

Supernumerary Chromosome Variants in Dichroplus elongatus (Acrididae): Fluorescent Banding and Cline Variation Pattern

Authors: Rosetti, Natalia, Rebagliati, Pablo, and Remis, Maria Isabel

Source: Journal of Orthoptera Research, 19(2): 261-265

Published By: Orthopterists' Society

URL: https://doi.org/10.1665/034.019.0212

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Supernumerary chromosome variants in Dichroplus elongatus (Acrididae): fluorescent banding and cline variation pattern

Submitted May 28, 2010, accepted September 6, 2010

NATALIA ROSETTI, PABLO REBAGLIATI AND MARIA ISABEL REMIS

Departamento de Ecología, Genética y Evolución, Facultad de Ciencias Exactas y Naturales, University of Buenos Aires, C1428EHA Buenos Aires, Argentina. Email: mariar@ege.fcen.com.ar

Abstract

Dichroplus elongatus is a South American grasshopper that shows simultaneous polymorphisms for B chromosomes and several supernumerary segments located in chromosome pairs S10, S9, and M6 (SS10, SS9 and SS6 respectively), in natural populations from Argentina. Heterochromatin characterization with DAPI-CMA, banding reveals that B chromosomes show an interstitial GC-rich band. The distal SS10 segment seems to also be GCrich, whereas the proximal supernumerary segments (SS6 and SS9) show two different heterochromatin regions: one GC-rich region (CMA3+/DAPI-) and the other that has no specificity to AT or CG base pairs (CMA,/DAPI dull). The results obtained permit us to distinguish two types of constitutive heterochromatin in the supernumerary segments of *D. elongatus*. The variation in the frequency of these chromosome polymorphisms was analysed with respect to some climatic and geographic variables. There is considerable B chromosome variation between populations, and this variation correlates negatively with latitude and positively with altitude and mean annual temperature. The incidence of the proximal SS9 segment is positively correlated with altitude. The pattern of chromosome variation is discussed in relation to the population's ability to maintain supernumerary DNA in more climatically favorable geographic regions.

Key words

B chromosomes, supernumerary segments, CMA₃/DAPI banding, clinal pattern

Introduction

The evolution of many Orthoptera groups is associated with chromosome rearrangements and in several cases, variation in the amount of heterochromatin has played an important role in these chromosome changes.

Constitutive heterochromatin can be observed in the genome during the cell cycle of all types of cells as C or fluorescent bands, which occur in different parts of individual chromosomes (Shibata et al. 1999). The pattern of heterochromatic bands is detected as differences in the number of bands and/or the amount of heterochromatin (Vershinin et al. 1996). Moreover, in Orthoptera, as in other animal and plant species, the genome also contains heterochromatic supernumerary elements. These chromosome variants are dispensable elements for the normal growth and development of the organism and may occur as B chromosomes or supernumerary segments.

B or supernumerary chromosomes are generally heterochromatic, fail to pair with the normal member of the complement (A chromosomes) during meiosis (unlike most chromosome mutations) and often show accumulation (*i.e.*, drive) and follow their own evolutionary pathway (Jones & Rees 1982, Beukeboom 1994, Camacho *et al.* 2000). Supernumerary segments are usually

heterochromatic elements integrated into the normal complement and according to their position within the involved chromosome, are usually nominated as distal, interstitial, proximal or extrinsic (Hewit 1979, John 1981). Simultaneous polymorphisms for B chromosomes and supernumerary segments were found in many grasshopper species (Hewitt 1979, Colombo 1989, Cabrero *et al.* 1997, Perfectti *et al.* 2000).

The DNA composition of heterochromatic regions has been intensively investigated in order to explain their genetic properties. Common procedure involves the use of different fluorochromes able to differentiate DNA regions based on their nucleotide composition. Notorious examples include 4'-6-diamidino-2-phenylindole (DAPI), which highlights AT-rich regions, and chromomycin A3 (CMA₃), which shows preference for GC-rich regions (Sumner 2003). These techniques have been used to cytogenetically analyze both intra- and interspecific differences in several Orthoptera species (Rufas *et al.* 1988; Bella *et al.* 1990, 1991, 1993; Lopez Fernandez *et al.* 1989, 1992; Loreto & de Souza 2000).

