



Cedrus atlantica Pollen Morphology and Investigation of Grain Size Variability Using Laser Diffraction Granulometry

Authors: Bell, Benjamin A., Bishop, Thomas H., Fletcher, William J., Ryan, Peter, and Ilmen, Rachid

Source: Palynology, 42(3) : 339-353

Published By: AASP: The Palynological Society

URL: <https://doi.org/10.1080/01916122.2017.1356760>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.



Cedrus atlantica pollen morphology and investigation of grain size variability using laser diffraction granulometry

Benjamin A. Bell ^a, Thomas H. Bishop ^a, William J. Fletcher ^a, Peter Ryan ^a and Rachid Ilmen ^b

^aQuaternary Environments and Geoarchaeology (QEG), Department of Geography, School of Environment, Education and Development, The University of Manchester, Oxford Road, Manchester, M13 9PL, UK; ^bDepartment of Hydraulic, Environment and Climate (HEC), Hassania School of Public Works (EHTP), Oasis-Casablanca, Morocco

ABSTRACT

The morphology and size variability of pollen grains of *Cedrus atlantica* were investigated using a novel approach employing laser diffraction granulometry. We provide new insights into size variability and present high-quality light microscopy (LM) and scanning electron microscopy (SEM) imagery of *Cedrus atlantica* pollen. Grains have an average size of $59.1 \pm 4.0 \mu\text{m}$, measured on millions of grains from 91 samples. Analysis showed there is high variability of grain size within individual samples, although variability between samples is not significant. We found no significant relationships between grain size and climate (including temperature, precipitation and aridity), and suggest that grain size of fossil *Cedrus* pollen would not be a good proxy for climate reconstruction. Grain size may be influenced by a number of complex factors such as genome size or adaptations to support wind pollination, while variability within individual samples may result from the irregular development of pollen. The laser diffraction method produced repeatable, robust measurements on millions of pollen grains which are highly correlated with measurements taken using LM ($r = 0.91$, $p = 0.002$). Where grain size information is crucial for pollen identification, for developing isolation techniques for geochemical analysis, for investigating climatic and environmental influence, or for investigating links between genomes and grain size, particle size analysis by laser diffraction provides a reproducible and robust method for quickly determining pollen grain size on many samples.

KEYWORDS

Cedrus atlantica; pollen morphology; pollen size; grain size methods; laser diffraction granulometry; climate influence; moisture availability

1. Introduction

Detailed information on pollen morphology and grain size is critical for palynologists to accurately identify vegetation from fossil pollen assemblages for pollen analysis. Grain size may be studied to improve the taxonomic resolution of pollen identification, for example with *Pinus* pollen (Desprat et al. 2015), *Poaceae* pollen (Radaeski et al. 2016), and indeed with *Cedrus* pollen (Fujiki et al. 2003). Pollen size has also been correlated with genome size and may indicate polyploidy (three or more chromosome sets) in plants (Gould 1957; Kapadia & Gould 1964; Bennett 1972; Tate et al. 2005; Knight et al. 2010; De Storme et al. 2013). Increasingly, geochemical studies utilising pollen, such as stable isotope analysis for palaeoclimate reconstructions (Amundson et al. 1997; Loader & Hemming 2004; Nelson et al. 2006, 2007; King et al. 2012; Nelson 2012; Bell et al. 2017) and biomarker analysis for UV-B reconstructions (Rozema et al. 2001, 2002; Fraser et al. 2011; Willis et al. 2011; Lomax et al. 2012; Jardine et al. 2017), require detailed knowledge of grain size for developing techniques to isolate specific grains from fossil assemblages. For example, the use of micro-sieving to concentrate pollen from sediment (Heusser & Stock 1984; Brown et al. 1989) can be modified to target grains within a specified size range to facilitate the isolation of pollen for specific species.

Traditional pollen analysis relies on visual identification of pollen grains to determine vegetation composition, while

geochemical studies undertake further analysis of the grains to gain insight into environmental or climate conditions. It has also been hypothesised that the size and shape of the pollen grain itself may be influenced by climate (Ejmsmond et al. 2011). Temperature has been previously linked to pollen grain size (Kurtz & Liverman 1958), and Schoch-Bodmer (1936) proposed that grain size variability in pollen is a result of fluctuations in moisture availability in ambient air during pollen development. Ejmsmond et al. (2011) analysed eight *Rosaceae* species and found grain size increased under desiccation stress (determined by temperature, potential evapotranspiration and altitude). A positive relationship was also found between temperature and pollen size in 232 plant species from 11 taxonomic groups, suggesting possible climatic influence on size during the flowering period (Ejmsmond et al. 2015) and offering potential for pollen grain size to aid in reconstructions of past environments.

Measurements of grain width on modern *Nothofagus* spp. pollen showed a significant relationship with mean annual precipitation (MAP), where grain size increased with reduced MAP ($r^2 = 0.66$, $p < 0.0001$). Applying this relationship to fossil pollen samples from Antarctica, where average grain size increased by 23% from the late Eocene to the mid Miocene, suggests a decrease in precipitation during this period (Griener & Warny 2015). This is supported by an earlier study on the same fossil samples which found stable carbon isotope discrimination

($\Delta^{13}\text{C}$) values of pollen decreased during the same period, which the authors also linked to a decrease in moisture availability (Griener et al. 2013). However, the validity of the relationship between grain size and moisture availability in this study has been questioned, citing a lack of theoretical and empirical support, with a view that it may be premature to use grain size as a moisture availability proxy (Jardine & Lomax 2017).

Atlas cedar (*Cedrus atlantica* (Endl.) Manetti ex Carrière) is a moisture-sensitive (Rhanem 2011; Linares et al. 2013; Ilmen et al. 2014) montane conifer endemic to semi-arid and humid areas of Morocco and Algeria (Farjon 1990), with pollen records indicating a presence in the area since at least the Last Glacial Maximum (Lamb et al. 1989; Magri et al. 2017; Zielhofer et al. 2017). The earliest work on *Cedrus atlantica* pollen morphology found the average size of the grain to be $61.5 \pm 1.8 \mu\text{m}$ based on 100 grain measurements of pollen from a single tree in Ifrane, Morocco (Aytug 1961), while a later study recorded a size of $45.8 \pm 2.4 \mu\text{m}$ measured on pollen collected from one tree in Marseille, France (Fujiki et al. 2003). In both studies, pollen samples had been collected and stored 15 years prior to

measurements being taken. *Cedrus atlantica* pollen grains have also been measured at $75 \mu\text{m}$ on samples from Turkey (Altuner et al. 2012), and $58 \mu\text{m}$ in Vancouver, Canada (Ho 1972). In Algeria, samples from the Tell Atlas ranged in size from 60 to $63 \mu\text{m}$, while samples from the eastern margins of the Saharan Atlas and Aurès Mountains measured 54 to $60 \mu\text{m}$ (Derridj et al. 1991).

The literature suggests that pollen from *Cedrus atlantica* varies in size significantly, which could possibly result from climate differences between sample locations. However, variations noted in these studies may also be due to different methodological approaches undertaken for measurement which could have affected grain size – e.g. changes resulting from chemical pre-treatments, pollen hydration, mounting media, cover-slip pressure and storage methods (Andersen 1960; Aytug 1960; Faegri & Deuse 1960; Cushing 1961; Reitsma 1969; Praglowski 1970) – so it is not possible to determine from current literature whether there is a climatic influence.

In this study, we provide a comprehensive analysis of grain size variability of *Cedrus atlantica* pollen from 91 modern samples across the Middle Atlas, Morocco (Figure 1), and a wider

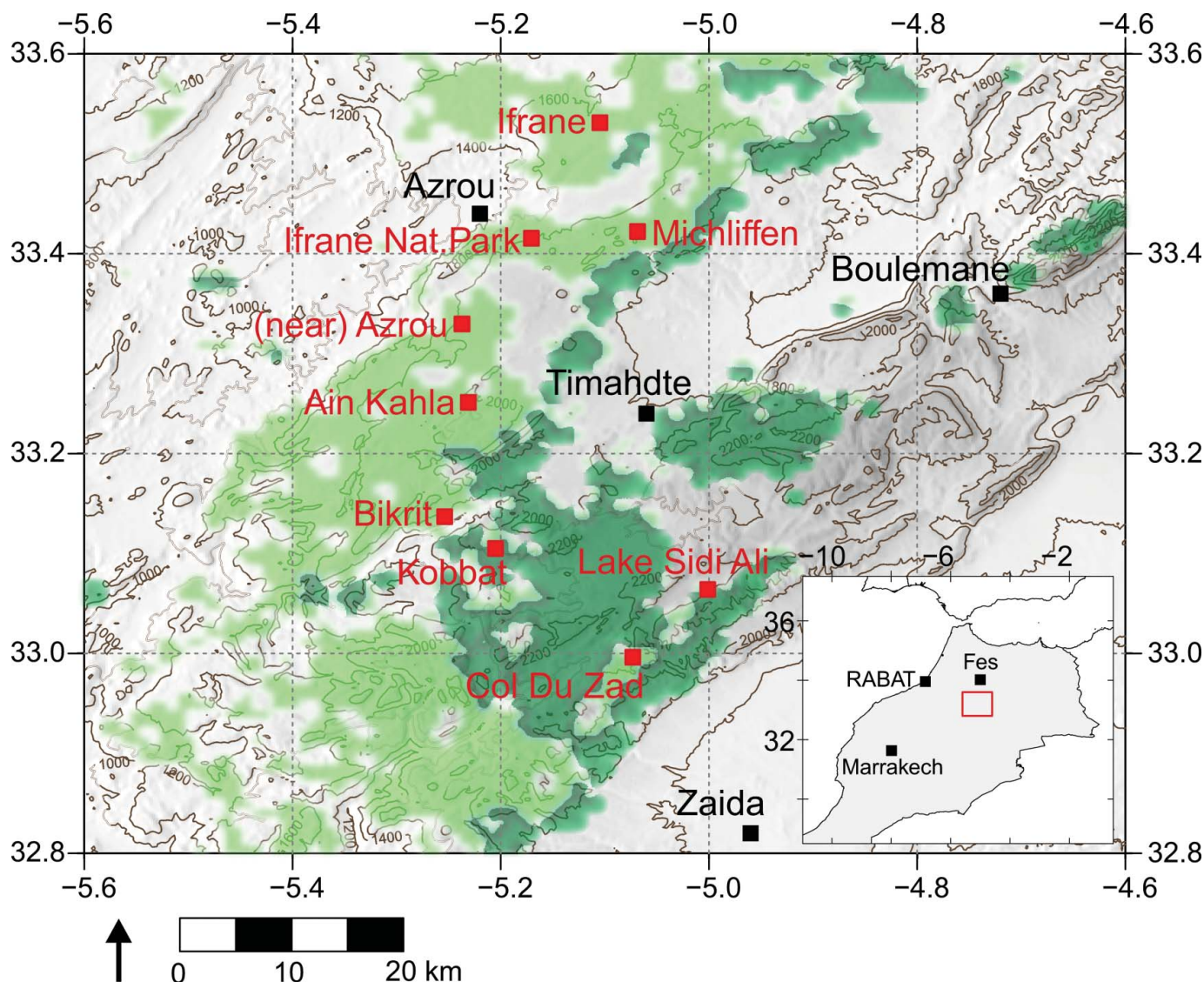


Figure 1. Map of the Middle Atlas and forest cover: dark green is predominately needleleaf evergreen, and light green is mixed broadleaf evergreen and needleleaf evergreen. Sample areas are shown in red and towns in black. Map created using Global Multi-resolution Terrain Elevation Data 2010 (GMTED2010) and forestry data extracted from Global Land Cover Characterization (GLCC) imagery. Data available from US Geological Survey (2017).

environmental gradient incorporating botanical garden samples from Europe and the USA. We employ a novel approach to determining pollen size by using non-destructive laser diffraction granulometry to consistently measure thousands to millions of individual pollen grains. We compare this technique to traditional measurements taken under light microscopy (LM), and we additionally measure samples following chemical treatment to analyse its effect on grain size. We describe the morphology and aim to determine the overall size of *Cedrus atlantica* pollen, and determine whether grain size variability is influenced by climate or environmental factors.

