

A Description of the First Instar of *Matus ovatus* (Coleoptera: Dytiscidae)

Authors: Dent, B. M., Barman, E. H., Shepley-James, T. A., and White, B. P.

Source: Florida Entomologist, 94(3) : 439-442

Published By: Florida Entomological Society

URL: <https://doi.org/10.1653/024.094.0308>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

A DESCRIPTION OF THE FIRST INSTAR OF *MATUS OVATUS*
(COLEOPTERA: DYTISCIDAE)B. M. DENT¹, E. H. BARMAN², T. A. SHEPLEY-JAMES¹ AND B. P. WHITE¹¹Natural Science Department, Georgia Military College, Warner Robins, GA 31093²Department of Biological & Environmental Science, Georgia College & State University, Milledgeville, GA 31061

ABSTRACT

The first instar of *Matus ovatus* Leech is described and illustrated. The analysis of the primary chaetotaxy of legs, head, and last abdominal segment of *M. ovatus* revealed no significant differences between this species and the previously described *M. bicarinatus*. Undescribed primary anatomical features of the tribe Matini presented herein include (1) a tentative identification of anterior tentorial pits and fragments of the anterior tentorial arms, (2) anterior protergal modifications and chaetotaxy, and (3) complete sclerotization of the seventh abdominal segment. Differences in mandibular morphology permit identification of first instars of *M. ovatus* and *M. bicarinatus* in the eastern United States.

Key Words: *Matus ovatus*, larval morphology, immature stage, Dytiscidae

RESUMEN

Primer estadio de *Matus ovatus* Leech se describe e ilustra. El análisis de la chaetotaxia primaria de las piernas, la cabeza, y el último segmento abdominal de *M. ovatus* no reveló diferencias significativas entre esta especie y la ya descrita anteriormente *M. bicarinatus*. No se describen características anatómicas primarias de la tribu Matini presentadas en el presente incluyen (1) una identificación provisional de pozos tentorial anterior y fragmentos de los brazos tentorial anterior, (2) modificaciones protergales anterior y chaetotaxia, y (3) esclerotización completa del séptimo segmento abdominal. Las diferencias en la morfología de la mandíbula permiten la identificación del primer estadio de *M. ovatus* y *M. bicarinatus* en el este de Estados Unidos.

Translation provided by the authors.

Matus Aubé, endemic to North America, includes 4 species, *M. bicarinatus* (Say), *M. leechi* Young, *M. ovatus* Leech, and *M. relictus* Young (Young 1953; Larson et al. 2000). Detailed descriptions of third instars of *Matus ovatus* Leech (Wolfe & Roughley 1985), second instars *M. leechi*, and first, second, and third instars of *M. bicarinatus* (Alarie 1995, 1998; Alarie et al. 2001) are available. Analyses of first instar (primary) morphology has provided information for the development and testing of systematic hypotheses (e.g., Nilsson 1988; Alarie 1995, 1998; Alarie et al. 2001, 2002, 2009). However, primary morphology is available only for *M. bicarinatus*. The objective of this study was to provide the results of an analysis of external morphology of the heretofore undescribed first instar of *M. ovatus* that is inclusive of, but not restricted to, chaetotaxy.

MATERIALS AND METHODS

Larvae described were collected between 31 V 2007 and 5 VI 2007 from a marsh site in Monroe County, (N33°6.47', W83°47.89'), GA, USA. Larvae were identified as *M. ovatus* on the basis of

distribution records (Turnbow & Smith 1983), associations with third instars described by Wolfe & Roughley (1985), and by comparison to descriptions of first instars of *M. bicarinatus* (Alarie 1995, 1998; Alarie et al. 2001).

Descriptions are based on 10 (unless otherwise specified) first instars randomly selected from a larger cohort preserved in 70% glycerated alcohol. Head lengths were measured dorsally from the posterior margin of the head capsule along the coronal suture to the anterior margin of the frontoclypeus, excluding the frontoclypeal sensilla. Head widths were measured dorsally from the widest point. Gape measurements were taken ventrally from the center of the ball of the right and left mandibular articulations, and mandibular length was measured ventrally from the apex of the mandible to the center of the ball of articulation (Wall et al. 2006). Because of the pseudochelate structural modifications, 2 lengths were taken for each pro- and mesothoracic tibia. One length was determined by measuring each from its articulation with the femur to its abutment with the tarsus, and a second length was taken that included the tibial extensions resulting in

the pseudo-chelate modifications. Measurements for all other structures were taken along either the greatest lengths or greatest widths. Total leg lengths were calculated based on the sums of the lengths of the coxae, femora, tibiae (excluding lengths that included pseudo-chelate modifications), and tarsi.

