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Resistance Level of Mosquito Species (Diptera: Culicidae) from Shandong Province, China



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ABSTRACT: This study describes the aquatic habitats, species composition, and the insecticide resistance level of the mosquito *Culex pipiens pallens* in Shandong Province, China. A cross-sectional survey of mosquito larval habitats was conducted from May to November 2014 to determine the species composition and larval abundance. Larvae were collected using the standard dipping technique, and a total of four habitat types were sampled. The fourth instar larvae of *Cx. pipiens pallens* collected in each habitat type were tested for resistance to five insecticides according to a WHO bioassay. A total of 7,281 mosquito larvae were collected, of which 399 (5.48%) were categorized as *Anopheles mosquito* larvae (*An. sinensis*), 6636 (91.14%) as culicine larvae (*Cx. pipiens pallens*, *Cx. tritaeniorhynchus*, *Cx. halifaxii*, and *Cx. bitaeniorhynchus*), 213 (2.93%) as *Armigeres* larvae, and 33 (0.45%) as *Aedes* larvae (*Aedes albopictus*). In addition, a total of 1,149 mosquito pupae were collected. *Culex* larvae were distributed in all habitats investigated. Tukeys HSD analysis showed that roadside drainages were the most productive habitat type for *Culex* larvae. *Armigeres* species were found only in drains, *Aedes* only in water tanks, and *Anopheles* in water that was comparatively clear and rich in emergent plants. Bioassay showed that the maximum resistance level of *Cx. pipiens pallens* was to deltamethrin, while it was lowest to plifenate. The productivity of various mosquitoes in different habitat types is very heterogeneous. It is particularly important to modify human activity and the environment to achieve effective mosquito vector control. For effective larval control, the type of habitat should be considered, and the most productive habitat type should be given priority in mosquito abatement programs.

KEYWORDS: mosquito, resistance level

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Introduction

As the most medically serious disease vector, the mosquito has abundant species and wide distribution.¹ Mosquitoes not only bite and harass humans but also spread various parasitic and viral diseases such as malaria, filariasis,² viral encephalitis, and dengue. In 2010, China reported 7,855 malaria cases with an incidence rate of 0.06/10,000. In addition, it reported 34,082 suspected cases and 19 deaths. For instance, Shandong Province reported 117 malaria cases in 2010, down 2.5% compared to the previous year, and among them 70% were imported cases. Furthermore, mosquito-borne viral diseases like epidemic encephalitis B happened occasionally.³ *Culex pipiens pallens* is also a potential vector of West Nile virus (WNV) in China.⁴ The control of mosquito populations is based on chemical insecticides, but, unfortunately, the rapid development of insecticide resistance undermines the effectiveness of control. Especially in *Cx. pipiens pallens*, long-term intensive and widespread use of pyrethroids has led to moderate or high resistance for DDVP, propoxur, acetofenate, cypermethrin, and deltamethrin, making the use of this class of insecticides ineffective and limiting the available options for mosquito control.^{5–7} Therefore, it is of great significance for planning control measures to counteract various

mosquito-borne diseases, to study the composition and distribution of the mosquito population, as well as their breeding places and disease vector control interventions.

The rapid development of tourism has increased sewage levels, thus making the lake environment more conducive for the growth of mosquitoes. The population of Shandong Province has reached about 95.79 million, and domestic sewage pools and above-ground pools of human excreta are widespread in urban areas, where they have become the main breeding ground of *Cx. pipiens pallens*.

We categorized four main mosquito breeding habitats in Shandong Province: irrigation ditches, roadside drainages, freshwater lake fringes, and water tanks. Then, we investigated the larvae according to their different ecological types to obtain a systematic understanding of the species and distribution of mosquitoes in Shandong Province, the relationship between the habits of the mosquitoes and the natural environment, and the insecticide resistance levels of *Cx. pipiens pallens*. A new mosquito database was established based on this investigation, which provided a scientific basis for both appropriate control measures targeting mosquito-borne diseases and efforts to prevent or slow down the development and spread of insecticide resistance in the mosquito population.

