

Genetic Diversity and Structure of Syagrus romanzoffiana (Cham.) Glassman (Arecaceae) in Southern Brazil

Authors: Laindorf, Bruna L., de Freitas, Karine E. J., Lucini, Fabiola, Stefenon, Valdir M., Küster, Mariele C. T., et al.

Source: Tropical Conservation Science, 11(1)

Published By: SAGE Publishing

URL: https://doi.org/10.1177/1940082918798330

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <u>www.bioone.org/terms-of-use</u>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Genetic Diversity and Structure of Syagrus romanzoffiana (Cham.) Glassman (Arecaceae) in Southern Brazil

Tropical Conservation Science Volume 11: 1–10 © The Author(s) 2018 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/1940082918798330 journals.sagepub.com/home/trc



Bruna L. Laindorf¹, Karine E. J. de Freitas¹, Fabiola Lucini¹, Valdir M. Stefenon², Mariele C. T. Küster¹, Adriano L. Schünemann¹, Jair Putzke¹, Filipe C. Victória¹, and Antonio B. Pereira¹

Abstract

The species Syagrus romanzoffiana is a monocot belonging to the family Arecaceae; it is a palm endemic to South America and is widely distributed throughout the southeast and south of Brazil, including the State of Rio Grande do Sul. To estimate the genetic diversity and structure of the species in the watersheds of Rio Grande do Sul, five simple sequence repeat markers were used to analyze 90 individuals, representing three populations. The genetic structure of *S. romanzoffiana* groups was evaluated through the analysis of molecular variance and a multivariate analysis based on Nei's genetic distance. The evaluated populations showed significant intrapopulation molecular variation ($F_{ST} = 0.11$). The observed heterozygosity ($H_O = 0.17$) was lower than the expected heterozygosity ($H_E = 0.75$). These data indicate that although the number of individuals is high, the diversity is low for some indices. Our findings suggest that further studies are needed, particularly on the genetics of natural populations of *S. romanzoffiana* in Rio Grande do Sul State, to fully characterize their genetic diversity and structure and determine strategies and priority areas for species conservation.

Keywords

molecular ecology, molecular markers, SSR, population genetics, palm, queen palm, pindó

Introduction

The family Arecaceae is listed as having 2,700 species globally, distributed into 240 genera (Dransfield et al., 2008; Lorenzi, Larry, Noblick, Kahn, & Ferreira, 2010). Predominantly present in the tropics, 270 species belonging to 38 genera can be found in Brazil (Lorenzi et al., 2010); this diversity has made palm trees symbols of tropical forests by endemism, wealth, or abundance (Galetti, Pizo, & Morellato, 2003; Jones & Dransfield, 1995).

The species *Syagrus romanzoffiana* (Cham.) Glassman (Figure 1), popularly known as Coqueiro or Jerivá, is an important species of the Brazilian flora (Lorenzi et al., 2010). Endemic to South America, it is found in Brazil from the south of Bahia to southern Rio Grande do Sul (Noblick, 2017; Reitz, 1974). Present even in archaeological records, this palm tree is widely used by humans since their arrival on the continent as a source of food and fermented beverages, and in dwellings, handicrafts, pigments, fuel, and traditional

medicines (Bonomo & Capeletti, 2014; Zambrana et al., 2007). In addition, it is one of the most used ornamental plants in urban landscaping in southern and southeastern Brazil (Lorenzi et al., 2010; Reitz, 1974).

S. romanzoffiana is characterized by the presence of a single stipe, with a diameter ranging from 14 to 45 cm and a height of up to 25 m. Leaves are pinnate, with leaflets up to 25 cm long arranged irregularly on the rachis, giving them a feathery appearance.

¹Núcleo de Estudos da Vegetação Antártica, Universidade Federal do Pampa, São Gabriel, RS, Brazil
²Núcleo de Ecologia Molecular e Micropropagação de Plantas,

Universidade Federal do Pampa, São Gabriel, RS, Brazil Received 25 March 2018; Accepted 9 August 2018

Corresponding Author:

Adriano L. Schünemann, Núcleo de Estudos da Vegetação Antártica (NEVA), Universidade Federal do Pampa (UNIPAMPA), Av. Antônio Trilha 1847, São Gabriel, RS, CEP: 973000-000, Brazil. Email: als@unipampa.edu.br

Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (http://www.creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.



Figure I. Habit of the palm Jerivá (Syagrus romanzoffiana).

Inflorescence is an interfoliate panicle of spiky measuring between 80 and 120 cm that produces light-yellow pistillate or staminate flowers. Blossom and fruiting in this species extend predominantly from October to March, and its fruits are globose-oval drupe with fibrous flesh and an orange-yellow color. The species exhibits remarkable resistance to cold, enduring temperatures of up to -9° C which allows for its distribution in temperate climates. It is also considered hydrophilic owing to its high resistance to extremely wet environments (Curcio, De Sousa, Bonnet, & Barddal, 2007; Glassman, 1972; Goudel, Shibata, Coelho, & Miller, 2013; Lorenzi et al., 2010; Rambo, 1958; Reis, 2006; Reitz, 1974; Soares, Longhi, Witeck Neto, & Assis, 2014; Sobral, 2006).

S. romanzoffiana has a fundamental ecological role as an important source of food for several species of mammals, birds, and insects, mainly related to its nutritional characteristics and wide spectrum of fruiting. In addition, this species constitutes an important component of natural ecosystems through interactions with its dispersers and pollinators (Carvalho, 2006; Galetti et al., 2013; Souza et al., 1993; Siqueira, 1989; Terborgh, 1986; Zona & Henderson, 1989).

