

# Twenty Novel Polymorphic Microsatellite Primers in the Critically Endangered Melastoma tetramerum var. tetramerum (Melastomataceae)

Authors: Narita, Ayu, Izuno, Ayako, Komaki, Yoshiteru, Tanaka, Takefumi, Murata, Jin, et al.

Source: Applications in Plant Sciences, 4(9)

Published By: Botanical Society of America

URL: https://doi.org/10.3732/apps.1600053

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <u>www.bioone.org/terms-of-use</u>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.



PRIMER NOTE

## Twenty novel polymorphic microsatellite primers in the critically endangered *Melastoma tetramerum* var. *tetramerum* (Melastomataceae)<sup>1</sup>

Ayu Narita<sup>2,4</sup>, Ayako Izuno<sup>2</sup>, Yoshiteru Komaki<sup>3</sup>, Takefumi Tanaka<sup>3</sup>, Jin Murata<sup>3</sup>, and Yuji Isagi<sup>2</sup>

<sup>2</sup>Graduate School of Agriculture, Kyoto University, Kitashirakawa Oiwake-cho, Sakyo-ku, Kyoto 606-8502, Japan; and <sup>3</sup>Botanical Gardens, Graduate School of Science, University of Tokyo, 3-7-1 Hakusan, Bunkyo-ku, Tokyo 112-0001, Japan

- *Premise of the study:* Microsatellite markers were identified for *Melastoma tetramerum* var. *tetramerum* (Melastomataceae), a critically endangered shrub endemic to the Bonin Islands, to reveal genetic characteristics in wild and restored populations.
- *Methods and Results:* Using next-generation sequencing, 27 microsatellite markers were identified. Twenty of these markers were polymorphic in *M. tetramerum* var. *tetramerum*, with two to nine alleles per locus and expected heterozygosity ranging from 0.10 to 0.71. Among the 20 polymorphic markers, 15 were applicable to other closely related taxa, namely *M. tetramerum* var. *pentapetalum*, *M. candidum* var. *candidum*, and *M. candidum* var. *alessandrense*.
- *Conclusions:* These markers can be potentially useful to investigate the genetic diversity, population genetic structure, and reproductive ecology of *M. tetramerum* var. *tetramerum* as well as of the three related taxa to provide appropriate genetic information for conservation.

Key words: Bonin Islands; conservation; Melastoma; Melastomataceae; microsatellites; next-generation sequencing.

Melastoma L. (Melastomataceae) is a genus comprising 50 species (Stevens, 2001) that are distributed from the Indo-Malesian region to the Pacific Ocean. Around the Japanese archipelago, 10 taxa (seven species and three varieties) grow wild, four of which are vulnerable, endangered, or critically endangered (Ministry of the Environment, Japan, 2015). In the Bonin Islands, which are typical oceanic islands with unique biota and which were designated as a UNESCO Natural World Heritage Site in 2011, three Melastoma taxa are of conservation concern. Melastoma tetramerum Hayata var. tetramerum is a critically endangered shrub, endemic to Chichi-jima Island in the Bonin Islands. Only one individual was known at Higashidaira (HD) until a population comprising 125 individuals was discovered in 1993 at Higashikaigan (HK), which is 2 km from HD (Shimizu, 1997). However, by 1995 the individual at HD had died (Toyoda, 2014). The HK population also declined with forest development because a forest containing fewer gaps was unsuitable for this light-demanding shrub. Several dozen wild individuals were remaining in HK in 2007 (Ministry of the Environment, Japan, 2015). Cuttings and seeds of the extinct individual in HD and 11 individuals in HK were collected for propagation at the Koishikawa Botanical Garden of the University of Tokyo before 1995 and in 2005–2008, respectively. The source seeds or cuttings of the restored plants were derived from individuals collected at the restoration sites. The garden has stored and propagated these

<sup>1</sup>Manuscript received 27 April 2016; revision accepted 31 May 2016.

This work was supported by the Grants-in-Aid for Scientific Research Program (KAKENHI grant no. 15H04414) from the Japan Society for the Promotion of Science.

