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NUCLEAR SSR MARKERS FOR *MISCANTHUS*, *SACCHARUM*, AND RELATED GRASSES (SACCHARINAE, POACEAE)¹

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- **Premise of the study:** We developed nuclear simple sequence repeat (SSR) markers for the characterization of the biomass crop *Miscanthus*, especially *M. sacchariflorus*, *M. sinensis*, and *M. ×giganteus*, and tested for cross-species amplification.
- **Methods and Results:** Twenty-nine SSR markers (di- and tetranucleotide repeats) were developed from DNA sequences obtained from 192 clones from an enriched genomic library of *M. sinensis*. All markers were successfully amplified in *M. sacchariflorus*, *M. sinensis*, and *M. ×giganteus*, and 19 amplified across a broad range of *Miscanthus* species. Polymorphism information content and expected heterozygosity values (19 locus sample) were 0.88 and 0.89, respectively, for *M. sinensis*, 0.48 and 0.54 for *M. sacchariflorus*, and were the lowest in *M. ×giganteus* (0.33, 0.41). Thirteen out of 19 primer pairs showed cross-species amplification in non-*Miscanthus* sensu stricto taxa.
- **Conclusions:** The new set of 29 SSR markers will be of high value for characterizing *Miscanthus* germplasm collections, for prebreeding, and for assessing variation in natural populations.

Key words: cross-species amplification; microsatellites; *Miscanthus*; Poaceae; *Saccharum*; SSRs.

Miscanthus Andersson is under development as a biomass crop and has been characterized by a wide range of markers including amplified fragment length polymorphism (AFLP; Hodkinson et al., 2002), restriction fragment length polymorphism (RFLP; Hernández et al., 2001), inter-simple sequence repeat (ISSR) PCR, and DNA sequences of nuclear and chloroplast regions generated using conventional (Hodkinson et al., 2002) and next-generation approaches including RNAseq and genotyping by sequencing (GBS; Ma et al., 2012). Simple sequence repeat (SSR) markers from maize and *Brachypodium distachyon* (L.) P. Beauv. (Hernández et al., 2001; Zhao et al., 2011) have been successfully applied to *Miscanthus*, and chloroplast SSRs have been developed by De Cesare et al. (2010).

Some nuclear SSR markers have also been developed, such as those for *M. sinensis* Andersson, *M. floridulus* (Labill.) Warb. (Ho et al., 2011), and several other *Miscanthus* species (Zhou et al., 2011). However, there is a need to develop additional SSR markers for *Miscanthus* as the total number of available markers is limited. There is also a need to test these markers on a range of species, especially *M. sacchariflorus* (Maxim.) Hack., *M. sinensis*, and *M. ×giganteus* Greef & Deuter ex Hodk. & Renvoize as these comprise the main species of germplasm collections. SSRs developed from *Saccharum officinarum* L. expressed sequence

tags (ESTs) have been recently used by Kim et al. (2012) to generate genetic maps of *M. sacchariflorus* and *M. sinensis* with genome coverage of 72.7% and 84.9%, respectively. The numbers of linkage groups found for the two maps (40 for *M. sacchariflorus* and 23 for *M. sinensis*) were higher than the basic chromosome number for *Miscanthus* ($x = 19$). Additional markers, such as those generated in this study, will be required to make more saturated maps, especially from noncoding regions that are underrepresented in current maps. Recently, single-nucleotide polymorphism (SNP) markers generated using GBS markers have been used for high-resolution mapping and identified all 19 linkage groups in *M. sinensis* (Ma et al., 2012).