The simultaneous analysis of B chromosomes and supernumerary segments variation in different populations offers the opportunity to analyze patterns of chromosome variation in nature. *Dichroplus elongatus* (Orthoptera: Acrididae) is a South American grasshopper that shows simultaneous polymorphisms for B chromosomes and several supernumerary segments located in the chromosome pairs S10, S9, and M6 in natural populations from Argentina (Remis & Vilardi 1986, Loray *et al.* 1991, Clemente *et al.* 1994). Both forms of supernumerary heterochromatin were found to influence chiasma frequency and distribution (Loray *et al.* 1991, Clemente *et al.* 1994). Moreover, B chromosomes also affect male fertility by increasing the production of macro- and microspermatids (Loray *et al.* 1991). They also affect other fitness components, such as mating success, viability and female fecundity (Rosetti *et al.* 2007, 2008).

Previous population cytogenetic studies on the grasshopper *D. elongatus* demonstrated different degrees of differentiation among populations for each heterochromatic variant (Remis *et al.* 1998, 2004). Furthermore, the hierarchical analyses of molecular differentiation at the mitochondrial DNA level, an essentially neutral marker, revealed a discrepancy with respect to chromosome differentiation and suggested that the pattern of B chromosomes might not be explained by historical factors (Clemente *et al.* 2000). These results suggest that the pattern of chromosome variation cannot be explained by the interaction between genetic drift and migration.

In the present work, we characterize different chromosome variants using fluorescent banding and analyze the pattern of simultaneous variation in relation to climatic and geographical factors in natural populations of *D. elongatus* from eastern Argentina.

Material and Methods

Biological material.—The standard karyotype of *D. elongatus* is composed of 22 telocentric autosomes and a XO/XX sex-determination system (Remis & Vilardi 1986). Natural Argentinean populations of this species are polymorphic for mitotically unstable B chromosomes and supernumerary segments in M6, S9 and S10 pairs (SS6, SS9 and SS10 respectively). Adult males of this grasshopper were collected from the Argentinean natural populations in Raco and Horco Molle from Tucumán province, Venado Tuerto located in Santa Fé province, Gualeguaychu and Colón from Entre Ríos province, and Mocoretá located in Corrientes. Specimens were collected during the years 2002, 2003, 2004, 2005 and 2007.

Chromosome preparations.—Males were dissected, and their testes fixed in 3:1 ethanol: acetic acid and stored at -20°C. Conventional chromosome preparations were performed by squashing several follicles in propionic hematoxylin (Remis & Vilardi 1986). A minimum of 10 meiotic cells per individual were examined to determine chromosome constitution.

Fluorescent staining with CMA₃ (chromomycin A3) and DAPI (4'-6-diamidino-2-phenylindole) was carried out on male individuals from Raco, Horco Molle and Venado Tuerto populations. For this, a few testes follicles were squashed in 45% acetic acid; the coverslip was then removed using the dry-ice method, and the slide air dried. Sequential DAPI-CMA₃ banding was performed, using the technique described by Rebagliati *et al.* (2003).

Photographs.—The best preparations were photographed using an analog Leica Wild MPS52 camera on a Leica DMLB fluorescence microscope, with filters for DAPI and CMA₃. Images were also taken using Kodak color ASA400 film and processed with Adobe Photoshop version 7.0.

Geographic chromosome pattern analysis.—B chromosome frequencies were transformed according to Christiansen et al.'s (1976) formula:

$$X_i = (p_1 - p_0) \sqrt{\frac{N_1}{p_0(1 - p_0)}}$$

where p_0 is the mean B-chromosome frequency, p_1 represents the B-chromosome frequency in the ith population and N_1 is the number of individuals sampled per population. This transformation normalizes the distribution and weights sampling sizes.

The relationships between chromosome frequencies and geographical (latitude, altitude and longitude) and climatic (mean annual temperature) variables were analyzed by means of nonparametric correlation using the STATISTICA program (STATISTICA STATSOFT Inc. 1996). Bonferroni's correction for

multiple testing was applied.

Temperature data were obtained using the Local Climate Estimator from www.fao.org/sd/dim_en3/en3_051002_en.

