar. ML bootstrap values (right numbers) and Bayesian posterior (left numbers) were presented above the branches. Different shapes on the right side indicated mites' economic importance. Strains with the ability of manipulating reproduction were marked with bold letters.

#### *Multiple infections of *Cardinium* and other endosymbionts in mites*

Apart from *Cardinium*, mites are hosts to other endosymbionts. For instance, Chaisiri *et al.* (2015) have revealed that mite species were associated with 85 bacterial genera (approximately 150 identified species) belonging to 7 phyla and 25 orders. Among these bacteria, *Cardinium*, *Wolbachia* and *Orientia* are the most prevalent bacteria. *Wolbachia* is another known reproductive parasite (Werren *et al.* 2008). Many mite species showed co-infected with *Cardinium* and *Wolbachia* (Table 1) and sometimes, the two endosymbionts were found co-infecting within the same individuals of particular species. Population differentiation itself may facilitate the persistence of individual endosymbionts within different populations, whereas the acquisition of superinfections through horizontal transmission may contribute to intra-individual co-infection.



The presence of competing endosymbiont strains is expected to influence the dynamics of the population as a whole, for example, if neighbouring populations are differently infected with one or more bacterial strains with different phenotypes or if a new strain invades one population. An example of this is observed in *Bryobia* spider mites where *Cardinium*, which causes CI, sometimes co-occurs with *Wolbachia* (Ros & Breeuwer 2009). Similarly, either or both *Wolbachia* and *Cardinium* induce CI, as *Wolbachia* induces weak CI, whereas *Cardinium* induces strong CI and doubly-infected ones showed the strongest CI in *T. piercei* (Zhu *et al.* 2012), *T. phaselus* (Zhao *et al.* 2013a) and *T. truncates* (Zhao *et al.* 2013b). In addition, Weeks *et al.* (2003) and Zchori-Fein & Perlman (2004) reported mixed infections with *Wolbachia* and *Cardinium* in the predatory mite *M. occidentalis*. The poultry red mite *Dermanyssus gallinae* was found to be infected with both *Cardinium* and *Spiroplasma* (De Luna *et al.* 2009). These observations indicated that *Cardinium* might shape its hosts' ecology and evolution in cooperation with other endosymbionts.

## Conclusions

Our literature review confirmed that *Cardinium* is widely distributed in mites. As a reproductive parasite, some *Cardinium* strains indeed manipulate their host reproduction. Comparisons of the evolutionary relationships between mites and associated *Cardinium* indicated that there was no complete association between the two, suggesting horizontal transmission of this symbiont, which possibly occurred more readily among species that have close ecological requirements. *Cardinium* is not alone in mites, as there are other symbionts; *Cardinium* may influence the host biology in combination with them. Overall, this work provided basic knowledge for understanding the interaction between endosymbionts and mites.

## Acknowledgements

This study was supported in part by a grant-in-aid from the Science and Technology Program of the National Public Welfare Professional Fund (201103020) from the Ministry of Agriculture of China, and a grant-in-aid for Scientific Research (31172131) from the National Natural Science Foundation of China. We are grateful to Xiao-Feng Xue and Jing-Tao Sun of the Department of Entomology, Nanjing Agricultural University for their valuable comments on the earlier version of this manuscript.

## References

- Breeuwer, H., Ros, V.I.D. & Groot, T.V.M. (2012) *Cardinium*: the next addition to the family of reproductive parasites. In: Zchori-Fein, E. & Bourtzis, K. (Eds), *Manipulative tenants: Bacteria associated with arthropods*, London, CRC Press, pp. 207–219.
- Chaisiri, K., McGarry, J.O.H.N., Morand, S. & Makepeace, B.L. (2015) Symbiosis in an overlooked microcosm: a systematic review of the bacterial flora of mites. *Parasitology*, 1–11.  
<http://dx.doi.org/10.1017/s0031182015000530>
- Chen, X.L., Xie, R.R., Li, G.Q. & Hong, X.Y. (2009) Simultaneous detection of endosymbionts *Wolbachia* and *Cardinium* in spider mites (Acari: Tetranychidae) by multiplex-PCR. *International Journal of Acarology*, 35 (5), 397–403.  
<http://dx.doi.org/10.1080/01647950903292756>
- Chigira, A. & Miura, K. (2005) Detection of 'Candidatus *Cardinium*' bacteria from the haploid host *Brevipalpus californicus* (Acari: Tenuipalpidae) and effect on the host. *Experimental and Applied Acarology*, 37(1–2), 107–116.  
<http://dx.doi.org/10.1007/s10493-005-0592-4>