2. Material and methods

2.1. Sample collection and preparation

Pollen samples were collected from *Cedrus atlantica* trees from nine locations (Table 1) across the Middle Atlas, Morocco ($n = 72$), with additional samples collected from sites in Spain, France, UK and USA ($n = 19$). Samples were collected between September and October 2015, apart from the UK samples which were collected in 2014. Multiple strobili from each tree were collected and placed in paper envelopes on site, and freeze-dried back in the laboratory. Grains were extracted by vigorous shaking of the strobili, and collecting in sieves. Non-pollen contaminants were removed by visual inspection, and grains were stored in glass vials at 4 °C.

Measurements were carried out on untreated samples using both high-power transmitted LM and laser diffraction granulometry, with additional measurements under LM on samples treated with 10% potassium hydroxide (KOH) in a water bath at 90 °C for 15 minutes.

2.2. Light microscopy

Pollen was mounted on glass slides with silicone oil following dehydration with ethanol and tert-butyl alcohol. Two hundred and forty grains from eight samples (30 grains per sample) were measured with a Zeiss Axioscope A1 microscope and Zeiss Axiocam ERC5S camera using AxioVision 4 software. Grains in equatorial view were selected randomly and measured live on-screen following microscope calibration at 400× magnification.

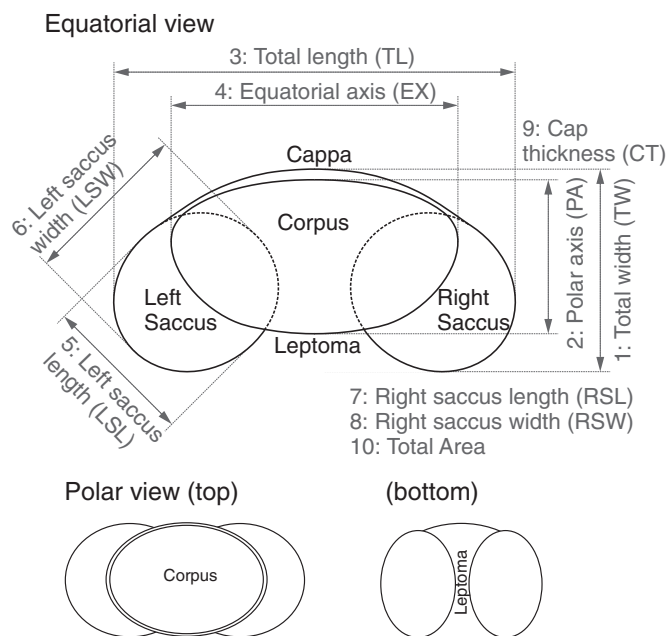


Figure 2. Schematic diagram of *Cedrus atlantica* pollen as it appears under light microscopy (LM), and the different measurements taken under LM.

Ten different properties of each grain were measured (Figure 2), adapted from Nakagawa et al. (1996) and Tiwari et al. (2012). Pollen terminology follows Erdtman (1943) and Punt et al. (2007).

2.3. Scanning electron microscopy

Scanning electron microscopy (SEM) images were obtained using a 120-kV FEI Tecnai 12 Twin Transmission Electron Microscope in The University of Manchester Life Sciences Faculty. Untreated and treated pollen samples were dehydrated in graded ethanol stages, then dried with Hexamethyldisilazane (HMDS) following (Chisoe et al. 1994). Grains were stuck to spurs with doubled-sided tape and sputter-coated with gold palladium for 5 minutes prior to SEM.

Table 1. Details of the sampling areas where pollen was collected.

Location ¹	No. of samples	Longitude	Latitude	Altitude (m asl) ²	Mean annual precipitation (mm) ³	Temperature range (°C) ⁴
Ifrane	2	−5.11	33.43	1653	474 (836)	−1.8 to 28.0
Michliffen	15	−5.08	33.34	1940	516 (714)	−3.0 to 28.5
Ifrane National Park	6	−5.17	33.39	1723	516 (747)	−1.7 to 29.8
Col Du Zad	7	−5.07	33.07	2106	470 (515)	−3.8 to 29.6
Lake Sidi Ali	10	−4.99	33.08	2150	470 (408)	−3.3 to 29.2
Bikrit	9	−5.26	33.24	1611	516 (738)	−1.0 to 30.5
Kobbat	11	−5.21	33.19	2074	516 (667)	−3.8 to 27.7
Ain Kahla	6	−5.23	33.30	1939	516 (695)	−3.0 to 28.5
(near) Azrou	6	−5.24	33.35	1814	516 (696)	−2.2 to 29.3
Westonbirt, UK	5	−2.21	51.61	132	854	0.9 to 20.7
Manchester, UK	3	−2.21	53.41	43	890	1.4 to 19.6
Boston, USA	2	−71.12	42.30	45	1236	−9.2 to 27.1
Paris, France	4	2.36	48.84	35	618	1.8 to 24.8
Bordeaux, France	3	−0.60	44.85	23	958	2.4 to 25.2
Pyrenees, Spain	2	−0.55	42.57	820	753	−1.9 to 26.0

¹Location within Morocco unless otherwise indicated.

²Average altitude (metres above sea level [asl]) of samples collected in the location.

³CRU (East Anglia Climate Research Unit) data averaged over 30 years (1986–2015). Values in parentheses indicate interpolated precipitation values (Bell et al. 2017).

⁴Mean annual minimums and maximums.

2.4. Laser diffraction particle size analysis

Untreated pollen samples were measured using a Malvern Mastersizer 2000, fitted with a Hydro 2000 μ P liquid sample dispersion unit for small particles (e.g. Sperazza et al. 2004) in The University of Manchester Geography laboratories. The machine was calibrated prior to measurements on pollen, using a spherical glass bead standard supplied by the manufacturer. The system was configured using the following method: Sample material was characterised as sporopollenin with a refractive index of 1.475 (Traverse 2007). Dispersant used was water with a refractive index set to 1.33 (Hecht 2002). Sample measurement time and background measurement time were set to 30 seconds. Pump speed was set to 2000 rpm, and ultrasonic treatment set to 75% with a pre-measurement period and delay of 30 seconds. Measurement repeats were set to three per aliquot with a 10-second delay. Using this method, the dispersion unit (Hydro 2000 μ P) was filled with deionised water using the anaerobic fill option (to remove air bubbles, which would otherwise affect the result). Once full, the pollen sample was added to the dispersion unit to reach a laser obscuration which ideally fell between 10 and 20% (this is shown on screen as the sample is added). Once the desired laser obscuration was reached, the measurement cycle was run. On completion of measurements, the system was drained, and flushed with deionised water 3 times to remove the sample. The sample can be recovered at this stage if required.

Particle size distribution of the sample was computed from the diffraction measurements using a model based on Fraunhofer diffraction theory (Syvitski 1991). The resulting analysis is reported as the relative distribution of the volume (%) of pollen grains by size (μ m) (Figure 3). Each grain measurement is placed within a size class (which can be defined by the user), and the distribution therefore shows the percentage of grains within a given size range. For example, in Figure 3, 16.8% of the individual grains measured within the entire sample were between 56 and 63 μ m in size. The D_{10} value represents the

size of the grains below which 10% of the sample lies (i.e. 10% of the individual grains measured in the sample are smaller than the D_{10} value), D_{90} is the size of the grains below which 90% of the sample lies, and D_{50} is the median of the particle size distribution. The D_{50} value is the measurement we use to describe the size of each pollen sample.

All laser particle size distribution models (Fraunhofer and Mie theory) assume the analyte particles are spherical (Syvitski 1991), and the machine will record a measurement of a particle in whichever orientation it passes through the laser. As *Cedrus atlantica* pollen grains are not completely spherical, and they may pass the laser in different orientations, the model will in practice provide a weighted average of the diameter of the pollen grain. Since the pollen morphology of *Cedrus atlantica* is consistent, an empirical relationship can be derived between the particle size measurement and LM measurements. For *Cedrus atlantica* pollen, the D_{50} value is effectively equivalent to the size of the equatorial axis (EX) as measured under LM (see Results section 3.2.2.).

2.5. Data processing and analysis

Data and statistical analyses were carried out using R (R Core Team 2016). Climate data was extracted from CRU TS v3.24.01 high-resolution (0.5°) gridded datasets (Harris et al. 2014). Precipitation for Middle Atlas locations was interpolated using local climate stations described in Bell et al. (2017). Aridity data was extracted from the self-calibrating Palmer Drought Severity Index (scPDSI) (Dai 2011).

3. Results

3.1. Description of grains (Plates 1 to 4)

Cedrus atlantica pollen are large diploxylonoid-type bisaccate grains. The grains are elongated, the corpus is typically prolate, and the cap wall is thick. Sacci are spheroidal in equatorial view, appearing more oblate in polar view. The junction of the sacci and corpus lacks a strong defining ridge, with the two parts appearing seamlessly together. Under LM the surface ornamentation of the corpus is a rough, reticulate pattern, while ornamentation is somewhat smoother on the sacci. Under SEM, surface ornamentation of the corpus is rough, appearing fossulate, with groups of irregular, spheroidal to elongated elements protruding, interspaced by deep grooves. The sacci appear smooth with a scabrate-perforate surface. Surface ornamentation appears more defined following chemical treatment under both LM and SEM.