Materials and Methods

Study area. Shandong Province lies in the coastal area of east China, 34°22'52"–38°15'02"N and 114°19'53"–122°43'E, covering an area of 157,100 km². The land is generally flat, with very few sections of undulating ground, with wheat, rice, and corn as the predominant crops. It has a temperate continental monsoon climate with concentrated rainfall during the hot, rainy summer season. The average annual rainfall for this region is approximately 550–950 mm (based on data of 1958–2008),⁸ with 60%–70% concentrated in the months of June, July, and August and an average temperature of 21.5°C–27°C, thus appropriate for the growth and reproduction of mosquitoes.

Mapping of study areas. Larvae were collected in larvae form from three separate districts in Jinan (36°38'N, 116°56'E), Qingdao (35°41'N, 119°44'E), and Jining (35°18'N, 116°29'E) in Shandong Province (Fig. 1). Coordinate readings (latitude and longitude) and altitude of major roads, freshwater lakes, aquatic habitats, and houses were taken once using a geographic positioning system (GPS) unit, and records were kept in order to repeat samplings.

Selection of aquatic habitats. All selected aquatic habitats were mapped in Shandong Province from May to November in 2014. These were sampled twice per month to determine the presence or absence of larvae. Habitats were grouped by the types of irrigation ditches, roadside drainages, freshwater lake fringes, and water tanks. These habitat types were described as follows: 1) irrigation ditches were those used to irrigate rice, wheat, corn, and other produces; 2) drains on roadsides were collections of rainwater and effluents from factories and houses; 3) lake fringes were edges of a freshwater lakes with

reeds (*Phragmites australis*), lotus (*Nelumbo nucifera*), and common duckweed (*Lemna minor*); and 4) water tanks were containers in which people collect rainwater.

The dissolved oxygen, pH, and NH₄ + N of all standing waters in habitats greater than 100 mm depth was evaluated using a YSI Professional multiparameter meter. When any breeding site was dry, an attempt was made to survey the vicinity.

Larval sampling. A larva dipper was used to collect mosquito larvae from various habitats, and the collected larvae were put in specimen boxes and taken to the laboratory for morphological identification.^{9,10} Every potential breeding site was surveyed for mosquito larvae and pupae using a standard dipper (~350 mL) and performing five dips per site (1750 mL). The specimen boxes (length = 25 cm, width = 15 cm, height = 20 cm) were made of plastic.

Mosquito larvae densities were calculated as the mean number of larvae per dip sample. Larvae with similar morphology were fed with pork liver powder and yeast powder until they reached adult stage. Care was taken to retain the fourth instar larvae of *Cx. pipiens pallens* for subsequent bioassay.

Bioassay. The susceptibility of fourth stage larvae of *Cx. pipiens pallens* collected was tested on site to five insecticides, namely cypermethrin, propoxur, deltamethrin, plifenate, and DDVP, and testing was done according to WHO bioassay procedure.¹¹ The fourth instar larvae (20–24 numbers) were exposed in plastic vials with 50 mL tap water, and the test concentrations were as follows: cypermethrin, 2–32 µg/L, propoxur, 0.28–1.35 mg/L, deltamethrin, 2.5–40 µg/L, plifenate, 2.5–40 µg/L, and DDVP, 0.25–4 mg/L. Susceptible



Figure 1. Map of China demonstrating the distribution of mosquito sampling sites. Jinan (36°38'N, 116°56'E), Qingdao (35°41'N, 119°44'E), and Jining (35°18'N, 116°29'E).

mosquitoes were exposed in the plastic cup with tap water as control. After 24 hours, the number of dead mosquitoes was recorded. Sensitive strains (protected from contact with insecticides for 20 years), routinely reared in our laboratory, were used as the control. The test was repeated thrice.

Statistical analysis. Statistical analyses were carried out using the SPSS software (Version 19 for Windows, SPSS Inc.). Logistic regression analysis¹² was used to test associations of the environmental variables with the occurrence of different mosquito larvae. Presence of larvae was categorized as 1, while the absence of larvae was categorized as zero. One-way analysis of variance (ANOVA) was used to compare the differences in the number of mosquito larvae in different habitats.