S. romanzoffiana is present throughout the State of Rio Grande do Sul, appearing in different forest habitats, both in the Pampa and Atlantic Forest biomes, predominantly along watercourses (Soares et al., 2014). The presence of this palm tree in areas of the periodic flooding was highlighted by Rambo (1958), and Leite and Klein (1990).

Clarifying genetic diversity of a species is essential to understanding its ecological characteristics such as the delimitations of groupings and the composition of populations. These factors are fundamental to the establishment of conservation strategies and determination of high-diversity priority areas for conservation (Torggler, 1995; Vinson, Kanashiro, Harris, & Boshier, 2015; Yeeh, Kang, & Chung, 1996).

In general, microsatellite markers provide essential information for estimating genetic parameters of interest in population genetics (Ferreira & Grattapaglia, 1998; Gupta et al., 2003); these are also known as simple sequence repeats (SSRs), which are short DNA sequences of 1 to 6 base pairs repeated in tandem that can be detected by polymerase chain reaction (PCR) using specific primers (Litt & Luty, 1989; Stefenon, 2010; J. L. Weber & May, 1989; Zucchi et al., 2003). Allelic and genotypic information obtained from these markers is useful for understanding the genetic diversity, genetic structure, and phylogeographic patterns of natural populations (e.g., Stefenon, Gailing, & Finkeldey, 2007), aiding in formulating conservation strategies for genetic resources (Gao, Schaal, Zhang, Jia, & Dong, 2002; Powell, Machray, & Provan, 1996; Victoria, Da Maia, & De Oliveira, 2011).

On the basis of the characteristics related to the occurrence of a species within an environment and the molecular analysis tools currently available, ecological studies on the relationship between a watershed and the molecular variability of a given species may be useful for inferring if hydrographic basins are limiting for species distribution. The use of the watershed approach intrinsically simulates the space, that is, the patterns and processes occurring within a spatial unit called the watershed (Schiavetti & Camargo, 2002). In the State of Rio Grande do Sul, *S. romanzoffiana* is found within three main watersheds, namely Guaíba, Litoral, and Uruguai; therefore, it may be a valuable model for testing this hypothesis. On the basis of these premises, this study aimed to evaluate the genetic diversity and structure of three populations of *S. romanzoffiana* in Rio Grande do Sul State, each growing within one of the three watersheds, to determine the influence of the hydrographic basin on the allelic distribution of this species.

Material and Methods

Study Area

This study was carried out in the State of Rio Grande do Sul in the southernmost region of Brazil. The state covers an area of approximately 281,748 km², with an altitude varying from sea level up to 1,398 m, and a climate classified predominantly as Cfa, with a restricted portion being classifies as Cfb (Koppen–Geiger). It comprises two distinct biomes, the Atlantic Forest and Pampa (Kuinchtner & Buriol, 2001; E. Weber et al., 1998).

In this study, the three watersheds dividing the territory of the Rio Grande do Sul State were used as limiting factors for determining the populations of S. romanzoffiana. The Litoral watershed consists of the easternmost and southernmost regions of the state, presenting altitudes from 0 to 100 m above sea level (ASL) and covering mainly the geomorphological province of Planície Costeira. The Guaíba watershed is located in the central-eastern part of the state, with altitudes ranging between 100 and 800 m ASL and covering the geomorphological regions of the Southern Plateau and Central Depression. The Uruguai watershed covers the extreme north and western regions of the state, with altitudes ranging between 0 and 1,200 m ASL and corresponds to the geomorphological unit of the Southern Plateau.

Sample DNA Isolation and PCR Essays

Foliar samples from 90 adult individuals of *S. romanzoffiana* (30 plants each watershed; Figure 2) were obtained on expeditions performed between March and December 2016. All specimens sampled were georeferenced using a Gamin 26 GPS; leaflets were identified with a reference number and then stored in an ultrafreezer at -80° C.

Total DNA was isolated from the sampled leaflets with a Qiagen Plant MiniKit[®], following the manufacturers' instructions. Quantity and quality of the isolated DNA were evaluated in a NanoVueTM Plus

spectrophotometer. The PCR conditions were adapted from Geethanjali, Rukmani, Rajakumar, Kadirvel, and Viswanathan (2017). Five molecular markers developed for Livistona chinensis (Jack.) R. Br. (Ohtani, Tani, & Yoshimaru, 2009), Cocos nucifera L. (Teulat et al., 2000), and *Phoenix dactylifera* L. (Elshibli & Korpelainen, 2008; Table 1) were used for genotyping the 90 samples of S. romanzoffiana, using 2 µL of DNA, 1 µL of oligonucleotide initiators, continuous and reverse; 1.5 μ L of Buffer (10×); 1 μ L MgCl₂ (5 U/ μ L); deoxyribonucleotide triphosphates 1 μ L (100 mM); 0.2 μ L Tag Polymerase buffer (5 U/ μ L), and 7.3 μ L of ultrapure water (Milli-Q) for a final volume of 20 μ L. Amplification was performed in a BIO-RAD C1000 TouchTM thermocycler, with an initial denaturation step of 94°C for 5 min, followed by 35 cycles of 94°C for 30 s for denaturation, 1 min at 51°C for annealing, 1 min extension at 72°C, and a final extension at 72°C for 10 min.