3.2. Grain size

3.2.1. Light microscope measurements

LM measurements of grain size on eight untreated pollen samples, with 240 individual grains measured (Table 2), found the average total grain size including sacci and corpus (TL) was $71.4 \pm 7.6 \mu$ m, while the EX measured an average $52.2 \pm 4.0 \mu$ m. There was large variation in the size of grains observed: For TL, there was a 40- μ m difference between the smallest and largest grains, and for EX, there was a 20- μ m difference. Full LM

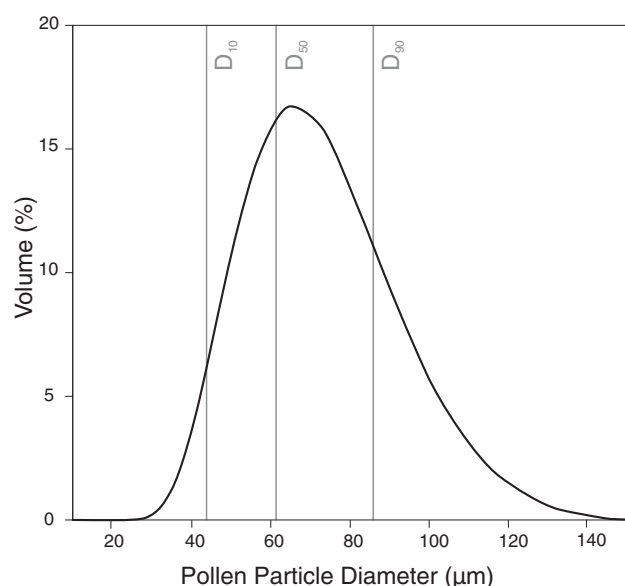


Figure 3. Results from laser diffraction granulometry. Size data for each sample is presented as the particle size distribution of the sample.

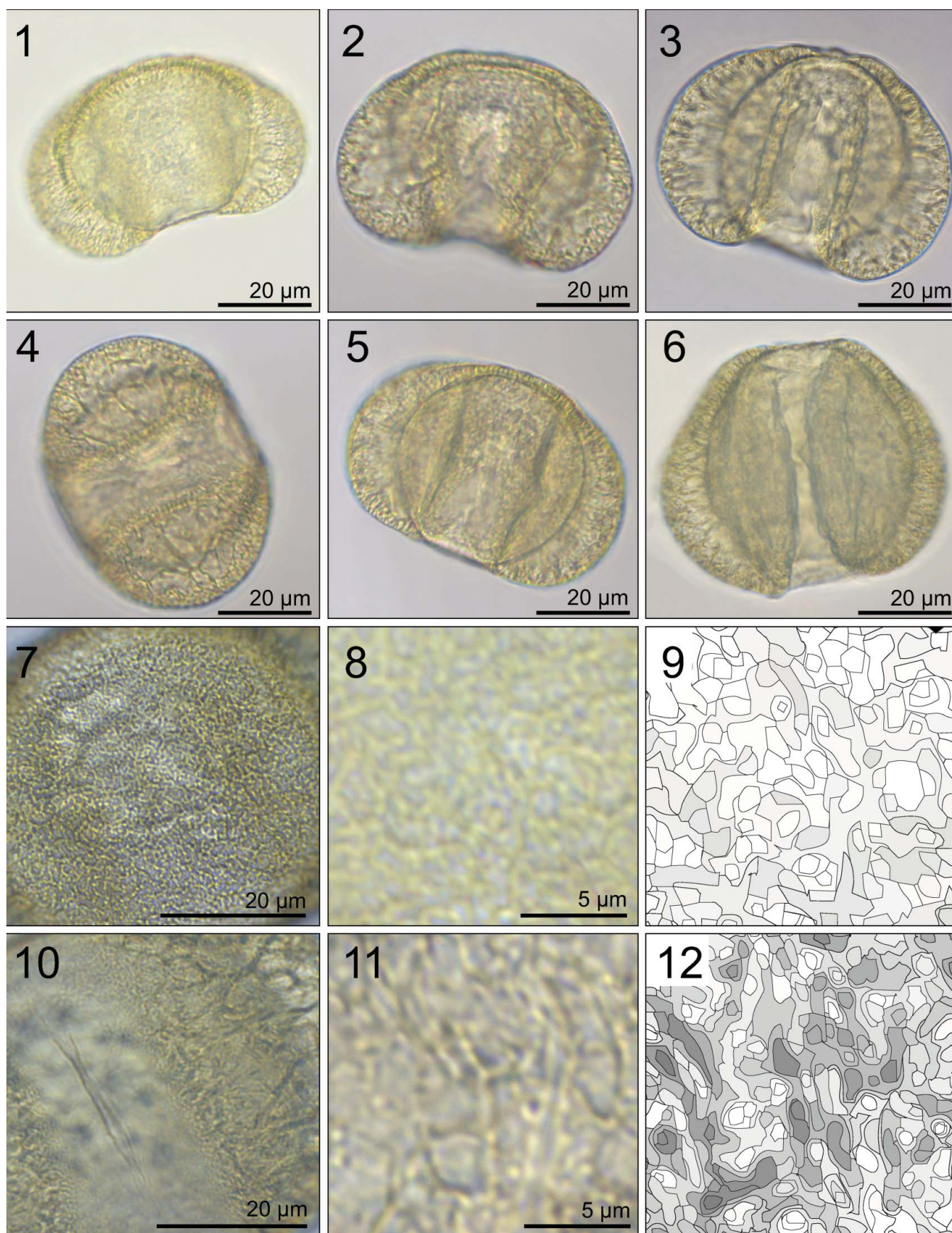


Plate 1. Photographs taken under light microscopy (LM) of *Cedrus atlantica* pollen grains. Scale indicated on each image. Figures 1–3. Equatorial view showing complete grain. Figures 4–5. Polar view of complete grain. Figures 7–8. Close-up of corpus surface. Figure 9. Corpus surface pattern (extracted using CorelDraw X8). Figures 10–11. Close-up of saccus surface. Figure 12. Saccus surface pattern.

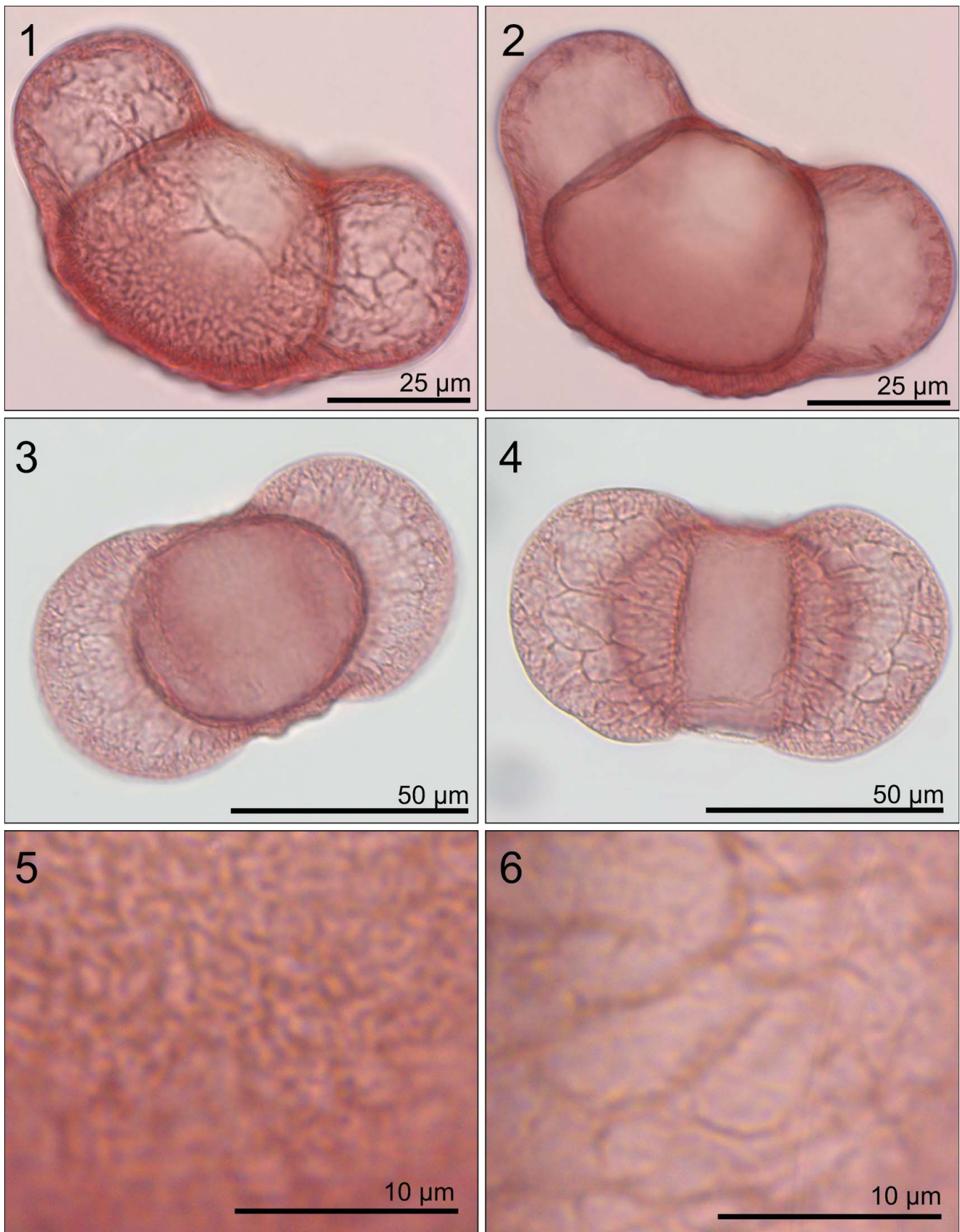


Plate 2. Photographs taken under light microscopy (LM) of treated *Cedrus atlantica* pollen grains stained with Safranin. Scale indicated on each image. Figures 1–2. Equatorial view showing complete grain at different foci. Figures 3–4. Polar view of complete grain. Figure 5. Close-up of corpus surface. Figure 6. Close-up of saccus surface.