Results

Habitat characterization, larval species, and their abundance. The habitat types sampled in Shandong Province during this survey, from May to November 2014, included irrigation ditches ($n = 3$), roadside drains ($n = 6$), freshwater lake fringes ($n = 4$), and water tanks ($n = 5$). Larvae were differentiated macroscopically according to whether they hung down from the water surface (*Culicines*), floated parallel to the surface (*Anophelines*),¹³ or had a distinguishing thin and bifurcated hair V¹⁴ or S-shaped movement in the water (*Aedes*).^{15,16} A total of 7,281 mosquito larvae were collected, of which 399 (5.48%) were categorized as *Anopheles*, 6636 (91.14%) as *Culex*, 213 (2.93%) as *Armigeres*, and 33 (0.45%) as *Aedes* larvae. In addition, a total of 1,149 mosquito pupae were collected (Table 1). In the investigation, culicine late instars were found in 34.29% (2,497) of all samples, and *Anopheles* late instars were found in 3.91% (285). Culicine early instars were found in 4,139 samples (56.85%), whereas *Anopheles* early instars were found in 114 samples (1.57%). The density of early instars was 1.66-fold higher than late instars for culicines, while for *Anopheles* the late instars were 2.50-fold higher than early instars. Furthermore, late instar larvae are frequently used to assess the dynamics of larval populations in habitats of different types and characteristics.^{17,18} Further identification based on morphology showed that the most abundant was

Cx. pipiens pallens (88.94%, $n = 6,476$), which was observed in all habitats investigated. *An. sinensis* (5.42%, $n = 399$), the second most abundant mosquito, was found in irrigation ditches and margins of a freshwater lake with reeds, lotus, and common duckweed. The third most important mosquito was *Aedes albopictus* (0.48%, $n = 33$), which was only found in water tanks. In addition, *Cx. tritaeniorhynchus* (0.81%, $n = 59$), *Cx. halifaxii* (0.16%, $n = 12$), *Ar. subalbatus* (2.93%, $n = 213$), and *Cx. bitaeniorhynchus* (1.22%, $n = 89$) were identified based on their morphology. *Ar. subalbatus* and *Cx. halifaxii*, which were sampled from roadside drains (collections of rainwater and effluent from factories or houses), had more niche overlap with *Cx. pipiens pallens* and were a predator of it. *Cx. tritaeniorhynchus* and *Cx. bitaeniorhynchus* were sampled from the margins of a freshwater lake.

Water samples from all these habitats were detected by a handheld multiparameter meter (Table 2). It is noteworthy that drains on the roadsides were the most turbid habitat type, as observed with the naked eye, and the most productive habitat type for *Culex* mosquitoes. Irrigation ditches and the margins of freshwater lakes were the most suitable habitat types for *Cx. pipiens pallens* and other clean-water mosquitoes such as *An. sinensis*. Water tanks found near homesteads, mostly used to harvest rainwater and containing fallen leaves, were the ideal habitat for *Ae. albopictus*. Obviously, productivity was not homogeneous for every habitat type. The highest density of larvae was 182.1 per dipper in roadside drainages, and the lowest was 26.4 per dipper in water tanks. The densities of larvae in irrigation ditches and freshwater lake fringes were 59 and 60.45 per dipper, respectively.

Culex was distributed in all habitats investigated in this study, but there was a significant difference in *Culex* production from the different larval habitat types ($F_{(3,14)} = 217.236$, $P < 0.001$). Tukeys HSD analysis further showed that roadside drainages were the most productive habitat type for *Culex* larvae. By contrast, *Armigeres* were found only in roadside drains, and *Aedes* were found only in water tanks (Table 1). Logistic regression showed that emergent plants ($P = 0.029$) were the best predictors of *Anopheles* larval abundance in the

Table 1. Mosquito larval abundance in the different habitat types in Shandong Province.

