



PATTERNS OF HEMORRHAGIC DISEASE IN WHITE-TAILED DEER (ODOCOILEUS VIRGINIANUS) IN THE GREAT PLAINS OF THE USA, 1982–2020

Authors: Kring, Emma K., Stallknecht, David E., D'Angelo, Gino J., Kohl, Michel T., Bahnson, Charlie, et al.

Source: Journal of Wildlife Diseases, 60(3) : 670-682

Published By: Wildlife Disease Association

URL: <https://doi.org/10.7589/JWD-D-23-00021>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Patterns of Hemorrhagic Disease in White-Tailed Deer (*Odocoileus virginianus*) in the Great Plains of the USA, 1982–2020

Emma K. Kring,^{1,2} David E. Stallknecht,¹ Gino J. D'Angelo,² Michel T. Kohl,² Charlie Bahnson,³ Christopher A. Cleveland,¹ Liliana C. M. Salvador,^{4,5} and Mark G. Ruder^{1,6}

¹ Southeastern Cooperative Wildlife Disease Study, Department of Population Health, College of Veterinary Medicine, 589 D. W. Brooks Dr., University of Georgia, Athens, Georgia 30602, USA

² Warnell School of Forestry and Natural Resources, 180 E. Green St., University of Georgia, Athens, Georgia 30602, USA

³ North Dakota Game and Fish Department, 100 N. Bismarck Expressway, Bismarck, North Dakota 58501, USA

⁴ Department of Infectious Diseases, College of Veterinary Medicine, Institute of Bioinformatics, 501 D. W. Brooks Dr., University of Georgia, Athens, Georgia 30602, USA

⁵ Current address: School of Animal and Comparative Biomedical Sciences, College of Agriculture, Life and Environmental Sciences, University of Arizona, 1117 E. Lowell St., Room 222, Tucson, Arizona 85721, USA

⁶ Corresponding author (email: mgruder@uga.edu)

ABSTRACT: Hemorrhagic disease (HD) of deer is caused by epizootic hemorrhagic disease virus (EHDV) or bluetongue virus (BTV) and is considered one of the most important viral diseases of white-tailed deer (*Odocoileus virginianus*). Despite evidence of changing patterns of HD in the northeastern and upper midwestern US, the historical and current patterns of HD in the Great Plains remain poorly described. We used results from an annual survey documenting HD mortality to characterize historic and current patterns of HD in the northern and central Great Plains (North Dakota, South Dakota, Nebraska, Kansas, and Oklahoma), US, between 1982 and 2020. Further, we assessed temporal change using linear regression to determine change in annual reporting intensity (percentage of counties in a state with reported HD) and change in reporting frequency (the number of years a county or state reported HD) during each decade between 1982 and 2020. Across the 38-yr study period, HD reports expanded northeast across latitude and longitude. Intensity of HD reports significantly increased during this period for three (North Dakota, South Dakota, Kansas) of five states examined. Frequency of reports also increased for all five states. Such changes in northern latitudes might lead to increased deer mortality in regions where HD epizootics have been historically less frequent. Understanding how patterns of HD are changing on the landscape is important when considering future deer management in the face of other mortality factors.

Key words: Bluetongue virus, BTV, epizootic hemorrhagic disease virus, EHDV, Great Plains, HD.

INTRODUCTION

Hemorrhagic disease (HD) of white-tailed deer (WTD, *Odocoileus virginianus*) is caused by epizootic hemorrhagic disease virus (EHDV) and bluetongue virus (BTV), which may also cause disease in some other ruminant species (Nettles et al. 1992). Both EHDV and BTV are in the genus *Orbivirus* (*Reoviridae* family) and are vectored by *Culicoides* biting midges (Stallknecht et al. 2002). Currently, *Culicoides sonorensis* and *Culicoides insignis* are the only confirmed vector species for EHDV and BTV, but other *Culicoides* species are suspected vectors (McGrogan et al. 2022). Hemorrhagic disease is considered one of the most important viral diseases of WTD in North America (Stallknecht and Howerth 2004). Hemorrhagic disease in deer ranges from inapparent infection

to acute and fatal febrile disease typically characterized by edema, effusion, and hemorrhage in multiple organs and tissues (Howerth et al. 2001). Some deer may survive acute HD infection to develop chronic sequelae, such as cracked hoof walls or scarred ruminal papillae (Couvillion et al. 1981).

Reports of cyclical WTD mortality events consistent with HD have been documented in the US since the late 1800s (Nettles and Stallknecht 1992), but it was not until the 1950s and 1960s that the viral causes of HD were confirmed (Shope et al. 1960; Stair et al. 1968). Following this, classical and molecular viral diagnostics enabled long-term monitoring of HD in the US and helped to characterize the diversity of viruses responsible for outbreaks (Stallknecht and Howerth 2004; Ruder et al. 2016). Annual HD surveillance indicates that

EHDV-2 is the predominant virus causing HD outbreaks among WTD in the US, but other EHDV and BTV serotypes are commonly involved with HD outbreaks in wild ruminants (Ruder et al. 2016; Rivera et al. 2021). Although severe disease associated with EHDV and BTV infection is most commonly reported in WTD, other wild ruminant species commonly affected during outbreaks include mule deer (*Odocoileus hemionus*), bighorn sheep (*Ovis canadensis*), and pronghorn (*Antilocapra americana*; Howerth et al. 2001).

Clinical disease associated with HD in individual deer is variable, but general patterns of infection and disease in WTD populations vary predictably depending on the geographic region (Stallknecht et al. 2002; Ruder et al. 2015). Historically, EHDV and BTV circulated broadly between the latitudes of 35° S and 40–45° N in areas with habitats and climatic conditions that support *Culicoides* spp. vector populations (Purse et al. 2005). In the US, geographic areas may be described as having either epizootic or enzootic disease patterns (Stallknecht et al. 2002). Regions with enzootic patterns are generally at more southern latitudes where EHDV and BTV frequently circulate but reports of HD in WTD are less common (e.g., 2–3-yr cycles), because inapparent infections or mild disease without mortality predominate (Couvillion et al. 1981; Nettles et al. 1992). Regions with epizootic patterns generally occur at more northern latitudes where virus circulation is intermittent, herd immunity is low, and intense HD epizootics with high mortality occur infrequently (e.g., 8–10-yr cycle historically) (Stallknecht et al. 1996; Ruder et al. 2015; Stilwell et al. 2021). These epidemiologic patterns have been best characterized for states in the eastern and southeastern US, ranging from enzootic patterns throughout much of the coastal plain physiographic region in the southeast, to epizootic patterns in the upper midwest, northeast, and Piedmont and Appalachian Mountain areas in the southeast (Stallknecht et al. 1991; Nettles and Stallknecht 1992; Stallknecht et al. 2002). Over the last two decades northward expansion of HD has occurred in parts of the northeastern

and midwestern US, highlighting the dynamic nature of this disease system (Stallknecht et al. 2015; Casey et al. 2021). In addition to these changing regional patterns, northward shifts in latitude of reported HD have occurred within individual states, such as Illinois (Boyer et al. 2007; Dorak et al. 2022). Much of the observed regional variation in infection outcome is related to differences in population immunity, including previous exposure to the viruses, passive antibody transfer to fawns, as well as genetic factors influencing innate immunity (Stallknecht et al. 1996; Gaydos et al. 2002a, b).