<sup>4</sup>Author for correspondence: narita.ayu.85m@st.kyoto-u.ac.jp

doi:10.3732/apps.1600053

cuttings and seeds and has restored 558 and 252 plants to HD and HK, respectively.

To determine the genetic variation and differentiation of in situ and ex situ populations and to maintain genetic diversity in future populations, we identified 20 polymorphic microsatellite loci to genotype all available individuals of *M. tetramerum* var. tetramerum. The broad applicability of these markers was examined in three closely related taxa, namely, M. tetramerum var. pentapetalum Toyoda, M. candidum D. Don var. candidum, and M. candidum var. alessandrense S. Kobay. Melastoma tetramerum var. *pentapetalum* is an endangered variety, endemic to two mountains on Haha-jima Island, which is 37 km from Chichi-jima Island. This variety has flowers with five petals, whereas *M. tetramerum* var. *tetramerum* has flowers with four petals. More than 100 individuals of *M. tetramerum* var. pentapetalum had been observed in 1994; however, only several dozen were found in 2007 (Shimizu, 1997; Ministry of the Environment, Japan, 2015). Melastoma candidum var. candidum is a common variety native to Okinawa, Taiwan, China, and Indochina (Ohwi, 1978). Melastoma candidum var. alessandrense is endemic to Kita-Iwo-To Island, an uninhabited island isolated 200 km from Chichi-jima Island. This variety is vulnerable, with a total of several hundred individuals known in 2007 (Ministry of the Environment, Japan, 2015).

### METHODS AND RESULTS

An individual of *M. tetramerum* var. *tetramerum* was collected at HK and stored at Koishikawa Botanical Garden (voucher: komaki201601; Appendix 1) for use in developing microsatellite loci. From silica gel–dried leaf tissues, total genomic DNA was extracted using the QIAGEN DNeasy Plant Mini Kit (QIA-GEN, Hilden, Germany), and a fragment DNA library was constructed using Ion

Applications in Plant Sciences 2016 4(9): 1600053; http://www.bioone.org/loi/apps © 2016 Narita et al. Published by the Botanical Society of America. This work is licensed under a Creative Commons Attribution License (CC-BY-NC-SA).

#### Applications in Plant Sciences 2016 4(9): 1600053 doi:10.3732/apps.1600053

Table 1.	. Characteristics of the 27 microsatellite loci amplified in Melastoma te	etramerum var. tetramerum.
----------	---	----------------------------