HABITAT TYPES	n*	ANOPHELES		CULEX		ARMIGERES		AEDES		PUPAE
		EARLY INSTARS**	LATE INSTARS†	EARLY INSTARS	LATE INSTARS	EARLY INSTARS	LATE INSTARS	EARLY INSTARS	LATE INSTARS	
Irrigation ditches	3	108	270	231	57	0	0	0	0	219
Roadside drains	6	0	0	2,535	2,109	101	112	0	0	819
Freshwater lake fringes	4	6	15	1,072	98	0	0	0	0	18
Water tanks	5	0	0	301	233	0	0	9	24	93
Total	18	114	285	4,139	2,497	101	112	9	24	1,149

Notes: *Number of habitats sampled; **Early instars (L1 and L2); †Late instars (L3 and L4).

Table 2. Characteristics of larval habitats.

HABITAT TYPES	n*	DO%**	pH	NH ₄ + N
Irrigation ditches	3	135.9–189.6	8.45–8.80	2.09–2.23
Roadside drainages	6	10.5–48.0	7.47–7.72	0.28–17.46
Freshwater lake fringes	4	161.0–206.4	9.38–9.56	0.69–0.72
Water tanks	5	80.9–84.4	8.84–9.84	0.16–1.01

Notes: *Number of habitats sampled; **Dissolved oxygen (%).

aquatic habitats, and putrilages ($P = 0.003$) were positively associated with *Aedes* larval abundance (Table 3).

Bioassay showed that the highest resistance levels were to deltamethrin and the highest resistance ratio was 112.37-fold in *Cx. pipiens pallens* collected in freshwater lake fringes, while the plifenate resistance level was the lowest in larvae from all of the habitat types (Table 4).

Discussion

This study was conducted to understand the ecologies of mosquito larvae and mosquito species composition in this area. A total of 7,281 mosquito larvae were collected in Jinan, Qingdao, and Jining, of which 5.48% were categorized as *Anopheles* mosquito larvae (*An. sinensis*), 91.14% as culicine larvae (*Cx. pipiens pallens*, *Cx. tritaeniorhynchus*, *Cx. halifaxii*, and *Cx. bitaeniorhynchus*), 2.93% as *Armigeres* larvae, and 33 (0.45%) as *Aedes* larvae (*Aedes albopictus*). To that end, the spatial distribution of the dominant populations and the resistance level in these populations were analyzed. The habitats sampled in the study included irrigation ditches, roadside drainages, freshwater lake fringes, and water tanks. The mosquito larvae species found in these samples belong to four genera *Culex* (four species), *Anopheles* (*An. sinensis*), *Armigeres* (*Ar. subalbatus*), and *Aedes* (*Ae. albopictus*).

Cx. pipiens pallens, *An. Sinensis*, and *Ae. albopictus* are medically important vectors of bancroftian filariasis, malaria, and yellow fever.

Culicine larvae were distributed in all habitats investigated, and roadside drains were their most productive habitat type. *Culex* mosquitoes are the predominant blood-sucking mosquitoes in the north of China and the main carrier of bancroftian filariasis: *Cx. tritaeniorhynchus* ($n = 59$, 0.81%), *Cx. halifaxii* ($n = 12$, 0.16%), and *Cx. bitaeniorhynchus* ($n = 89$, 1.22%). In this study, *Cx. pipiens pallens* was very common in polluted, stinking waters and was the most abundant overall. The result indicated that the species composition and quantity were the same as earlier results.¹⁹ As *Cx. pipiens pallens* is the main mosquito in Shandong, its insecticide resistance levels are particularly important. The resistance levels of *Cx. pipiens pallens* to cypermethrin and deltamethrin were viewed as middle and high, which were similar to the national levels. In 1997, the resistance ratios of Guangzhou, Shenzhen, Shantou, Shaoguan, Maoming, Foshan and Jiangmen City, and Guangdong Province were 222.0, 146.5, 149.0, 91.0, 26.9, 26.9, and 48.1, respectively.²⁰ Monitoring in the above-mentioned cities 12 years later has found that the resistance ratio had varied by as much as two-fold.²¹ Since pyrethroid insecticides had been widely applied in preventing and controlling disease vectors, the problem of resistance was very important.²² Plifenate (resistance ratio 0.06–0.62) is currently the only organochlorine insecticide allowed for production in China.²³ Currently, the mosquitoes are susceptible to it.