The PCR products were resolved on a 3% agarose gel at 80 V, stained with GelRedTM (Biotium[®]) and visualized under ultraviolet light. A 100-base pair marker ladder (Norgen) was used for sizing the alleles. Images were generated with the LPix EX-2.6-PGR program (Locus Biotecnologia) and fragment size was determined using the TotalLab TL120 gel analysis program.

Data Analysis

Total number of alleles (A), effective number of alleles (A_e) , observed heterozygosity (H_O) , expected heterozygosity (H_E) , and inbreeding coefficient, $F = (H_E - H_O)/H_E$, were estimated for each locus and multiloci for each population. The genetic structure of *S. romanzoffiana* populations were evaluated through the analysis of molecular variance (AMOVA; Excoffier, 1993) and a multivariate analysis (PCoA) based on Nei's (1978) genetic distance. All analysis were performed using the GenAlEx 6.4 software (Peakall & Smouse, 2012).

Results

Levels of Genetic Diversity

Overall, the five loci evaluated in the 90 individuals of *S. romanzoffiana* were polymorphic and generated 82 alleles. Locus MSr27 comprised the highest number of alleles (22), while the smallest number (14 alleles) was observed for the MSr01 and MSr02 loci.

At the population level, all loci were polymorphic in the Guaíba and Uruguai populations, whereas locus MSr02 was monomorphic and presented an elevated level of missing data in the Litoral population. The number of alleles per population ranged from 40 in the Guaíba population to 28 in the Litoral population

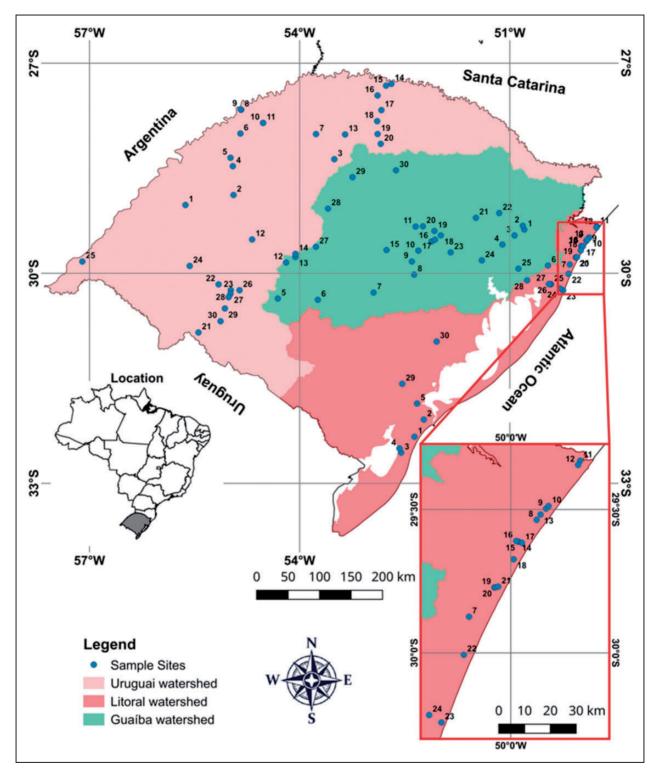


Figure 2. Map of the State of Rio Grande do Sul, with details of the three watersheds: Guaíba (Green), Litoral (Red), and Uruguai (Pink). The blue dots indicate the sample sites.

(Table 2). Private alleles were found in all populations: five in Guaíba, three in Litoral, and three in Uruguai.

Multilocus estimations of observed heterozygosity ranged from $H_O = 0.10$ (Guaíba population) to

 $H_O = 0.28$ (Uruguai population), while estimations of expected heterozygosity ranged from $H_E = 0.59$ (Litoral population) to $H_E = 0.83$ (Guaíba population). Consequently, estimations of fixation index were

Marker	Sequence	Product size (pb) 206–290	
MSr01ª	F:AACTGCAGGAACAAAGACGATT		
	R:TCACGTCCACGTATTCGTACTC		
MSr02 ^ª	F:CATGGAATTGTAATCCCCACTT	170–230	
	R:TATCCACTTGTCGGAGTTTTCC		
MSr03 ^ª	F:CATAGGCAGTCACAGATGGTTT	240–296	
	R:CCAGCTAATTTCCCCTTCTTT		
MSr27 ^b	F:AGCAAGGTCAGCGAA	220–300	
	R:ACGAGAAGGGAACAAAC		
MSr28 ^c	F: ATTCATTATTCAACACCAAC	78–124	
	R:GGTCTCTCTCTCTCTCTC		

 Table I. SSR Markers Used in This Study.

Note. SSR = simple sequence repeat.

^aLivistona chinensis (Jack) B.Br.

^bCocos nucifera L.

^cPhoenix dactylifera L.

Table 2. Genetic Diversity Parameters Estimated for Syagrus romanzoffiana in the Rio Grande doSul watersheds.