Locus <sup>a,b</sup>		Primer sequences $(5'-3')$	Repeat motif	Allele size (bp) <sup>c</sup>	Multiplex PCR set <sup>d</sup>	Fluorescent tage	GenBank accession no.
Mte002*	F:	GCACCTCCACACATTGCTC	(GA) <sub>9</sub>	258	_	6-FAM	KX394447
	R:	ACGCACGTCCTGTTAGGG					
Mte003	F:	ATTTGCATGGCCAGTTGCG	(AG) <sub>10</sub>	171-173	С	6-FAM	KX394448
	R:	TGACAATCAGTTCCAACACGTC					
Mte005	F:	CTCCTTCCGATCGTCGTTATG	$(AG)_{10}$	212-230	С	VIC	KX394449
16.007		CGATGGTGTCTAACTAAGCTTCC		177 101	D	NED	113120 1 1 50
Mte007		CATCTCTCCGGATCCAATTCC	$(GA)_{12}$	177–181	В	NED	KX394450
Mt=009		GGTCACCGCGATAAACGAC	(TA)	166 169	٨	6 EAM	VV204451
Mte008	F: R:		$(TA)_9$	166–168	А	6-FAM	KX394451
Mte012		GAGTATCAGAGTTGTTAGCTGATCG CAAAGCCCAACATCGGGAC	(TG) <sub>12</sub>	163-178	А	NED	KX058005
WIIC012		ACTAATGGAGTACGAATAGCAACG	$(10)_{12}$	105-178	Λ	NLD	<b>K</b> X050005
Mte014		ACCTTGGCATCTTCACAAAGG	(GAA) <sub>15</sub>	209-237	D	NED	KX394452
		TGGGCAACACTGGGATCTG	(0111)]]	207 207	2	1122	
Mte015*		ACCTCGGAAGTGTCCATGAG	(CT) <sub>13</sub>	193		VIC	KX394453
		GAAGTGCTGAAGCGTCTCG	( )15				
Mte017	F:	ACTCGCCTTATTTGAGTATCCG	(CT) <sub>9</sub>	203-209	Е	6-FAM	KX394454
	R:	AAGTGGCCATTCATCCACG					
Mte018	F:	CGGAGAAGGACGAATGTGC	(CAA) <sub>9</sub>	184–186	С	NED	KX058008
	R:	TTGAAGGGAGGTGGCAGAC					
Mte019		AGTTTGGACCCATCCCATTTG	$(AG)_{10}$	167-170	В	6-FAM	KX058009
16.000*		AGGGTAGTGAAACAGCTAAGG		202		NED	111000000
Mte022*		CACCCGAAGCACGAATCAC	(AAT) <sub>9</sub>	203	—	NED	KX237522
Mte023		TCGGGAAAGACCCAGTTCG	(CT)	225 227	В	6-FAM	KX058010
MICO25		CCCTTCATCCCAAGCAACG	(CT) <sub>11</sub>	225–227	D	0-FAM	KA036010
Mte025*		GGACCGTCGATTGAGTCCG TTTCCCGCCAACTTCATCG	$(AG)_9$	215	_	6-FAM	KX237523
WICO25		GCTCGAAATCTTCCAGGCG	(AO)	215		0-174101	KA257525
Mte026*		TCACACCCATGGCACTCTG	$(AG)_8$	191	_	NED	KX237524
		AACTGCATCACCAGGCAAG	()8				
Mte027*		GGAGAAAGAAAGGATATCATCTGTCG	(CT) <sub>10</sub>	245	_	NED	KX237525
		ACGTCTATTTGGGCCTCGG	( )10				
Mte029	F:	CATCGTCCCACATGCTGTC	(CTT) <sub>8</sub>	264-279	С	6-FAM	KX058011
	R:	GCTTTGATCCCAATCCGCC					
Mte030		ACGGCTTTGCAGTTAAGGTC	(AG) <sub>9</sub>	240-241	E	NED	KX058012
		GCATCAGATCCCACAGGAG			_		
Mte032		GGACACTTGCATCACCCTTC	(GT) <sub>9</sub>	229–233	D	VIC	KX058013
N/ 022		GTCGAGGTAAATCTCAATCGCC	(CTTT)	000 005	D	MIC	1232050014
Mte033		GTTGAAACGGGATTGTTTAGCG	$(CTT)_{10}$	223–235	В	VIC	KX058014
Mte034		AACGGTACCACGGACATCG	(TTC) <sub>8</sub>	225-228	А	6-FAM	KX058015
MICO34		CGATCATGCAAAGGATATCTGC AGCCAGGCAGCCAATTTAC	$(11C)_{8}$	223-228	A	0-I'AIVI	KA036013
Mte035		CCCAGTGGATGGAGTTTGC	(AG) <sub>13</sub>	262-264	Е	6-FAM	KX058016
Micoss		TCCTTAATTGGGTTTAGGGACAAC	(10)]3	202 204	L	017101	1111050010
Mte038*	F:		$(GA)_8$	187	_	VIC	KX237526
		GAAGGGAGGCATTCATGGG	(- )0				
Mte039	F:	CTTCCCTGAGCTGCAATCC	(CT) <sub>12</sub>	238-240	Е	VIC	KX058017
	R:	AGACAGGCTACAAGGCTCC					
Mte040	F:	CCCAGGAATCCCAAATCCAG	$(GAA)_8$	185-188	D	6-FAM	KX058018
	R:	CTGATTCGCTTGCCGACAG					
Mte041		TCGGAGCAGCCATAGAACG	(TCG) <sub>10</sub>	272-299	D	NED	KX058019
		TGCCTCGACCTCCTGAAATC					
Mte042		GTTTGAAGCTTATGGCCAAGAC	$(AG)_8$	170-174	А	VIC	KX058020
	R:	GCAGGAAAGTGTTGAGAAGACC					

<sup>a</sup>Monomorphic loci are marked with an asterisk.