Sustained and long-term monitoring of HD in wild ruminant populations by wildlife management agencies is important for understanding epidemiological patterns in the US and how cyclical variation in outbreaks might affect population dynamics and management goals. Given the cyclical nature of HD in areas where deer are immunologically naïve to EHDV or BTV infection (e.g., northern latitudes), it is difficult to predict when outbreaks might occur. However, passive surveillance and outbreak investigations have helped to identify new virus serotypes (Ruder et al. 2017), novel patterns of disease (Stallknecht et al. 2015; Dorak et al. 2022), and potential risk factors such as severe drought (Christensen et al. 2020). Although investigation of an EHDV-2 outbreak in West Virginia revealed an estimated 20% mortality rate in the affected WTD population (Gaydos et al. 2004), questions remain regarding the impact of more frequent HD outbreaks on a population. Sustained monitoring of HD outbreaks is fundamentally important for understanding the potential population impacts of HD, especially in areas where deer populations are immunologically naïve and there is evidence of northern expansion and increased frequency and intensity of HD outbreaks (Stallknecht et al. 2015; Dorak et al. 2022).

Historically, severe HD outbreaks have been reported in parts of the Great Plains of the US, including the states of North Dakota, South Dakota, Nebraska, and Kansas (Stevens et al. 2015). However, HD in the Great Plains remains poorly characterized. Describing historic and

current patterns of reported HD is the first step in recognizing changes in this disease system and how HD patterns might change in the future. Anecdotally, patterns of HD in this region may be changing, similar to the documented northern expansion and increased frequency and intensity of HD in parts of the eastern US (Stallknecht et al. 2015). The objectives of our study were 1) to broadly describe historic patterns of HD in the Great Plains through the use of annual survey data between 1982 and 2020; and 2) to determine if patterns of reported HD changed in frequency, intensity, or geographic distribution during this time period.

MATERIALS AND METHODS

Study area

The Great Plains encompasses approximately 2.17 million square kilometers across the US states of Montana, Wyoming, Colorado, New Mexico, Texas, Oklahoma, Missouri, Iowa, Minnesota, North Dakota, South Dakota, Nebraska, Kansas, and Oklahoma (Zhu et al. 2011). The Great Plains encompasses highly diverse habitats and spans the latitudinal gradient that has been significant in determining risk of HD outbreaks in other states (Omernik 1987; Christensen et al. 2020; Casey et al. 2021). Our study focused on the interior portion of the region that includes the five states of North Dakota, South Dakota, Nebraska, Kansas, and Oklahoma (hereafter referred to as the Great Plains), because HD patterns have yet to be described in this region (Supplementary Material Fig. S1). Our study area ranged from 33–49° N latitude and 104–96° W longitude, including approximately 958,300 km².

Annual HD questionnaire

The reports of HD between 1982 and 2020 were obtained from a long-term (1982–present) survey of all state wildlife management agencies conducted by the Southeastern Cooperative Wildlife Disease Study (SCWDS; Nettles et al. 1992; Ruder et al. 2016). This survey is completed annually by state wildlife agency personnel in all 50 states who are responsible for the conservation and management of deer populations in the US (Ruder et al. 2016). The survey uses syndromic surveillance to report HD based on characteristic

field signs and postmortem lesions associated with HD in WTD populations, as well as diagnostic testing of select mortality events. Data are collected at the county level in a binary format of 1 or 0 to represent presence or absence of HD, respectively, based on one or more of the four criteria established at the inception of the survey in 1982. These criteria have remained consistent throughout the entire study and include: 1) sudden and unexplained deer mortality that occurs during late summer and early autumn; 2) necropsy-based diagnosis of HD based on lesions; 3) isolation or molecular detection of EHDV or BTV from postmortem tissues; and 4) observation of deer with sloughing hooves (Nettles et al. 1992; Ruder et al. 2016). Depending on the outbreak, a report of HD in a county might involve a small cluster of dead deer or potentially dozens to hundreds of dead deer. For our project, the presence of HD in counties was determined using the first three reporting criteria, to focus on HD mortality. The fourth criterion was excluded from the data set because it focuses more on HD morbidity and was less consistently used across all five states over the 38-yr study period.

Virus detection data (criterion 3) was also compiled for other wild ruminant species in the study area. These data, combined with criterion 3 for WTD, help to corroborate the spatial and temporal patterns of suspected HD (criteria 1 and 2) in WTD for the Great Plains between 1982 and 2020. Use of confirmatory diagnostic testing on all dead wild ruminants during outbreaks is not possible, but their use on a subset provides confidence in HD reports based on criteria 1 and 2 during outbreaks. Virus detections from postmortem tissues (e.g., spleen, lung, lymph node) were performed at SCWDS, as described (Kienzle et al. 2017), or at state or federal veterinary diagnostic laboratories, and reported in the annual HD survey. Reported virus detection data were organized by state, species, serogroup, and serotype, along with the number of years the specific serotypes were reported.

Data analysis

We estimated the potential change in reported HD patterns through data visualization, evaluation of intensity, and frequency of HD reports. To visually observe potential changes in reported frequency of HD trends in the Great Plains, we created county-level maps split into four decades: 1982–

1991, 1992–2001, 2002–2011, and 2012–2020. We define a decade as a set of 9 yr, with the last set resulting in 8 yr, 2012–2020. Reporting frequency was defined as the number of years a county or state reported HD within each of these four decades.

For our temporal analysis, we used linear regression to estimate the change in HD reporting intensity from 1982–2020. We defined reporting intensity as the percentage of counties in a state with reported HD each year (i.e., number of counties with reported HD divided by total number of counties in the state \times 100). Representing reporting intensity as a percentage provided a scale for all counties across the Great Plains states. Scaling the counties allowed us to compare the temporal change in HD reporting intensity from 1982 to 2020 for all five states without bias towards states that have more counties than others. For simplicity, we excluded virus detection data (criterion 3) from wild ruminant species other than WTD from statistical analyses, because the data did not provide new county detections in addition to counties where HD was reported in WTD populations. Therefore, all statistical analyses were conducted on WTD (deer) reports only. Finally, we used linear regression over all counties, regardless of state, to determine a statistical significance in change in HD reporting frequency from 1982 to 2020 in the Great Plains.