Results

Chromosome polymorphisms.—Frequencies of simultaneous polymorphisms for B chromosomes and supernumerary segments in the chromosome pairs M6 (SS6), S9 (SS9) and S10 (SS10) were analyzed in four populations of the grasshopper *D. elongatus* from Eastern Argentina, together with their incidences. They are summarized in Table 1.

Fluorescent banding patterns.—Sequential banding with CMA₃/DAPI revealed positive CMA₃ (CMA₃+) and negative DAPI (DAPI-) bands in most of the autosomes. The comparison of the two different fluorochrome banding patterns showed that CMA₃+ and DAPI- bands were colocalized in terminal regions (Fig. 1A, B). In some bivalents, terminal CMA₃+/DAPI- bands are displayed in both centromeric and telomeric positions.

The sex chromosome (X) showed proximal and distal CMA₃+/DAPI- bands (Fig. 1C, D and Fig. 2A, B), whereas B chromosomes are characterized by showing an interstitial CMA₃+/DAPI- band (Fig. 1C, D).

Heterozygote individuals for the distal SS10 segment showed a heteromorphic bivalent with telomeric CMA₃+/DAPI- bands of different sizes (Fig. 2A, B). Most of the SS10 segment is CMA₃+/DAPI-, revealing a bigger bright band in the heteromorphic bivalent.

Heterozygote individuals for the proximal SS9 and SS6 segments showed a bigger CMA₃+/DAPI- band than standard bivalents (Fig. 2C-F). Both proximal segments are composed of a CMA₃+/DAPI-region more proximal to the centromere and a CMA₃+/DAPI- region.

Geographical distribution pattern.—The geographical distribution of chromosome polymorphisms was analyzed from 2002 to 2007 in six *D. elongatus* populations, four of which are analyzed in the present paper, two being analyzed previously (Rosetti *et al.* 2007) (Table 1).

B chromosomes and proximal SS6 segments are geographically widespread in the studied area, whereas distal SS10 and proximal SS9 were absent in some populations. Supernumerary segments are always present in heterozygous condition, probably due to their low frequency in the population. SS6 and SS10 segments may occur simultaneously in the same individual. Three out of 246 analyzed males were double heterozygote for these variants, exceeding the expected frequency of this karyotype.

The geographic distribution of chromosome polymorphisms was analysed with respect to some climatic and geographic variables. There is considerable B-chromosome variation between populations and this variation correlates negatively with latitude (r = -0.943,

Table 1. Frequencies of B chromosomes (B) and supernumerary segments in M6 (SS6) S9 (SS9) and S10 (SS10), along with geographic and climatic data from natural populations of *D. elongatus* collected in Argentina. N= sample size. * Data from Rosetti *et al.* 2007.

Location	В	SS10	SS6	SS9	N	Latitude	Longitude	Altitude	Mean °C
Venado Tuerto	0.029	0.014	0.043	0.014	35	33°45′	61°57′	40	16.83
Gualeguaychú	0.091	0.015	0.030	0.00	33	33°06′	58°32′	40	17.18
Colón	0.032	0.00	0.016	0.00	31	32°13′	58°09′	40	17.88
Mocoreta	0.111	0.019	0.019	0.019	27	30°38′	57°58′	60	19.01
Raco- Nogues*	0.132	0.026	0.046	0.026	76	26°40′	65°10′	650	19.19
Horco Molle-Yerba Buena*	0.136	0.045	0.034	0.023	44	26°49′	65°19`	550	18.83

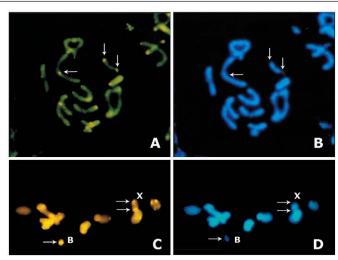


Fig. 1. *D. elongatus*. CMA₃ staining (A, C); DAPI staining (B, D). A, B) Standard complement at late diplotene. C, D) Metaphase I showing a B chromosome (B). White arrows show CMA₃ bright - DAPI negative. Bar= 10μm. For color version, see Plate IV.