The second most abundant mosquito was *An. sinensis* ($n = 399$, 5.42%), which was found in water comparatively clear and rich in emergent plants. *An. sinensis* is the predominant malaria vector. There were 1,130 reported malaria cases in Shandong Province during 1989–2007, among

Table 3. Results of logistic regression showing the association between different environmental factors and larval abundance.

	B	df	SIG	OR	LOWER CI	UPPER CI
Anopheles						
DO% ²	0.745	1	0.998	2.107	0	1.75E + 238
PH	-0.913	1	1.000	0.401	0	–
NH ₄ + N	1.465	1	0.999	4.326	0	–
Emergent plant	0.732	1	0.029	2.978	1.104	8.084
Armigeres						
DO% ²	0.013	1	0.922	1.013	0.786	1.305
PH	-5.984	1	0.609	0.003	0	2.28E + 07
NH ₄ + N	0.176	1	0.671	1.193	0.529	2.69
Aedes						
DO% ²	-0.032	1	0.180	0.969	0.925	1.015
PH	0.583	1	0.625	1.792	0.173	18.505
NH ₄ + N	-0.451	1	0.284	0.637	0.279	1.453
Putrilages	0.021	1	0.003	1.132	1.042	1.229

Table 4. Resistance level of *Cx. pipiens pallens* larvae to five insecticides.

INSECTICIDE	HABITAT TYPES	LC ₅₀	REGRESSION EQUATION	LOWER CI	UPPER CI	RR
Cypermethrin (µg/L)	Irrigation ditches	7.8654	$y = 1.5240 + 3.8807x$	6.5611	9.4289	39.33
	Roadside drains	5.6701	$y = 3.3758 + 2.1553x$	4.6238	6.9532	28.35
	Freshwater lake fringes	18.7944	$y = 2.5721 + 1.9057x$	15.1977	23.2423	93.97
	Water tanks	8.6892	$y = 2.8702 + 2.2682x$	6.6487	11.356	43.45
	Sensitive strain	0.2000	$y = 7.0360 + 2.2141x$			1.00
Propoxur (µg/L)	Irrigation ditches	0.3387	$y = 8.9683 + 8.4407x$	0.3069	0.3739	3.55
	Roadside drains	0.4786	$y = 7.6252 + 8.2039x$	0.4474	0.5121	5.02
	Freshwater lake fringes	0.4255	$y = 7.8012 + 7.5491x$	0.3934	0.4603	4.46
	Water tanks	0.2687	$y = 8.9939 + 6.9970x$	0.2337	0.3089	2.82
	Sensitive strain	0.0954	$y = 7.2198 + 2.1750x$			1.00
Deltamethrin (µg/L)	Irrigation ditches	6.5054	$y = 2.5916 + 2.9613x$	5.4141	7.8166	54.21
	Roadside drains	10.829	$y = 2.1144 + 2.7891x$	9.2380	12.694	90.24
	Freshwater lake fringes	13.4843	$y = 2.5341 + 2.1825x$	11.0174	16.5035	112.37
	Water tanks	4.2230	$y = 3.7539 + 1.9918x$	3.1355	5.6877	35.19
	Sensitive strain	0.1200	$y = 7.6256 + 2.8696x$			1.00
Plifenate (µg/L)	Irrigation ditches	132.1538	$y = -5.7165 + 5.0524x$	117.0621	149.191	0.62
	Roadside drains	13.7931	$y = 1.3943 + 3.1639x$	11.6432	16.3400	0.07
	Freshwater lake fringes	13.4583	$y = 2.5719 + 2.1506x$	10.8820	16.6466	0.06
	Water tanks	115.5092	$y = 0.1515 + 2.3506x$	94.4657	141.2404	0.54
	Sensitive strain	212.0000	$y = 5.3560 + 0.5256x$			1.00
DDVP (µg/L)	Irrigation ditches	0.5339	$y = 5.9360 + 3.4344x$	0.4594	0.6204	4.7
	Roadside drains	0.5982	$y = 6.0484 + 4.6979x$	0.5266	0.6796	5.27
	Freshwater lake fringes	0.4654	$y = 6.4992 + 4.5129x$	0.4033	0.5369	4.10
	Water tanks	0.4070	$y = 5.6444 + 1.6505x$	0.3176	0.5215	3.59
	Sensitive strain	0.1135	$y = 7.6650 + 2.9986x$			1.00