Identification and coding of primary sensilla was based on Nilsson (1988) and Alarie (1995, 1998). Other morphometric terminology utilized is based on Snodgrass (1935), Bousquet & Goulet (1984), and Larson et al. (2000).

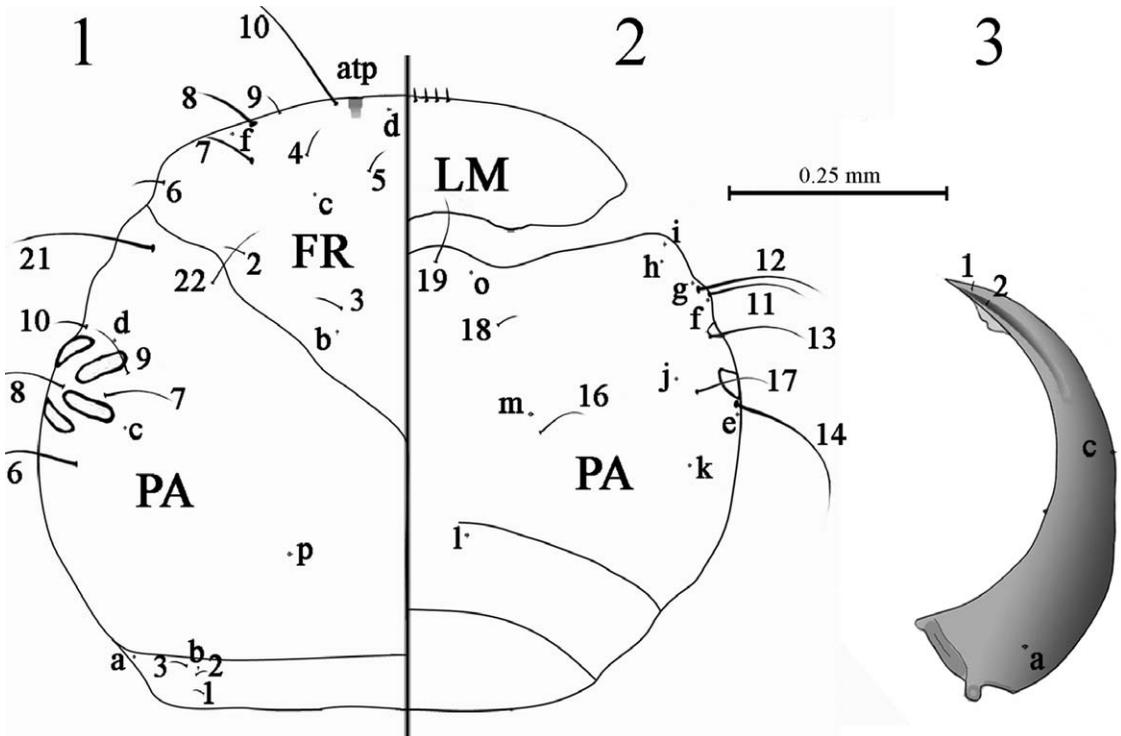
Description of the First Instar of *M. ovatus* (Figs. 1-3)

Body. General shape subcylindrical, vermiform with membranous areas generally translucent and sclerotized areas reddish-brown (alcohol preserved specimens).

Head (Figs. 1 and 2). Ovoid with lateral margins inflated laterally, resulting in a continuous arc between bases of antennae and postoccipital suture, occipital region short, defined anteriorly by a prominent occipital suture, posteroventral emargination subtle, length 0.66-0.72 mm ($\bar{x} = 0.70 \pm 0.02$ mm); maximum width, 0.66-0.78 mm ($\bar{x} = 0.72 \pm 0.04$ mm), near cranial midpoint; gape (Wall et al. 2006), 0.46-0.68 mm ($\bar{x} = 0.55 \pm 0.07$);

coronal suture, length 0.24-0.44 mm ($\bar{x} = 0.31 \pm 0.05$ mm); frontoclypeus broadly triangular with pair of eggburstors, length 0.36-0.44 ($\bar{x} = 0.40 \pm 0.02$ mm); 8 small spiniform frontoclypeal sensilla present anteriorly; anterior tentorial pits (Fig. 1) (tentative) on or near the clypeolabral suture (Snodgrass 1935), contiguous with remnants of anterior arms (Beutel 1994); adnasale absent (Alarie 1998; Alarie et al 2001); extensively sclerotized labrum directed posteroventrally; capsule with numerous spinulae both dorsally and ventrally; temporal spines absent; posterior tentorial pits visible ventrally; corneal lenses (Shepley-James et al. 2009) arranged in 2 columns of 3 each, lenses nearly equal in size. Antenna. Length, 0.46-0.50 mm ($\bar{x} = 0.47 \pm 0.02$ mm); antennomere (AN); AN1, 0.08-0.10 mm ($\bar{x} = 0.09 \pm 0.01$ mm), AN2, 0.12-0.15 mm ($\bar{x} = 0.14 \pm 0.01$ mm), AN3, 0.17-0.21 mm ($\bar{x} = 0.18 \pm 0.01$ mm), with prominent finger-like accessory appendage (A3'), AN4, 0.06-0.08 mm ($\bar{x} = 0.07 \pm 0.01$ mm).

