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Bartonella henselae as a cause of acute-onset febrile illness in cats

Edward B Breitschwerdt[1], Jack J Broadhurst[2] and Natalie A Cherry[3]

Abstract

Case series summary At different time points spanning 6 months, three adopted feral flea-infested cats, residing in the household of a veterinary technician, became acutely anorexic, lethargic and febrile. Enrichment blood culture/PCR using *Bartonella* alpha Proteobacteria growth medium (BAPGM) confirmed initial infection with the same *Bartonella henselae* genotype in all three cases. With the exception of anemia and neutropenia, complete blood counts, serum biochemical profiles and urinalysis results were within reference intervals. Also, tests for feline leukemia virus, feline immunodeficiency virus, *Toxoplasma gondii* and feline coronavirus antibodies were negative. Serial daily temperature monitoring in one case confirmed a cyclic, relapsing febrile temperature pattern during 1 month, with resolution during and after treatment with azithromycin. *Bartonella henselae* Western immunoblot (WB) results did not consistently correlate with BAPGM enrichment blood culture/PCR results or *B henselae* indirect fluorescent antibody (IFA) titers, and WB titration results were not informative for establishing antibiotic treatment failure. During the respective follow-up periods, no illnesses or additional febrile episodes were reported, despite repeat documentation of *B henselae* bacteremia in two cats available for follow-up (one with the same genotype and the other with a different *B henselae* genotype); one cat was, unfortunately, killed by dogs before follow-up testing.

Relevance and novel information We conclude that microbiological diagnosis and treatment of *B henselae* infection in cats can be challenging, that antibody titration results and resolution of clinical abnormalities may not correlate with a therapeutic cure, and that fever and potentially neutropenia should be differential diagnostic considerations for young cats with suspected bartonellosis.

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Cases 1 and 2, male domestic shorthair cats, were rescued from a shopping center parking area on 16 May 2012. Fleas were removed manually while bathing. On 16 June 2012, physical examinations (estimated age of the cats was 8 weeks and their body weight was 1 kg) were unremarkable. FRCP vaccination (Purevax Feline 3; Merial), empirical treatment for intestinal parasites (Profender; Bayer Animal Health) and a parasiticide (Frontline Plus; Merial) was applied every 25–30 days. Castrations, scheduled for 21 August 2012, were uneventful until 3 weeks later when case 1 became acutely lethargic with an elevated temperature (39.7°C).

Case 1's complete blood count (CBC; Irvine, CA), serum chemistry profile and urinalysis values (ANTECH Diagnostics) were negative or were within the reference intervals (RIs). Feline leukemia virus (FeLV; indirect fluorescent antibody [IFA; National Veterinary Laboratory, Franklin Lakes, NJ]), feline immunodeficiency virus (FIV; Western blot [WB; National Veterinary Laboratory]), *Toxoplasma gondii* (ELISA; ANTECH Diagnostics), feline coronavirus (IFA; ANTECH Diagnostics) and a *Bartonella henselae* WB test (National Veterinary Laboratory) were

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negative. As summarized in Table 1, *Bartonella* alpha Proteobacteria growth medium (BAPGM) enriched blood culture/PCR (Galaxy Diagnostics, Research Triangle Park, NC) results were negative (ie, no *Bartonella* species DNA amplification) and *B henselae* WB (National Veterinary Laboratory) was again negative.[1–3] Because of these negative test results, antibiotics were not administered and the owner (veterinary technician) recorded daily aural temperatures (Baby Braun ThermoScan 5 ear thermometer; Kaz US) for the next 30 days. Fever was defined as a temperature recording at or above 38.3°C. A relapsing, cyclic fever pattern was documented (Figure 1) with inactivity accompanying fever spikes.