<sup>b</sup>Annealing temperature for all loci was 57°C.

<sup>c</sup>Allele size indicates the size of PCR products identified in 30 individuals from the wild population on Chichi-jima Island and restored or collected individuals originating from the extinct population on Chichi-jima Island.

<sup>d</sup>Loci with identical letters were amplified in the same PCR.

<sup>c</sup>Amplified with M13 primers with fluorescent dye at each 5' end: 6-FAM = 5'-CACGACGTTGTAAAACGAC-3', VIC = 5'-TGTGGAATTGTGAGCGG-3', NED = 5'-CTATAGGGCACGCGTGGT-3'.

Shear Plus Reagents (Life Technologies, Carlsbad, California, USA) and the Ion Plus Fragment Library Kit (Life Technologies) according to the manufacturer's protocol. After dilution to 15 pM, the library was sequenced using the Ion PGM system (Life Technologies) with Ion PGM Template OT2 400 Kit (Life Technologies), Ion PGM Sequencing 400 Kit (Life Technologies), and Ion 318 Chip

Kit v2 (Life Technologies) according to the manufacturer's protocols. We obtained 546,612 reads with an average length of 184 bp.

Microsatellite loci were located and primers were designed using MSAT-COMMANDER version 0.8.2 (Faircloth, 2008) under the following conditions: more than eight microsatellite motif repeats, 40–60% of GC content in the PCR

	Total A	M. tetramerum					M. candidum						
		var. <i>tetramerum</i> $(n = 30)$			var. $pentapetalum (n = 12)$			var. <i>candidum</i> $(n = 12)$			var. <i>alessandrense</i> $(n = 3)$		
Locus		Ā	$H_{\rm o}$	$H_{\rm e}$	A	$H_{\rm o}$	$H_{\rm e}$	Α	$H_{\rm o}$	H <sub>e</sub>	Α	$H_{\rm o}$	$H_{\rm e}$
Mte003	3	2	0.333	0.420	1	0.000	0.000	1	0.000	0.000	2	0.333	0.278
Mte005	7	6	0.467	0.708*	2	0.167	0.153	4	0.167	0.625	1	0.000	0.000
Mte007	5	3	0.367	0.653*	1	0.000	0.000				_	_	
Mte008	3	2	0.567	0.486	1	0.000	0.000				_		
Mte012	4	3	0.333	0.352	1	0.000	0.000	_			_		
Mte014	11	9	0.733	0.676	1	0.000	0.000	2	0.417	0.469	1	0.000	0.000
Mte017	4	2	0.300	0.339	1	0.000	0.000	2	0.417	0.469	1	0.000	0.000
Mte018	4	2	0.233	0.299	1	0.000	0.000	1	0.000	0.000	1	0.000	0.000
Mte019	2	2	0.167	0.495*	1	0.000	0.000	_			_		
Mte023	4	2	0.267	0.480*	1	0.000	0.000	3	0.000	0.542	1	0.000	0.000
Mte029	7	3	0.367	0.485	1	0.000	0.000	3	0.667	0.625	1	0.000	0.000
Mte030	6	2	0.100	0.095	1	0.000	0.000	5	0.583	0.507	1	0.000	0.000
Mte032	4	3	0.633	0.523	1	0.000	0.000	_			_		
Mte033	7	3	0.533	0.509	1	0.000	0.000	3	0.167	0.434	2	1.000	0.500
Mte034	4	2	0.100	0.095	1	0.000	0.000	2	0.083	0.219	1	0.000	0.000
Mte035	5	3	0.233	0.415*	2	0.000	0.153	4	0.333	0.573	1	0.000	0.000
Mte039	4	2	0.133	0.124	2	0.000	0.153	4	0.500	0.549	1	0.000	0.000
Mte040	4	2	0.167	0.153	1	0.000	0.000	1	0.000	0.000	1	0.000	0.000
Mte041	8	5	0.400	0.607*	2	0.167	0.153	2	0.000	0.153	1	0.000	0.000
Mte042	4	3	0.367	0.363	1	0.000	0.000	3	0.667	0.559	2	1.000	0.500