- Cruickshank, R.H. (2002) Molecular markers for the phylogenetics of mites and ticks. *Systematic and Applied Acarology*, 7, 3–14.  
<http://dx.doi.org/10.11158/saa.7.1.1>
- Darriba, D., Taboada, G.L., Doallo, R. & Posada, D. (2012) jModelTest 2: more models, new heuristics and parallel computing. *Nature Methods*, 9(8), 772.  
<http://dx.doi.org/10.1038/nmeth.2109>
- De Luna, C.J., Moro, C.V., Guy, J.H., Zenner, L. & Sparagano, O.A. (2009) Endosymbiotic bacteria living inside the poultry red mite (*Dermanyssus gallinae*). *Experimental and Applied Acarology*, 48(1–2), 105–113.  
<http://dx.doi.org/10.1007/s10493-008-9230-2>
- Duron, O., Bouchon, D., Boutin, S., Bellamy, L., Zhou, L., Engelstädter, J. & Hurst, G.D. (2008) The diversity of reproductive parasites among arthropods: *Wolbachia* do not walk alone. *BMC Biology*, 6(1), 27.  
<http://dx.doi.org/10.1186/1741-7007-6-27>
- Enigl, M. & Schausberger, P. (2007) Incidence of the endosymbionts *Wolbachia*, *Cardinium* and *Spiroplasma* in phytoseiid mites and associated prey. *Experimental and Applied Acarology*, 42, 75–85.  
<http://dx.doi.org/10.1007/s10493-007-9080-3>
- Gerson, U., Smiley, R.L. & Ochoa, R. (2003) *Mites (Acari) for pest control*. John Wiley & Sons.  
<http://dx.doi.org/10.1002/9780470750995>
- Gerth, M., Saeed, A., White, J.A. & Bleidorn, C. (2015) Extensive screen for bacterial endosymbionts reveals taxon-specific distribution patterns among bees (Hymenoptera, Anthophila). *FEMS Microbiology Ecology*, 91(6), fiv047.  
<http://dx.doi.org/10.1093/femsec/fiv047>
- Giorgini, M., Monti, M.M., Caprio, E., Stouthamer, R. & Hunter, M.S. (2009) Feminization and the collapse of haplodiploidy in an asexual parasitoid wasp harboring the bacterial symbiont *Cardinium*. *Heredity*, 102(4), 365–371.  
<http://dx.doi.org/10.1038/hdy.2008.135>
- Gotoh, T., Noda, H. & Ito, S. (2007) *Cardinium* symbionts cause cytoplasmic incompatibility in spider mites. *Heredity*, 98, 13–20.  
<http://dx.doi.org/10.1038/sj.hdy.6800881>
- Groot, T. (2007) Cause and consequence of haploid females in *Brevipalpus* mites. *Entomologische Berichten* (Amsterdam), 67(5), 165–169.
- Groot, T.V.M. & Breeuwer, J.A.J. (2006) *Cardinium* symbionts induce haploid thelytoky in most clones of three closely related *Brevipalpus* species. *Experimental and Applied Acarology*, 39, 257–271.  
<http://dx.doi.org/10.1007/s10493-006-9019-0>
- Haine, E.R., Pickup, N.J. & Cook, J.M. (2005) Horizontal transmission of *Wolbachia* in a *Drosophila* community. *Ecological Entomology*, 30(4), 464–472.  
<http://dx.doi.org/10.1111/j.0307-6946.2005.00715.x>
- Halliday, R.B., O'Connor, B.M. & Baker, A.S. (2000) Global diversity of mites. In: Raven P.H. (Eds), *Nature and Human Society—the Quest for a Sustainable World*. National Academy Press, Washington DC, pp. 192–203.
- Helle, W. & Sabelis, M.W. (1985) *Spider mites: their biology, natural enemies and control*. Elsevier, Amsterdam, the Netherlands.
- Hess, R.T. & Hoy, M.A. (1982) Microorganisms associated with the spider mite predator *Metaseiulus* (=Typhlodromus) *occidentalis*: electron microscope observations. *Journal of Invertebrate Pathology*, 40, 98–106.  
[http://dx.doi.org/10.1016/0022-2011\(82\)90041-6](http://dx.doi.org/10.1016/0022-2011(82)90041-6)
- Hoy, M.A. & Jeyaprakash, A. (2005) Microbial diversity in the predatory mite *Metaseiulus occidentalis* (Acari: Phytoseiidae) and its prey, *Tetranychus urticae* (Acari: Tetranychidae). *Biological Control*, 32(3), 427–441.  
<http://dx.doi.org/10.1016/j.biocontrol.2004.12.012>
- Hubert, J., Kopecký, J., Perotti, M.A., Nesvorná, M., Braig, H.R., Ságová-Marečková, M., Macovei, L. & Zurek, L. (2012) Detection and identification of species-specific bacteria associated with synanthropic mites. *Microbial Ecology*, 63(4), 919–928.  
<http://dx.doi.org/10.1007/s00248-011-9969-6>
- Hunter, M.S., Perlman, S.J. & Kelly, S.E. (2003) A bacterial symbiont in the bacteroidetes induces cytoplasmic incompatibility in the parasitoid wasp *Encarsia pergandiella*. *Proceedings of the Royal Society of London B: Biological Sciences*, 270(1529), 2185–2190.  
<http://dx.doi.org/10.1098/rspb.2003.2475>
- Jeong, G., Lee, K., Choi, J., Hwang, S., Park, B., Kim, W., Choi, Y., Park, I. & Kim, J. (2009) Incidence of *Wolbachia* and *Cardinium* endosymbionts in the *Osmia* community in Korea. *The Journal of Microbiology*, 47(1), 28–32.