P= 0.0048) and positively with altitude (r = 0.941, P = 0.005). The frequency of the proximal SS9 segment is positively correlated with the altitude, whereas the SS10 distal segment seems to be positively associated with longitude, though no significant correlation was detected (r = 0.77, P = 0.07). No significant associations were detected between geographic variables and the frequency of the SS6 segment. With respect to climatic variables, B chromosomes showed a significant positive correlation with the mean annual temperature (r = 0.88, P = 0.018).

The analysis of geographic chromosome patterns revealed that B chromosomes tend to decrease from the North towards the South, while both B chromosomes and SS9 have a tendency to increase with altitude.

Discussion

Natural populations of *D. elongatus* from Argentina are polymorphic for the presence of several additional dispensable elements.

Our results, based on fluorescence banding, demonstrate heterogeneity in the constitution of the supernumerary heterochromatin. In insects, especially grasshoppers, chromosomal regions with differential fluorescence may be rich in GC base pairs. All fluorescent bands are rich in GC base pairs, a condition commonly found in grasshoppers (Bella et al. 1991, 1993; Loreto & de Souza 2000). The distal SS10 segment is mainly composed of GC-rich heterochromatin, whereas proximal supernumerary segments (SS6 and SS9) showed two different heterochromatic regions: one GC-rich (CMA,+/DAPI-) and another that has no specificity to AT or CG base pairs (CMA₂/DAPI dull). The centromeric region of the S6 and S9 autosomes, as well as the telomeric region of the S10 pair, showed a bright CMA2+/DAPI- band. These results are in agreement with the tentative hypothesis arguing that supernumerary chromosomal segments in the genome of D. elongatus may have originated by duplication of pre-existing heterochromatic regions.

Different heterochromatin constitutions were also detected in the supernumerary segments of another grasshopper, *Dociostaurus genei*, which appear differentially stained with DAPI banding and thus seem to be AT-rich (Rodriguez Iñigo *et al.* 1993, 1996, 1998).

Previous studies in *D. elongatus* showed that the nucleolus is always associated with the proximal end of the S10 pair (Remis 1989). An association between CMA₃+ bands and NORs is rather

common in insects, especially grasshoppers (Camacho *et al.* 1991; Bella *et al.* 1990, 1993; Loreto & de Souza 2000; Bakkali *et al.* 2001). However, in the present paper there is no evidence of any differential positive CMA, band in the proximal region of the S10 pair.

B chromosomes showed an interstitial GC-rich band. However no standard chromosome of the complement exhibited CMA₃ positive regions in interstitial position. It is widely accepted that B chromosomes are derived from polysomic A chromosomes (Jones & Rees 1982, Camacho *et al.* 2000). Contemplating this hypothesis of the origin of such supernumerary chromosomes, our results would seem to indicate that B chromosomes should have suffered posterior rearrangements to explain their banding pattern, compared to that of the chromosomes of the normal complement.

The prevalence of supernumerary variants within a species depends on the degree to which the species tolerates these extra genetic elements and the probable existence of an accumulation mechanism of the supernumerary variants (Remis *et al.* 2004). In the present paper some supernumerary variants of *D. elongatus* exhibited chromosome interpopulation variation in their frequencies. Here, as in previous population cytogenetic studies (Remis *et al.* 1998, 2004), we detected different degrees of differentiation among populations for each heterochromatic variant. Under a neutral scenario (interaction between genetic drift and migration) the differentiation degree is expected to be similar between distinct markers. Therefore, different factors may be moulding chromosome frequencies in natural populations of *D. elongatus*. There is no reason to reject the most likely hypothesis to explain SS9 chromosome variation patterns through historical factors in the studied area.

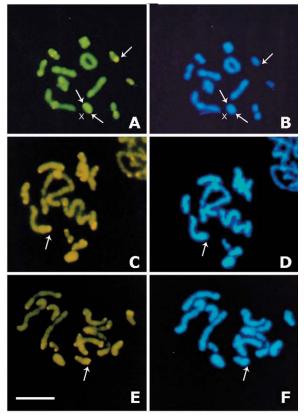


Fig. 2. *D. elongatus.* CMA₃ staining (A, C, E); DAPI staining (B, D, F). A, B) Late diakinesis showing distal SS10 segment. C, D) Diplotene showing proximal SS6 segment. E, F) Diplotene showing proximal SS9 segment. White arrows show CMA₃ bright - DAPI negative. Bar= 10μm. For color version, see Plate IV.