Mouthparts. Mandible, (Fig. 3), length 0.41-0.57 mm ($\bar{x} = 0.45 \pm 0.05$ mm) with pronounced lateral and medial arcs, angle of attack $\approx 45^\circ$ (Wall et al. 2006), well-developed mandibular channel, a single prominent distal tooth on the anteroventromedial edge, minute proximal tooth or teeth on some specimens. Maxilla. Cardio plate-like and



Figs. 1-3. *Matus ovatus*. 1, Cranium dorsal view; 2, Cranium, ventral view; 3, Mandible, ventral view. Legend: atp, anterior tentorial pit; FR, frontoclypeus; LM, labrum; PA, parietal. Spiniform and setiform sensilla are identified by numbers with lower case letters used identify campaniform (pore-like) sensilla (Alarie 1998)

triangular; stipes prominent and sub-triangular; finger-like galea subequal in length to that of palpomere (PL); PL1, 0.10- 0.13 mm ($\bar{x} = 0.11 \pm 0.01$ mm), PL2, 0.10- 0.12 mm ($\bar{x} = 0.10 \pm 0.01$ mm), PL3, 0.14- 0.18 mm ($\bar{x} = 0.16 \pm 0.01$ mm). Labium. Prementum short, concave medially, width approximately twice its length; PL1, 0.10- 0.21 mm ($\bar{x} = 0.17 \pm 0.04$ mm), PL2, 0.17- 0.21 mm ($\bar{x} = 0.19 \pm 0.01$ mm).

Thorax ($n = 9$). Protergum, medial dorsal length 0.36-0.58 mm ($\bar{x} = 0.49 \pm 0.06$ mm), widest posteriorly, bearing 4 prominent sensilla anterolaterally and posterior to antecostal suture, with a precostal inflection widest laterally (Fig. 4); respective meso-, and metatergal dorsal lengths, 0.27-0.38 mm ($\bar{x} = 0.34 \pm 0.04$ mm), 0.27-0.38 mm ($\bar{x} = 0.33 \pm 0.05$ mm); meso- and metaterga widest medially with prominent sensilla; spiracles absent. Legs ($n = 8$). Respective total lengths of pro-, meso-, and metalegs, 1.25-1.44 mm ($\bar{x} = 1.33 \pm 0.06$ mm), 1.20- 1.46 mm ($\bar{x} = 1.35 \pm 0.09$ mm), 1.70- 2.09 mm ($\bar{x} = 1.90 \pm 0.12$ mm); metaleg anterior claw shorter than posterior; metatibia and metatarsus with discreet anteroventral spinulae; respective lengths of pro-, meso-, and meta individual segments: coxae, 0.44-0.50 mm ($\bar{x} = 0.47 \pm 0.02$ mm), 0.42-0.55 mm ($\bar{x} = 0.49 \pm 0.05$ mm), 0.52-0.67 mm ($\bar{x} = 0.61 \pm 0.05$ mm); trochanters, 0.13-0.20 mm ($\bar{x} = 0.18 \pm 0.02$ mm), 0.18-0.22 mm ($\bar{x} = 0.20 \pm 0.01$ mm), 0.19-0.26 mm ($\bar{x} = 0.22 \pm 0.02$ mm); femora, 0.39-0.48 mm ($\bar{x} = 0.44 \pm 0.03$ mm), 0.44-0.51 mm ($\bar{x} = 0.47 \pm 0.02$ mm), 0.44-0.56 mm ($\bar{x} = 0.50 \pm 0.04$ mm); tibiae (excluding pseudo-chelate anomaly), 0.13-0.20 mm ($\bar{x} = 0.17 \pm 0.02$ mm), 0.11-0.23 mm ($\bar{x} = 0.17 \pm 0.04$ mm), 0.37-0.46 mm ($\bar{x} = 0.41 \pm 0.03$ mm); tarsi, 0.20-0.24 mm ($\bar{x} = 0.23 \pm 0.01$ mm), 0.19-0.34 mm ($\bar{x} = 0.24 \pm 0.05$ mm), 0.26-0.44 mm ($\bar{x} = 0.39 \pm 0.06$ mm); respective pro and mesotibial lengths including pseudo-chelate anomaly, 0.23-0.39 mm ($\bar{x} = 0.35 \pm 0.05$ mm), 0.26-0.51 mm ($\bar{x} = 0.38 \pm 0.07$ mm).