- <http://dx.doi.org/10.1007/s12275-009-0198-3>
- Johanowicz, D.L. & Hoy, M.A. (1998) Experimental induction and termination of non-reciprocal reproductive incompatibilities in a parhaploid mite. *Entomologia Experimentalis et Applicata*, 87, 51–58.  
<http://dx.doi.org/10.1046/j.1570-7458.1998.00303.x>
- Kageyama, D., Narita, S. & Watanabe, M. (2012) Insect sex determination manipulated by their endosymbionts: incidences, mechanisms and implications. *Insects*, 3(1), 161–199.  
<http://dx.doi.org/10.3390/insects3010161>
- Kitajima, E.W., Groot, T.V., Novelli, V.M., Freitas-Astúa, J., Alberti, G. & de Moraes, G.J. (2007) In situ observation of the *Cardinium* symbionts of *Brevipalpus* (Acari: Tenuipalpidae) by electron microscopy. *Experimental and Applied Acarology*, 42(4), 263–271.  
<http://dx.doi.org/10.1007/s10493-007-9090-1>
- Kopecky, J., Nesvorna, M., Mareckova-Sagova, M. & Hubert, J. (2014) The effect of antibiotics on associated bacterial community of stored product mites. *PLoS ONE*, 9(11), e112919.  
<http://dx.doi.org/10.1371/journal.pone.0112919>
- Kopecky, J., Perotti, M.A., Nesvorna, M., Erban, T., & Hubert, J. (2013) *Cardinium* endosymbionts are widespread in synanthropic mite species (Acari: Astigmata). *Journal of Invertebrate Pathology*, 112(1), 20–23.  
<http://dx.doi.org/10.1016/j.jip.2012.11.001>
- Krantz, G.W. & Walter, D.E. (2009) *A manual of acarology*. 3rd. Edn. Texas Tech University Press, Texas, 807 pp.
- Lachowska, D., Kajtoch, E. & Knutelski, S. (2010) Occurrence of *Wolbachia* in central European weevils: correlations with host systematics, ecology, and biology. *Entomologia Experimentalis et Applicata*, 135(1), 105–118.  
<http://dx.doi.org/10.1111/j.1570-7458.2010.00974.x>
- Liberti, J., Sapountzis, P., Hansen, L.H., Sørensen, S.J., Adams, R.M. & Boomsma, J.J. (2015) Bacterial symbiont sharing in *Megalomyrmex* social parasites and their fungus-growing ant hosts. *Molecular Ecology*, 24(12), 3151–3169.  
<http://dx.doi.org/10.1111/mec.13216>
- Liu, Y. Miao, H. & Hong, X.Y. (2006) Distribution of the endosymbiotic bacterium *Cardinium* in Chinese populations of the carmine spider mite *Tetranychus cinnabarinus* (Acari: Tetranychidae). *Journal of Applied Entomology*, 130(9–10), 523–529.  