For spatial analysis, we used generalized linear mixed models (GLMM) with a binomial distribution to analyze changes in the probability of detecting HD for the Great Plains (Johnson et al. 2006). We developed univariate and multivariate models with independent variables of latitude, longitude, and year to assess how the dependent variable (HD report probability for a county each year) changed over time. We included interaction terms in the multivariate models to determine how the probability of detecting HD changed relative to other independent variables. To account for the numerical differences between the ranges of the independent variables of time and location, we centered and scaled latitude, longitude, and year by standard deviation prior to running models.

Model selection was not used for the temporal linear regression for each state, but we used Akaike information criterion corrected for small sample size (AIC_c) for model selection to determine the top model for spatial changes in HD patterns (Burnham and Anderson 2002). We used

our top spatial model to determine how patterns of HD reports changed both for counties in the Great Plains (excluding state boundaries) and for each state. We ran the top model by each state to account for reporting differences that may have occurred between states between 1982 and 2020. In the model used for all counties in the Great Plains, state was used as a random effect to account for the variation in annual HD reports from all states (Gillies et al. 2006). All linear regression and GLMM models were run using the lme4 package in R version 4.0.3, and statistical significance was determined with $P \leq 0.05$ (Bates et al. 2015; R Core Team 2022).

RESULTS

Over the 38 yr of the study, all states submitted HD reports for each county regardless of HD status. In total, 1,262 county-level HD reports were submitted between 1982 and 2020 across all five states. Counties with the majority of HD reports followed a diagonal pattern from southeast to the northwest corners of the region, similar to larger HD patterns previously described by Stallknecht et al. (2015; Fig. 1A). The counties where EHDV and/or BTV was confirmed through diagnostic testing (criterion 3) during the entire study period are shown in Figure 1B, revealing those counties in the study area where virus confirmation was achieved throughout the distribution of suspected HD (criteria 1 and 2). Virus detections from WTD and other free-ranging ruminant species including mule deer, elk, bighorn sheep, and pronghorn, are shown in Table 1.

Changes in report frequency and report intensity varied for each state. Report frequency increased for all states throughout the decades, but Nebraska experienced a slight decrease in report frequency between the third and fourth decade (Table 2). Within the five states, Oklahoma had the lowest reporting frequency throughout all decades (Table 2). South Dakota had the highest reporting frequency in counties in the fourth decade. In addition to observed increased report frequency, our spatial map of reported HD by

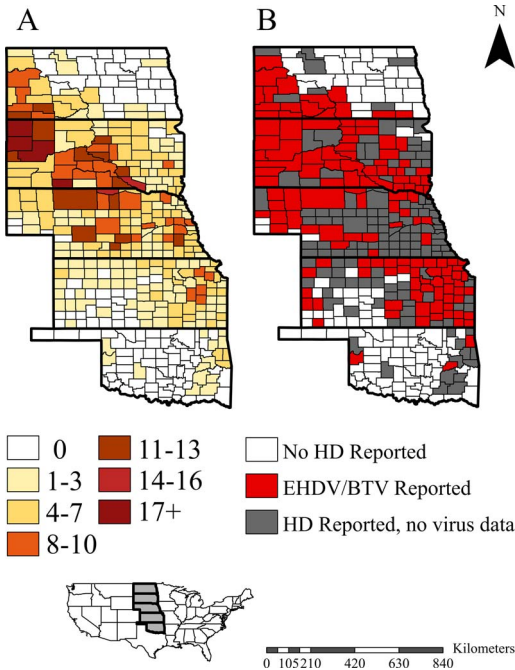


FIGURE 1. (A) Map showing the total years hemorrhagic disease (HD) was reported in white-tailed deer (*Odocoileus virginianus*) by county between 1982 and 2020 in the Great Plains states of (from north to south) North Dakota, South Dakota, Nebraska, Kansas, and Oklahoma, USA. Shading corresponds to the number of years HD was reported in each county during the 38-yr study period. County-level reports of HD were collected for each state annually based upon three specific criteria: 1) sudden and unexplained deer mortality that occurs during late summer and early autumn, 2) necropsy-based diagnosis of HD based on lesions, and 3) isolation or molecular detection of EHDV or BTV. (B) Map showing county reports of HD between 1982 and 2020 (gray) and those counties that reported confirmation of epizootic hemorrhagic disease virus (EHDV) or bluetongue virus (BTV) by laboratory testing (red) 1982–2020. Inset map of the USA shows the five states in the study area.

decade revealed the observed change in geographic distribution of reported HD for all states (Fig. 2).

Reporting intensity also varied between the states over the 38 yr. At the state level, North Dakota, South Dakota, and Kansas had a significant increase ($P \leq 0.05$) in HD reporting intensity, but reporting intensity in Nebraska and Oklahoma did not significantly increase from 1982 to 2020 (Fig. 3). South Dakota

experienced the largest increase in report intensity, with a probability of 0.58, whereas Oklahoma experienced the lowest increase in reporting intensity, with a probability of 0.041 (Table 3). When all counties were combined, reporting intensity did significantly increase ($P \leq 0.05$) from 1982 to 2020 for the Great Plains region (Table 3). Overall, each state experienced at least 1 yr without any counties reporting HD, but North Dakota, South Dakota, Nebraska, and Kansas experienced peaks ($\geq 25\%$) in the percent of counties reporting HD in some years, suggesting geographically large-scale outbreaks (e.g., 2012 in North Dakota, South Dakota, Nebraska, and Kansas; 2020 in North Dakota and South Dakota; Fig. 3).

Lastly, our top spatial model suggested that the probability of detecting HD through the county-level reports increased in both latitude and longitude from 1982 to 2020. Our top spatial model included an interaction with all three variables of year, latitude, and longitude (Supplementary Material Table S1). Latitude and the interaction of latitude and year were not important predictors of spatial change in HD reports, but longitude, year, the interactions of latitude and longitude, and the three-way interaction of year, latitude, and longitude were significant predictors of changing patterns of HD reports (Supplementary Material Table S2). We observed an increase in the probability of detecting HD through a report for every 1.2° increase in longitude but did not observe a significant change in the probability of detecting HD through reports in latitude (Fig. 4; Supplementary Material Table S2).