Case 1 remained intermittently febrile through 11 October 2012, when lethargy, anorexia and fever (40.8°C) necessitated antibiotic therapy. CBC, serum

Table 1 Sequential clinical findings. Western immunoblot (WB), PCR, *Bartonella* alpha Proteobacteria growth medium (BAPGM) blood culture/PCR/DNA sequencing results and post-treatment Western blot antibody titration titers for three febrile *Bartonella henselae*-infected kittens

	Date	Clinical findings	*Bartonella* WB and titration titers	Blood PCR	BAPGM culture PCR
Case 1	3 September 2012	Temperature 39.8°C, lethargy	Negative	Negative	Negative
	5 September 2012			Negative	Negative
	7 September 2012			Negative	Negative
	11 October 2012	Temperature 40.8°C, lethargy Azithromycin 10 mg/kg for 21 days	Negative	*Bh** *Bh**	*Bh**
	12 November 2012		1:256,000		
	12 May 2013	Titration testing	1:16,000 (16-fold decrease)		
	28 August 2014	Follow-up testing	ND	Negative	*Bh*†
	30 August 2014		ND	Negative	Negative
	1 September 2014		1:16,000 (no change)	Negative	Negative
Case 2	21 August 2012	Presurgery	1:512,000		
	3 September 2012	40.0°C, lethargy	ND	*Bh**	*Bh**
	5 September 2012		ND	*Bh**	*Bh**
	7 September 2012	Azithromycin 10 mg/kg for 21 days	ND	*Bh**	*Bh**
	2 March 2013	39.8°C, lethargy anorexia Pradofloxacin 7.5 mg/kg x 21 days	1:512,000 (no change)	*Bh** *Bh**	*Bh**
	5 September 2013	Titration testing	1:64,000 (eight-fold decrease)		
	30 May 2014	Killed by dogs and lost to follow-up			
Case 3	23 August 2012	37.7°C, lethargy anorexia neutropenia Azithromycin 10 mg/kg for 21 days	1:64,000	*Bh**	*Bh**
	7 May 2013	Titration testing	1:8,000 (eight-fold decrease)		
	28 August 2014	Follow-up BAPGM triple blood draw and titration testing	ND	*Bh**	*Bh**
	30 August 2014		ND	*Bh**	*Bh**
	1 September 2014		1:32,000 (four-fold decrease)	Negative	*Bh**

**Bartonella henselae* Houston-1 strain type based upon 16S-23S intergenic spacer DNA sequences
†*Bartonella henselae* San Antonio-2 strain type based upon 16S-23S intergenic spacer DNA sequences
ND = not determined

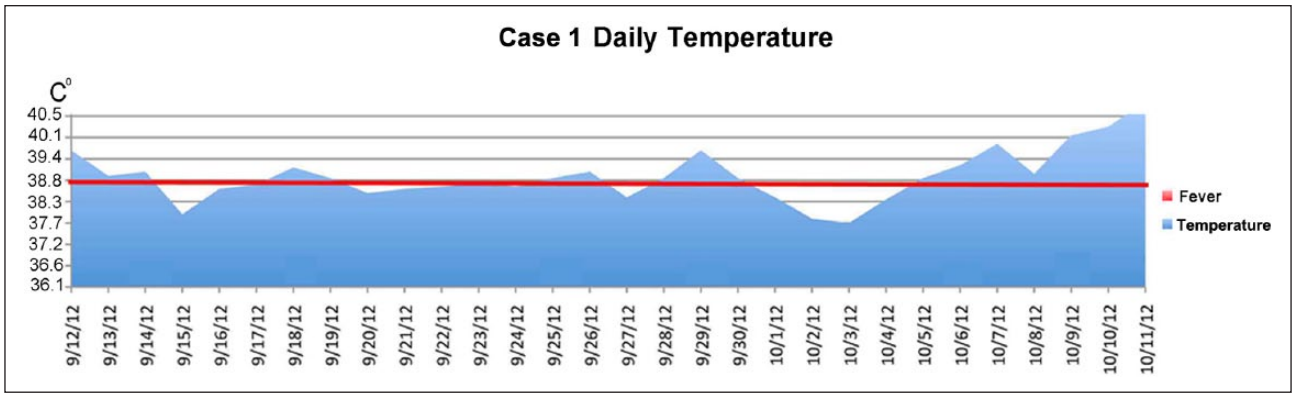

Figure 1 Sequential aural temperature recordings from case 1 prior to azithromycin therapy, documenting a fluctuating, cyclic fever pattern