METHODS AND RESULTS

DNA samples were either freshly extracted or obtained from the DNA bank at Trinity College, Dublin. Fresh leaves were frozen in liquid nitrogen and ground manually to a fine powder. Total genomic DNA was extracted following a modified cetyltrimethylammonium bromide (CTAB) method (Hodkinson et al., 2007). Total genomic DNA from the *M. sinensis* clone SW217 was used by ATG Genetics (Vancouver, British Columbia, Canada) to build a nuclear microsatellite-enriched library. After digestion with multiple 4-cutter restriction enzymes, enrichment for SSRs containing fragments was obtained through biotinylated TC_n, TG_n, and GATA_n simple sequence motifs. The selected fragments were cloned into the EcoRI site of the plasmid pUC19 and screened for positive clones using ³²P-labeled TC_n, CA_n, and GATA_n simple sequence motifs. Two 96-well microtiter plates containing single positive bacterial colonies, one selected for the presence of dinucleotide repeats and the second for the presence of tetranucleotide repeats, were produced. The 192 clones were sequenced by AGOWA GmbH (Berlin, Germany), and SSRs were identified in the clones using 'find microsat Win32' (Salamin, unpublished). All 192 clones contained SSRs (96 dinucleotides and 96 tetranucleotides). Eighty primer pairs were designed equally among these sets using Primer3 software (Rozen and Skaletsky, 2000; <http://frodo.wi.mit.edu/primer3/>) and tested with PCR. Selection of the final sample of 29 primers was based on clarity of product on an agarose gel. Primer details and GenBank numbers are provided in Table 1.

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TABLE 1. Characteristics of 29 primer pairs developed for microsatellite genotyping.

Locus	Clone, GenBank accession no.	Repeat motif	Fluorescent dye	Forward primer sequence (5'–3')	Reverse primer sequence (5'–3')	T _a (°C)	Sequence length (bp)	SSR size (bp)
Mis-1	SSR1A10, KF130838	(TCTA) ₂₀	FAM	CAGTCCTTGGAGCAGGCTAT	AAGATCTCAAACCTATAGTC	54	202	80
Mis-13	SSR1F10, KF130839	(TAGA) ₁₉	ROX	CGGACTAACTTGTGAATCTT	GTCCTTGGAGCAGGCTATGA	54	230	76
Mis-14	SSR1F12, KF130840	(GATA) ₁₅	FAM	GTAGCTGCAACTGCTAGTGT	ACTCGCATTGGTTGGTATGA	59	141	60
Mis-15	SSR1F2, KF130841	(ATCT) ₁₆	FAM	ACTACTGCATGCATCATGATG	TGCTTCGCGGCGAAGTTCA	59	195	64