which 456 cases (40.4%) were infected locally within the province, indicating that there were still indigenous cases. An earlier study indicated that a potentially important malaria vector control strategy was to target the immature stages of *An. sinensis*.²⁴ Therefore, realization and modification of larval habitats have been important aspects for malaria control. *An. sinensis* larvae were collected in temporary pools located in irrigation ditches on farms and freshwater lake fringes. It appears that conditions in these habitats may be very heterogeneous. Irrigation ditches are small, shallow habitats, while freshwater lake fringes were wide and deep, but they were both comparatively clear and rich in emergent plants. The presence of emergent plants in aquatic habitats has been, in different studies, positively, negatively, or not associated with that of *Anopheles* mosquitoes.^{25,26} That might be because *Anopheles* mosquitoes are known to oviposit in habitats with a certain degree of shade and light. Bryson considered that the presence of emergent plants in habitats may be describing a wide variety of plants at varying coverage levels, and the conflicting results may be due to this.²⁶ Lastly, *Ae. albopictus*, as the major vector

of yellow fever, was found only in water tanks that had putrilages.

Conclusion

In conclusion, the productivity of different habitat types for various mosquitoes was found to be heterogeneous. However, it is of particular importance to modify human activity and the environment to enhance the effects of mosquito vector control. For achieving this target, the type of habitat should be considered and the most productive habitat type²⁷ should be given priority in the mosquito abatement program.

Author Contributions

Conceived and designed the experiments: HML, MQG. Analyzed the data: PPY, PC, HFW, HWW. Wrote the first draft of the manuscript: HML. Contributed to the writing of the manuscript: XH, CXZ, LJJ. Agree with manuscript results and conclusions: MQG. Jointly developed the structure and arguments for the paper: YQZ. Made critical revisions and approved final version: MQG. All authors reviewed and approved of the final manuscript.