Both the size and frequency of these supernumerary segments may be shaped by their age, so that the older the segment is, the more likely it is to have experienced duplications and thus increase in size, and the closer its frequency will be to equilibrium.

The frequency of the B chromosomes in *D. elongatus* seem to decrease with latitude and increase with altitude and temperature. Supernumerary chromosome frequency often shows geographical variation which may be attributed to the geographic origin and evolutionary history of the B chromosome and/or the ability of the populations to maintain B chromosomes in climatically more favorable regions (Camacho *et al.* 2000). Indeed, several reports show spatial chromosome differentiation with B-chromosome clines (Hewitt & Brown 1970; Parker *et al.* 1991; Zima & Macholan 1995; Cabrero *et al.* 1997; Tsurusaki & Shimada 2004; Bakkali *et al.* 1999, 2002).

Some studies analyzed the relative significance of these factors for the patterns of geographic variation (Hewitt & Brown 1970; Bougourd & Parker 1979; Cabrero et al. 1997; Colombo 1989; Tsurusaki & Shimada 2004; Colombo & Confalonieri 2004; Perfectti et al. 2004a, 2004b; Bakkali et al. 1999, 2002; Bakkali & Camacho 2004). In the grasshopper *Myrmeleotettix maculatus*, B chromosomes were common in climatically more favorable regions (Hewitt & Brown 1970). The distribution of the B chromosomes in British populations may be attributable to the overall level of stringency or favorability of the environment. Cytogenetic population analysis of the grasshopper Eyprepocnemis plorans along four rivers belonging to the Segura basin (Spain), demonstrated that the geographical distribution of B chromosomes was molded mainly by historical nonselective events (Cabrero et al. 1997), a conclusion supported by similar studies on the B chromosomes of the same species in Moroccan populations (Bakkali et al. 1999, 2002; Bakkali & Camacho 2004). Clinal variation of B-chromosome frequency was also reported in the harvestman Psathyropus tenuipes from Japanese Islands (Tsurusaki & Shimada 2004). Previous studies demonstrated an association between the number of B chromosomes and resistance to gregarine infection in natural populations of this species (Gorlov & Tsurusaki 2000). The authors suggested that the clinal variation in B-chromosome frequency might be attributed to variability in the geographic abundance of gregarines.

Given that we have a negative correlation between latitude and altitude in the studied area and that the mean temperature is higher at higher altitudes, we suspect that the effects of these three variables on B-chromosome frequency in *D. elongatus* is due to the effect of just one of them. *A priori*, the mean temperature would be a good candidate, since at higher, closer to optimal, temperatures any negative effect of the B chromosome on the fitness of the carrier would be less pronounced. In other words, the harmful effects of the B chromosome on the fitness of the carrier are probably worse at nonoptimal temperatures.

The pattern of B-chromosome distribution in *D. elongatus* may be more complex and further studies, to relate B chromosome distribution to their geographic distribution in the light of different geographic/climatic variables, may help us understand the maintenance of such chromosome variants in nature.

Acknowledgments

Funding provided by Universidad de Buenos Aires (X-186) through grants to Dr M. I. Remis is gratefully acknowledged.