Mis-16	SSR1F5, KF130842	(TATC) _{13/16}	VIC	ATCTTGCCCTAGGATGCATTAG	TGGTCTATTACAACAAGGCT	60	264	52+64*
Mis-20	SSR1G12, KF130843	(TCTA) ₁₇	TAMRA	TAGCTGAGCTGTCTATGGTA	TAGCCATTGAGGCTAAGGAT	54	249	68
Mis-22	SSR1G8, KF130844	(TAGA) ₁₇	VIC	CGAGCGAGCCTGCATGTGTG	TTGACGTCAGCAAGATATTG	54	173	68
Mis-23	SSR1G9, KF130845	(ATCT) ₁₅	TAMRA	CACGAAGTGAATCAGCATGC	GTAGCTGCAACTGCTAGTGT	60	240	60
Mis-24	SSR1H10, KF130846	(AGAT) ₁₅	VIC	AGGCAGCATCCAAACATGTC	ATGCTGCTACCCAAGAGATG	60	324	60
Mis-33	SSR2B7, KF130847	(CT) ₂₀	TAMRA	TGACATAGGGCTACACATAT	CGAGTGAGGCAGCTAGTTCA	48	242	40
Mis-37	SSR2D9, KF130848	(TC) ₃₄	FAM	GAATGCAGTCATCAGCAGCT	TGGACATCTCTAGGTTGATC	54	218	68
Mis-41	SSR2F5, KF130849	(GA) ₂₄	ROX	ATAATGCAGGTCAGTTCAAC	CGCAGCTAGCTGCTTGTGAC	54	226	48
Mis-42	SSR2F6, KF130850	(AG) ₃₁	FAM	GCCGAGCCTCCCAAGCCT	ATCCGAGCCATGTATGCACG	54	206	62
Mis-50	SSR2H9, KF130851	(GA) ₂₁	ROX	TACGGACGATTAACCAAGCC	CGCAAGGTGCAGGACCATCA	54	230	42
Mis-51	SSR2G4, KF130852	(TC) ₂₀	FAM	GATCCATCACGGATTCATCA	ATCATAGGCAAAACGGATCG	60	164	40
Mis-52	SSR2C11, KF130853	(GA) ₁₉	NED	TTATTGGTGCCCAAAGGTGT	AACAAGCCCTCAAGCTTCTCT	60	370	38
Mis-53	SSR2G10, KF130854	(GA) ₁₉	FAM	AGATGGCAGCTCACAAAAC	GGTGGAGATGCTCTTCTTGC	60	173	38
Mis-54	SSR2A11, KF130855	(CT) ₁₈	NED	TAAGAAACGCAGCAGCAGAA	AGTCTCCGGCTTTCTCACAA	60	226	36
Mis-55	SSR2B9, KF130856	(GA) ₁₈	VIC	CGGCTTCGAGTGATACCTTT	TACCGGATTTAAGGGGCTTT	60	250	36
Mis-59	SSR2B3, KF130857	(GA) ₁₆	FAM	GAGCTGATCGCGTAGCAAG	TTGATAAAGCAGGGGATTTG	60	152	32
Mis-60	SSR2C3, KF130858	(GA) ₁₆	FAM	AGATGGCAGCTTGTCTTGT	CCATTTGTTGAGCACGATGT	60	190	32
Mis-63	SSR1G3, KF130859	(TCTA) ₁₄	VIC	AGGCTAGCACTTCTCCAAA	CTGCCTGGTGACCCCTATAA	60	234	56
Mis-64	SSR1G6, KF130860	(AGAT) ₁₄	NED	TCCCTTAGTGTCCGTGAAG	GAGGCAGGTGTAGTCGGAGA	60	236	56
Mis-66	SSR1D5, KF130861	(CTAT) ₁₃	VIC	CATGGCTACAGGCACCTAAAA	ATAACGAGAAATGGCCGATG	60	165	52
Mis-69	SSR1F4, KF130862	(TCTA) ₁₃	NED	CCTCTGCGGATATGAGGTGT	GAAAGTACAACATGCGATGG	60	175	52
Mis-70	SSR1B10, KF130863	(TATC) ₁₂	NED	TGCGACCTTAATTTTTCAT	TTATGAACCCGACAGGGAGA	60	249	48
Mis-71	SSR1D3, KF130864	(TAGA) ₁₂	VIC	CAACCATGAGCACTTCTCCA	AACATAGGAGGCCAAGCAAA	60	179	48
Mis-78	SSR2G11, KF130865	(CT) ₁₅	NED	TCTGCAGGTGACAAGGAAGA	GTCAACCAGGCATAGTTTCAT	60	167	30
Mis-79	SSR2G9, KF130866	(CT) ₁₅	VIC	GCCAACTCGTGGATTTGAGT	CGTAGCAAGAGGGGAACAAA	60	248	30