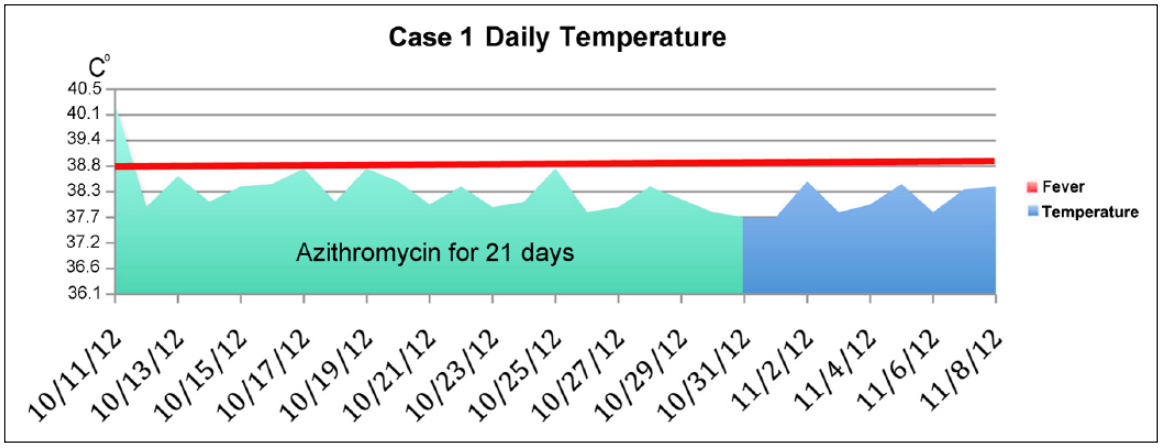

Figure 2 Sequential aural temperature recordings from case 1 during and after azithromycin therapy, documenting resolution of the fluctuating, cyclic fever pattern

biochemical profile and urinalysis results (ANTECH Diagnostics) were within the RIs. *B henselae* DNA was amplified and sequenced from the kitten's blood in two laboratories (Galaxy Diagnostics and Vector Borne Disease Diagnostic Laboratory). By aligning DNA sequences, both laboratories amplified the same *B henselae* genotype. WB remained negative. Azithromycin suspension (40 mg/cc, 10 mg/kg/day) was administered orally for 21 days. Temperature, behavior and food intake normalized within 24 h. Aural temperatures remained normal for the next 30 days (Figure 2). On 12 November 2012, WB documented seroconversion. Six months later (7 May 2013), WB titration documented a 16-fold decrease in the titer of *B henselae* antibodies. Despite remaining healthy, *B henselae* DNA was again PCR amplified and sequenced from a BAPGM enrichment blood culture obtained on 28 August 2014, and WB titration indicated no decrease in antibodies, whereas a *B henselae* IFA titer (Vector Borne Disease Diagnostic Laboratory) was <1:16.

During the examinations of 15 June and 21 August 2012, case 2, presumably case 1's littermate, was vaccinated, treated and tested identically. A *B henselae* WB was positive. There were no intra- or immediate postoperative complications. The owner was instructed to monitor and measure aural temperature if signs of illness occurred. Anorexia, lethargy and fever (40.0°C) developed 10 days postcastration. Laboratory abnormalities (ANTECH Diagnostics; 6 September 2012) included normocytic, normochromic anemia (hematocrit 26.1% [RI 29–48%], hemoglobin 8.2 g/dl [RI 9.3–15.9 g/dl]) and neutrophilia (12,456/µl [RI 2500–8500/µl]). *B henselae* DNA was amplified (Galaxy Diagnostics) and sequenced (GENEWIZ, Research Triangle Park, NC) from three BAPGM enrichment blood cultures. Azithromycin suspension (40 mg/cc, 10 mg/kg/day) was administered orally for 21 days. Temperature and food intake normalized within 24 h.