References

- Bakkali M., Cabrero J., Lopez-Leon M.D., Perfectti F., Camacho J.P. 1999. The B chromosome polymorphism of the grasshopper *Eyprepocnemis plorans* in North Africa. I. B variants and frequency. Heredity 83: 428-34.
- Bakkali M., Cabrero J., López-León M.D., Perfectti F., Camacho J.P. 2001. Population differences in the expression of nucleolus organizer regions in the grasshopper *Eyprepocnemis plorans*. Protoplasma 217: 185-190.
- Bakkali M., Camacho J.P. 2004. The B chromosome polymorphism of the grasshopper *Eyprepocnemis plorans* in North Africa. IV. Transmission of rare B chromosome variants. Cytogenetic and Genome Research 106: 332-337.
- Bakkali M., Perfectti F., Camacho J.P. 2002. The B-chromosome polymorphism of the grasshopper *Eyprepocnemis plorans* in North Africa: II. Parasitic and neutralized B1 chromosome. Heredity 88: 14-18.
- Bella J.L., Westerman M., López-Fernández C., de la Torre J., Rubio J.M., Gosálvez J. 1991. Sex chromosome and autosome divergence in *Podisma* (Orthoptera) in western Europe. Genetics Selection Evolution 23: 5-13.
- Bella J.L., Serrano L., Hewitt G.M., Gosálvez J. 1993 Heterochromatin heterogeneity and rapid divergence of the sex chromosomes in *Chorthippus parallelus* and *C. perythropus* (Orthoptera). Genome 36: 542-547.
- Beukeboom L.W. 1994. Bewildering Bs: an impression of the 1st B chromosome conference. Heredity 73: 328-336.
- Bougourd S.M., Parker J.S. 1979. The B-chromosome system of *Allium schoenoprasum* II. Stability, inheritance and phenotypic effects. Chromosoma 75: 369-383.
- Cabrero J., Lopez M.D., Leon Gomez R., Castro A.J., Martin Alganza A., Camacho, J.P M. 1997. Geographical distribution of B chromosome in the grasshopper *Eyprepocnemis plorans*, along a river basin, is mainly shaped by non-selective historical events. Chromosome Research 5: 194-198.
- Camacho J.P.M., Cabrero J., Viseras E., Lopez-Leon M.D., Navas-Castillo J. 1991. G banding in two species of grasshopper and its relationship to C, N and fluorescent banding techniques. Genome 34: 638-643.
- Camacho J.P.M., Sharbel T.F., Beukeboom L.W. 2000. B-chromosome evolution. Philosophical Transactions Royal Society London 355: 163-178
- Clemente M., Remis M.I., Vilardi J.C. 2000. Mitochondrial DNA variation in the South American grasshopper *Dichroplus elongatus* (Orthoptera: Acrididae). Annals Entomological Society of America 93: 653-662.
- Colombo P.C. 1989. Chromosome polymorphisms affecting recombination and exophenotypic traits in *Leptysma argentina* (Orthoptera): a populational survey. Heredity 62: 289-299.
- Colombo P., Confalonieri. V. 2004. Cytogeography and the evolutionary significance of B chromosomes in relation to inverted rearrangements in a grasshopper species. Cytogenetic and Genome Research 106: 351-358.
- Gorlov I.P., Tsurusaki N. 2000. Morphology and meiotic/mitotic behavior of B chromosomes in a Japanese harvestman, *Metagagrella tenuipes* (Arachnida: Opiliones): no evidence for B accumulation mechanisms. Zoological Science 17: 349-355.
- Hewitt G.M. 1979. Animal Cytogenetics, pp. 1-170. In: Bernard J. et al. (Eds) Orthoptera. Gebruder Borntraeger, Berlin.
- Hewitt G.M., Brown F.M. 1970. The B-chromosome system of *Myrmeleotettix maculatus* V. A steep cline in East Anglia. Heredity 25: 363-371.
- *maculatus* V. A steep cline in East Anglia. Heredity 25: 363-371.

 Jones R.N., Rees H. 1982. B chromosomes. Academic Press, New York.
- López-Fernández C., Gosälvez J., Mezzanotte R. 1989. Heterochromatin heterogeneity in *Oedipoda germanica* (Orthoptera) detected by *in situ* digestion with restriction endonucleases. *Heredity* 62: 269-277.
- López-Fernández C., Mezzanotte R., Gosálvez J. 1992. Autosomal, sex and B chromosomes in *Eyprepocnemis plorans* (Orthoptera) viewed with restriction endonuclease *in situ* digestion. Heredity 68: 365-372.
- Loray M.A., Remis M.I., Vilardi J.C. 1991. Parallel polymorphism for supernumerary heterochromatin in *Dichroplus elongatus*. Genetica 84: 155-163
- Loreto V., Souza M.J. 2000. Karyotype, constitutive heterochromatin and nucleolar organizer regions (NORs) in *Belosacris coccineipes* (Acrididae-Leptysminae). Genetics and Molecular Biology 23: 575-579.