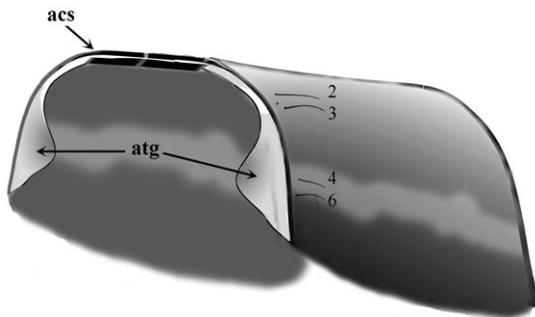


Fig. 4. *Matus ovatus*. Protergum, anterolateral view. Legend: atg, acrotergite; acs, antecostal suture. Sensilla are identified by numbers (Bousquet & Goulet 1984).

Abdomen: Segments 1-5 sclerotized dorsally, segment 6 sclerotized dorsally and laterally below dorsopleural line, otherwise membranous; segments 1-6 each with prominent antecostal suture and bearing long, broadly dispersed, robust sensilla; segments 7 and 8 heavily and completely sclerotized, dorsal length of segment 8, 0.92-1.30 mm ($\bar{x} = 1.18 \pm 0.12$ mm), siphon length 0.38-0.62 mm ($\bar{x} = 0.54 \pm 0.08$ mm). Urogomphus One-segmented, length 0.48- 0.60 mm ($\bar{x} = 0.55 \pm 0.05$ mm).

Chaetotaxy. Primary ancestral and additional sensilla in general as attributed to *M. bicarinatus* on legs, last abdominal segments, urogomphi (Alarie 1995; Alarie et al. 2001), cranium, and cranial appendages (Alarie 1998; Alarie et al. 2001). Differences between *M. ovatus* and *M. bicarinatus* include: Mandible, MN_b not found; femur with 2 additional AV primary sensilla; mesofemoral FE_b short and spine-like, *vs.* elongate and hair-like (Alarie 1995), or coded for phylogenetic analyses as short and spine-like (Alarie et al. 2001).

Bionomics. First instars of *M. ovatus* were present in small numbers between 25 IV 2008 and 7 VI 2008 in localized areas of a relatively large anthropomorphic marsh. Mature larvae of *Hydrovatus* sp. indet. were frequently collected along with the *Matus* larvae. This lentic system supported dense populations of aquatic macrophytes, including *Egeria densa* Planchon and *Alternanthera philoxeroides* (Mart.) Griseb., with large amounts of sedimentary and detrital materials suspended in the water column. The temporal distribution of larvae indicates that *M. ovatus* commenced oviposition at this site in late Apr and that this activity continued until at least early Jun.

DISCUSSION

Structures present on first instars of *M. ovatus* but not reported for *M. bicarinatus* because of experimental design or objectives (Alarie 1995, 1998; Alarie et al 2001) include (1) apparent tentorial pits (Fig. 1) on or near the clypeolabral suture (Snodgrass 1935), (2) an anterior protergal inflection (Fig. 4) that appears to accommodate an articulation of the roughly ovoid cranium with the protergum; and (3) robust anterolateral protergal sensilla that may be homologous to those reported for first instars of Carabidae (Bousquet & Goulet 1984). The seventh abdominal segment of *M. ovatus* is completely sclerotized. However, the seventh abdominal segment of Matini larvae, including *M. bicarinatus*, was described (Alarie et al. 2001) as having a ventral plate distinct from the tergite.

In southeastern Canada and the U.S. east of the Mississippi River (Larson et al. 2000; Epler 2010), larvae with pseudo-chelate tibial modifica-

tions, crania that are less than 0.8 mm in width, and frontoclypeal egg bursters, but with neither temporal spines nor spiracles, will be first instars of either *M. ovatus* or *M. bicarinatus*. Mandibles of first instars of *M. bicarinatus* possess multiple teeth on the ventromedial surface (Alarie 1998; Alarie et al. 2001), whereas first instars of *M. ovatus* possess a single prominent tooth with an occasional minute proximate tooth or teeth on some specimens (Fig. 3). This provides an accessible and objective non-chaetotaxal character for identification of first instars of *M. ovatus* and *M. bicarinatus* within the geographic limits noted above.