Population	%Polym.	Locus	А	Ae	Ho	H _E	F
Guaíba	100%	MSr01	8.00	4.17	0.00	0.77	1.00
		MSr02	6.00	3.92	0.10	0.78	0.87
		Msr03	12.00	8.23	0.42	0.92	0.53
		MSr27	8.00	5.92	0.00	0.84	1.00
		MSr28	8.00	5.70	0.00	0.82	1.00
		Multiloci	8.40	5.59	0.10	0.83	0.88
Litoral	80%	MSr01	6.00	4.91	0.00	0.82	1.00
		MSr02	1.00	1.00	0.00	0.00	-
		Msr03	6.00	3.27	0.50	0.76	0.28
		MSr27	9.00	3.73	0.03	0.75	0.95
		MSr28	6.00	2.74	0.14	0.75	0.78
		Multiloci	5.60	3.13	0.13	0.59	0.75
Uruguai	100%	MSr01	9.00	6.25	0.00	0.68	1.00
Ū		MSr02	8.00	8.00	1.00	1.00	-0.14
		Msr03	5.00	2.98	0.40	0.70	0.40
		MSr27	9.00	6.54	0.00	0.86	1.00
		MSr28	6.00	3.08	0.00	0.69	1.00
		Multiloci	7.40	5.37	0.28	0.82	0.65

%Polym. = percentage of polymorphism per population; A = number of alleles per locus; Ae = effective number of alleles; $H_o =$ observed heterozygosity; $H_E =$ expected heterozygosity; F = fixation index.

relatively high, ranging from F = 0.65 (Uruguai population) to F = 0.88 (Guaíba population), indicating a high level of inbreeding (Table 2).

Population Genetic Structure

The AMOVA based on the five polymorphic loci revealed that 11% of the observed genetic variability was distributed among populations, 82% among individuals within the populations, and 7% among individuals (Table 3). Estimations of *F* statistics (Weir &

Cockerham, 1984) were high and statistically significant (p < .001) among populations ($F_{ST} = 0.110$), within populations ($F_{IS} = 0.925$), and overall ($F_{IT} = 0.933$).

The estimated gene flow based on pairwise F_{ST} ranged from Nm = 1.73 to Nm = 2.34 migrants per generation. As expected, these values are directly correlated to the estimations of Nei's genetic distance (Table 4).

The PCoA of the individuals of the three populations of *S. romanzoffiana*, revealed an overlap between populations (Figure 3). However, the individual assignment analysis revealed a clear population signature. Among the 90 genotyped individuals, only 6 presented tendencies to belong to a genetic population different from that of origin (Table 5).

Discussion

There are several ways to explain the current distribution pattern of a species. Although *S. romanzoffiana* is not considered a threatened species, the populations evaluated in this study presented moderate levels of allelic diversity, but markedly low levels of H_O . Habitat fragmentation and opening of space for agriculture and urbanization, with consequent loss of dispersers and

 Table 3. AMOVA for Populations of Syagrus romanzoffiana in Rio

 Grande do Sul.

Source	df	SS	MS	VT%
Among pops	2	30.31	30.31	11
Among indiv	87	271.30	6.24	82
Within indiv	90	11.00	0.12	7
Total	179	312.61		100

Note. $F_{ST} = 0.110$ (p < .001), $F_{IS} = 0.925$ (p < .001), $F_{IT} = 0.933$ (p < .001). df = degrees of freedom; SS = sum of squares; MS = mean squares; VT% = total variance; F_{ST} = genetic divergence among populations.

Table 4. Number of Migrants (Below Diagonal) Between Populations Per Generation, and Nei's Genetic Distance (Above Diagonal).

	Guaíba	Litoral	Uruguai
Guaíba	-	1.05	1.27
Litoral	1.73	-	1.09
Uruguai	2.34	1.81	_

pollinators in the Pampa and Atlantic Forest biomes, threatens the viability of the species mainly because of loss of genetic diversity (Murcia, 1995; Roesch et al., 2009; Wright & Duber, 2001; Wright et al., 2000).

The low levels of H_O estimated in this study may be an effect of the transferability of the employed SSR loci. Significantly lower estimations of H_O (0.148) in relation to H_E (0.695) were observed in *Acromia emensis* (Tol.) Lorenzi (Arecaceae) in an analysis based on SSR markers transferred from other species (Neiva et al., 2016). In addition, lower observed heterozygosity ($H_O = 0.164$) was observed in comparison to expected heterozygosity ($H_E = 0.262$) in *Elaeis oleifera* (Kunth) Cortés (Arecaceae) genotyped with species-specific SSR markers (Zaki, Singh, Rosli, & Ismail, 2012).

This low level of H_O is detrimental to a species as a low input from heterozygotes makes populations vulnerable to environmental changes, limiting the ability to respond and adapt (Avise, Bowen, Lamb, Meylan, &

Table 5. Assignment Analysis of Individual Samples.

Sample	Origin population	Guaíba	Litoral	Uruguai	Assigned population
24	Guaíba	-6.64	- 6.63	-12.00	Litoral
25	Guaíba	-7.76	-9.56	- 6.64	Uruguai
35	Litoral	- 3.69	-4.12	-6.75	Guaíba
40	Litoral	-6.77	-7.82	-10.45	Guaíba
67	Uruguai	-4.74	-4.95	-5.95	Guaíba
78	Uruguai	-9.20	-9.56	-10.72	Guaíba

Note. Only samples assigned to a population different from the population origin are presented.

The numbers in bold represents the genotyoped individuals that presented tendencies to belong to a genetic population diffrent from the original are sampled (p>0,01).

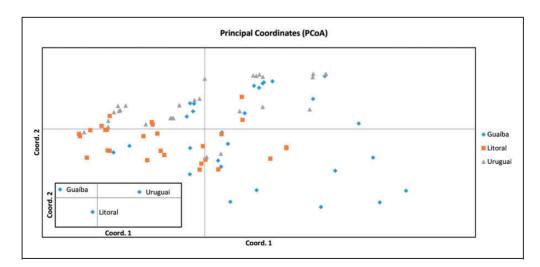


Figure 3. Analysis of Principal Components for individuals of three populations of Syagrus romanzoffiana in Rio Grande do Sul.