<http://dx.doi.org/10.1111/j.1439-0418.2006.01112.x>
- Liu, Y., Xie, R.R. & Hong, X.Y. (2010) Manipulation of symbiont *Cardinium* on the reproduction of the carmine spider mite, *Tetranychus cinnabarinus* (Boisduval) (Acari: Tetranychidae). *Acta Entomologica Sinica*, 53(11), 1233–1240.
- Lo, N., Bandi, C., Watanabe, H., Nalepa, C. & Beninati, T. (2003) Evidence for cocladogenesis between diverse dictyopteran lineages and their intracellular endosymbionts. *Molecular Biology and Evolution*, 20(6), 907–913.  
<http://dx.doi.org/10.1093/molbev/msg097>
- Matsuda, T., Morishita, M., Hinomoto, N. & Gotoh, T. (2014) Phylogenetic analysis of the spider mite sub-family Tetranychinae (Acari: Tetranychidae) based on the mitochondrial COI gene and the 18S and the 5' end of the 28S rRNA genes indicates that several genera are polyphyletic. *PLoS ONE*, 9(10), e108672.  
<http://dx.doi.org/10.1371/journal.pone.0108672>
- Moran, N. & Baumann, P. (1994) Phylogenetics of cytoplasmically inherited microorganisms of arthropods. *Trends in Ecology & Evolution*, 9(1), 15–20.  
[http://dx.doi.org/10.1016/0169-5347\(94\)90226-7](http://dx.doi.org/10.1016/0169-5347(94)90226-7)
- Moro, C.V., De Luna, C.J., Tod, A., Guy, J.H., Sparagano, O.A.E. & Zenner, L. (2009a) The poultry red mite (*Dermanyssus gallinae*): a potential vector of pathogenic agents. *Experimental and Applied Acarology*, 48, 93–104.  
<http://dx.doi.org/10.1007/s10493-009-9248-0>
- Moro, C.V., Thioulouse, J., Chauve, C., Normand, P. & Zenner, L. (2009b) Bacterial taxa associated with the hematophagous mite *Dermanyssus gallinae* detected by 16S rRNA PCR amplification and TTGE fingerprinting. *Research in Microbiology*, 160(1), 63–70.  
<http://dx.doi.org/10.1016/j.resmic.2008.10.006>
- Mounsey, K.E., McCarthy, J.S. & Walton, S.F. (2013) Scratching the itch: new tools to advance understanding of scabies. *Trends in Parasitology*, 29(1), 35–42.  
<http://dx.doi.org/10.1016/j.pt.2012.09.006>
- Nakamura, Y., Kawai, S., Yukuhiro, F., Ito, S., Gotoh, T., Kisimoto, R., Yanase, T., Matsumoto, Y., Kageyama, D. & Noda, H. (2009) Prevalence of *Cardinium* bacteria in planthoppers and spider mites and taxonomic re-