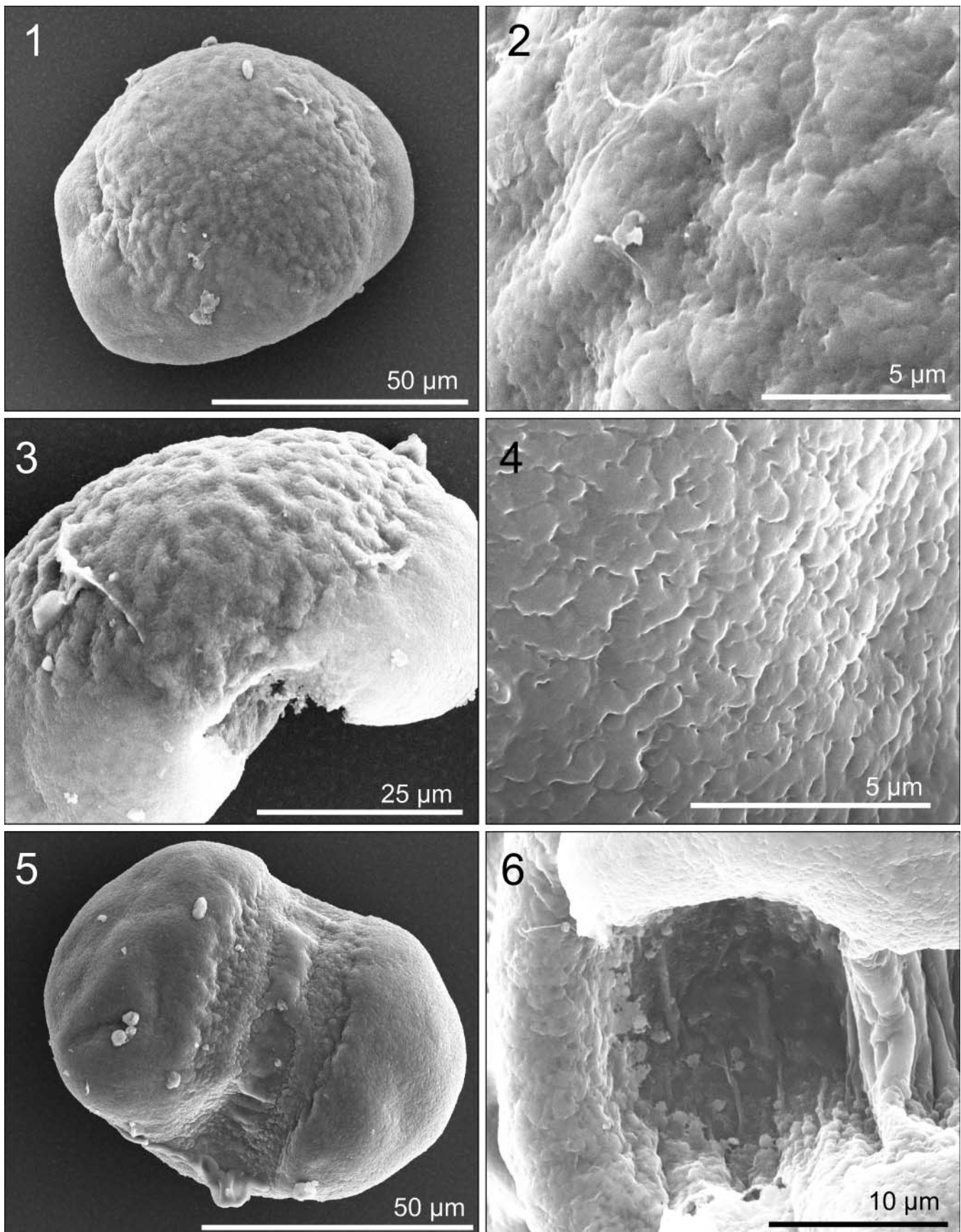


Plate 3. Scanning electron microscopy (SEM) images of *Cedrus atlantica* pollen grains. Scale indicated on each image. Figure 1. Polar view from the top. Figure 2. Close-up of corpus surface. Figure 3. Equatorial view. Figure 4. Close-up of saccus surface. Figure 5. Polar view from underneath the grain showing the leptoma. Figure 6. Close-up of leptoma, pollen wall visible to the left.

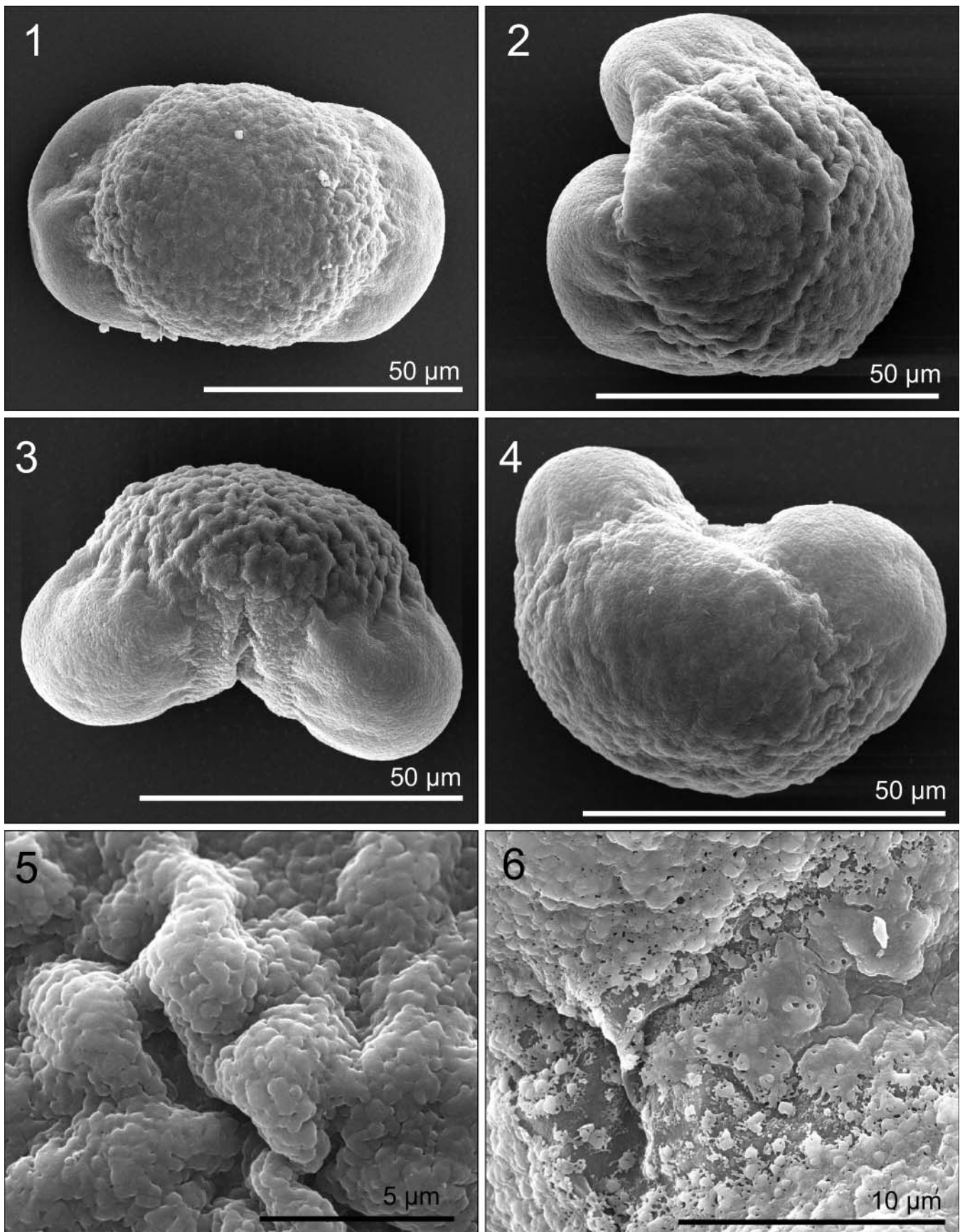


Plate 4. Scanning electron microscopy (SEM) images of treated *Cedrus atlantica* pollen grains. Scale indicated on each image. Figures 1–2. Polar view from the top and side. Figures 3–4. Equatorial view. Figure 5. Close-up of corpus surface. Figure 6. Close-up of leptoma.

Table 2. Grain size of each measured property (please refer to Figure 2) under light microscopy (LM; eight samples, 240 grains measured).

	1: TW (μm)	2: PA (μm)	3: TL (μm)	4: EX (μm)	5: LSL (μm)	6: LSW (μm)	7: RSL (μm)	8: RSW (μm)	9: CT (μm)	10: area (μm^2)	P/E ratio (EX/PA)
Mean	53.6	38.8	71.4	52.2	31.8	28.6	31.3	27.9	3.1	2774.9	1.4
Standard deviation	4.8	5.5	7.6	4.0	3.9	3.6	3.9	3.4	0.7	472.6	0.2
Minimum	35.5	25.5	49.8	43.8	19.1	20.1	21.5	19.7	1.4	1580.5	0.9
Maximum	68.0	54.0	89.1	63.8	42.5	39.7	43.0	37.3	5.2	4180.9	1.9

Table 3. Percentage change (%) in size for different properties (please refer to Figure 2) of the grain measured under light microscopy (LM) following chemical treatment with KOH.

1: TW	2: PA	3: TL	4: EX	5: LSL	6: LSW	7: RSL	8: RSW	9: CT	10: area	P/E ratio (EX/PA)
−2.8	8.5	9.3	−1.5	4.1	12.5	5.9	16.2	9.0	7.6	−7.1

measurement data can be found in the online Supplementary material.

Treated pollen samples measured under LM were on average 6.9% larger in size overall compared to untreated grains (Table 3). However, the increase in size was not consistent across all the measured properties of the grain, with the largest increases to the sacci width and cap thickness (CT), while the total grain width (TW) and EX decreased in size. This resulted in the shape of the grain changing slightly, becoming subprolate.

3.2.2. Laser diffraction particle size analysis

Pollen grain size measurements taken using laser diffraction granulometry on 91 untreated pollen samples, where millions of individual grains were measured, recorded an average grain size of $59.1 \pm 4.0 \mu\text{m}$. Particle size distribution data for each sample can be found in the online Supplementary material.

The reliability of the laser diffraction particle size measurements was tested by correlation analysis with the LM measurements (Figure 2) for the samples measured using both methods (Table 4). The strongest and most significant correlation was found between the D_{50} median size value (laser diffraction) and the LM measurement for the EX ($r = 0.91$, $p = 0.002$). During the measurement cycle for laser diffraction granulometry, pollen grains float freely in water as they pass the laser beam and are not subject to external pressure (e.g. from a cover-slip); consequently, the 'natural shape' of the grain is measured. In *Cedrus atlantica* pollen, the sacci lie underneath the corpus, protruding a short distance; effectively, the total length of the grain is slightly bigger than the equatorial axis of the corpus. Under LM, the protrusion of the sacci can appear to be greatly inflated (particularly in equatorial view) resulting in the total length of the grain appearing larger. Consequently, the best correlation with the grain size reported by laser diffraction granulometry for *Cedrus atlantica* pollen is with the equatorial axis as measured under a light microscope.