On 1 March 2013, case 2 became anorexic, lethargic and febrile (39.8°C). Body weight had increased (3.5 kg). Neutropenia (986/µl [RI 2500–8500/µl], confirmed by blood smear examination [ANTECH Diagnostics]) was the only hematological abnormality. Serum biochemical profile and urinalysis values were within the RIs. BAPGM enrichment blood culture/PCR (Galaxy Diagnostics) and FastPanel PCR Feline Flea and Tick Borne Profile (ANTECH Diagnostics) amplified *B henselae* DNA, whereas *Anaplasma, Ehrlichia* and hemotropic *Mycoplasma* species were not amplified from the cat's blood. Azithromycin suspension (40 mg/cc, 10 mg/kg/day) was administered orally for 21 days, and again temperature and food intake normalized within 24 h. *B henselae* WB titration antibody levels (August 2012 vs March 2013) were unchanged. On 26 April 2013, case 2 was again febrile and lethargic and was treated with pradofloxacin (Veraflox, 7.5 mg/kg per day as an oral suspension; Bayer Animal Health) for 21 days. Within 6 h the cat appeared clinically improved. CBC values (5 September 2013) were within the RIs (neutrophil count 3060/µl) by WB titration (pretreatment and 5 months post-treatment), serum samples identified an eight-fold decrease in *B henselae* antibody levels; whereas, a *B henselae* IFA titer (Galaxy Diagnostics) was <1:16. Case 2 remained healthy until 30 May 2014, when the cat was killed by two dogs after escaping from the house. A necropsy was not performed.

On 15 August 2012, case 3, a female domestic shorthair cat, was found hiding under a car. Prior to introduction into the home, the flea-infested cat (weight 1.4 kg, aural temperature 37.8°C) was bathed, Frontline-Plus was applied, ear mites were treated (Acarexx; Boehringer Ingelheim Vetmedica) empirically with Profender and a vaccine (Nobivac: 1-HCP + FeLV; Merck Animal Health) was administered. Prior to comingling, case 3 was isolated from the other two kittens for 7 days. Cases 3 and 2 began playing and interacting immediately, whereas case 1 interacted minimally with case 3 for the initial month after introduction, after which all three cats interacted frequently.

On 23 August 2012, case 3 was lethargic and anorexic (aural temperature 37.7°C, body weight 1.38 kg). Physical examination was unremarkable. Neutropenia (930/µl [RI 2500–8500/µl]) was the only hematological abnormality. A serum biochemistry profile and urinalysis results were within the RIs. FeLV (National Veterinary Laboratory), FIV (National Veterinary Laboratory), *T gondii*, and feline coronavirus tests were negative. WB was positive and *B henselae* DNA was amplified (Galaxy Diagnostics) and sequenced (GENEWIZ) via BAPGM enrichment blood culture. Azithromycin suspension was administered (40 mg/cc, 10 mg/kg/day) for 21 days. Normal behavior and activities resumed within 5 h. WB titration (August 2012 and May 2013 serum) documented an eight-fold decrease in *B henselae* antibodies. Despite remaining clinically healthy, case 3 was bacteremic in August 2014, accompanied by a four-fold increase in *B henselae* WB antibodies, whereas a *B henselae* IFA titer (Galaxy Diagnostics) was <1:16.

Fleas were not seen on the three kittens or the owner's only dog following initial adoption. The time course for each of the three cases is summarized in Figure 3.

Identical pretreatment *B Henselae* DNA sequences were obtained from all three cats. For each amplicon, there was 100% 16S–23S intergenic spacer region (ITS) sequence identity (ie, 360/360 nucleotides aligned [National Center for Biotechnology Information]), supporting infection with the same *B Henselae* ITS genotype (Houston-1, Genbank Accession L35101). This Houston-1 genotype was again documented at fever recrudescence 112 days later from case 2, and from case 3 2 years later, supporting failure of azithromycin to eliminate either infection. In contrast, case 1 was infected with a different ITS genotype (415/415 base pair identity to *B Henselae* San Antonio-2, Genbank accession AF369529) 2 years later, supporting co-infection with more than one genotype or a newly acquired infection.