- sion of “*Candidatus Cardinium hertigii*” based on detection of a new *Cardinium* group from biting midges. *Applied and Environmental Microbiology*, 75(21), 6757–6763.  
<http://dx.doi.org/10.1128/AEM.01583-09>
- Novelli, V.M., Freitas-Astúa, J., Arrivabem, F., Locali-Fabris, E.C., Hilf, M.E., Gottwald, T.R. & Machado, M.A. (2007) Effects of temperature, storage period and the number of individuals on the detection of the false spider mite *Cardinium* endosymbiont. *Experimental and Applied Acarology*, 42(1), 17–21.  
<http://dx.doi.org/10.1007/s10493-007-9070-5>
- Novelli, V.M., Freitas-Astúa, J., Segatti, N., Mineiro, J.L., Arthur, V., Bastianel, M., Hilf, M.E., Gottwald, T.R. & Machado, M.A. (2008) Effects of radiation (Cobalt-60) on the elimination of *Brevipalpus phoenicis* (Acari: Tenuipalpidae) *Cardinium* endosymbiont. *Experimental and Applied Acarology*, 45(3–4), 147–153.  
<http://dx.doi.org/10.1007/s10493-008-9176-4>
- Palyvos, N.E., Emmanouel, N.G. & Saitanis, C.J. (2008) Mites associated with stored products in Greece. *Experimental and Applied Acarology*, 44(3), 213–226.  
<http://dx.doi.org/10.1007/s10493-008-9145-y>
- Penz, T., Schmitz-Esser, S., Kelly, S.E., Cass, B.N., Müller, A., Woyke, T., Malfatti S.A. & Horn, M. (2012) Comparative genomics suggests an independent origin of cytoplasmic incompatibility in *Cardinium hertigii*. *PLoS Genetics*, 8, e1003012.  
<http://dx.doi.org/10.1371/journal.pgen.1003012>
- Ronquist, F. & Huelsenbeck, J.P. (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics*, 19(12), 1572–1574.  
<http://dx.doi.org/10.1093/bioinformatics/btg180>
- Ros, V.I. (2010) Evolutionary consequences of reproductive parasites in spider mites. *Entomologische Berichten*, 70(5), 154–161.
- Ros, V.I., Fleming, V.M., Feil, E.J. & Breeuwer, J.A. (2012) Diversity and recombination in *Wolbachia* and *Cardinium* from *Bryobia* spider mites. *BMC Microbiology*, 12(1), 1.  
<http://dx.doi.org/10.1186/1471-2180-12-s1-s13>
- Ros, V.I.D. & Breeuwer, J.A.J. (2009) The effects of, and interactions between, *Cardinium* and *Wolbachia* in the doubly infected spider mite *Bryobia sarothamni*. *Heredity*, 102(4), 413–422.  
<http://dx.doi.org/10.1038/hdy.2009.4>
- Sourassou, N.F., Hanna, R., Breeuwer, J.A.J., Negloh, K., de Moraes, G.J. & Sabelis, M.W. (2014) The endosymbionts *Wolbachia* and *Cardinium* and their effects in three populations of the predatory mite *Neoseiulus paspalivorus*. *Experimental and Applied Acarology*, 64(2), 207–221.  
<http://dx.doi.org/10.1007/s10493-014-9820-0>
- Stahlhut, J.K., Desjardins, C.A., Clark, M.E., Baldo, L., Russell, J.A., Werren, J.H. & Jaenike, J. (2010) The mushroom habitat as an ecological arena for global exchange of *Wolbachia*. *Molecular Ecology*, 19(9), 1940–1952.  
<http://dx.doi.org/10.1111/j.1365-294X.2010.04572.x>
- Tamura, K., Stecher, G., Peterson, D., Filipski, A. & Kumar, S. (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, mst197.  
<http://dx.doi.org/10.1093/molbev/mst197>
- Weeks, A., Marec, F. & Breeuwer, J.A.J. (2001) A mite species that consists entirely of haploid females. *Science*, 29, 2479–2482.  
<http://dx.doi.org/10.1126/science.1060411>
- Weeks, A.R. & Breeuwer, J.A.J. (2003) A new bacterium from the Cytophaga-Flavobacterium-Bacteroides phylum that causes sex-ratio distortion. *Insect Symbiosis*, 165–176.  
<http://dx.doi.org/10.1201/9780203009918.ch11>
- Weeks, A.R. & Stouthamer, R. (2004) Increased fecundity associated with infection by a cytophaga-like intracellular bacterium in the predatory mite, *Metaseiulus occidentalis*. *Proceedings of the Royal Society of London B: Biological Sciences*, 271(Suppl 4), S193–S195.  
<http://dx.doi.org/10.1098/rsbl.2003.0137>
- Weeks, A.R., Velten, R. & Stouthamer, R. (2003) Incidence of a new sex-ratio-distorting endosymbiotic bacterium among arthropods. *Proceedings of the Royal Society of London B: Biological Sciences*, 270, 1857–1865.  
<http://dx.doi.org/10.1098/rspb.2003.2425>
- Weinert, L.A., Araujo-Jnr, E.V., Ahmed, M.Z. & Welch, J.J. (2015) The incidence of bacterial endosymbionts in terrestrial arthropods. *Proceedings of the Royal Society of London B: Biological Sciences*, 282(1807), 20150249.