Bermingham, 1992). Excessive inbreeding resulting either from habitat fragmentation or a species characteristic, leads to significant loss of diversity by a decrease or isolation of a population; to redress this balance, a large input of individuals over a long period of time is needed as inclusion of an allele occurs by random crossing (Hamrick, Godt, & Sherman-Broyles, 1992).

Palynological studies in the study area have indicated that *S. romanzoffiana* emerged approximately 940 radiocarbon years ago and is associated with the emergence of ciliary vegetation. In the same period, the forest diversity of the gallery to the southeast of South America expanded exponentially (Mourelle, Prieto, & García-Rodríguez, 2017).

In a study of communities of palm trees in the western Amazon, Kristiansen et al. (2012) verified that there was a correlation between species distribution and the flood environment. Consequently, it was possible to classify palm trees according to hydrology, in addition to the dispersion environment where geographical distance was the main limiting factor. In addition, events such as flood times and resurgence of land areas (Crisp, Trewick, & Cook, 2011), which determined the distribution pattern of the hydrophilic terrace, may be related to the current distribution pattern of the palm.

Factors contributing to these population dynamics may be explained by relationships with pollinators. According to Nogueira-Neto (2002), pollination of *S. romanzoffiana* is carried out by *Apis melífera*, several species of the tribe Meliponini, and Jataí bees attracted by its flowers containing abundant pollen, in addition to species of Coleoptera that reproduce in the flowers of the palm tree and consequently contribute to dispersal. Moreover, large mammals are also important dispersers as they normally occupy large areas. In this sense, the extent of dispersal may also contribute to the spatial organization of *S. romanzoffiana* populations (Budke, Athayde, Giehl, Záchia, & Eisinger, 2005; Freire, Closel, Hasui, & Ramos, 2013; Giombini, Bravo, & Martíinez, 2009; Messias & De Assis Alves, 2009).

Adjacent area studies by Terral et al. (2012) suggested that routes of human dispersion may have contributed to the geographical distribution pattern of *P. dactylifera* L. (Arecaceae), as the indigenous populations made daily use of the palm. Human dispersion may also have contributed to the distribution of *S. romanzoffiana* as indigenous populations in South America traveled long distances along riverbeds and may have transported the seeds; indeed, several archaeological studies have reported palm use by indigenous groups in food, house covering, and traditional medicines (Bonomo & Capeletti, 2014). The ornamental use of this species in urban areas such as in roads, squares, and gardens of southern Brazil (Lorenzi et al., 2010; Reitz, 1974) may also affect its natural organization, although to a lesser extent.

The level of genetic divergence among populations estimated as F_{ST} =0.11 indicates moderate differentiation (Slatkin, 1987). Interpopulation gene flow is important for avoiding significant genetic distinctions within populations because they would be subject to evolutionary pressures, endogamy, and genetic drift (Nagel & Stefenon, 2013; Serrote, Reiniger, & Stefanon, 2016; Slatkin, 1987). Considering that 11% of the observed differentiation occurred among populations, as well as the significant geographical distribution of plants along the three watersheds, we propose that gene flow is efficient among populations. Nevertheless, approximately 84% of the plants had a genetic identity different to their population of origin, resulting from differences in allelic frequencies among populations and the presence of private alleles.

Finally, the results of this study were inconclusive in relation to the influence of the hydrographic basin on the distribution of alleles. A larger number of molecular markers are needed to elucidate this issue. However, our results shed some light with regard to conservation as they provided the first estimate of genetic diversity in the S. romanzoffiana palm. Although the risk of extinction is low, according to the Red List-International Union for Conservation of Nature (Govaerts & Dransfield, 2005), our analysis showed low levels of H_0 and extremely high inbreeding in the evaluated populations. This outcome suggests that further studies on the genetic characteristics of natural populations of S. romanzoffiana in Rio Grande do Sul State are needed to fully characterize their genetic diversity and structure, and to determine strategies and priority areas for species conservation.

Implications for Conservation

This study suggests that the assessment criteria used to determine the vulnerability this species may not reflect the true state of conservation; a large population may not guarantee the survival of *S. romanzoffiana* as our results demonstrated the presence of low diversity.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work had the support of the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior—CAPES, the Conselho Nacional de Desenvolvimento Científico—CNPq (grant number 446234/2015-0), and from the Universidade Federal do Pampa—UNIPAMPA.