DISCUSSION

Hemorrhagic disease in the Great Plains generally followed the pattern of enzootic stability in the south transitioning to patterns of epizootic mortality moving north. Consistent with our observation of very few HD reports in Oklahoma, deer populations in Oklahoma have a high population immunity, evidenced

TABLE 1. Diversity of epizootic hemorrhagic disease virus (EHDV) and bluetongue virus (BTV) detections in free-ranging wild ruminants from the Great Plains of the USA, including North Dakota, South Dakota, Nebraska, Kansas, and Oklahoma, between 1982–2020. For each state, the virus detected is presented followed by the years detected. Tissue samples from free-ranging ruminant mortality events were collected by state wildlife management agency personnel during suspected hemorrhagic disease outbreaks and submitted for laboratory confirmation. Viral diagnostics (reverse transcriptase PCR and orbivirus isolation and neutralization) were performed at either the Southeastern Cooperative Wildlife Disease Study (University of Georgia, Athens, Georgia, USA), National Veterinary Services Laboratories (US Department of Agriculture, Ames, Iowa, USA), Michigan State University (East Lansing, Michigan, USA), North Dakota State University (Fargo, North Dakota, USA), or the South Dakota State University (Brookings, South Dakota, USA).

| State | Species ^a | EHDV or BTV serotype (years detected) |
|--------------|----------------------|---|
| North Dakota | White-tailed deer | BTV-17 (1987, 1988); EHDV ^b (1988, 2013, 2017, 2020); EHDV-2 (1987, 1988, 2018–2020) |
| | Mule deer | EHDV ^b (2017); EHDV-2 (2018, 2020) |
| | Elk | EHDV-2 (2020) |
| | Pronghorn | EHDV ^b (2017); EHDV-2 (2020) |
| South Dakota | Bighorn sheep | BTV ^b (2016); EHDV-2 (1997) |
| | White-tailed deer | BTV ^b (1988); BTV-3 (2012); BTV-13 (2012); BTV-17 (2013); EHDV ^b (2016); EHDV-1 (2013, 2016); EHDV-2 (2007–2008, 2011–2013, 2015–2017, 2019–2020); EHDV-6 (2011–2013) |
| | Mule deer | EHDV-2 (2020) |
| | Elk | BTV-13 (2012); EHDV-2 (2012–2013) |
| Nebraska | Pronghorn | BTV-11 (2013); BTV-17 (2016); EHDV-2 (2017) |
| | Bighorn sheep | BTV ^b (2020); BTV-17 (2020); EHDV-2 (2019) |
| | White-tailed deer | BTV ^b (2019); BTV-17 (2016); EHDV ^b (2018, 2020); EHDV-2 (2016–2018, 2020); BTV-5 (2020) |
| | Mule deer | BTV-17 (2016); EHDV-1 (2016); BTV ^b (2018); EHDV-2 (2018) |
| Kansas | Pronghorn | BTV-17 (2016) |
| | White-tailed deer | BTV ^b (2020); BTV-17 (2000, 2003, 2006, 2011); EHDV ^b (2019–2020); EHDV-1 (2012, 2017); EHDV-2 (1994, 1998, 2000–2009, 2011–2013, 2015–2019); EHDV-6 (2017) |
| | Mule deer | BTV-17 (2020); EHDV ^b (2019) |
| Oklahoma | Pronghorn | BTV-1 (2020) |
| | White-tailed deer | BTV ^b (2012); EHDV ^b (2012) |

^a White-tailed deer (*Odocoileus virginianus*), mule deer (*Odocoileus hemionus*), elk (*Cervus canadensis*), pronghorn (*Antilocapra americana*), and bighorn sheep (*Ovis canadensis*).

^b Serotype not determined.

by high EHDV or BTV antibody prevalence with minimal HD mortality (Kocan et al. 1987). Moving north, the percent of counties with reported HD increased in Kansas, especially in the most recent decade (Table 2). Similar to Oklahoma, a pattern of enzootic stability has been identified in western Kansas (Flacke et al. 2004). However, Kansas appears

to be an epidemiological transition state, with evidence of enzootic stability in the western counties transitioning to epizootic mortality in the east (Flacke et al. 2004). In our study, most Kansas counties reporting HD were primarily in the eastern half of the state (Figs. 1 and 2). Patterns of epizootic mortality intensified moving north from Kansas into Nebraska

TABLE 2. Total positive county reports of hemorrhagic disease (HD) in white-tailed deer (*Odocoileus virginianus*) in Oklahoma, Kansas, Nebraska, South Dakota, and North Dakota, USA, by decade, submitted to the Southeastern Cooperative Wildlife Disease Study (SCWDS) between 1982 and 2020. The study period was divided into four decades: 1982–1991 (1), 1992–2001 (2), 2002–2011 (3), and 2012–2020 (4). County-level reports of suspected or confirmed HD were collected for each state annually based on three specific criteria: 1) sudden and unexplained deer mortality that occurs during late summer and early autumn, 2) necropsy-based diagnosis of HD based on lesions, and 3) isolation or molecular detection of epizootic hemorrhagic disease virus or bluetongue virus. Positive county reports represent the number of counties with reported HD in each decade. The total potential reports represent the number of opportunities to report HD in each state by decade (number of counties multiplied by number of years).

| State (number of counties) | Decade | Positive county reports | Total potential reports | Percent positive counties |
|----------------------------|--------|-------------------------|-------------------------|---------------------------|
| North Dakota (53) | 1 | 16 | 530 | 3.0% |
| | 2 | 17 | 530 | 3.2% |
| | 3 | 39 | 530 | 7.4% |
| | 4 | 37 | 477 | 7.8% |
| South Dakota (66) | 1 | 71 | 660 | 10.8% |
| | 2 | 83 | 660 | 12.6% |
| | 3 | 74 | 660 | 11.2% |
| | 4 | 191 | 594 | 32.2% |
| Nebraska (93) | 1 | 74 | 930 | 8.0% |
| | 2 | 101 | 930 | 10.9% |
| | 3 | 164 | 930 | 17.6% |
| | 4 | 111 | 837 | 13.3% |
| Kansas (105) | 1 | 12 | 1050 | 1.1% |
| | 2 | 23 | 1050 | 2.2% |
| | 3 | 59 | 1050 | 5.6% |
| | 4 | 157 | 945 | 16.6% |
| Oklahoma (77) | 1 | 2 | 770 | 0.3% |
| | 2 | 10 | 770 | 1.3% |
| | 3 | 6 | 770 | 0.8% |
| | 4 | 15 | 693 | 2.2% |

where HD reports have remained relatively consistent from 1982 to 2020, although Nebraska experienced peaks in mortality similar to the other four states (Fig. 3). In South Dakota over the past decade there is evidence of increasing outbreak intensity, as the percentage of counties reporting HD increased from 11.2% to 32.2% (Table 2). Furthermore, we showed an increase in the geographic distribution of reported HD in South Dakota, which was corroborated by postmortem diagnostic data (i.e., virus detections), from 1982 to 2020 (Table 1).

Although frequency of reported HD increased over the four decades, the geographic distribution of HD also changed in the Great Plains. Unlike northern expansion of HD previously observed in states east of the Great Plains

(Stallknecht et al. 2015; Christensen et al. 2020; Dorak et al. 2022), our findings indicated that HD has expanded longitudinally in an eastern direction from 1982 to 2020 (Fig. 4). Hemorrhagic disease has been reported in North Dakota during all four decades of the study period, therefore we could not observe significant change in reported HD relative to latitude alone, but both visual observation and linear regression suggest that counties reporting HD have expanded eastward and northward across the Great Plains (Fig. 2).