Discussion

Although involving only three cases, these findings are unique for several reasons. The owner, a veterinary technician, worked for a veterinarian who systematically pursued an infectious disease diagnosis, in association with acute-onset lethargy and febrile illness. Additionally, there was consistency in preventive healthcare (vaccines and parasiticides), diagnostic testing, and the dose and duration of azithromycin administration. BAPGM enrichment blood culture/PCR followed by DNA sequencing documented *B henselae* infection in cases 2 and 3 at illness onset and at the time of case 2's fever recrudescence, whereas case 1 initially tested PCR negative. Two possibilities for the negative BAPGM enrichment blood culture/PCR results seem most likely. Following experimental blood transfusion transmission of *B henselae* to cats, Kordick and Breitschwerdt documented prolonged periods of abacteremia (weeks to months) in some cats.[4] Thus, it is possible that case 1 did not have circulating *B henselae* organisms at the time of specimen collections in September 2012. Alternatively, case 1 could have been infected by a flea, unknowingly introduced into the household, following case 3's adoption in August 2012. This possibility could explain the negative WB results, the negative PCR results and the subsequent PCR positive results and documentation of seroconversion by WB. Based upon 16S–23S ITS DNA sequence alignments, all three kittens were initially infected with the same *B henselae* genotype.

Short-duration fever has been reported in cats experimentally infected with *B henselae*,[4–10] and the bacterium