ORCID iD

Adriano L. Schünemann D http://orcid.org/0000-0001-7227-7074

Filipe C. Victória (http://orcid.org/0000-0002-8580-1813

References

- Avise, J. C., Bowen, B. W., Lamb, T., Meylan, A. B., & Bermingham, E. (1992). Mitochondrial DNA evolution at a turtle's pace: Evidence for low genetic variability and reduced microevolutionary rate in the testudines. *Molecular Biology and Evolution*, 9(3), 457–473.
- Bonomo, M., & Capeletti, L. E. (2014). Uso prehispánico de las palmeras Syagrus romanzoffiana y Butia yatay en el Nordeste argentino: Aportes desde la etnografía y la biometría [Prehispanic use of the palm trees Syagrus romanzoffiana and Butia yatay in the northeast of Argentina: Contributions of ethnography and biometrics]. Revista Del Museo de Antropología, 7(2), 227–234.
- Budke, J. C., Athayde, E. A., Giehl, E. L. H., Záchia, R. A., & Eisinger, S. M. (2005). Composição florística e estratégias de dispersão de espécies lenhosas em uma floresta ribeirinha, arroio Passo das Tropas, Santa Maria, RS, Brasil [Floristic composition and strategies of dispersion of woody species in a riverine forest, Passo das Tropas stream, Santa Maria, RS, Brazil]. *Iheringia. Série Botânica*, 60(1), 17–24.
- Carvalho, P. E. R. (2006). Espécies arbóreas brasileiras [Brazilian tree species.] (Vol. 1). Distrito Federal, Brasília: Embrapa Informação Tecnológica.
- Crisp, M. D., Trewick, S. A., & Cook, L. G. (2011). Hypothesis testing in biogeography. *Trends in Ecology & Evolution*, 26(2), 66–72.
- Curcio, G. R., de Sousa, L. P., Bonnet, A., & Barddal, M. L. (2007). Recomendação de espécies arbóreas nativas, por tipo de solo, para recuperação ambiental das margens da Represa do rio Iraí, Pinhais, PR [Recommendation of native tree species, by soil type, for environmental recovery of the banks of the Rio Iraí dam, Pinhais, PR]. *Floresta*, 37(1), 113–122. Retrieved from https://revistas.ufpr.br/flor esta/article/view/7847/5539
- Dransfield, J., Uhl, N. W., Asmussen, C. B., Baker, W. J., Harley, M. M., & Lewis, C. E. G. (2008). *Genera Palmarum: The evolution and classification of palms* (2nd ed.). Richmond, England: Royal Botanic Gardens.
- Elshibli, S., & Korpelainen, H. (2008). Microsatellite markers reveal high genetic diversity in date palm (*Phoenix dactylifera* L.) germplasm from Sudan. *Genetica*, 134(2), 251–260.
- Excoffier, L. (1993). Analysis of molecular variance (AMOVA) (Version 1.55) [Computer software]. Geneva, Switzerland: Genetics and Biometry Laboratory, University of Geneva.
- Ferreira, M. E., & Grattapaglia, D. (1998). *Introducción al uso de marcadores moleculares en el análisis genético*. Federal District, Brazil: Embrapa.

- Freire, C. C., Closel, M. B., Hasui, E., & Ramos, F. N. (2013). Reproductive phenology, seed dispersal and seed predation in *Syagrus romanzoffiana* in a highly fragmented landscape. *Annales Botanici Fennici*, 50(4), 220–228.
- Galetti, M., Guevara, R., Côrtes, M. C., Fadini, R., Von Matter, S., Leite, A. B., & Pires, M. M. (2013). Functional extinction of birds drives rapid evolutionary changes in seed size. *Science*, 340(6136), 1086–1090.
- Galetti, M., Pizo, M. A., & Morellato, P. C. (2003). Fenologia, frugivoria e dispersão de sementes [Phenology, frugivory and seed dispersal]. In L. Cullen, C. Rudram, & C. Padua (Eds), Métodos de estudos em biologia da conservação e manejo da vida Silvestre (pp. 395–422). Corumbá, Brazil: Fundação O Boticário de Proteção à Natureza.
- Gao, L. Z., Schaal, B., Zhang, C. H., Jia, J. Z., & Dong, Y. S. (2002). Assessment of population genetic structure in common wild rice *Oryza rufipogon* Griff. using microsatellite and allozyme markers. *TAG Theoretical and Applied Genetics*, 106(1), 173–180.
- Geethanjali, S., Rukmani, J. A., Rajakumar, D., Kadirvel, P., & Viswanathan, P. L. (2017). Genetic diversity, population structure and association analysis in coconut (Cocos nucifera L.) germplasm using SSR markers. *Plant Genetic Resources*, 16(2), 1–13.
- Giombini, M. L., Bravo, S. P., & Martíinez, M. F. (2009). Seed dispersal of the palm *Syagrus romanzoffiana* by tapirs in the semi-deciduous Atlantic forest of Argentina. *Biotropica*, 41(4), 408–413.
- Glassman, S. (1972). Studies in the palm genus Syagrus Mart. American Journal of Botany, 363–400.
- Goudel, F., Shibata, M., Coelho, C. M. M., & Miller, P. R. M. (2013). Fruit biometry and seed germination of *Syagrus romanzoffiana* (Cham.) Glassm. *Acta Botanica Brasilica*, 27(1), 147–154.
- Govaerts, R., & Dransfield, J. (2005). World checklist of palms. *Royal Botanic Gardens*, 14(1), 1–223. Retrieved from https://www.cabdirect.org/cabdirect/abstract/ 20053205919
- Gupta, P. K., Rustgi, S., Sharma, S., Singh, R., Kumar, N., & Balyan, H. S. (2003). Transferable EST-SSR markers for the study of polymorphism and genetic diversity in bread wheat. *Molecular Genetics and Genomics*, 270(4), 315–323.
- Hamrick, J. L., Godt, M. J. W., & Sherman-Broyles, S. L. (1992). Factors influencing levels of genetic diversity in woody plant species. *New Forests*, 6(1), 95–124.
- Jones, D. L., & Dransfield, J. (1995). Palms throughout the world. Smithsonian, 410, 31.
- Kristiansen, T., Svenning, J. C., Eiserhardt, W. L., Pedersen, D., Brix, H., Munch Kristiansen, S., & Balslev, H. (2012). Environment versus dispersal in the assembly of western Amazonian palm communities. *Journal of Biogeography*, 39(7), 1318–1332.
- Kuinchtner, A., & Buriol, G. A. (2001). The climate of the Rio Grande do Sul State according to the Köppen e Thornthwaite systems. *Disciplinarum Scientia*, 2, 171–182.
- Leite, P. F., & Klein, R. M. (1990). Vegetação [Vegetation]. Geografia Do Brasil: Região Sul, 2, 113–150.
- Litt, M., & Luty, J. A. (1989). A hypervariable microsatellite revealed by in vitro amplification of a dinucleotide repeat

within the cardiac muscle actin gene. *American Journal of Human Genetics*, 44(3), 398–401.