While not investigated in our study, the overall shift in the distribution, intensity, and frequency of reported HD for the Great Plains may be related to recent changes in climate conditions (i.e., increasing annual temperatures)

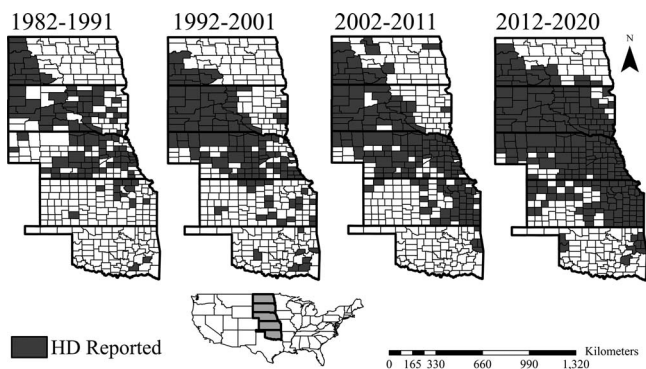


FIGURE 2. Maps showing reports of hemorrhagic disease (HD) grouped by decade between 1982 and 2020 for the Great Plains states of (from north to south) North Dakota, South Dakota, Nebraska, Kansas, and Oklahoma, USA. Shaded counties represent those counties where HD was reported at least once during the decade. County-level reports of HD were collected for each state annually based upon three specific criteria: 1) sudden and unexplained deer mortality that occurs during late summer and early autumn, 2) necropsy-based diagnosis of HD based on lesions, and 3) isolation or molecular detection of epizootic hemorrhagic disease virus or bluetongue virus. Inset map of the USA shows the five states in the study area.

and subsequent impact on vector populations and virus transmission (Purse et al. 2005). Temperature impacts vectorial capacity by dictating vector developmental rates, biting rates, and virogenesis (Mullens et al. 2004). As temperature increases between 27 C and 30 C, the *Culicoides* generation time and extrinsic incubation period decreases, resulting in more infected adult *Culicoides* on the landscape (Wittman et al. 2002). In regions with patterns of epizootic mortality such as North Dakota and South Dakota, changing climatic variables may enhance vectorial capacity (Sleeman et al. 2009; Salley et al. 2016). Although we acknowledge that certain climate components such as temperature and moisture influence vector-virus interactions, further research is warranted to understand how habitat in relation to climate impacts HD reports in the Great Plains.

A noteworthy change in HD reporting intensity and frequency in the Great Plains occurred between South Dakota and North Dakota. In the northeastern US, the effect of latitude was significant in predicting HD mortality relative to drought index, but reported HD generally decreased north of 40° N latitude (Stallknecht et al. 2015; Christensen et al. 2020). Similar to the trend of reduced HD reporting frequency at northern latitudes, HD reporting frequency and

intensity increased at a smaller rate in North Dakota versus South Dakota, potentially related to the effect of local climate conditions on vectorial capacity. Populations of *C. sonorensis* (a confirmed vector of EHDV and BTV), were found during 2002 in regions of western North Dakota, western South Dakota, and Nebraska where soils are nonglaciated and annual evaporation exceeds annual precipitation (Schmidtman et al. 2011). Whether *C. sonorensis* populations have since expanded into areas east of the Missouri River, or whether other resident *Culicoides* spp. are responsible for transmission of EHDV and BTV in eastern North Dakota and South Dakota remains unclear because of the paucity of *Culicoides* surveillance in the region. Other *Culicoides* spp. that are suspected vectors of EHDV and BTV and could play a role in the region include *Culicoides stellifer*, *Culicoides paraenesis*, *Culicoides venustus*, and *Culicoides haematopotus*, among others (McGregor et al. 2022). Regardless of the underlying factors involved, the differences in reported HD intensity and frequency between South Dakota to North Dakota might demarcate the northeastern boundary of HD in the Great Plains and changing climatic conditions might continue to shift the leading edge of HD in a northeasterly direction.

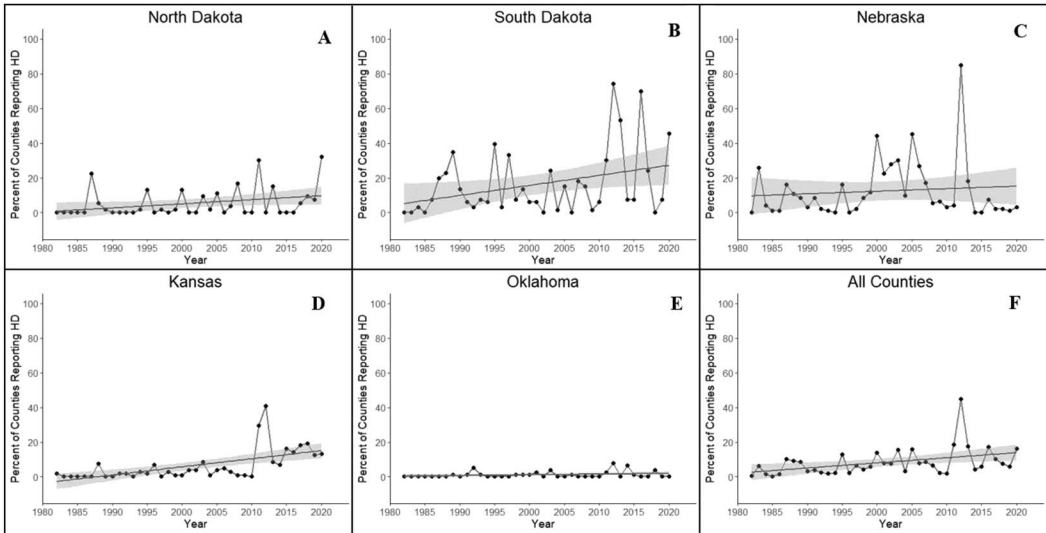


FIGURE 3. Graphs showing reporting intensity of hemorrhagic disease (HD) between 1982 and 2020 for the Great Plains states of (from north to south) North Dakota, South Dakota, Nebraska, Kansas, and Oklahoma, USA. County-level reports of HD were collected for each state annually based upon three specific criteria: 1) sudden and unexplained deer mortality that occurs during late summer and early autumn, 2) necropsy-based diagnosis of HD based on lesions, and 3) isolation or molecular detection of epizootic hemorrhagic disease virus or bluetongue virus. The percentage of counties with reported HD in each state (A–E) or for all five states combined (F) represent the reporting intensity for each year in the study. Ninety-five percent confidence intervals are represented by the shaded area in each panel. The states of North Dakota, South Dakota, and Kansas had significant increases ($P < 0.05$) in HD reporting intensity from 1982 to 2020 as determined by linear regression.