Note: T_a = annealing temperature.

* Compound SSR separated by a nonpolymorphic region.

Twenty-nine primer sets provided reliable amplification, and 19 of these were selected to have a mixture of di- and tetranucleotide SSRs. A template DNA volume of 1 μL (40 ng μL⁻¹) was amplified with an initial denaturation of 5 min at 95°C followed by 35 cycles each with a denaturation of 1 min at 95°C,

1 min at a primer-specific annealing temperature (Table 1), and an extension of 1 min at 72°C, followed by a final extension at 72°C for 10 min. The reaction mixture (final volume) contained 1× reaction buffer containing 2 mM MgSO₄, 0.125 μM dNTPs, 0.25 μM of each primer, and 0.5 U of *Taq* DNA polymerase

TABLE 2. Genetic properties of the newly developed markers for three *Miscanthus* species.^a

Locus	<i>M. sacchariflorus</i> (n = 9)				<i>M. sinensis</i> (n = 73)				<i>M. xgiganteus</i> (n = 15)			
	A	Size range (bp)	H _e	PIC	A	Size range (bp)	H _e	PIC	A	Size range (bp)	H _e	PIC
Mis-1	2	127–161	0.375	0.305	19	125–256	0.904	0.896	3	125–161	0.370	0.340
Mis-14	2	87–119	0.663	0.604	25	87–208	0.928	0.924	2	99–119	0.500	0.375
Mis-15	3	144–148	0.620	0.548	20	144–205	0.862	0.852	2	146–148	0.500	0.375
Mis-20	2	200–234	0.320	0.269	28	197–300	0.907	0.901	2	200–234	0.499	0.375
Mis-22	1	124	0.000	0.000	14	103–174	0.837	0.818	1	124	0.000	0.000
Mis-23	3	191–223	0.625	0.555	27	191–314	0.935	0.932	2	203–223	0.499	0.375
Mis-24	1	331	0.000	0.000	21	283–361	0.905	0.899	1	331	0.000	0.000
Mis-37	5	160–200	0.789	0.756	27	160–222	0.938	0.935	3	160–226	0.531	0.420
Mis-41	2	214–215	0.444	0.346	35	197–512	0.924	0.919	1	214	0.000	0.000
Mis-42	3	206–247	0.560	0.499	21	163–247	0.909	0.903	4	183–236	0.574	0.500
Mis-50	2	207–256	0.408	0.325	25	199–260	0.869	0.859	2	207–256	0.497	0.373
Mis-51	2	136–140	0.463	0.356	24	132–176	0.887	0.879	1	140	0.000	0.000
Mis-52	6	177–207	0.806	0.777	18	170–207	0.863	0.850	3	177–207	0.557	0.457
Mis-54	5	213–236	0.796	0.763	18	207–244	0.860	0.848	4	213–224	0.647	0.586
Mis-59	7	135–155	0.840	0.820	10	123–160	0.792	0.766	4	148–155	0.678	0.618
Mis-64	4	214–258	0.740	0.692	30	194–286	0.923	0.918	2	232–258	0.476	0.363
Mis-69	3	130–143	0.612	0.541	17	105–197	0.861	0.848	2	130–138	0.500	0.375
Mis-70	3	219–237	0.595	0.526	26	211–328	0.903	0.897	2	219–225	0.500	0.375
Mis-79	3	242–266	0.540	0.466	22	235–274	0.904	0.897	4	224–252	0.479	0.427
Mean			0.537	0.481			0.890	0.881			0.411	0.333

Note: A = number of alleles; H_e = expected heterozygosity; PIC = polymorphism information content.

^a Statistics provided for species where sample size (n) was 9 or greater.

(New England BioLabs, Herts, United Kingdom). Five different fluorescent dyes were used for primer labeling to allow multiplexing, in pools (Table 1). A polyA treatment at 65°C was applied for 30 min to the PCR products. Undiluted PCR products were then sized using an ABI 3130xl automated DNA sequencer (Applied Biosystems, Carlsbad, California, USA) and the resulting peaks were scored with GeneMapper version 4.0 software (Applied Biosystems). All 29 primer pairs produced good amplification on eight test genotypes of *M. sacchariflorus*, *M. sinensis*, and *M. ×giganteus*, but 11 loci were not consistently amplified across our entire collection and were discarded from further analyses. Our final analysis therefore included 19 SSR markers. Allele number, size range, expected heterozygosity (H_e), and polymorphism information content (PIC) were calculated using PIC Calculator Extra (<http://www.genomics.liv.ac.uk/animal/pic.html>). H_e and PIC values were only calculated for *M. sacchariflorus*, *M. sinensis*, and *M. ×giganteus* because of sample size (Table 2).

Polymorphism at 19 microsatellite loci was studied in a collection of 166 individual grasses (Appendix 1), mostly belonging to the species *M. sinensis*, *M. sacchariflorus*, and *M. ×giganteus*. Fourteen individuals belonging to closely related genera were also included. All markers revealed considerable length polymorphism, with the number of alleles ranging from 13 to 44 per locus, with an average of 27.5 (Table 3). The loci amplified included a tetranucleotide repetition in nine cases and a dinucleotide repetition in the remaining 10. No major difference was observed between di- and tetranucleotide microsatellite loci in their ability to detect variation. Thirteen out of 19 primer pairs showed cross-amplification in non-*Miscanthus* species (Table 3). Average allele number was higher than the value of 12 found by Hernández et al. (2001) in a previous study using SSRs from maize. The higher number of clones used in our study (166 against 16 clones) and the introduction of species other than *M. sinensis*, *M. sacchariflorus*, and *M. ×giganteus* could account for the difference in allele number.

PIC and H_e values varied considerably among species (Table 2) and were the highest (0.88 and 0.89, respectively) for *M. sinensis*, 0.48 and 0.54 for *M. sacchariflorus*, and the lowest (0.33 and 0.41) in *M. ×giganteus*. The PIC value of *M. sinensis* (0.88) was consistent with the value of 0.83 in Hernández et al. (2001), both are higher than the average PIC value recently found by Zhao et al. (2011) in a study examining transferability of 49 microsatellite markers from *Brachypodium distachyon* to *M. sinensis*.