- Lorenzi, H., Larry, R., Noblick, Kahn, F., & Ferreira, E. (2010). *Brazilian flora Lorenzi: Arecaceae (Palms)* [Brazilian flora: Arecaceae (Palms)]. Nova Odessa, Brazil: Instituto Plantarum de Estudos da Flora.
- Messias, A. D., & de Assis Alves, F. (2009). Jerivá (Syagrus romanzoffiana–Arecaceae) como oferta de alimento para fauna silvestre em fragmentos de mata ciliar, em período de outono-inverno [Jerivá (Syagrus romanzoffiana-Arecaceae) as food supply for wildlife in fragments of ciliary forest, in autumn–winter period]. Revista Eletrônica de Biologia (REB), 2(1), 36–50.
- Mourelle, D., Prieto, A. R., & García-Rodríguez, F. (2017). Riparian woody vegetation history in the campos region, southeastern South America, during two time windows: Late Pleistocene and late Holocene. *Quaternary Science Reviews*, 167, 14–29.
- Murcia, C. (1995). Edge effects in fragmented forests: Implications for conservation. *Trends in Ecology & Evolution*, 10(2), 58–62.
- Nagel, J. C., & Stefenon, V. M. (2016). Gestão da Espécie Açoita Cavalo no Bioma Pampa: Diversidade e Estrutura Genética Populacional [Species management Açoita cavalo in the Pampa biome: Diversity and genetic population structure.]. Novas Edições Acadêmicas, 1(59), 1–64.
- Nei, M. (1987). *Molecular evolutionary genetics*. New York, NY: Columbia University Press.
- Neiva, D. S., Júnior, A. M., Oliveira, D. A., Royo, V. A., Brandão, M. M., & Menezes, E. V. (2016). Acrocomia emensis (Arecaceae) genetic structure and diversity using SSR molecular markers. Genetics and Molecular Research, 15(1), 1–11.
- Noblick, L. R. (2017). A revision of the genus Syagrus (Arecaceae). Phytotaxa, 294(1), 1–262.
- Nogueira-Neto, P. (2002). Inbreeding and building up small populations of stingless bees (*Hymenoptera, Apidae*). *Revista Brasileira de Zoologia*, 19(4), 1181–1214.
- Ohtani, M., Tani, N., & Yoshimaru, H. (2009). Isolation of polymorphic microsatellite loci in *Livistona chinensis* (Jacq.)
 R. Br. ex Mart. var. *boninensis* Becc. an endemic palm species of the oceanic Bonin Islands, Japan. *Conservation Genetics*, 10(4), 997–999.
- Peakall, R., & Smouse, R. (2012). GenAlEx 6.5: Genetic analysis in excel. Population genetic software for teaching and research an update. *Bioinformatics*, 28, 2537–2539.
- Powell, W., Machray, G. C., & Provan, J. (1996). Polymorphism revealed by simple sequence repeats. *Trends Plant Science*, 1, 215–222.
- Rambo, B. (1958). A fisionomia do Rio Grande do Sul: Ensaio de monografia natural [The physiognomy of Rio Grande do Sul: Essay of natural monograph]. *Livraria Selbach*, 71, 1–6.
- Reis, R. C. C. (2006). Palmeiras (*Arecaceae*) das restingas do Estado do Rio de Janeiro, Brasil [Palm trees (*Arecaceae*) from the restingas of the Rio de Janeiro, State, Brazil]. *Acta Botânica Brasilica*, 20(3), 501–512.
- Reitz, R. (1974). Flora Ilustrada Catarinense: Palmeiras (parte I). Herbário Barbosa Rodrigues, Itajaí. 189(1), 143–150.
- Roesch, L. F. W., Vieira, F. C. B., Pereira, V. A., Schünemann, A. L., Teixeira, I. F., Senna, A. J. T., & Stefenon, V. M.

(2009). The Brazilian Pampa: A fragile biome. *Diversity*, *1*(2), 182–198.