TABLE 2. Genetic properties of 20 polymorphic microsatellite loci in Melastoma tetramerum var. tetramerum and their applicability to three related taxa.<sup>a</sup>

*Note:* A = number of alleles per locus;  $H_e =$  expected heterozygosity;  $H_o =$  observed heterozygosity; n = number of individuals sampled. <sup>a</sup>Voucher and locality information are provided in Appendix 1.

\*Significant deviation from Hardy–Weinberg equilibrium (P < 0.05).

products, annealing temperature of 57–60°C, and 150–400 bp of product sizes. Any primers with self-annealing sequences or hairpin structures were removed using Primer3 (Rozen and Skaletsky, 1999). Any primer pairs were removed if melting temperatures of forward and reverse primers differed more than 1°C. After filtering the candidate microsatellite regions, 42 microsatellite regions were retained for primer testing. M13 sequences, which enable fluorescent labeling of PCR products, were ligated to the 5' end of forward primers (Table 1).

The primers were tested using eight individuals of *M. tetramerum* var. *tetramerum*, consisting of one restored individual at HD (voucher: 04110; Appendix 1) and seven wild individuals at HK (voucher: komaki201601; Appendix 1). PCR was performed in a final reaction volume of 5  $\mu$ L, which included 4–240 ng of template DNA, 0.01  $\mu$ M of each forward primer, 0.2  $\mu$ M of each reverse primer, 1  $\mu$ M of each M13 primer with fluorescent labels, and 2.5  $\mu$ L of QIAGEN Multiplex PCR MasterMix (QIAGEN). The PCR conditions were as follows: initial denaturation at 95°C for 15 min; followed by 33 denaturation cycles at 94°C for 30 s, annealing at 57°C for 90 s, extension at 72°C for 90 s; and a final extension at 72°C for 10 min. One microliter of the PCR product was electrophoresed using a 3130*xl* Genetic Analyzer (Applied Biosystems) and 0.15  $\mu$ L of GeneScan 500 LIZ Size Standard (Applied Biosystems).

Out of 42 microsatellite loci tested, 27 loci were amplified and characterized in all eight individuals of *M. tetramerum* var. *tetramerum*, and 20 were found to be polymorphic in *M. tetramerum* var. *tetramerum* (Table 1). The applicability of these 20 loci was examined in the four *Melastoma* taxa: 30 individuals of *M. tetramerum*, including 11 (10 restored and one stocked) individuals from HD and 19 (eight wild and 11 stocked) individuals from HK; 12 individuals of *M. tetramerum* var. *pentapetalum* from two populations on Hahajima Island; 12 individuals of *M. candidum* var. *candidum* from one population in Aha, Okinawa; and three individuals of *M. candidum* var. *alessandrense* from one population on Kita-Iwo-To Island (Appendix 1). The genotypes were identified using GeneMapper version 4.1 (Applied Biosystems). The number of alleles and observed and expected heterozygosities were calculated using GenAlEx version 6.5 (Peakall and Smouse, 2012). Linkage disequilibrium (LD) and deviation from Hardy–Weinberg equilibrium (HWE) were tested using GENEPOP version 4.2 (Rousset, 2008).