In the past few years, the first nuclear microsatellite markers for *Miscanthus* have been developed (Hung et al., 2009; Ho et al., 2011; Zhou et al., 2011). Both studies from Zhao et al. (2011) on transferability from *Brachypodium distachyon* P. Beauv. and from Hung et al. (2009) on nine new microsatellite loci specific for *Miscanthus*, were limited to *M. sinensis*, thus explaining the low level of polymorphism found compared to the markers in this study. Zhou et al. (2011) extended the test for their 14 newly developed markers to *M. floridulus*, *M. lutaripariensis* L. Liu ex S. L. Chen & Renvoize, and *M. sacchariflorus*, increasing the average number of alleles found to 16.1 and the PIC value to 0.76. A different approach was used by Ho et al. (2011) to develop 12 new SSR primer pairs for *Miscanthus*. They designed primers based on genic microsatellite loci (EST-SSRs) obtained through transcriptome sequencing and detected an average of 7.9 alleles per locus when tested on *M. floridulus* and *M. sinensis*.

CONCLUSIONS

The newly developed primers presented here were found to cross-amplify not only within *Miscanthus* species but also in other members of the Saccharinae, Andropogoneae, and

Panicaceae. They amplified DNA in *Zea* L. (Tripsacinae), *Sorghum* Moench (Sorghinae), *Cymbopogon* Spreng. (Andropogoninae), and *Pennisetum* Rich. (Panicaceae). The primers are of high value for characterization of *Miscanthus* species and can be applied to other closely related genera including *Saccharum* L.

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APPENDIX 1. List of all accessions used in the study, source, and herbarium voucher number. All taxa are Andropogoneae subtribe Saccharinae unless indicated otherwise.