Table 4. Correlation analysis between the laser diffraction granulometry D_{50} grain size value, and light microscopy (LM) measurements.

Grain property	Pearson's r	p
Total width (TW)	0.68	0.062
Polar axis (PA)	0.39	0.345
Total length (TL)	0.66	0.076
Equatorial axis (EX)	0.91	0.002
Total area	0.67	0.069

3.3. Grain size variability

The eight samples measured under LM show large variability in grain size (in all measured properties) between individual grains within the same sample (Table 2). Between samples, analysis of variability (ANOVA) of the equatorial axis (measured under LM) suggests this is significant ($df = 7$, $F = 14.97$, $p < 0.0001$); however, Tukey's test reveals that six of these samples do not vary significantly in size from each other.

Laser diffraction granulometry results likewise show large grain size variability between grains within the same individual sample. Samples from the same geographical area also show variability in grain size between samples, and, lastly, there is variability of grain size between sampling areas (Figure 4). Pollen samples within the same geographical area had an average grain size range of $5.7 \mu\text{m}$. The largest range was in the Pyrenees ($12.2 \mu\text{m}$), followed by Michliffen ($9.2 \mu\text{m}$), and the smallest range was found within Paris samples ($2.0 \mu\text{m}$). However, ANOVA shows that the variation in grain size between different sampling areas is not significant ($df = 14$, $F = 1.054$, $p = 0.412$).

3.3.1. Climatic controls on grain size variability

Regression analysis was performed on pollen grain size and possible influences including climate (temperature, precipitation, aridity and potential evapotranspiration), altitude, and carbon isotope discrimination values, which are an indicator of environmental moisture availability (Bell et al. 2017). Given that samples from within the same geographical area will experience the same climate conditions, and fossil pollen assemblages will comprise grains from several trees within the surrounding area (Bell & Fletcher 2016), analysis was performed testing the average grain size for each geographical sampling area, as well as individual sample grain size values (Table 5).

The regression analysis found no significant relationships with any of the climate variables or altitude to suggest a link to grain size. Similarly, multiple-regression analysis testing every possible combination of climate predictors using all subsets regression found no significant relationships or models to support a climate influence on grain size. Furthermore, there was no significant relationship with stable carbon isotope discrimination when samples were averaged by geographical location ($r^2 = 0.10$, $p = 0.234$) although there was a very weak significant association with individual samples ($r^2 = 0.05$, $p = 0.029$). However, 10-fold cross-validation of this model found the r^2 value could be as low as 0.003.

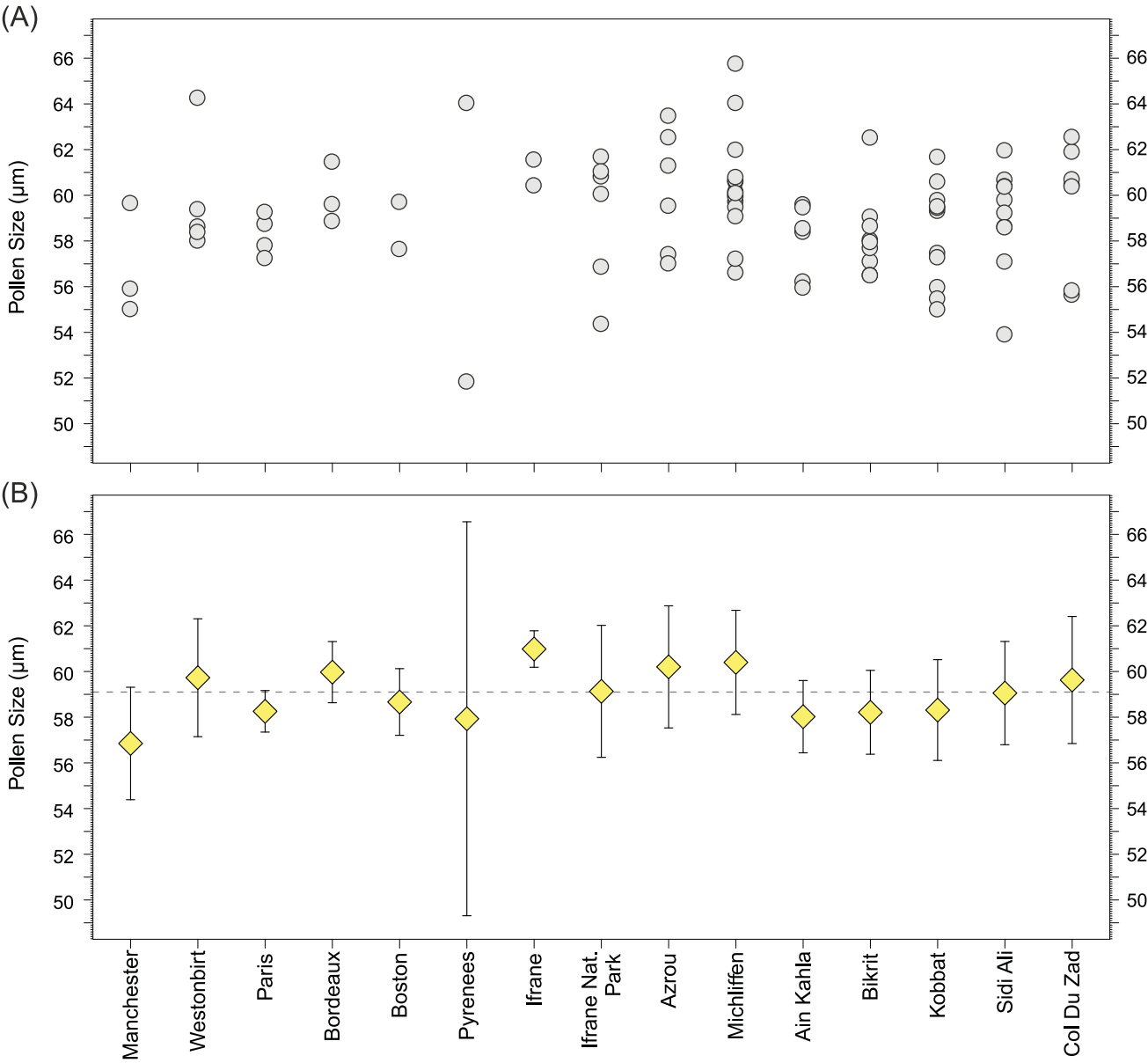


Figure 4. Results of laser diffraction granulometry showing: (A) Dot plot of D₅₀ values for each sample by geographical sampling area, and (B) mean D₅₀ values for each area. Error bars show standard deviation, and dashed horizontal line represents the mean grain size of all samples.

Table 5. Results of regression analysis on pollen grain size. Analysis was tested using individual sample grain size values, and the average grain size for each geographical sampling area.

Variable	Data source	Individual sample		Location average	
		r ²	p	r ²	p
Summer precipitation (2015) ¹	CRU TS v3.24.01	0.00	0.812	0.03	0.515
Mean annual precipitation (2015) ²	CRU TS v3.24.01	0.00	0.441	0.10	0.246
Mean annual precipitation (30-year average) ³	CRU TS v3.24.01	0.00	0.509	0.07	0.331
Mean annual precipitation (interpolated values)	Bell et al. (2017)	0.00	0.890	0.00	0.789
Mean annual temperature (2015)	CRU TS v3.24.01	0.00	0.747	0.06	0.380
Mean summer temperature (2015)	CRU TS v3.24.01	0.00	0.598	0.07	0.319
Mean annual temperature (30-year average)	CRU TS v3.24.01	0.00	0.779	0.06	0.375
Mean summer temperature (30-year average)	CRU TS v3.24.01	0.00	0.628	0.06	0.352
Aridity (scPDSI) ⁴	Dai (2011)	0.00	0.586	0.07	0.324
Annual potential evapotranspiration (PET) (2015)	CRU TS v3.24.01	0.00	0.396	0.01	0.396
Summer PET (2015)	CRU TS v3.24.01	0.01	0.551	0.00	0.551
Carbon isotope discrimination (Δ ¹³ C) ⁵	Bell et al. (2017)	0.05	0.029	0.10	0.234
Altitude	This study	0.00	0.439	0.07	0.316

¹Summer corresponds to the development period for *Cedrus atlantica* pollen.
²Values for the year of pollen collection.
³Values based on a 30-year average between 1986–2015.
⁴Aridity values from self-calibrating Palmer Drought Severity Index (30-year average).
⁵Carbon isotope discrimination calculated from δ¹³C values measured directly on the same pollen samples.

4. Discussion

4.1. Influences on pollen grain size variability

Our study has found no relationship between climate and the grain size of modern *Cedrus atlantica* pollen. This is supported by the lack of relationship between grain size and $\Delta^{13}\text{C}$ when grain size was averaged by geographic location ($r^2 = 0.10$, $p = 0.234$), where the $\Delta^{13}\text{C}$ value of the same pollen samples were previously shown to be significantly affected by moisture availability (Bell et al. 2017). However, there was a very weak association between individual sample grain size measurements and $\Delta^{13}\text{C}$ ($r^2 = 0.05$, $p = 0.029$). Further cross-validation of this model shows that less than 5% (in some cases, as little as 0.3%) of the variance in grain size is explained by carbon isotope discrimination. As no relationship was found between $\Delta^{13}\text{C}$ and grain size averaged by geographical area, this suggests that factors other than climate must influence grain size, as these samples would experience the same climate conditions. Although there may be micro-scale climate variations in these areas (Bell et al. 2017), variations in isotopic composition of samples from the same area may also be associated with the intrinsic water-use efficiency of the tree (Körner et al. 1991). Accordingly, the results suggest that while samples have a strong geochemical climate signature, there is no equivalent strong morphological climate signature (Figure 5).

In another study, summer moisture availability was also ruled out as influencing pollen grain size on *Cedrus atlantica* populations in Algeria. Analysis showed that pollen grain size discriminated trees into different genetic and ecological groups, suggesting genetics influenced pollen size (Derridj et al. 1991). The genetic control on pollen size may stem from genome size (Bennett 1972; De Storme et al. 2013), in particular the effect of an increased number of chromosomes due to polyploidy (Kapadia & Gould 1964; Dyer et al. 2013). Knight et al. (2010) found a significant positive trend between pollen width and genome size across 464 species. However, the trend was not significant when phylogeny was taken into account, suggesting that pollen size would not be a good proxy for genome size in the fossil record. The phylogenetic relationships among *Cedrus* have been debated (Bou Dagher-Kharrat et al. 2001), but DNA evidence suggests *Cedrus atlantica* separated from a common ancestor of *Cedrus libani* and *Cedrus brevifolia* around 23 to 18 Ma BP (Qiao et al. 2007), and DNA analysis of all four *Cedrus* (the 'true' cedars) species (including *Cedrus deodara*) found that

genome size is homogeneous among the species (Bou Dagher-Kharrat et al. 2001). Genetic diversity of *Cedrus atlantica* trees from Morocco was shown to be high within populations and between populations (Renau-Morata 2005; Terrab et al. 2006), particularly between populations from the Rif, and High Atlas, when compared to Middle Atlas populations (Cheddadi et al. 2009). It is possible that the variations in grain size we observe between samples could relate to genome size; however, further research is needed to confirm this in *Cedrus atlantica* pollen.