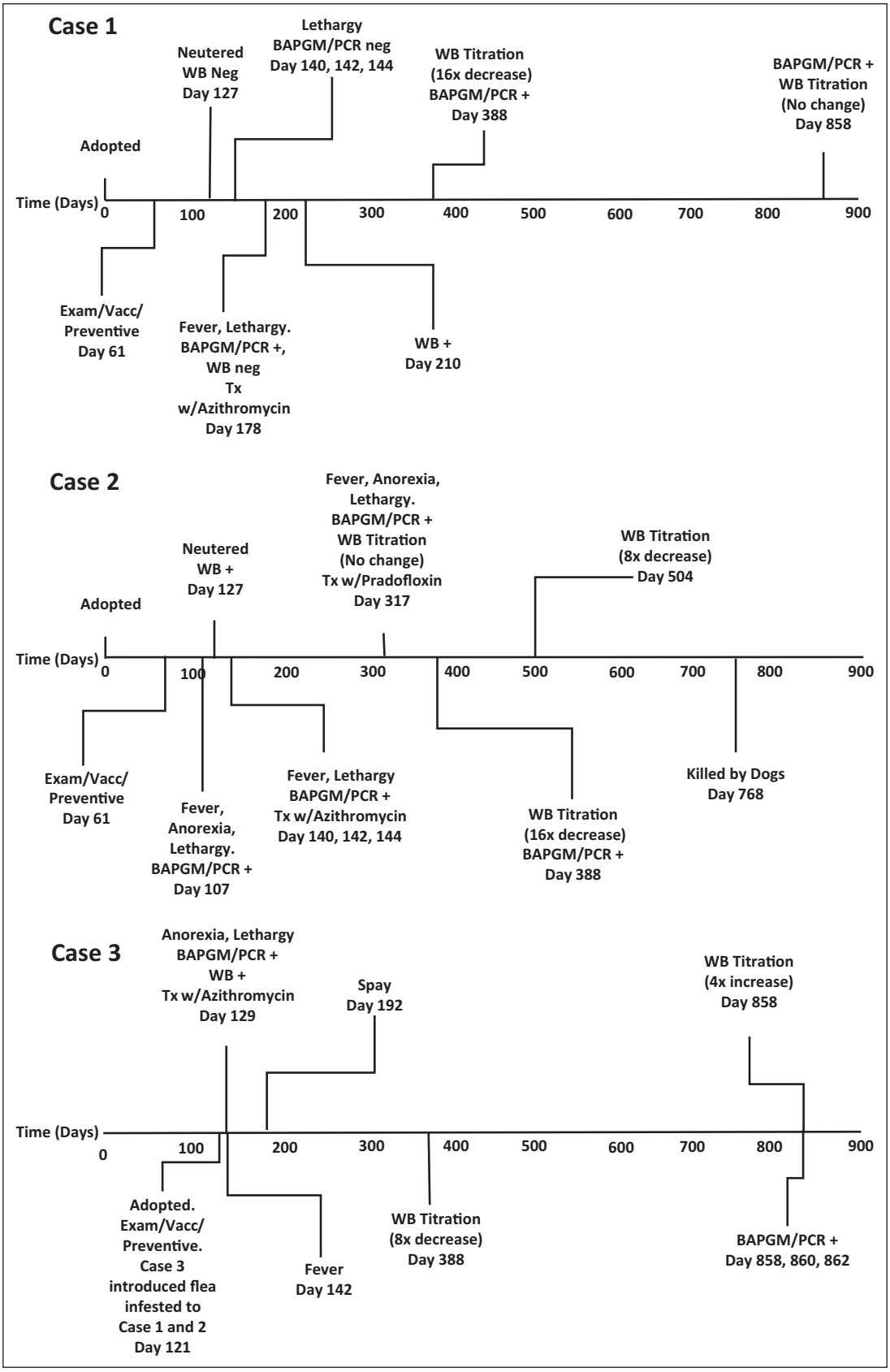

Figure 3 Summary of the time course for each of the three cases. WB Neg = Western blot negative; BAPGM = *Bartonella* alpha Proteobacteria growth medium; PCR neg = PCR negative; WB = Western blot; PCR (+) = PCR positive; Exam = examination; Vacc = vaccination; Tx w/ = treatment with; WB (+) = Western blot positive

causes relapsing fever in humans, particularly children.[11–13] Cat scratch disease, caused by *B henselae*, is characterized by fever and lymphadenopathy,[12,14] and recently *B henselae* bacteremia was documented in a dog with fever of unknown origin.[15] Thus virulent strains of *B henselae* cause fever in cats, dogs and humans. Diagnostically, there were no consistent hematological, biochemical or urinalysis abnormalities, despite acute-onset febrile illness; however, case 2 was initially mildly anemic and had neutrophilia but was neutropenic when febrile illness reoccurred. The neutropenia in these two cats may have been related to *B henselae* infection or infection with another organism that was not tested for diagnostically. Unique sequential temperature data in case 1 documented a fluctuating, relapsing febrile pattern prior to illness onset.

An obvious limitation of this study is the inability to determine when and how *B henselae* transmission initially occurred and whether there was subsequent bacterial transmission within the household. Importantly, the onset of illness in cases 1 and 2 occurred months after presumptive flea transmission of *B henselae*, but shortly after the introduction of case 3. It is possible that fleas were unknowingly introduced into the household with case 3. In the context of disease causation, immediately following administration of azithromycin, all three cases became afebrile, supporting a diagnosis ex juvantibus ('from that which helps'; medical definition: 'a response to a narrow, specific therapy that strengthens the association with a suspected etiology'). Although *B henselae* bacteremia was documented in all three cats in conjunction with resolution of clinical signs following initiation of antibiotic therapy, it is possible that this response was related to another infectious agent (co-infection), or that spontaneous resolution of clinical signs occurred independently of antibiotic administration. However, based upon the results of this study and others published previously,[4–9] *B henselae* may be an underestimated cause of fever of unknown origin in flea-exposed cats, particularly those with immature or naive immune systems.

In a study by Lappin et al, feline *B henselae* ELISA seroreactivity did not correlate with fever, whereas PCR positivity approached statistical significance ($P = 0.0571$).[16] Also, *Bartonella* ELISA and WB did not correlate with positive *Bartonella* PCR.[16] Because cats are natural reservoir hosts for *Bartonella clarridgeiae, B henselae* and *Bartonella koehlerae*, investigators have experienced difficulties establishing whether these *Bartonella* species are feline pathogens. Experimentally, the route of infection (intravenous or subcutaneous) and inoculum (culture grown, blood transfusion or flea transmission) substantially influence the duration and patterns of bacteremia (relapsing or non-relapsing), whether clinical signs were observed and, in those studies in which cats were necropsied, whether pathological lesions were found.[4–9] Owing to the high seroprevalences among cats in flea-endemic regions, seroepidemiological studies have, for the most part, not found statistical associations between *B henselae* seroreactivity and disease manifestations.[16–21] However, Whittemore et al found a correlation between *Bartonella* species antibodies and hypergammaglobulinemia in cats.[22] Sykes et al found an association between gingivitis and stomatitis and culturing *Bartonella* species from bacteremic cats, whereas there was no statistical difference in seroprevalence between cases and controls.[21] Also, a study from Switzerland, where the overall *B henselae* seroprevalence in 728 cats was low (8.3%) compared with many regions in North America where cat population seroprevalences are >40%, found a statistical correlation between seroreactivity and stomatitis.[23] Collectively, these and other published studies serve to illustrate the evolutionary adaptation of most *B henselae* strains among cats as a natural reservoir host and the inherent difficulty in establishing disease correlations.