As climate continues to change, vector–virus interactions and their impact on HD morbidity and mortality in northern deer populations will also change. Continued surveillance is essential for monitoring such impacts, understanding their drivers, and anticipating implications for deer management. One notable HD outbreak

occurred in 2012 across most of the Midwest, impacting both free-ranging and domestic ruminant species (Stevens et al. 2015; Ruder et al. 2015). Data from our study clearly show prominent peaks of HD reports during 2012, in Kansas, Nebraska, and South Dakota, consistent with that outbreak (Fig. 3). The 2012

TABLE 3. Results of linear regression analysis to determine change in annual hemorrhagic disease reporting intensity for white-tailed deer (*Odocoileus virginianus*) in Great Plains states of the US from 1982 to 2020. Linear regression was run separately for each state to account for reporting bias between different state wildlife management agencies. All counties were combined for linear regression with state as the random effect to account for state variation in reporting effort. Shown are coefficient estimates \pm standard error (SE), P -values, and 95% confidence intervals (CI). Statistical significance of $P \leq 0.05$.

| USA state | Estimate \pm SE | Pr(> z) | Lower 95% CI | Upper 95% CI |
|--------------|-------------------|----------|--------------|--------------|
| North Dakota | 0.235 \pm 0.115 | 0.048 | 0.002 | 0.468 |
| South Dakota | 0.578 \pm 0.255 | 0.029 | 0.061 | 1.094 |
| Nebraska | 0.147 \pm 0.241 | 0.545 | –0.341 | 0.635 |
| Kansas | 0.468 \pm 0.102 | 4.81e-05 | 0.262 | 0.674 |
| Oklahoma | 0.041 \pm 0.027 | 0.138 | –0.014 | 0.095 |
| All Counties | 0.960 \pm 0.105 | 0.007 | 8.41e-02 | 0.507 |

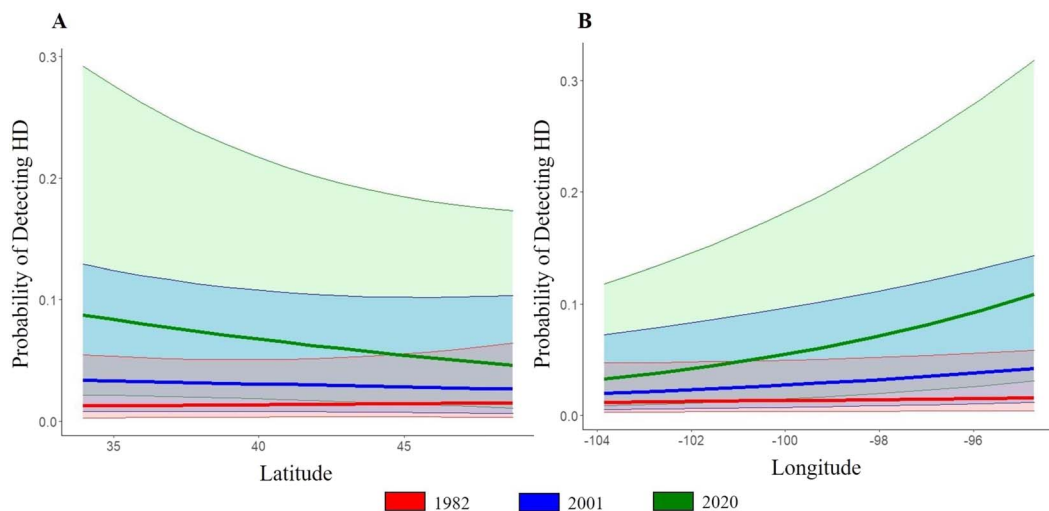


FIGURE 4. Probability of detecting hemorrhagic disease (HD) through county-level reports relative to latitude (A) and longitude (B) for the Great Plains area of the USA between 1982 and 2020. County-level reports of HD were collected for each state annually based upon three specific criteria: 1) sudden and unexplained deer mortality that occurs during late summer and early autumn, 2) necropsy-based diagnosis of HD based on lesions, and 3) isolation or molecular detection of epizootic hemorrhagic disease virus or bluetongue virus. Latitude and longitude ranges in the graphs represent the geographic boundaries of the Great Plains. Colored lines denote the probability of detecting HD through reports for the respective year. Shaded areas represent 95% confidence intervals, and lines represent the linear regression for the years 1982 (red), 2001 (blue), and 2020 (green). Years were selected to denote the probability of detecting HD at the beginning, middle, and end point of the study. Latitude or longitude is represented on the x-axis and the probability of detecting HD through a county report is represented on the y-axis.

Midwest HD outbreak was predominantly attributed to EHDV-2, and the severe drought conditions and high temperatures during the summer months probably contributed to the intensity of the outbreak (Stevens et al. 2015). Given that the Great Plains encompasses a gradient of different climate conditions, considerable changes in climate might affect how vector-borne diseases, such as HD, present on the landscape. Thus, it is plausible that HD may become an increasingly common mortality factor among some deer populations.

One important aspect of the complex HD system is host diversity and how different hosts for EHDV or BTV affect HD distribution on the landscape. Although WTD are considered a primary host for EHDV and BTV in North America, other free-living species such as mule deer, elk, and pronghorn also are frequently involved in outbreaks in the Great Plains (Howerth et al. 2001). Furthermore, livestock hosts for EHDV and

BTV, such as cattle and sheep, are common in the Great Plains, are often sympatric with wild ruminants, and are known to affect populations of certain *Culicoides* species (Hardy and Price 1952; Ruder et al. 2015; McGregor et al. 2022). Although wild ruminant species often suffer severe or fatal disease, cattle infected with EHDV and BTV typically develop inapparent or mild disease. Both domestic and wild ruminant species are vertebrate hosts defining this disease system, and the presence and movement of susceptible livestock species may potentially influence the occurrence and movement of EHDV and BTV on the landscape (Rivera et al. 2021). To understand the complexities of this vector-borne disease system involving multiple ruminant hosts and multiple viruses, future research and surveillance efforts should incorporate both domestic and free-living ruminant hosts, as well as *Culicoides* spp. vectors. Specifically, long-term studies that spatially and

temporally combine investigations of free-living ruminant mortality; serological testing of harvested free-living ruminants; serologic and virologic testing of sentinel livestock; and *Culicoides* spp. surveillance across diverse land and animal use settings, would collectively help inform the understanding of orbivirus epidemiology in the Great Plains and elsewhere.