Out of the 20 loci, 15 were amplified in all four taxa tested, whereas the remaining five only amplified in the two varieties of *M. tetramerum* (Table 2). The total number of alleles per locus observed across the four taxa ranged from two to 11 (Table 2). In *M. tetramerum* var. *tetramerum*, the observed and expected heterozygosities were in the ranges of 0.10–0.73 and 0.10–0.71, respectively (Table 2). A total of six loci (Mte005, Mte007, Mte019, Mte023, Mte035, and Mte041) significantly deviated from HWE (P < 0.05) in *M. tetramerum* var. *tetramerum*. The extremely small population size of *M. tetramerum* var. *tetramerum* could have strongly affected its genetic variation and caused HWE deviation by genetic drift. A significantly high LD was found between Mte019 and Mte023, Mte007 and Mte019, Mte007 and Mte014, and Mte007 and Mte012 (P < 0.05) in every taxon.

#### CONCLUSIONS

Twenty novel polymorphic loci identified in the critically endangered *M. tetramerum* var. *tetramerum* will be used for the assessment of the genetic diversity, effective population size, inbreeding coefficient, and population genetic structure of the extant and extinct populations of this variety and three related taxa, which will enable evaluation and improvement of the ongoing conservation management programs.

#### LITERATURE CITED

- FAIRCLOTH, B. C. 2008. MSATCOMMANDER: Detection of microsatellite repeat arrays and automated, locus-specific primer design. *Molecular Ecology Resources* 8: 92–94.
- MINISTRY OF THE ENVIRONMENT, JAPAN. 2015. Red Data Book 2014– Vascular plants, Threatened wildlife of Japan, vol. 8, 94–456. Gyosei, Tokyo, Japan [in Japanese].
- OHWI, J. 1978. Revised and enlarged Flora of Japan, 946. Shibundo, Tokyo, Japan.
- PEAKALL, R., AND P. E. SMOUSE. 2012. GenAlEx 6.5: Genetic analysis in Excel. Population genetic software for teaching and research–an update. *Bioinformatics (Oxford, England)* 28: 2537–2539.
- ROUSSET, F. 2008. GENEPOP'007: A complete reimplementation of the GENEPOP software for Windows and Linux. *Molecular Ecology Resources* 8: 103–106.
- ROZEN, S., AND H. J. SKALETSKY. 1999. Primer3 on the WWW for general users and for biologist programmers. *In S. Misener and S. A. Krawetz [eds.]*, Methods in molecular biology, vol. 132:

Bioinformatics methods and protocols, 365–386. Humana Press, Totowa, New Jersey, USA.

- SHIMIZU, Y. 1997. Ecology of genus *Melastoma* in the Bonin (Ogasawara) Islands: Present situation of a *M. tetramerum* community discovered in 1993. *Komazawa Geography* 33: 49–76 [in Japanese].
- STEVENS, P. F. 2001. Angiosperm Phylogeny Website. Version 12, July 2012. Website http://www.mobot.org/MOBOT/research/APweb/ [accessed 12 May 2016].
- TOYODA, T. 2014. The endemic plants of the Bonin Islands, 209–214. Woodspress, Kanagawa, Japan [in Japanese].

APPENDIX 1. Voucher information for *Melastoma* taxa examined in this study.

Taxon	Voucher specimen accession no. (Herbarium) <sup>a</sup>	Collection locality	Geographic coordinates
M. tetramerum Hayata var. tetramerum	komaki201601 (TI)	Higashikaigan (HK), Chichi-jima Island, Bonin Islands	27°24'08.1″N, 142°13'35.9″E
M. tetramerum var. tetramerum	04110 (TI)	Higashidaira (HD), Chichi-jima Island, Bonin Islands	27°04'30.8″N, 142°13'24.8″E
M. tetramerum var. pentapetalum Toyoda	komaki201602 (TI)	Mt. Sakaigatake, Haha-jima Island, Bonin Islands	26°40'26.2″N, 142°09'13.8″E
M. candidum D. Don var. candidum	KYO 00019995 (KYO)	Aha, Kunigami, Okinawa	26°42'30.46″N, 128°16'6.89″E
M. candidum var. alessandrense S. Kobay.	komaki201603 (TI)	Kita-Iwo-To Island, Bonin Islands	25°26'16.22″N, 141°16'56.91″E

<sup>a</sup>Vouchers are deposited at the herbarium of the University of Tokyo (TI) and the herbarium of Kyoto University (KYO).