- Paker J.S., Lozano R., Taylor S., Ruiz Rejon M. 1991. Chromosome structure of populations of *Silla autumnalis* in the Iberian Peninsula. Heredity 67: 287-297.
- Perfectti F., Pita M., De La Vega C.G., Gosalvez J., Camacho J.P.M. 2004a. Spatio-temporal dynamics of a neutralizad B chromosome in the grasshopper *Eyprepocnemis plorans*. Cytogenetic and Genome Research 106: 376-385.
- Perfectti F., Cabrero J., Lopez-Leon M.D. Muñoz E., Pardo M.C., Camacho J.P.M. 2000. Fitness effect analysis of a heterochromatic supernumerary segment in the grasshopper *Eyprepocnemis plorans*. Chromosome Research 8: 425-433.
- Perfectti F., Corral J.M., Mesa J.A., Cabrero J., Bakkali M., López-León M.D., Camacho J.P. 2004b. Rapid suppression of drive for a parasitic B chromosome. Cytogenetic and Genome Research 106: 338-343.
- Rebagliati P.J., Papeschi A.G., Mola L.M. 2003. Meiosis and fluorescent banding in *Edessa meditabunda* and *Edessa rufomarginata* (Heteroptera: Pentatomidae: Edessinae). European Journal of Entomology 100: 11-18.
- Remis M.I., Vilardi J.C. 1986. Meiotic behaviour and dosage effect of B chromosomes on recombination in *Dichroplus elongatus* (Orthoptera: Acrididae). Caryologia 39: 287-308.
- Remis M.I. 1989. Effects of supernumerary heterochromatin on chiasma condition in two species of Acrididae (Orthoptera). Genetica 79: 53-61
- Remis M.I., Clemente M., Pensel S., Vilardi J.C. 1998. Non random distribution patterns of supernumerary segments and B chromosomes in *Dichroplus elongatus* (Orthoptera). Hereditas 129: 207-213.
- Remis M.I., Pensel S., Rosetti N. 2004. Interaction among different heterochromatic variants in the grasshopper *Dichroplus elongatus*. Hereditas 141: 181-185.
- Rodriguez Iñigo E., Bella J.L., Garcia de La Vega C. 1993. Heterochromatin differential between two species of the genus *Dociostaurus* (Orthoptera-Acrididae). Heredity 70: 458-465.
- Rodríguez Iñigo E., Fernández Calvín B., Capel J., García de la Vega C. 1996. Equilocality and heterogeneity of constitutive heterochromatin: *in situ* localization of two families of highly repetitive DNA in *Dociostaurus genei* (Orthoptera). Heredity 76: 70-76.
- Rodríguez Iñigo E., Mason P.L., Rufas J.S., García de la Vega C. 1998. Effects of supernumerary heterochromatin on chiasma formation and chromosome segregation in *Dociostaurus genei* (Orthoptera). Heredity 80: 353-360.
- Rosetti N., Vilardi J.C., Remis M.I. 2007. Effects of B chromosomes and supernumerary segments on morphometric traits and adult fitness components in the grasshopper *Dichroplus elongatus* (Acrididae). Journal of Evolutionary Biology 20: 249-259.
- Rufas J.S., Esponda P., Gosalvez J. 1985. NOR and nucleolus in the spermatogenesis of acridoid grasshoppers. Genetica 66: 139-144.
- Shibata F., Masahiro H., Yutzo K. 1999. Molecular cytogenetic analysis of supernumerary heterochromatic segments in *Rumex acetosa*. Genome 43: 391-397.
- Statistica Statsoft Inc. 1996. Statistica 5 for Windows (Computer Program Manual). Statistica, Tulsa, OK.
- Sumner A.T. 2003. Chromosomes: Organization and Function. Blackwell Publishing, Oxford.
- Tsurusaki N., Shimada T. 2004. Geographic and seasonal variations of the number of B chromosomes and external morphology in *Psathyropus tenuipes* (Arachnida: Opiliones). Cytogenetic and Genome Research 106: 365-375.
- Vershinin A.V., Alkhimova A.G., Heslop-Harrison J.S. 1996. Molecular diversification of tandemly organized DNA sequences and heterochromatic chromosome regions in some Triticeae species. Chromosome Research 4: 517-525.
- Zima J., Macholán M. 1995. B chromosomes in the wood mice (genus *Apodemus*). Acta Theriologica (Suppl) 3: 75-86.