To our knowledge, no other HD data set exists for a similar length of time and spatial coverage. The HD survey relies on the continued participation of state wildlife management agency personnel, but we acknowledge certain levels of bias exist within the data set. Disease events in wildlife may be difficult to observe, thus potentially leading to underreporting of events such as HD. The HD survey primarily captures outbreaks with mortality, but small-scale mortality in dense vegetation or where humans are not present, for example, may not be detected. In enzootically stable regions, virus transmission events do not result in significant morbidity or mortality, and these inapparent infections are therefore not accounted for in this survey (e.g., in Oklahoma). Further, reports of HD were received across the 38-yr period from independent sources that could not be standardized between states (Stallknecht et al. 2015). Nevertheless, reliability of these data is made possible by the diligence and continued support of state agency personnel, well-characterized and consistent field signs and necropsy lesions of HD in WTD, consistent criteria for reporting HD by state agencies, the relative visibility of deer to the public, and the value of deer as a natural resource (Ruder et al. 2016). The validity of our survey is further supported by lack of outlier reports in areas with patterns of epizootic mortality (e.g., northeast US); consistency in spatial and temporal reporting of HD during outbreaks regardless of state borders; and through confirmatory diagnostics throughout the study (criterion 3; Stallknecht et al. 2015). Given the duration and consistent case criteria, this survey provides unique opportunities to study spatial and temporal

changes in patterns of HD that short term projects could not capture.

The general pattern of increase in the frequency and intensity of reported HD in parts of the central and northern Great Plains, and the significant latitude and longitude interaction supports the concept that conditions have changed in favor of HD outbreaks from 1982 to 2020. Additionally, our finding that counties reporting HD expanded eastward in North Dakota and South Dakota suggests that this region may represent the leading edge of HD in the northern Great Plains. Continued monitoring of EHDV and BTV in the multihost system of the Great Plains is important to document future changes in the face of changing climate.

ACKNOWLEDGMENTS

This study was made possible through long-term financial support from state member wildlife agencies of Southeastern Cooperative Wildlife Disease Study (SCWDS) provided by the Federal Aid to Wildlife Restoration Act (50 Stat. 917). We also thank the US Fish and Wildlife Service National Wildlife Refuge System and the US Geological Survey Ecosystems Mission Area for their long-term support of SCWDS. Furthermore, this research depends on the continuous efforts of the numerous wildlife biologists, veterinarians, and technicians from state wildlife agencies that contribute to this long-term survey and provide diagnostic samples for testing. We are especially grateful to current and former staff with the North Dakota Game and Fish Department, South Dakota Game, Fish & Parks, Nebraska Game and Parks Commission, Kansas Department of Wildlife & Parks, and Oklahoma Department of Wildlife Conservation for providing data through the annual survey. We thank D. Barber, S. Hesting, T. Nordeen, and S. Griffin for reviewing this manuscript. Additional virus detections were made by the US Department of Agriculture's National Veterinary Services Laboratories and the South Dakota State University Animal Disease Research and Diagnostic Laboratory. Laboratory and data management support at SCWDS were provided by V. Nettles, J. Fischer, R. Poulson, C. Kienzle, N. Stilwell, A. Allison, S. Vigil, and C. McElwee.

SUPPLEMENTARY MATERIAL

Supplementary material for this article is online at <http://dx.doi.org/10.7589/JWD-D-23-00021>.

LITERATURE CITED

- Bates D, Mächler M, Bolker B, Walker S. 2015. Fitting linear mixed-effects models using lme4. *J Stat Software* 67(1):1–48.
- Boyer TC, Ward MP, Wallace RL, Singer RS. 2007. Regional seroprevalence of bluetongue virus in cattle in Illinois and western Indiana. *Am J Vet Res* 68:1212–1219.
- Burnham KP, Anderson DR. 2002. Information and likelihood theory: A basis for model selection and inference. *Model selection and multimodel inference: A practical information-theoretic approach*. 2nd Ed. Springer, New York, New York. pp. 49–97.
- Casey CL, Rathbun SL, Stallknecht DE, Ruder MG. 2021. Spatial analysis of the 2017 outbreak of hemorrhagic disease and physiographic region in the eastern United States. *Viruses* 13:550.
- Christensen SA, Ruder MG, Williams DM, Porter WF, Stallknecht DE. 2020. The role of drought as a determinant of hemorrhagic disease in the eastern United States. *Glob Chang Biol* 26:3799–3808.
- Couvillion C, Nettles V, Davidson W, Pearson J, Gustafson G. 1981. Hemorrhagic disease among white-tailed deer in the Southeast from 1971 through 1980. In: *Proceedings of the 85th Annual Meeting of the United States Animal Health Association*, St. Louis, Missouri, 11–16 October. United States Animal Health Association, Saint Joseph, Missouri, pp. 522–537.
- Dorak SJ, Varga C, Ruder MG, Gronemeyer P, Rivera NA, Dufford DR, Skinner DJ, Roca AL, Novakofski J, Mateus-Pinilla NE. 2022. Spatial epidemiology of hemorrhagic disease in Illinois wild white-tailed deer. *Sci Rep* 12:6888.
- Flacke GL, Yabsley MJ, Hanson BA, Stallknecht DE. 2004. Hemorrhagic disease in Kansas: Enzootic stability meets epizootic disease. *J Wildl Dis* 40:288–293.
- Gaydos JK, Crum JM, Davidson WR, Cross SS, Owen SF, Stallknecht DE. 2004. Epizootology of an epizootic hemorrhagic disease outbreak in West Virginia. *J Wildl Dis* 40:383–393.
- Gaydos JK, Davidson WR, Elvinger F, Mead DG, Howerth EW, Stallknecht DE. 2002a. Innate resistance to epizootic hemorrhagic disease in white-tailed deer. *J Wildl Dis* 38:713–719.
- Gaydos JK, Stallknecht DE, Kavanaugh D, Olson RJ, Fuchs ER. 2002b. Dynamics of maternal antibodies to hemorrhagic disease viruses (Reoviridae: Orbivirus) in white-tailed deer. *J Wildl Dis* 38:253–257.
- Gillies CS, Hebblewhite M, Nielsen SE, Krawchuk MA, Aldridge CL, Frair JL, Saher DJ, Stevens CE, Jerde CL. 2006. Application of random effects to the study of resource selection by animals. *J Anim Ecol* 75:887–898.
- Hardy WT, Price DA. 1952. Soremuzzle of sheep. *J Am Vet Med Assoc* 120:23–25.
- Howerth EW, Stallknecht DE, Kirkland PD. 2001. Bluetongue, epizootic hemorrhagic disease, and other Orbivirus-related diseases. In: *Infectious diseases of wild mammals*. 3rd Ed. Williams ES, Barker IK, editors. Iowa State University Press, Iowa, pp. 77–97.
- Johnson CJ, Nielsen SE, Merrill EH, McDonald TL, Boyce MS. 2006. Resource selection functions based on use–availability data: Theoretical motivation and evaluation methods. *J Wildl Manage* 70:347–357.
- Kienzle C, Poulson RL, Ruder MG, Stallknecht DE. 2017. Virus isolation and molecular detection of bluetongue and epizootic hemorrhagic disease viruses from naturally infected white-tailed deer (*Odocoileus virginianus*). *J Wildl Dis* 53:843–849.
- Kocan AA, Castro AE, Shaw MG, Rogers SJ. 1987. Bluetongue and epizootic hemorrhagic disease in white-tailed deer from Oklahoma: Serologic evaluation and virus isolation. *Am J Vet Res* 48:1048–1049.
- McGregor BL, Shults PT, McDermott EG. 2022. A review of the vector status of North American *Culicoides* (Diptera: Ceratopogonidae) for bluetongue virus, epizootic hemorrhagic disease virus, and other arboviruses of concern. *Curr Trop Med Rep* 9:130–139.
- Mullens BA, Gerry AC, Lysyk TJ, Schmidtman ET. 2004. Environmental effects on vector competence and virogenesis of bluetongue virus in *Culicoides*: Interpreting laboratory data in a field context. *Vet Ital* 40:160–166.
- Nettles VF, Davidson WR, Stallknecht DE. 1992. Surveillance for hemorrhagic disease in white-tailed deer and other wild ruminants, 1980–1989. In: *Proceedings of the 46th Annual Conference of the Southeastern Association of Fish and Wildlife Agencies*, Corpus Christi, Texas, 25–28 October 1992, pp. 138–146.
- Nettles VF, Stallknecht DE. 1992. History and progress in the study of hemorrhagic disease of deer. In: *Transactions of the 57th North American Wildlife and Natural Resources Conference*, Charlotte, North Carolina, 27 March–1 April, pp. 499–516.
- Omernik JM. 1987. Ecoregions of the conterminous United States. *Ann Assoc Am Geogr* 77:118–125.
- Purse BV, Mellor PS, Rogers DJ, Samuel AR, Mertens PPC, Baylis M. 2005. Climate change and the recent emergence of bluetongue in Europe. *Nat Rev Microbiol* 3:171–181.
- R Core Team. 2022. R: A language and environment for statistical computing. R Foundation, Vienna, Austria.
- Rivera NA, Varga C, Ruder MG, Dorak SJ, Roca AL, Novakofski JE, Mateus-Pinilla NE. 2021. Bluetongue and epizootic hemorrhagic disease in the United States of America at the wildlife–livestock interface. *Pathogens* 10:915.
- Ruder MG, Fischer JR, Stallknecht DE. 2016. Monitoring hemorrhagic disease: What every wildlife professional needs to know. In: *Transactions of the 81st North American Wildlife and Natural Resources Conference*, Pittsburgh Pennsylvania, 13–18 March 2016, pp. 121–131.
- Ruder MG, Johnson D, Ostlund E, Allison AB, Kienzle C, Phillips JE, Poulson RL, Stallknecht DE. 2017. The first 10 years (2006–15) of epizootic hemorrhagic disease virus serotype 6 in the USA. *J Wildl Dis* 53:901–905.
- Ruder MG, Lysyk TJ, Stallknecht DE, Foil LD, Johnson DJ, Chase CC, Dargatz DA, Gibbs EPJ. 2015. Transmission and epidemiology of bluetongue and epizootic hemorrhagic disease in North America: Current