There is substantial genetic variation among *B henselae* strains,[24] potentially contributing to different virulence characteristics, most likely mediated by genetic variation among *B henselae* type IV secretion systems,[25] Bad A adhesion proteins and potentially other unknown virulence factors.[26] Also, the strains most often isolated from healthy, non-clinical cats are not the genetic strains most often found in humans with bartonellosis.[24,27] Thus, similar to other bacteria such as *Escherichia coli* (*E coli* 0157:H7 can induce hemolytic uremic syndrome and death), there are differences in virulence among *B henselae* strains. One might speculate that virulence differences, in conjunction with the robustness of the host immune response, determine the extent to which a *B henselae* genotype induces disease. For example, the CSU-1 strain, originally isolated from a shelter cat in Florida and used experimentally by Bradbury and Lappin in flea transmission studies, has induced acute myocarditis in young experimentally infected cats.[9] Other feline experimental infection studies have also provided evidence to support virulence differences among *B henselae* strains.[4,9] Granulomatous myocarditis was reported in young, naturally flea-infested cats, by direct bacterial visualization and PCR amplification of *B henselae* from the myocardial lesions.[28] Thus, future field-based studies designed to address disease causation should incorporate bacterial isolation, *B henselae* genotyping, sequential serological testing, an enrichment culture/PCR approach to enhance the sensitivity of documenting infection with one or more *Bartonella* species and documentation of each cat's clinical response to a defined antibiotic treatment regimen.

As recently reviewed, there is no 'gold standard' test for the diagnosis of bartonellosis in cats, dogs, horses or humans.[29] A study involving canine vector-borne

diseases (CVBDs) diagnosis provided support for obtaining serological, microbiological culture and PCR data when attempting to confirm a CVBD diagnosis.[30] With the advent of PCR testing, some clinicians have abandoned serology in favor of vector-borne disease molecular diagnostic panels. As PCR testing has substantial sensitivity limitations (ie, false-negative test results),[30] both modalities are recommended for diagnosis of feline vector-borne diseases, including bartonellosis.

In this study, WB results did not correlate with *B henselae* IFA titers, whereas in previous studies there were no correlations between ELISA and WB.[9,16] Why WB, ELISA and IFA titers do not correlate deserves future research consideration. Also, WB titration results from case 3 were not informative for establishing antibiotic treatment failure. As cats (an unknown percentage), dogs (50–75% depending upon infecting *Bartonella* species),[32] horses (100%)[32] and humans (approximately 50–100% depending upon infecting *Bartonella* species)[33,34] can be *Bartonella* bacteremic without detectable IFAs, IFA serology has substantial lack of sensitivity across several animal species. Similarly, case 1 was initially WB negative despite documentation of bacteremia, whereas case 2 was not initially treated (owing to lack of illness), despite being WB positive. As a result of stress-induced interference with premonition (infection immunity), one might speculate that hospitalization and castration contributed to the onset of febrile illness in cases 1 and 2, 10–21 days later. At the time of illness onset and at the time of recrudescent febrile illness, case 2 was BAPGM blood culture/PCR positive. Also, in order to determine antimicrobial susceptibility and resistance patterns, it is important for diagnostic laboratories to culture *Bartonella* species from cats and other animals whenever possible.

While there are multiple serologic assays, PCR assays and culture methods available for use to aid in the diagnosis of bartonellosis in cats, studies proving which are optimal are lacking. Despite these limitations, we recommend the use of serology, a *Bartonella* enrichment culture/PCR platform and subculture isolation to aid clinicians in making antibiotic treatment decisions or to advise clients relative to zoonotic concerns.