- <http://dx.doi.org/10.1098/rspb.2015.0249>
- Weinert, L.A., Werren, J.H., Aebi, A., Stone, G.N. & Jiggins, F.M. (2009) Evolution and diversity of *Rickettsia* bacteria. *BMC Biology*, 7(1), 6.  
<http://dx.doi.org/10.1186/1741-7007-7-6>
- Werren, J.H., Baldo, L. & Clark, M.E. (2008) *Wolbachia*: master manipulators of invertebrate biology. *Nature Review Microbiology*, 6, 741–751.  
<http://dx.doi.org/10.1038/nrmicro1969>
- Wu, K. & Hoy, M.A. (2012) *Cardinium* is associated with reproductive incompatibility in the predatory mite *Metaseiulus occidentalis* (Acari: Phytoseiidae). *Journal of Invertebrate Pathology*, 110(3), 359–365.  
<http://dx.doi.org/10.1016/j.jip.2012.03.027>
- Xie, R.R., Zhou, L.L., Zhao, Z.J. & Hong, X.Y. (2010) Male age influences the strength of *Cardinium*-induced cytoplasmic incompatibility expression in the carmine spider mite *Tetranychus cinnabarinus*. *Applied Entomology and Zoology*, 45(3), 417–423.  
<http://dx.doi.org/10.1303/aez.2010.417>
- Zchori-Fein, E. & Perlman, S.J. (2004) Distribution of the bacterial symbiont *Cardinium* in arthropods. *Molecular Ecology*, 13, 2009–2016.  
<http://dx.doi.org/10.1111/j.1365-294X.2004.02203.x>
- Zchori-Fein, E., Gottlieb, Y., Kelly, S.E., Brown, J.K., Wilson, J.M., Karr, T.L. & Hunter, M.S. (2001) A newly discovered bacterium associated with parthenogenesis and a change in host selection behavior in parasitoid wasps. *Proceedings of the National Academy of Sciences*, 98(22), 12555–12560.  
<http://dx.doi.org/10.1073/pnas.221467498>
- Zhang, Z.Q. (2013) Phylum Arthropoda. In: Zhang, Z.Q. (Ed.) *Animal Biodiversity: An Outline of Higher-level Classification and Survey of Taxonomic Richness (Addenda 2013)*. *Zootaxa*, 3703, 17–26.
- Zhao, D.X., Chen, D.S., Ge, C., Gotoh, T. & Hong, X.Y. (2013a) Multiple infections with *Cardinium* and two strains of *Wolbachia* in the spider mite *Tetranychus phaselus* Ehara: revealing new forces driving the spread of *Wolbachia*. *PLoS ONE*, 8, e54964.  
<http://dx.doi.org/10.1371/journal.pone.0054964>
- Zhao, D.X., Zhang, X.F. & Hong, X.Y. (2013b) Host-symbiont interactions in spider mite *Tetranychus truncatus* doubly infected with *Wolbachia* and *Cardinium*. *Environmental Entomology*, 42(3), 445–452.  
<http://dx.doi.org/10.1603/EN12354>
- Zhu, L.Y., Jiang, X.Y., Yang, S.X., Xu, M. & Hong, X.Y. (2012a) Reproductive effects and localization of *Wolbachia* and *Cardinium* in the spider mite *Tetranychus piercei* (Acari: Tetranychidae). *Acta Entomologica Sinica*, 55(6), 684–693.
- Zhu, L.Y., Zhang, K.J., Zhang, Y.K., Ge, C., Gotoh, T. & Hong, X.Y. (2012b) *Wolbachia* strengthens *Cardinium*-induced cytoplasmic incompatibility in the spider mite *Tetranychus piercei* McGregor. *Current Microbiology*, 65, 516–523.  
<http://dx.doi.org/10.1007/s00284-012-0190-8>
- Zindell, R. (2012) *Mites and endosymbionts: towards improved biological control*. Université de Neuchâtel, Faculté des sciences.

Submitted: 3 Feb. 2016; accepted by Zhi-Qiang Zhang: 14 May 2016; published: 13 Jun. 2016