- perspectives, research gaps, and future directions. *Vector Borne Zoonotic Dis* 15:348–363.
- Salley SW, Sleezer RO, Bergstrom RM, Martin PH, Kelly EF. 2016. A long-term analysis of the historical dry boundary for the Great Plains of North America: Implications of climatic variability and climatic change on temporal and spatial patterns in soil moisture. *Geoderma* 274:104–113.
- Schmidtman ET, Herrero MV, Green AL, Dargatz DA, Rodriguez JM, Walton TE. 2011. Distribution of *Culicoides sonorensis* (Diptera: Ceratopogonidae) in Nebraska, South Dakota, and North Dakota: Clarifying the epidemiology of bluetongue disease in the northern Great Plains region of the United States. *J Med Entomol* 48:634–643.
- Shope RE, MacNamara LG, Mangold R. 1960. A virus-induced epizootic hemorrhagic disease of the Virginia white-tailed deer (*Odocoileus virginianus*). *J Exp Med* 111:155–170.
- Sleeman JM, Howell JE, Knox WM, Stenger PJ. 2009. Incidence of hemorrhagic disease in white-tailed deer is associated with winter and summer climatic conditions. *Ecohealth* 6:11–15.
- Stair EL, Robinson RM, Jones IP. 1968. Spontaneous bluetongue in Texas white-tailed deer. *Path Vet* 5:164–173.
- Stallknecht DE, Allison AB, Park AW, Phillips JE, Goekjian VH, Nettles VF, Fischer JR. 2015. Apparent increase of reported hemorrhagic disease in the midwestern and northeastern United States. *J Wildl Dis* 51:348–361.
- Stallknecht DE, Blue JL, Rollor EA, Nettles VF, Davidson WR, Pearson JE. 1991. Precipitating antibodies to epizootic hemorrhagic disease and bluetongue viruses in white-tailed deer in the southeastern United States. *J Wildl Dis* 27:238–247.
- Stallknecht DE, Howerth EW. 2004. Epidemiology of bluetongue and epizootic hemorrhagic disease in wildlife: Surveillance methods. *Vet Ital* 40:203–207.
- Stallknecht DE, Howerth EW, Gaydos JK. 2002. Hemorrhagic disease in white-tailed deer: Our current understanding of risk. In: *Transactions of the 67th Annual North American Wildlife and Natural Resources Conference*, pp. 75–86.
- Stallknecht DE, Luttrell MP, Smith KE, Nettles VF. 1996. Hemorrhagic disease in white-tailed deer in Texas: A case for enzootic stability. *J Wildl Dis* 32:695–700.
- Stevens G, McCluskey B, King A, O’Hearn E, Mayr G. 2015. Review of the 2012 epizootic hemorrhagic disease outbreak in domestic ruminants in the United States. *PLoS One* 10:e0133359.
- Stilwell NK, Clarke LL, Howerth EW, Kienzle-Dean C, Fojtik A, Hollander LP, Carter D, Osborn DA, D’Angelo GJ, Miller KV, et al. 2021. The effect of maternal antibodies on clinical response to infection with epizootic hemorrhagic disease virus in white-tailed deer (*Odocoileus virginianus*) fawns. *J Wildl Dis* 57:189–193.
- Wittman EJ, Mellor PS, Baylis M. 2002. Effect of temperature on the transmission of orbiviruses by the biting midge, *Culicoides sonorensis*. *Med Vet Entomol* 16:147–156.
- Zhu Z, Bouchard M, Butman D, Hawbaker T, Li Z, Liu J, Liu S, McDonald C, Reker R, Sayler K, et al. 2011. *Baseline and projected future carbon storage and greenhouse-gas fluxes in the Great Plains region of the United States. Professional Paper 1787*. US Geological Survey, Reston, Virginia.

Submitted for publication 14 February 2023.

Accepted 13 February 2024.