The influence of temperature on grain size as demonstrated by Ejsmond et al. (2015) was suggested to relate to pollen performance, whereby a trade-off exists between the size of the grain and the quantity of grains produced (Vonhof & Harder 1995). The competitive ability of pollen increases with temperature during the flowering period, and this increased competition promotes larger pollen grains (Ejsmond et al. 2015). The lack of any temperature influence on *Cedrus atlantica* pollen size may be due to it being a wind-pollinating species. The apparent trade-off between pollen size and quantity may not be necessary, as priority is on the quantity of grains produced in order to increase the chances of successful pollination (Whitehead 1983; Cruden 2000). Larger grains are heavier, giving increased chance of reaching ovules as they can more easily break from the airstream, while smaller, lighter grains can travel greater distances (Niklas 1985). Pollen size in wind-pollinated species consequently reflects an equilibrium between these demands (Lu et al. 2011). Large grain size variation within individual samples could therefore possibly reflect an adaptation to facilitate both demands. The observed grain size variation also implies that smaller grains found within fossil assemblages from geological archives might be an indicator of long-distance pollen transport, with larger grains indicating nearby pollen sources rather than reflecting a climate signal.

Smith (1923) noted 'irregularity' with the development of *Cedrus atlantica* pollen, where newly forming grains exist alongside mature grains, and long retention of grains after they reached maturity. Strobili typically form 2–3 months prior to pollen release, with grains developing in the later parts of this period. Pollen release is a phenological response lasting a few days, with the timing varying between individual trees and between geographical locations, depending on optimal environmental conditions including temperature, humidity levels and wind speed (Whitehead 1983; Khanduri & Sharma 2009).

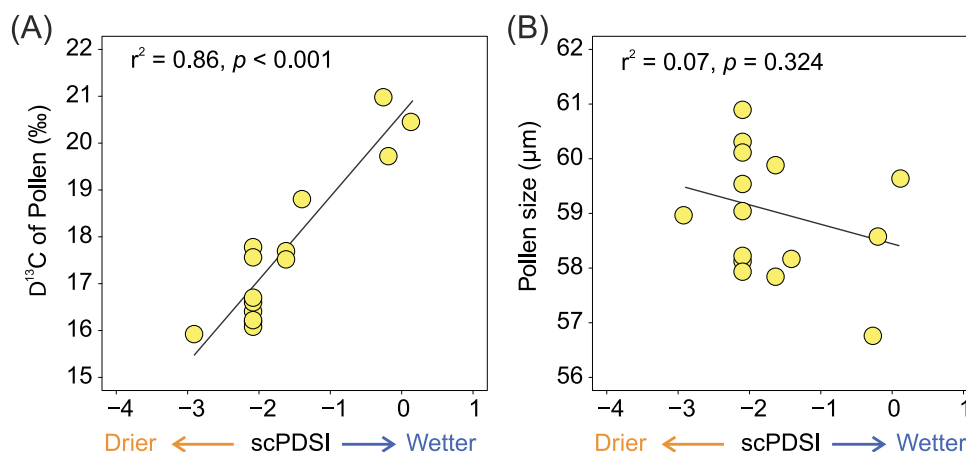


Figure 5. Biplots showing: (A) carbon isotope discrimination ($\Delta^{13}\text{C}$) (Bell et al. 2017) versus aridity (self-calibrating Palmer Drought Severity Index, scPDSI) (Dai 2011), and (B) pollen grain size versus aridity (scPDSI), with values averaged by each sampling area.

For *Cedrus atlantica* this typically occurs from early to late September and early October. This irregular development of grains may also contribute to the size variation observed, if pollen release occurs while some grains are still developing and others are fully developed.

Soil nutrient availability has also been linked to pollen grain size and production, in squash (*Cucurbita pepo*) plants. Grain size and the number of grains produced both increased where plants were in soil which had higher nutrient contents; this effect was greatest with increased nitrogen (Lau & Stephenson 1993), but also evident with increased phosphorus (Lau & Stephenson 1994). If soil nutrient availability influenced grain size for *Cedrus atlantica* pollen, then we might expect to see larger grains in the botanical garden sites and smaller grains in the relatively nutrient-poor Middle Atlas locations. However, grain size is smaller than average at two of the three botanical garden sites (Westonbirt, Paris and Boston), while grain size is larger than the average at the most southerly Middle Atlas site (Col Du Zad), an environment characterised by sparse open forest and semi-arid conditions where nutrient availability is poor. This suggests that nutrient availability does not influence grain size for *Cedrus atlantica* pollen.

Overall, the large variability in pollen grain size we observe within individual samples, in some cases by as much as 20 μm , suggests that grain size is influenced by a number of complex factors. The grain size variability is also not unexpected, and is well documented in other pollen (e.g. Bell 1959; Clausen 1962; Bragg 1969; Cruzan 1990; Desprat et al. 2015). Consequently, we propose that due to the size variability in *Cedrus atlantica* pollen, and lack of evidence for climatic influence, it would not be possible to use this as a proxy for climate or environmental reconstruction, as differences in the size of fossil pollen may simply result from the observed natural variation in size between pollen grains. Our findings contrast the suggested climatic influence on grain size observed in other species (Ejsmond et al. 2011 2015; Griener & Warny 2015), and underscore the need for further investigation of the complex controls on pollen grain size (Jardine & Lomax 2017).

4.2. Methodological approaches to grain size measurements

We demonstrate the importance of a consistent methodological approach to grain size measurement, through the differences in grain size reported in the literature, as a result of the technique used, and between untreated and chemically treated pollen grains. It is notable that the size changes we observe in *Cedrus atlantica* pollen grains treated with KOH are not uniform across the entire grain, suggesting that morphological differences in pollen have different susceptibilities to chemical treatment. For example, Reitsma (1969) found the size-altering effects of KOH treatment depended on pollen type and exposure time to the treatment, while effects on pollen size from chemical pre-treatment have also been noted in Faegri & Deuse (1960), Praglowski (1970) and Charman (1992). In *Cedrus atlantica* the largest size increase in the pollen grain occurred in the sacci, which have a perforate surface in contrast to the rest of the grain. The small apertures allow the pollen to quickly dehydrate following pollen release (Tekleva et al. 2007), reducing its

weight to assist long-distance transport (Lu et al. 2011). In a reversal of this process, the apertures may allow greater penetration of chemical treatment into the sacci compared to other parts of the grain, and may explain why they exhibited a greater increase in size. This suggests that morphological variations within pollen grains and between pollen types are affected by treatments in different ways, so relationships of grain size should always be established to specific pollen types following the same preparation and methodological protocols.

Fossil pollen grain size may also be affected by diagenesis. Although potential size-altering effects are not fully known, they are likely to differ depending on species and sedimentary setting (Mäkelä 1996), with resistance to these effects likely stemming from morphological traits of the pollen grains. Due to these effects, it may be difficult to compare grain size between fossil pollen from different sedimentary settings, implying that apparent grain size changes in a fossil sequence could result from changes to the sedimentary setting, unless it remains homogeneous throughout. Consequently, pollen grain size relationships and comparisons between pollen from a fossil setting and modern pollen samples may not be straightforward (Mäkelä 1996). Indeed, before comparison, the effects of diagenesis and sedimentary setting on fossil pollen size should first be established. Overall, all possible influences on grain size should be considered in the interpretation of results (Desprat et al. 2015), and a consistent methodological approach to measurements should be taken.

4.3. Practical considerations for laser diffraction particle size analysis of pollen

We have shown that laser diffraction granulometry provides a reliable and consistent method of determining pollen grain size, and produces results in line with existing LM methods of grain size measurement. The main benefit of laser diffraction granulometry is that it can measure thousands to millions of pollen grains (dependent on sample size) in just a few minutes, providing a very robust assessment of the typical grain size (here, D_{50} or median) which can be repeated on multiple samples quickly. The method is also non-destructive, allowing recovery of the sample after the measurement cycle. Measurement of grains is not affected by operator subjectivity or potential inaccuracies between measurements, nor is it affected by size changes caused by mounting medium or cover-slip pressure on the grain, which can affect LM measurements. Provided the morphology of the pollen grains is consistent, an empirical relationship between laser diffraction granulometry measurements and light microscope measurements can be established. In additional testing of the method, we found similar results with *Pinus* pollen between the grain size reported by laser diffraction granulometry and LM measurements; however, further testing of the method is needed on other pollen types with different morphologies, and pollen types with smaller grains.

The primary use of laser diffraction granulometry for pollen size determination would be on modern samples, due to the relatively large quantity of sample material required. There is a range of possible applications, including investigating the influence of climate and environmental factors, nutrient availability, and genome size. In testing, approximately 15 mg of pollen

was used to reach 10% laser obscuration, which could easily equate to millions of grains (dependent on the grain size/weight of the specific pollen type), based on estimates of pollen weight. For example, Brown & Irving (1973) weighed several pollen types, including *Quercus robur*, suggesting 93 grains weigh 1 μg , or that there are 1,395,000 grains per 15 mg. There are reportedly 404 maize grains per 1 μg , or 6,072,874 grains per 15 mg (Miller 1982). Bunderson & Levetin (2015) weighed different *Juniperus* species where approximately 273 grains weigh 1 μg , or there are 4,109,589 grains per 15 mg (averaged across species at 60% relative humidity).

We found that it is possible to use a reduced sample size, where laser obscuration only reached 0.4%, which produced grain size results in line with those at 10% laser obscuration. This equates to approximately 0.6 mg of pollen material required, which could equate to several thousand grains. Though perhaps this is still out of reach for measurements on fossil samples, it does allow measurements to be taken on modern pollen for a range of species, from small insect-pollinated flowers to large wind-pollinated trees.

5. Conclusions

Our study finds that while there is large variability in the pollen size of *Cedrus atlantica*, it is not significant between samples. We found no significant relationships between climate and grain size, testing temperature, precipitation, aridity and potential evapotranspiration (PET). Grain size of *Cedrus atlantica* may be influenced by a number of complex factors, and variability within individual samples may result from the irregular development of pollen. Our study confirms that a consistent methodological approach to grain size measurement must be taken, as there are many aspects of pollen handling and preparation stages that may influence the observed size. Size comparisons between values reported in literature, between different species/pollen types, and from different sedimentary settings may not always be possible or advisable. We have also shown a method utilising laser diffraction granulometry which can be used on modern pollen samples to accurately determine pollen size, and which produces results consistent with LM measurements. This non-destructive method of determining pollen size provides reproducible, robust results, from potentially millions of pollen grains, quickly and easily.