Taxon ^a	Source ^b	Voucher ^c
<i>M. sacchariflorus</i> 1	TCD Bot. Gardens	TCD P15
<i>M. sinensis</i> 'Zebrinus' 2	TCD Bot. Gardens	TCD P20
<i>M. sinensis</i> 'Zebrinus' 3	TCD Bot. Gardens	TCD P21
<i>M. ×giganteus</i> 4	TCD Bot. Gardens	TCD P34
<i>M. ×giganteus</i> 5	TCD Bot. Gardens	TCD P36
<i>Miscanthus</i> sp. 6	TCD Bot. Gardens	Tea-6
<i>M. sinensis</i> 7	TCD Bot. Gardens	TCD P48
<i>Miscanthus</i> sp. 8	TCD Bot. Gardens	TCD P50
<i>M. sinensis</i> 9	TCD Bot. Gardens	TCD P51
<i>M. sacchariflorus</i> 10	TCD Bot. Gardens	TCD P58
<i>Miscanthus</i> sp. 11	TCD Bot. Gardens	Tea-11
<i>M. sinensis</i> 13	TCD Bot. Gardens	TCD P73
<i>M. sinensis</i> 14	TCD Bot. Gardens	TCD P75
<i>Miscanthus</i> sp. 15	TCD Bot. Gardens	TCD P104
<i>M. transmorrisonensis</i> 16	TCD Bot. Gardens	TCD P105
<i>M. ×giganteus</i> 17	TCD Bot. Gardens	TCD P108
<i>Miscanthus</i> sp. 18	TCD Bot. Gardens	Tea-18
<i>M. sinensis</i> 'Goliath' 19	TCD Bot. Gardens	TCD P110, SIN-H6
<i>M. ×giganteus</i> 20	TCD Bot. Gardens	TCD P114
<i>Miscanthus</i> sp. 21	TCD Bot. Gardens	Tea-21
<i>Miscanthus</i> sp. 22	TCD Bot. Gardens	Tea-22
<i>Miscanthus</i> sp. 23	TCD Bot. Gardens	Tea-23
<i>M. sinensis</i> 24	TCD Bot. Gardens	TCD P11
<i>M. sinensis</i> 25	TCD Bot. Gardens	TCD P11
<i>M. sinensis</i> 26	TCD Bot. Gardens	TCD P11
<i>Miscanthus</i> sp. 27	TCD Bot. Gardens	Tea-27
<i>Miscanthus</i> sp. 28	TCD Bot. Gardens	Tea-28
<i>Miscanthus</i> sp. 29	TCD Bot. Gardens	Tea-29
<i>M. sinensis</i> 30	TCD Bot. Gardens	Tea-30
<i>M. ×giganteus</i> 31	TCD Bot. Gardens	Tea-31
<i>M. ×giganteus</i> 32	TCD Bot. Gardens	Tea-32
<i>M. sinensis</i> 'Zebrinus' 33	TCD Bot. Gardens	TCD P20
<i>Miscanthus</i> sp. 34	TCD Bot. Gardens	Tea-34
<i>M. sinensis</i> 'Gross Fontane' 35	TCD Bot. Gardens	TCD P30
<i>M. sinensis</i> 'Gross Fontane' 36	TCD Bot. Gardens	Tea-36
<i>Miscanthus</i> sp. 37	TCD Bot. Gardens	Tea-37
<i>Miscanthus</i> sp. 38	TCD Bot. Gardens	Tea-38
<i>Miscanthus</i> sp. 39	TCD Bot. Gardens	Tea-39
<i>M. sinensis</i> 40	TCD Bot. Gardens	TCD P62
<i>Miscanthus</i> sp. 42	TCD Bot. Gardens	Tea-42
<i>Miscanthus</i> sp. 43	TCD Bot. Gardens	Tea-43
<i>M. sinensis</i> subsp. <i>condensatus</i> 44	TCD Bot. Gardens	TCD P94
<i>Miscanthus</i> sp. 45	TCD Bot. Gardens	Tea-45
<i>Miscanthus</i> sp. 46	TCD Bot. Gardens	Tea-46
<i>Miscanthus</i> sp. 47	TCD Bot. Gardens	Tea-47
<i>Miscanthus</i> sp. 48	TCD Bot. Gardens	Tea-48
<i>Miscanthus</i> sp. 49	TCD Bot. Gardens	Tea-49
<i>Miscanthus</i> sp. 50	TCD Bot. Gardens	Tea-50
<i>Miscanthus</i> sp. 51	TCD Bot. Gardens	Tea-51
<i>Miscanthus</i> sp. 52	TCD Bot. Gardens	Tea-52
<i>Miscanthus</i> sp. 53	TCD Bot. Gardens	Tea-53
<i>Miscanthus</i> sp. 54	TCD Bot. Gardens	Tea-54
<i>Miscanthus</i> sp. 55	TCD Bot. Gardens	Tea-55
<i>M. sinensis</i> 'Goliath' 56	Teagasc Oak Park	Tea-56
<i>M. sinensis</i> 'Goliath' 57	TCD Bot. Gardens	Tea-57
<i>M. sinensis</i> 'Sirene' 58	Teagasc Oak Park	Tea-58
<i>M. sinensis</i> 'Strictus' 59	TRH garden	Tea-59
<i>M. sinensis</i> 'Strictus' 60	TCD Bot. Gardens	Tea-60
<i>M. sinensis</i> 'Malapartus' 61	TRH Garden	Tea-61
<i>M. sinensis</i> 62	TRH Garden	Tea-62
<i>M. sinensis</i> 'Sirene' 63	TCD Bot. Gardens	Tea-63
<i>M. ×giganteus</i> 64	TCD Bot. Gardens	Tea-64
<i>M. ×giganteus</i> 65	TCD Bot. Gardens	Tea-65
<i>M. ×giganteus</i> 66	TRH Garden	Tea-66
<i>Miscanthus</i> sp. 68	TCD Bot. Gardens	Tea-68
<i>Miscanthus</i> sp. 69	TCD Bot. Gardens	Tea-69
<i>Miscanthus</i> sp. 70	TCD Bot. Gardens	Tea-70
<i>Miscanthus</i> sp. 71	TCD Bot. Gardens	Tea-71

APPENDIX 1. Continued.