An optimal bartonellosis treatment regimen has not been established for bacteremic animals or humans.[29,35] In a study involving naturally infected febrile cats, both doxycycline and orbifloxacin induced resolution of clinical abnormalities by day 5 of treatment; however, despite a 28 day treatment course, most cats remained *Bartonella* species PCR positive after antibiotic administration.[10] Experimentally, cats inoculated with *B henselae*-infected blood remained culture positive despite treatment with 2 or 4 weeks of doxycycline.[36] Relapses after antibiotic withdrawal have also been reported in *B henselae*-infected dogs and human patients treated with extended courses of doxycycline.[37–39] Doxycycline treatment failure (6 week course) was also suspected in a febrile dog from France.[15] Based upon the limited clinical and experimental evidence, we do not recommend doxycycline treatment protocols published to date as the sole antibiotic for treatment of feline bartonellosis, if therapeutic elimination is the primary goal.

By E-test and disk diffusion assays, pradofloxacin had greater antimicrobial activity against feline *B henselae* isolates than enrofloxacin and azithromycin[40] and appeared to be an effective treatment for case 2; however, post-treatment BAPGM enrichment blood culture was not performed. Based upon negative culture and PCR results, cats treated with orbifloxacin appeared to clear their *B henselae* infections.[9] A combination of antibiotics with different modes of action, such as doxycycline and a fluoroquinolone, may be required for therapeutic elimination of *B henselae* in cats; however, data confirming that a combination of doxycycline and pradofloxacin (as used in case 2) eliminates *B henselae* infections, rather than suppressing the infection, are currently lacking. Azithromycin has often been used to treat feline bartonellosis; however, based upon in vitro testing, cat and human *B henselae* strains rapidly developed genetic resistance to macrolides through a single nucleotide mutation in the 23S rRNA gene.[41] Currently, no studies have investigated antimicrobial resistance in *B henselae* isolates obtained from azithromycin-treated cats, and there are no studies confirming the efficacy of azithromycin as a sole antibiotic for the treatment of feline bartonellosis.

Optimal monitoring of cats previously diagnosed with bartonellosis has not be established and further studies are needed. However, we recommend that follow-up serology and enrichment culture/PCR testing be considered to confirm therapeutic elimination of the infection. Post-treatment decreases in antibody levels seem to support elimination of infection in dogs.[42] However, in a dog experimentally infected with *B henselae*, IFAs were no longer detectable after administration of corticosteroids (no antibiotic treatment administered), yet *B henselae* was isolated from the bone marrow.[43] Based upon human case experiences, obtaining three specimens for enrichment blood culture/PCR within a 7 day period increased sensitivity of *Bartonella* species detection (odds ratio 3.4; $P = 0.02$),[44] and in no instance was a patient enrichment culture/PCR positive in all three specimens and only 3/12 patients were positive for two specimens. This finding, potentially related to a relapsing bacteremia as reported in cats,[4] indicates that diagnostic confirmation and subsequently proving therapeutic elimination of bartonellosis remains clinically and diagnostically challenging. Until additional studies define an optimal treatment regimen for feline bartonellosis, combination therapy is recommended for culture

or PCR confirmed cases. As optimal treatment regimens, including dual therapy, are not proven to eliminate *Bartonella* species bacteremia in cats, it is important to attempt to lessen potential transmission by maintaining stringent flea control.[8,9]

Conclusions

We conclude that *B henselae* causes acute febrile illness in cats that resolves in conjunction with antibiotic therapy, which may or may not represent elimination of infection. Neutropenia may be an accompanying haematological abnormality in a subset of cats with bartonellosis. Additional laboratory and field studies are needed to define optimal treatment regimens and optimal follow-up testing procedures for cats with bartonellosis.

Supplementary material Indirect fluorescent antibody testing using a panel of *Bartonella* species antigens and the BAPGM enrichment blood culture/PCR platform, with DNA sequence confirmation of the infecting *Bartonella* species, is available for clinical diagnostic and research study purposes through Galaxy Diagnostics, Inc, Research Triangle Park, NC.

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Conflict of interest In conjunction with Dr Sushama Sontakke and North Carolina State University, Dr Breitschwerdt holds US Patent No. 7,115,385; Media and Methods for cultivation of microorganisms, which was issued 3 October 2006. He is the Chief Scientific Officer for Galaxy Diagnostics, a company that provides diagnostic testing for the detection of *Bartonella* species infection in animals and human patients. Dr Natalie Cherry is the Laboratory Supervisor/Research Specialist for Galaxy Diagnostic, Inc.

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