Acknowledgements

Fieldwork was carried out with assistance from the Haut Commissariat aux Eaux et Forêts et à la Lutte Contre la Désertification, Morocco, with thanks to Said Hajib. The authors thank Jenifer Campbell, Rachel Hurley, John Moore and Jonathan Yarwood (University of Manchester) for field and laboratory assistance. We thank Tobias Starborg (University of Manchester) for assistance with SEM imagery. Additional pollen samples were kindly supplied by Penny Jones (Westonbirt Arboretum), Martin Gardner (Royal Botanical Gardens, Edinburgh), Kathryn Richardson (Arnold Arboretum, Harvard University), Stéphanie Desprat (EPHE, Bordeaux), Daniel Gómez (Instituto Pirenaico de Ecología, Spain) and the Paris Botanical Gardens. Lastly, we thank Phil Jardine and an anonymous reviewer for their constructive comments and useful discussion to improve the final manuscript.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was supported by an Engineering Physical Sciences Research Council (EPSRC) studentship award [grant number 1478466].

Supplementary materials

Explanatory notes for the supplementary data

Cedrus atlantica pollen size measurement data under light microscopy.

Cedrus atlantica particle size distribution data from laser diffraction granulometry.

Notes on contributors



BENJAMIN A. BELL is a PhD student at The University of Manchester whose work focuses on the application of geochemical techniques to palynology and quaternary science, with a particular interest in *Cedrus atlantica*, its role within the environment and how it adapts to climate change in Northwest Africa.



THOMAS H. BISHOP is a research and teaching technician within the Geography Laboratories at The University of Manchester. He specialises in analytical techniques for palaeoenvironmental research, including chemical, physical and biological methods. His PhD research project was a palaeolimnological investigation of central Patagonia focussing on the Holocene epoch, which he completed at the University of Southampton. His current research interests are in developing novel techniques for collecting and manipulating environmental data.



WILLIAM J. FLETCHER is a palynologist whose specialist area of interest is the detection and characterisation of abrupt environmental and climatic changes in the Mediterranean region using vegetation records from Quaternary terrestrial and marine sediment archives. He obtained his PhD at the University of Cambridge and is a senior lecturer in physical geography and quaternary science at The University of Manchester.



PETER RYAN is a palynologist at The University of Manchester and is currently a teaching fellow in geography. His work focuses on the environmental impact of prehistoric people through the use of palaeoecology.



RACHID ILMEN is a professor in the Department of Hydraulic, Environment and Climate (HEC) at Hassania School of Public Works (EHTP)-Casablanca, Morocco. He received an engineer diploma from the National School of Forest Engineers, Morocco (2004), an International Certificate on Environmental Education from Shiga University, Japan (2010), a master degree on biological sciences (2009) and a PhD in climatology and climate change (2014)

from the Mohammed V University, Morocco. He specialises on water, environment and sustainable development and is an author and co-author of more than 30 papers.

ORCID

Benjamin A. Bell  <http://orcid.org/0000-0002-0899-9280>
 Thomas H. Bishop  <http://orcid.org/0000-0001-8253-9293>
 William J. Fletcher  <http://orcid.org/0000-0000-0001-8918-0690>
 Peter Ryan  <http://orcid.org/0000-0003-3841-7736>
 Rachid Ilmen  <http://orcid.org/0000-0003-2174-8301>

References

- Altuner EM, Çeter T, Alpas H. 2012. High hydrostatic pressure processing: a method having high success potential in pollen protein extraction. *High Pressure Research* 32:291–298.
- Amundson R, Evett RR, Jahren AH, Bartolome J. 1997. Stable carbon isotope composition of Poaceae pollen and its potential in paleovegetational reconstructions. *Review of Palaeobotany and Palynology* 99:17–24.
- Andersen ST. 1960. Silicone oil as a mounting medium for Pollen Grains. *Danmarks Geologiske undersøgelse* 4:1–24.
- Aytug B. 1960. Quelques mensurations des pollens de *Pinus silvestris* L. *Pollen et Spores* 2:305–309.
- Aytug B. 1961. Etude Des Pollens Du Genre Cedre (*Cedrus* Link.). *Pollen et spores* 3:47–54.
- Bell BA, Fletcher WJ. 2016. Modern surface pollen assemblages from the Middle and High Atlas, Morocco: insights into pollen representation and transport. *Grana* 55:286–301.
- Bell BA, Fletcher WJ, Ryan P, Grant H, Ilmen R. 2017. Stable carbon isotope analysis of *Cedrus atlantica* pollen as an indicator of moisture availability. *Review of Palaeobotany and Palynology* 244:128–139.
- Bell CR. 1959. Mineral nutrition and flower to flower pollen size variation. *American Journal of Botany* 46:621–624.
- Bennett MD. 1972. Nuclear DNA content and minimum generation time in herbaceous plants. *Proceedings of the Royal Society B: Biological Sciences* 181:109–135.
- Bou Dagher-Kharat M, Grenier G, Bariteau M, Brown S, Siljak-Yakovlev S, Savouré A. 2001. Karyotype analysis reveals interspecific differentiation in the genus *Cedrus* despite genome size and base composition constancy. *Theoretical and Applied Genetics* 103:846–854.
- Bragg LH. 1969. Pollen size variation in selected grass taxa. *Ecology* 50:124–127.
- Brown HM, Irving KR. 1973. The size and weight of common allergenic pollens. *Allergy* 28:132–137.
- Brown TA, Nelson ED, Mathewes RW, Vogel JS, Southon JR. 1989. Radiocarbon dating of pollen by accelerator mass spectrometry. *Quaternary Research* 32:205–212.
- Bunderson LD, Levetin E. 2015. Hygroscopic weight gain of pollen grains from *Juniperus* species. *International Journal of Biometeorology* 59:533–540.
- Charman DJ. 1992. The effects of acetylation on fossil *Pinus* pollen and Sphagnum spores discovered during routine pollen analysis. *Review of Palaeobotany and Palynology* 72:159–164.
- Cheddadi R, Fady B, François L, Hajar J, Suc JP, Huang K, Demarteau M, Vendramin GG, Ortu E. 2009. Putative glacial refugia of *Cedrus atlantica* deduced from Quaternary pollen records and modern genetic diversity. *Journal of Biogeography* 36:1361–1371.
- Clausen KE. 1962. Size variation in pollen of three taxa of *Betula*. *Pollen et Spores* 4:169–174.
- Chissoe WF, Vezey EL, Skvarla JJ. 1994. Hexamethyldisilazane as a drying agent for pollen scanning electron microscopy. *Biotechnic & Histochemistry* 69:192–198.
- Cruden RW. 2000. Pollen grains: Why so many? *Plant Systematics and Evolution* 222:143–165.
- Cruzan MB. 1990. Variation in pollen size, fertilization ability, and postfertilization siring ability in *erythronium grandiflorum*. *Evolution* 44:843–856.
- Cushing EJ. 1961. Size increase in pollen grains mounted in thin slides. *Pollen et Spores* 3:265–274.
- Dai A. 2011. Characteristics and trends in various forms of the palmer drought severity index during 1900–2008. *Journal of Geophysical Research Atmospheres* 116.
- De Storme N, Zamariola L, Mau M, Sharbel TF, Geelen D. 2013. Volume-based pollen size analysis: an advanced method to assess somatic and gametophytic ploidy in flowering plants. *Plant Reproduction* 26:65–81.
- Derridj A, Cadeac F, Durrieu G. 1991. Etude de la variabilité géographique des dimensions des pollens du cèdre de l'Atlas (*Cedrus atlantica* Manetti) en Algérie. *Bulletin de la Société Botanique de France. Lettres Botanique* 138:215–230.
- Desprat S, Diaz Fernandez PM, Coulon T, Ezzat L, Pessarossi-Langlois J, Gil L, Morales-Molino C, Sanchez Goni MF. 2015. *Pinus nigra* (European black pine) as the dominant species of the last glacial pinewoods in southwestern to central Iberia: A morphological study of modern and fossil pollen. *Journal of Biogeography* 42:1998–2009.
- Dyer RJ, Pellicer J, Savolainen V, Leitch IJ, Schneider H. 2013. Genome size expansion and the relationship between nuclear DNA content and spore size in the *Asplenium monanthes* fern complex (Aspleniaceae). *BMC Plant Biology* 13:219.
- Ejsmond MJ, Ejsmond A, Banasiak Ł, Karpińska-Kołaczek M, Kozłowski J, Kołaczek P. 2015. Large pollen at high temperature: an adaptation to increased competition on the stigma? *Plant Ecology* 216:1407–1417.
- Ejsmond MJ, Wrońska-Pilarek D, Ejsmond A, Dragosz-Kluska D, Karpińska-Kołaczek M, Kołaczek P, Kozłowski J. 2011. Does climate affect pollen morphology? Optimal size and shape of pollen grains under various desiccation intensity. *Ecosphere* 2:1–15.
- Erdtman G. 1943. An introduction to pollen analysis. Massachusetts: The Chronica Botanica Company.
- Faegri K, Deuse P. 1960. Size variations in pollen grains with different treatment. *Pollen et spores* 2:293–298.
- Farjon A. 1990. Pinaceae: drawings and descriptions of the genera *Abies*, *Cedrus*, *Pseudolarix*, *Keteleeria*, *Nothotsuga*, *Tsuga*, *Cathaya*, *Pseudotsuga*, *Larix* and *Picea*. Königstein: Koeltz Scientific Books.
- Fraser WT, Sephton MA, Watson JS, Self S, Lomax BH, James DI, Wellman CH, Callaghan TV, Beerling, DJ. 2011. UV-B absorbing pigments in spores: Biochemical responses to shade in a high-latitude birch forest and implications for sporopollenin-based proxies of past environmental change. *Polar Research* 30:1.
- Fujiki T, Inoue T, Yasuda Y. 2003. Pollen morphology of *Cedrus*. *Japanese Journal of Palynology* 49:21–24.
- Gould FW. 1957. Pollen size as related to polyploidy and speciation in the andropogon saccharoides-A. Barbinodis Complex. *Brittonia* 9:71–75.
- Griener KW, Nelson DM, Warny S. 2013. Declining moisture availability on the Antarctic Peninsula during the Late Eocene. *Palaeogeography Palaeoclimatology Palaeoecology* 383–384:72–78.
- Griener KW, Warny S. 2015. *Nothofagus* pollen grain size as a proxy for long-term climate change: an applied study on Eocene, Oligocene, and Miocene sediments from Antarctica. *Review of Palaeobotany and Palynology* 221:138–143.
- Harris I, Jones PD, Osborn TJ, Lister DH. 2014. Updated high-resolution grids of monthly climatic observations - the CRU TS3.10 Dataset. *International Journal of Climatology* 34:623–642.
- Hecht E. 2002. Optics. 4th ed. San Francisco: Addison-Wesley.
- Heusser LE, Stock CE. 1984. Preparation techniques for concentrating pollen from marine sediments and other sediments with low pollen density. *Palynology* 8:225–227.
- Ho R. 1972. Studies on pollen of selected species in Pinaceae. Vancouver: University of British Columbia.
- Ilmen R, Sabir A, Benziane M, Karrouk MS. 2014. Variability and dynamic response of the cedar to climate change in the Eastern Middle Atlas Mountains, Morocco. *Moroccan Journal of Chemistry* 2:512–516.
- Jardine PE, Lomax BH. 2017. Is pollen size a robust proxy for moisture availability? *Review of Palaeobotany and Palynology* <https://doi.org/10.1016/j.revpalbo.2017.06.013>
- Jardine PE, Abernethy FAJ, Lomax BH, Gosling WD, Fraser WT. 2017. Shedding light on sporopollenin chemistry, with reference to UV reconstructions. *Review of Palaeobotany and Palynology* 238:1–6.
- Kapadia ZJ, Gould FW. 1964. Biosystematic Studies in the *Bouteloua Curtipendula* Complex. III. Pollen Size As Related To Chromosome Numbers. *American Journal of Botany* 51:166–172.
- Khanduri VP, Sharma CM. 2009. Cyclic pollen production in *Cedrus deodara*. *Sexual Plant Reproduction* 22:53–61.
- King DC, Schubert BA, Jahren AH. 2012. Practical considerations for the use of pollen $\delta^{13}C$ value as a paleoclimate indicator. *Rapid Communications in Mass Spectrometry* 26:2165–2172.