Taxon ^a	Source ^b	Voucher ^c
<i>Miscanthus</i> sp. 72	TCD Bot. Gardens	Tea-72
<i>Miscanthus</i> sp. 73	TCD Bot. Gardens	Tea-73
<i>M. ×giganteus</i> 74	Germany—from Denmark	Tea-M1 Lasei 1
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 75	Germany	Tea-M81 RH 81
<i>M. sinensis</i> 76	Germany—from Japan	Tea-88-110
<i>M. sinensis</i> 77	Germany—from Japan	Tea-88-111
<i>M. sinensis</i> 78	Germany—from Japan	Tea-90-5
<i>M. sinensis</i> 79	Germany—from Japan	Tea-90-6
<i>M. sinensis</i> 80	Germany—from Sweden	Tea-SW 217
<i>M. ×giganteus</i> 81	Germany—from Denmark	Tea-M53 IPL 53
<i>M. ×giganteus</i> 82	Germany	Tea-M56 HAGA 56
<i>M. ×giganteus</i> 83	Germany	Tea-M63 GREIF 63
<i>M. sacchariflorus</i> 84	Germany—from Japan	Tea-M11 MATEREC 11
<i>M. sinensis</i> 'Goliath' 85	Germany	Tea-M7 GOFAL 7
<i>M. sinensis</i> hybrid 86	Germany	Tea-M42 BERBO 42
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 87	Germany	Tea-M43RH43
<i>M. sinensis</i> hybrid 88	Germany	Tea-M78 JESEL 78
<i>Miscanthus</i> sp. 89	Oak Park	Tea-89
<i>Miscanthus</i> sp. 90	Oak Park	Tea-90
<i>Miscanthus</i> sp. 91	Oak Park	Tea-91
<i>Miscanthus</i> sp. 92	Oak Park	Tea-92
<i>M. ×giganteus</i> 93	IGER/TinPlant/Oak Park	Tea-93
<i>M. ×giganteus</i> 94	Old Trial Teagasc Oak Park	Tea-94
<i>M. sinensis</i> 95	Sweden	Tea-95
<i>M. sinensis</i> 96	Sweden	Tea-96
<i>M. sinensis</i> 97	Sweden	Tea-97
<i>M. sinensis</i> 98	Sweden	Tea-98
<i>M. sinensis</i> 99	Sweden	Tea-99
<i>M. sinensis</i> 100	Sweden	Tea-100
<i>M. sinensis</i> 101	Sweden	Tea-101
<i>M. sinensis</i> 102	Sweden	Tea-102
<i>M. sinensis</i> 103	Sweden	Tea-103
<i>M. sinensis</i> 104	Sweden	Tea-104
<i>M. sinensis</i> 105	Sweden	Tea-105
<i>M. sinensis</i> 106	Sweden	Tea-106
<i>M. sinensis</i> 107	Sweden	Tea-107
<i>M. sinensis</i> 108	Sweden	Tea-108
<i>M. sinensis</i> 109	Sweden	Tea-109
<i>M. sinensis</i> 110	Sweden	Tea-110
<i>M. sinensis</i> 111	Sweden	Tea-111
<i>M. sinensis</i> 112	Sweden	Tea-112
<i>M. sinensis</i> 113	Sweden	Tea-113
<i>M. sinensis</i> 114	Sweden	Tea-114
<i>M. sinensis</i> 115	Sweden	Tea-115
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 116	Sweden	Tea-116
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 117	Sweden	Tea-117
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 118	Sweden	Tea-118
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 119	Sweden	Tea-119
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 120	Sweden	Tea-120
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 121	Sweden	Tea-121
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 122	Sweden	Tea-122
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 123	Sweden	Tea-123
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 124	Sweden	Tea-124
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 125	Sweden	Tea-125
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 126	Sweden	Tea-126
<i>M. sacchariflorus</i> × <i>M. sinensis</i> 127	Sweden	Tea-127
<i>M. sacchariflorus</i> 128	TCD Bot. Gardens	Tea-128
<i>M. sacchariflorus</i> 129	TCD Bot. Gardens	Tea-129
<i>Miscanthus</i> sp. 130	TCD Bot. Gardens	Tea-130
<i>Miscanthus</i> sp. 131	TCD Bot. Gardens	Tea-131
<i>Saccharum officinarum</i>	TCD Bot. Gardens	TCD TRH s.n.
<i>Cymbopogon citratus</i> ^d	TCD Bot. Gardens	TCD TRH s.n.
<i>Zea diploperennis</i> ^e	TCD Bot. Gardens	TCD TRH s.n.
<i>Sorghum halepense</i> 6 ^f	RBG Kew 151 01	Kew 1966-54209
<i>Pennisetum</i> sp. ^g	TCD Bot. Gardens	TCD TRH s.n.
<i>M. sinensis</i> var. <i>variegatus</i> 1	RBG Kew 154 04	Kew 1969-19093
<i>M. sinensis</i> subsp. <i>condensatus</i> 7	RBG Kew 151	Kew 1969-19091
<i>M. oligostachyus</i> 16	RBG Kew 151 (pot)	Kew 1995-1864
<i>M. nepalensis</i> 25	RBG Kew TH 4	Kew 1985-8388

APPENDIX 1. Continued.