REFERENCES

1. Abad-Franch F, Zamora-Perea E, Ferraz G, Padilla-Torres SD, Luz SL. Mosquito-disseminated pyriproxyfen yields high breeding-site coverage and boosts juvenile mosquito mortality at the neighborhood scale. *PLoS Negl Trop Dis*. 2015;9(4):e0003702.
2. Pedersen EM, Stolk WA, Laney SJ, Michael E. The role of monitoring mosquito infection in the global programme to eliminate lymphatic filariasis. *Trends Parasitol*. 2009;25(7):319–327.
3. Zhou CL, Zhou HN. Progress in the study of the prevalence of encephalitis B in Shandong Province. *Zhongguo Bing Yuan Sheng Wu Xue Za Zhi*. 2010;4:301–303.
4. Jiang S, Wang Z, Guo X, et al. Infection and dissemination of West Nile virus in China by the potential vector, *Culex pipiens pallens*. *J Vector Ecol*. 2014;39(1):78–82.
5. Rivero A, Vezilier J, Weill M, Read AF, Gandon S. Insecticide control of vector-borne diseases: when is insecticide resistance a problem? *PLoS Pathog*. 2010;6(8):e1001000.
6. Liu HM, Cheng P, Huang X, et al. Identification of TCT, a novel knockdown resistance allele mutation and analysis of resistance detection methods in the voltage-gated Na(+) channel of *Culex pipiens pallens* from Shandong Province, China. *Mol Med Rep*. 2013;7(2):525–530.
7. Chen L, Zhong D, Zhang D, et al. Molecular ecology of pyrethroid knockdown resistance in *Culex pipiens pallens* mosquitoes. *PLoS One*. 2010;5(7):e11681.
8. Ma L, Zuo CQ, Yin ZD, Qiu GY. Analysis on variation characteristics of the rainfall ersivity during last 58 years in Shandong Province. *Sci Soil Water Conservat*. 2010;8(4):79–85.
9. Edwards F. *Mosquitoes of the Ethiopian Region III: Culicine Adults and Pupae*. London: British Museum (Nat Hist); 1941.
10. Gillies MT, Coetzee M. A supplement to anophelinae of Africa south of Sahara (Afro-tropical region). *Publ South Afr Inst Med Res*. 1987;55:1–143.
11. Rey D, Despres L, Schaffner F, Meyran JC. Mapping of resistance to vegetable polyphenols among *Aedes* taxa (diptera, culicidae) on a molecular phylogeny. *Mol Phylogenet Evol*. 2001;19(2):317–325.
12. Mwangangi JM, Mbogo CM, Muturi EJ, et al. Spatial distribution and habitat characterisation of *Anopheles* larvae along the Kenyan coast. *J Vector Borne Dis*. 2007;44(1):44–51.
13. Rozendaal JA. *Vector Control—Methods for use by Individuals and Communities*. Geneva: World Health Organization; 1997:411.
14. Liu S, Kelvin DJ, Leon AJ, Jin L, Farooqui A. Induction of Fas mediated caspase-8 independent apoptosis in immune cells by *Armigeres subalbatus* saliva. *PLoS One*. 2012;7(7):e41145.
15. Hill LA, Davis JB, Haggood G, et al. Rapid identification of *Aedes albopictus*, *Aedes scutellaris*, and *Aedes aegypti* life stages using real-time polymerase chain reaction assays. *Am J Trop Med Hyg*. 2008;79(6):866–875.
16. Huang YM. The subgenus stegomyia of *Aedes* in Southeast Asia. I—the scutellaris group of species. *Contrib Am Entomol Inst*. 1972;9:1–109.
17. Majambere S, Fillinger U, Sayer DR, Green C, Lindsay SW. Spatial distribution of mosquito larvae and the potential for targeted larval control in the Gambia. *Am J Trop Med Hyg*. 2008;79(1):19–27.
18. Fillinger U, Kannady K, William G, et al. A tool box for operational mosquito larval control: preliminary results and early lessons from the Urban malaria control programme in Dar es Salaam, Tanzania. *Malar J*. 2008;7:20.
19. Zhang XZ, Huang KJ, Wang LK, Peng XW, Fu XZ, Liu HL. Mosquito species (diptera: culicidae) reported from Shandong Province, China. *J Entomol Sci*. 2011;46(3):247–255.
20. Liu LP, Lin LF, Zhang ZH. Survey on the resistance of *Culex quinquefasciatus* several common insecticides in cities of guangdong province. *Chin J Vector Biol Control*. 1999;6:410–413.
21. Liu SL, Cui F, Yan SG, Qiao CL. Investigation of organophosphate and pyrethroid resistance in vector mosquitoes in China. *Chin J Vector Biol Control*. 2011;2:184–188.
22. Wu FZ, Wang YY, Gu BG. The present of health insecticide and the development prospect in China. *Pestic Sci Admin*. 2011;5:16–19.
23. *Ministry of Agriculture of the People's Republic of China*. 2011 the Pesticide Management Information Compilation. Beijing: Chinese pesticide press; 2011:1325–1326.
24. Fillinger U, Knols BG, Becker N. Efficacy and efficiency of new *Bacillus thuringiensis* var israelensis and *Bacillus sphaericus* formulations against afro-tropical anophelines in Western Kenya. *Trop Med Int Health*. 2003;8(1):37–47.
25. Howard AF, Omlin FX. Abandoning small-scale fish farming in western Kenya leads to higher malaria vector abundance. *Acta Trop*. 2008;105(1):67–73.
26. Ndenga BA, Simbauni JA, Mbugi JP, Githeko AK, Fillinger U. Productivity of malaria vectors from different habitat types in the Western Kenya highlands. *PLoS One*. 2011;6(4):e19473.
27. Gu W, Novak RJ. Habitat-based modeling of impacts of mosquito larval interventions on entomological inoculation rates, incidence, and prevalence of malaria. *Am J Trop Med Hyg*. 2005;73(3):546–552.