- Knight CA, Clancy RB, Götzenberger L, Dann L, Beaulieu JM. 2010. On the relationship between pollen size and genome size. *Journal of Botany* 2010:1–7.
- Körner C, Farquhar GD, Wong SC. 1991. Carbon isotope discrimination by plants follows latitudinal and altitudinal trends. *Oecologia* 88:30–40.
- Kurtz EB, Liverman JL, Tucker H. 1958. Some effects of temperature on pollen characters. *Bulletin of the Torrey Botanical Club* 87:85–94.
- Lamb HF, Eicher U, Switsur V. 1989. An 18,000-year record of vegetation, lake-level and climatic change from Tigalmamine, Middle Atlas, Morocco. *Journal of Biogeography* 16:65–74.
- Lau T, Stephenson A. 1994. Effects of soil phosphorus on pollen production, pollen size, pollen phosphorus content, and the ability to sire seeds in *Cucurbita pepo* (Cucurbitaceae). *Sexual Plant Reproduction* 7:215–220.
- Lau T, Stephenson A. 1993. Effects of soil nitrogen on pollen production, pollen grain size, and pollen performance in *Cucurbita pepo* (Cucurbitaceae). *American Journal of Botany* 80:763–768.
- Linares JC, Taïqui L, Sangüesa-Barreda G, Seco JI, Camarero JJ. 2013. Age-related drought sensitivity of Atlas cedar (*Cedrus atlantica*) in the Moroccan Middle Atlas forests. *Dendrochronologia* 31:88–96.
- Loader NJ, Hemming DL. 2004. The stable isotope analysis of pollen as an indicator of terrestrial palaeoenvironmental change: a review of progress and recent developments. *Quaternary Science Reviews* 23:893–900.
- Lomax BH, Fraser WT, Harrington G, Blackmore S, Sephton MA, Harris NBW. 2012. A novel palaeoaltimetry proxy based on spore and pollen wall chemistry. *Earth and Planetary Science Letters* 353:22–28.
- Lu Y, Jin B, Wang L, Wang Y, Wang D, Jiang X, Chen P. 2011. Adaptation of male reproductive structures to wind pollination in gymnosperms: cones and pollen grains. *Canadian Journal of Plant Science* 91:897–906.
- Magri D, Di Rita F, Aranbarri J, Fletcher W, González-Sampériz P. 2017. Quaternary disappearance of tree taxa from Southern Europe: timing and trends. *Quaternary Science Reviews* 163:23–55.
- Mäkelä EM. 1996. Size distinctions between *Betula* pollen types — A review. *Grana* 35:248–256.
- Miller PD. 1982. Maize Pollen: collection and enzymology. In: Sheridan WF, editor. *Maize for biological research*. Charlottesville: Plant Molecular Biology Association; p. 279–293.
- Nakagawa T, Yasuda Y, Tabata H. 1996. Pollen morphology of Himalayan *Pinus* and *Quercus* and its importance in palynological studies in Himalayan area. *Review of Palaeobotany and Palynology* 91:317–329.
- Nelson DM. 2012. Carbon isotopic composition of Ambrosia and Artemisia pollen: Assessment of a C3-plant paleophysiological indicator. *New Phytologist* 195:787–793.
- Nelson DM, Hu FS, Michener RH. 2006. Stable-carbon isotope composition of Poaceae pollen: an assessment for reconstructing C3 and C4 grass abundance. *The Holocene* 16:819–825.
- Nelson DM, Hu FS, Mikucki JA, Tian J, Pearson A. 2007. Carbon-isotopic analysis of individual pollen grains from C3 and C4 grasses using a spooling-wire microcombustion interface. *Geochimica et Cosmochimica Acta* 71:4005–4014.
- Niklas KJ. 1985. The aerodynamics of wind pollination. *The Botanical Review* 51:328–386.
- Praglowksi J. 1970. The effects of pre-treatment and the embedding media on the shape of pollen grains. *Review of Palaeobotany and Palynology* 10:203–208.
- Punt W, Hoen PP, Blackmore S, Nilsson S, Le Thomas A. 2007. Glossary of pollen and spore terminology. *Review of Palaeobotany and Palynology* 143:1–81.
- Qiao CY, Ran JH, Li Y, Wang XQ. 2007. Phylogeny and biogeography of *Cedrus* (Pinaceae) inferred from sequences of seven paternal chloroplast and maternal mitochondrial DNA regions. *Annals of Botany* 100:573–580.
- R Core Team. 2016. R: a language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing.
- Radaeski JN, Bauermann SG, Pereira AB. 2016. Poaceae Pollen from Southern Brazil: distinguishing grasslands (Campos) from Forests by Analyzing a Diverse Range of Poaceae Species. *Frontiers in Plant Science* 7:1–18.
- Reitsma T. 1969. Size modification of recent pollen grains under different treatments. *Review of Palaeobotany and Palynology* 9:175–202.
- Renau-Morata B, Nebauer SG, Sales E, Allainguillaume J, Caligari P, Segura J. 2005. Genetic diversity and structure of natural and managed populations of *Cedrus atlantica* (Pinaceae) assessed using random amplified polymorphic DNA. *American Journal of Botany* 92:875–884.
- Rhanem M. 2011. Aridification du climat régional et remontée de la limite inférieure du cèdre de l'Atlas (*Cedrus atlantica* Manetti) aux confins de la plaine de Midelt (Maroc). *Physio-Géo* 5:143–165.
- Rozema J, Noordijk AJ, Broekman RA, van Beem A, Meijkamp BM, de Bakker NVJ, van de Staaij JWM, Stroetenga M, Bohncke SJP, Konert M, et al. 2001. (Poly)phenolic compounds in pollen and spores of Antarctic plants as indicators of solar UV-B: a new proxy for the reconstruction of past solar UV-B? *Plant Ecology* 154:9–26.
- Rozema J, van Geel B, Björn LO, Lean J, Madronich S. 2002. Toward solving the UV puzzle. *Science* 296:1621–1622.
- Schoch-Bodmer H. 1936. Zur Methodik der Größenbestimmung von Pollenkörner, mit besonderer Berücksichtigung von *Corylus avellana*. *Berichte Schweizerische Botanische Gesellschaft* 45:62–70.
- Smith RW. 1923. Life history of *cedrus Atlantica*. *Botanical Gazette* 75:203–208.
- Sperazza M, Moore JN, Hendrix MS. 2004. High-Resolution particle size analysis of naturally occurring very fine-grained sediment through laser diffractometry. *Journal of Sedimentary Research* 74:736–743.
- Syvitski JPM. 1991. Principles, methods and application of particle size analysis. Cambridge: Cambridge University Press.
- Tate JA, Soltis DE, Soltis PS. 2005. Polyploidy in plants. In: Ryan GT, editor. *The evolution of the genome*. Boston: Academic Press.
- Tekleva MV, Polevova SV, Zavialova NE. 2007. On some peculiarities of sporoderm structure in members of the Cycadales and Ginkgoales. *Paleontological Journal* 41:1162–1178.
- Terrab A, Paun O, Talavera S, Tremetsberger K, Arista M, Stuessy TF. 2006. Genetic diversity and population structure in natural populations of Moroccan Atlas cedar (*Cedrus atlantica*; Pinaceae) determined with cpSSR markers. *American Journal of Botany* 93:1274–1280.
- Tiwari SP, Yadav D, Chauhan DK. 2012. Scanning Electron microscopic study of some saccate pollen grains of Pinaceae and Podocarpaceae. *National Academy Science Letters* 35:415–419.
- Traverse A. 2007. *Paleopalynology*. 2nd ed. Dordrecht: Springer Netherlands.
- U.S. Geological Survey. 2017. Global land cover characterization (GLCC) [Internet]. Available from: <https://lta.cr.usgs.gov/GLCC>
- Vonhof MJ, Harder LD. 1995. Size-number trade-offs and pollen production by papilionaceous legumes. *American Journal of Botany* 82:230–238.
- Whitehead DR. 1983. Wind pollination: ecological and evolutionary perspectives. In: Real L, editor. *Pollination biology*. Orlando: Academic Press.
- Willis KJ, Feurdean A, Birks HJB, Björne AE, Breman E, Broekman R, Grytnes JA, New M, Singarayer JS, Rozema J. 2011. Quantification of UV-B flux through time using UV-B-absorbing compounds contained in fossil *Pinus* sporopollenin. *New Phytologist* 192:553–560.
- Zielhofer C, Fletcher WJ, Mischke S, De Batist M, Campbell JFE, Joannin S, Tjallingii R, El Hamouti N, Junginger A, Stele A, et al. 2017. Atlantic forcing of Western Mediterranean winter rain minima during the last 12,000 years. *Quaternary Science Reviews* 157:29–51.