Taxon ^a	Source ^b	Voucher ^c
<i>M. sinensis</i> 'Goliath' 27	ADAS Steinmann nurseries	Kew MB93/02
<i>M. sinensis</i> 'Gracillimus' 28	ADAS Piccoplant, Germany	Kew MB94/05
<i>M. sinensis</i> 'Roland' 29	ADAS Piccoplant, Germany	Kew MB94/06
<i>M. sinensis</i> Anderss. 30	ADAS Wye College	Kew MB94/07
<i>M. sinensis</i> 'Gross Fontane' 31	ADAS Genft Dogels, Germany	Kew PN95/01
<i>M. sacchariflorus</i> 61	RBG Kew	Kew 1987-2727
<i>M. sinensis</i> 'Yakushmanum' 63	RBG Kew	Kew 1987-1148
<i>M. transmorrisonensis</i> 65	RBG Kew	Kew1990-2748
<i>M. fuscus</i> 82	RBG Kew	Kew 590
<i>M. violaceus</i> 84	RBG Kew	Kew 7437
<i>M. ecklonii</i> 86	RBG Kew	Kew 2347
<i>M. junceus</i> 88	RBG Kew	Kew 1060
<i>M. junceus</i> 89	RBG Kew	Kew 2309
<i>M. ecklonii</i> 105	RBG Kew	Kew 2929
<i>M. ecklonii</i> 106	RBG Kew	Kew 247
<i>M. yunnanensis</i> 107	RBG Kew	Kew 30689
<i>M. nudipes</i> 109	RBG Kew	Kew 2007
<i>M. tinctorius</i> 112	RBG Kew	Kew 1466
<i>Saccharum spontaneum</i> 117	RBG Kew	Kew Butt, 1977
<i>Narenga porphyrocoma</i> 120	RBG Kew	Kew 2092
<i>Saccharum contortum</i> 121	RBG Kew	Kew 3797
<i>Spodiopogon rhizophorus</i> 125	RBG Kew	Kew 283
<i>Spodiopogon sibiricus</i> 128	RBG Kew	Kew 210
<i>Eulalia quadrinervis</i> 134	RBG Kew	Kew 3294
<i>M. sinensis</i> 'Morning Light' 155	RBG Kew	Kew 1996 821
<i>M. sacchariflorus</i> 159	RBG Kew	Kew 3598 1935
<i>M. sacchariflorus</i> 160	RBG Kew	Kew 1984
<i>M. tinctorius</i> 'Nana Variegata' 161	RBG Kew	Kew 1996 1065
<i>M. sinensis</i> 'Goliath' 194	ADAS	Kew PN96/30

^a Numbers accompanying species names represent the DNA extraction identifier for this study.

^b Source abbreviations: ADAS = Agricultural Development Advisory Service (now Agriculture and Environmental Consultancy); IGER = Institute of Grassland and Environmental Research (now Institute of Biological, Environmental and Rural Sciences [IBERS]); RBG Kew = Royal Botanic Gardens, Kew, Richmond, Surrey, United Kingdom; TCD Bot. Gardens = Trinity College Dublin Botanical Garden, Dublin, Ireland; Teagasc Oak Park = Teagasc Oak Park Research Centre, Carlow, Ireland; TRH Garden = personal garden of first author.

^c Voucher abbreviations: Kew = Herbarium of the Royal Botanic Gardens, Kew, Richmond, Surrey, United Kingdom; TCD = Trinity College Dublin Herbarium, Ireland; Tea = Teagasc Oak Park Research Centre, Carlow, Ireland.

^d Andropogoninae, Andropogoneae (subtribe/tribe).

^e Tripsacinae, Andropogoneae.

^f Sorghinae, Andropogoneae.

^g Cenchrinae, Paniceae.