ACKNOWLEDGMENTS

William P. Wall Museum of Natural History; Aquatic Coleoptera Laboratory Contribution No. 75. James Bailey and Jan R. Oliva, students at Georgia Military College (Warner Robins), were helpful in collecting the larval material. This project was supported in part by a Faculty Research Grant awarded by the Office of Research Services, Georgia College & State University.

REFERENCES CITED

- ALARIE, Y. 1995. Primary setae and pores on the legs, the last abdominal segment, and the urogomphi of larvae of Nearctic Colymbetinae (Coleoptera: Adephegata: Dytiscidae) with an analysis of their phylogenetic relationship. *Can. Entomol.* 127: 913-943.
- ALARIE, Y. 1998. Phylogenetic relationships of Nearctic Colymbetinae (Coleoptera: Adephegata: Dytiscidae) based on chaetotaxal and porotaxal analysis of head capsule and appendages of larvae. *Can. Entomol.* 130: 803-824.
- ALARIE, Y., WATTS, C. H. S., AND NILSSON, A. N. 2001. Larval morphology of the tribe Matini (Coleoptera: Dytiscidae: Colymbetinae): descriptions of *Batrachomatus daemeli*, *Matus bicarinatus*, and *Allomatus nannup* and phylogenetic relationships. *Can. Entomol.* 133: 165-196.
- ALARIE, Y., ARCHANGELSKY, M., NILSSON, A. N., AND WATTS, C. H. S. 2002. Larval morphology of the genus *Lancetes* (Coleoptera: Adephegata: Dytiscidae): the hypothesis of sister-group relationship with the subfamily Dytiscinae revisited. *Can. Entomol.* 134: 467-501.
- ALARIE, Y., MICHA, M. C., NILSSON, A. N., ARCHANGELSKY, M., AND HENDRICH, L. 2009. Larval morphology of *Rhantus* Dejean, 1833 (Coleoptera: Dytiscidae: Colymbetinae): descriptions of 22 species and phylogenetic considerations. *Zootaxa* 2317: 1-102.
- BEUTEL, R. G. 1994. On the systematic position of *Hydrotrupes palpalis* Sharp (Coleoptera: Dytiscidae). *Aquatic Insects* 16: 157-164.
- BOUSQUET, Y., AND GOULET, H. 1984. Notation of primary setae and pores on larvae of Carabidae (Coleoptera: Adephegata). *Can. J. Zool.* 62: 573-588.
- LARSON, D. J., ALARIE, Y., AND ROUGHLEY, R. E. 2000. Predaceous diving beetles (Coleoptera: Dytiscidae) of the Nearctic Region, with an emphasis on the fauna of Canada and Alaska. NRC Research Press, Ottawa, 982 pp.
- NILSSON, A. N. 1988. A review of primary setae and pores on legs of larval Dytiscidae (Coleoptera). *Can. J. Zool.* 66: 2283-2294.
- SNODGRASS, R. E. 1935. Principles of Insect Morphology. New York: McGraw-Hill, 667 pp.
- SHEPLEY-JAMES, T. A., WHITE, B. P., BARMAN, E. H., BINKOWSKI, J., AND TREAT, A. 2009. Variation in stemmatid morphology of larvae of *Liodes noviaffinis* Miller (Dytiscidae: Hydroporinae: Bidessini). *Georgia J. Sci.* 67: 72-74.
- TURNBOW, R., AND SMITH, C. L. 1983. An annotated checklist of the Hydradephaga (Coleoptera) of Georgia. *J. Georgia Entomol. Soc.* 18: 429-443.
- WALL, W. P., BARMAN E. H., AND BEALS, C. M. 2006. A description and functional interpretation of the mandibular geometry of *Agabus punctatus* Melsheimer, 1844, *Rhantus calidus* (Fabricius, 1792) and *Acilius mediatius* (Say, 1823), (Coleoptera: Dytiscidae). *Aquatic Insects* 28: 277-289.
- WOLFE, G. W., AND ROUGHLEY, R. E. 1985. Description of the pupa and mature larva of *Matus ovatus ovatus* Leech (Coleoptera: Dytiscidae) with a chaetotaxal analysis emphasizing mouthparts, legs, and urogomphus. *Proc. Acad. Nat. Sci. Philadelphia* 137: 61-79.
- YOUNG, F. N. 1953. Two new species of *Matus*, with a key to the known species and subspecies of the genus (Coleoptera: Dytiscidae). *Ann. Entomol. Soc. America* 46: 49-55.