- Schiavetti, A., & Camargo, A. F. (2002). Conceitos de bacias hidrográficas: teorias e aplicações [Watershed concepts: theories and applications]. EUSC: Editus–Ilhéus. 289 pp.
- Serrote, C. M. L., Reiniger, L. R. S., & Stefenon, V. M. (2016). Simulações em Genética de Populações e Conservação de Recursos Florestais [Simulations in genetics of populations and conservation of forest resources] (Vol. 1, pp. 52–56). Jundiaí, Brazil: Paco Editorial.
- Siqueira, J. C. (1989). Considerações sobre a biologia da polinização em palmeira gerivá (*Arecastrum romanzoffianum* - Palmae) [Considerations on the biology of pollination in gerivá palm (*Arecastrum romanzoffiana* – Palmae)]. *Acta Biologica Leopoldensia*, 11, 203–212.
- Slatkin, M. (1987). Gene flow and the geographic structure of natural populations. *Science (Washington)*, 236(4803), 787–792.
- Soares, K. P., Longhi, S. J., Witeck Neto, L., & Assis, L. C. D. (2014). Palms (*Arecaceae*) from Rio Grande do Sul, Brazil. *Rodriguésia*, 65(1), 113–139.
- Sobral, M. (2006). Flora arbórea e arborescente do Rio Grande do Sul, Brasil [Tree and arborescent flora of Rio Grande do Sul, Brazil]. *RiMa*, *8*, 1–86.
- Souza, V. C., Cortopassi-Laurino, M., Simão-Bianchini, R., Pirani, J. R., Azoubel, M. L., Guibu, L. S., & Cortopassi-Laurino, M. (1993). Plantas apícolas de São Paulo e arredores [Apiculture plants of São Paulo and surroundings]. In R. L. Pirani & M. Cortopassi-Laurino (Eds), *Flores e abelhas em São Paulo* (pp. 43–179). São Paulo, Brazil: EDUSP/FAPESP.
- Stefenon, V. M. (2010). Methodological aspects of gene flow estimation in natural populations. In K. V. Urbano (Ed.). *Advances in genetic research* (Vol. 3, 1st ed., pp. 264–277). New York, NY: Nova Science Publisher.
- Stefenon, V. M., Gailing, O., & Finkeldey, R. (2007). Genetic structure of *Araucaria angustifolia* (Araucariaceae) populations in Brazil: Implications for the in situ conservation of genetic resources. *Plant Biology*, 9(4), 516–525.
- Terborgh, J. (1986). Community aspects of frugivory in tropical forests. In A. Estrada & T. H. Fleming (Eds), *Frugivores* and seed dispersal (pp. 371–384). Dordrecht, the Netherlands: Springer.
- Terral, J. F., Newton, C., Ivorra, S., Gros-Balthazard, M., de Morais, C. T., . . . Pintaud, J. C. (2012). Insights into the historical biogeography of the date palm (*Phoenix dactylifera* L.) using geometric morphometry of modern and ancient seeds. *Journal of Biogeography*, 39(5), 929–941.
- Teulat, B., Aldam, C., Trehin, R., Lebrun, P., Barker, J. H., Arnold, G. M., & Rognon, F. (2000). An analysis of genetic diversity in coconut (*Cocos nucifera*) populations from across the geographic range using sequence-tagged microsatellites (SSRs) and AFLPs. *Theoretical and Applied Genetics*, 100(5), 764–771.
- Torggler, M. G. F. (1995). *Isoenzimas variabilidade genética em plantas* [Isoenzymes genetic variability in plants] (Vol. 39, Chap. 5, 175 pp.). Ribeirao Preto, Brazil: Sociedade Brasileira de Genética.

- Victoria, F. C., da Maia, L. C., & de Oliveira, A. C. (2011). In silico comparative analysis of SSR markers in plants. *BMC Plant Biology*, 11(1), 15.
- Vinson, C. C., Kanashiro, M., Harris, S. A., & Boshier, D. H. (2015). Impacts of selective logging on inbreeding and gene flow in two Amazonian timber species with contrasting ecological and reproductive characteristics. *Molecular Ecology*, 24(1), 38–53.
- Weber, E., Duarte, G. F., Frank, M., Hoff, R., Zomer, S., Bassani, E., & Junqueira, I. (1998). *Estruturação de sistemas de informação ambiental em bacias hidrográficas: O caso da bacia hidrográfica do rio Caí–RS* [Structuring of environmental information systems in watershed: The case of the Caí-RS watershed]. Retrieved from http://www.ecologia. ufrgs.br/labgeo/arquivos/Publicacoes/Congressos/1998/ Weber et al 1998 Estruturacao SIG bacia rio Cai.pdf.
- Weber, J. L., & May, P. E. (1989). Abundant class of human DNA polymorphisms which can be typed using the polymerase chain reaction. *American Journal of Human Genetics*, 44(3), 388.
- Weir, B. S., & Cockerham, C. C. (1984). Estimating F-statistics for the analysis of population structure. *Evolution*, 38(6), 1358–1370.
- Wright, S. J., & Duber, H. C. (2001). Poachers and forest fragmentation alter seed dispersal, seed survival, and seedling

recruitment in the palm *Attalea butyraceae*, with implications for tropical tree diversity. *Biotropica*, 33(4), 583–595.

- Wright, S. J., Zeballos, H., Domínguez, I., Gallardo, M. M., Moreno, M. C., & Ibáñez, R. (2000). Poachers alter mammal abundance, seed dispersal, and seed predation in a Neotropical forest. *Conservation Biology*, 14(1), 227–239.
- Yeeh, Y., Kang, S. S., & Chung, M. G. (1996). Evaluations of the natural monument populations of *Camellia japonica* (Theaceae) in Korea based on allozyme studies. *Botanical Bulletin of Academia Sinica*, 37(1), 141–146.
- Zaki, N. M., Singh, R., Rosli, R., & Ismail, I. (2012). Elaeis oleifera genomic-SSR markers: Exploitation in oil palm germplasm diversity and cross-amplification in Arecaceae. International Journal of Molecular Sciences, 13(4), 4069–4088.
- Zambrana, N. Y. P., Byg, A., Svenning, C. C., Moraes, M., Grandez, C., & Balsley, H. (2007). Diversity of palm uses in the western Amazon. *Biodiversity and Conservation*, *16*(10), 2771–2787.
- Zona, S., & Henderson, A. (1989). A review of animalmediated seed dispersal of palms. *Selbyana*, 11, 6–21.
- Zucchi, M. I., Brondani, R. P. V., Pinheiro, J. B., Chaves, L. J., Coelho, A. S. G., & Vencovsky, R. (2003). Genetic structure and gene flow in *Eugenia dysenterica* DC in the Brazilian Cerrado utilizing SSR markers. *Genetics and Molecular Biology*, 26(4), 449–457.