Molecular Identification and Phylogeny of Bactrocera Species (Diptera: Tephritidae)


Source: Florida Entomologist, 94(4) : 1026-1035

Published By: Florida Entomological Society

URL: https://doi.org/10.1653/024.094.0441

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne’s Terms of Use, available at [www.bioone.org/terms-of-use](http://www.bioone.org/terms-of-use).

Usage of BioOne Complete content is strictly limited to personal, educational, and nonprofit, non-commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.
MOLECULAR IDENTIFICATION AND PHYLOGENY OF BACTROCERA SPECIES (DIPTERA: TEPHRITIDAE)

R. ASOKAN1, K. B. REBIJITH1, SHAKTI K. SINGH1, A. S. SIDHU2, S. SIDDHARTHAN3, PRAVEEN. K. KARANTH3, R. ELLANGO1 AND V. V. RAMAMURTHY2

1Division of Biotechnology, Indian Institute of Horticultural Research (IIHR), Bangalore, India
2Network Project on Insect Biosystematics, Division of Entomology, Indian Agricultural Research Institute (IARI), New Delhi, India
3Centre for Ecological Sciences, Indian Institute of Science (IISc), Bangalore, India

Corresponding authors e-mail: asokaniihr@gmail.com, rebijith@gmail.com

ABSTRACT

Fruit flies that belong to the genus Bactrocera (Diptera: Tephritidae) are major invasive pests of agricultural crops in Asia and Australia. Increased transboundary movement of agricultural produce has resulted in the chance introduction of many invasive species that include Bactrocera mainly as immature stages. Therefore quick and accurate species diagnosis is important at the port of entry, where morphological identification has a limited role, as it requires the presence of adult specimens and the availability of a specialist. Unfortunately when only immature stages are present, a lacuna in their taxonomy impedes accurate species diagnosis. At this juncture, molecular species diagnostics based on COX-I have become handy, because diagnosis is not limited by developmental stages. Yet another method of quick and accurate species diagnosis for Bactrocera spp. is based on the development of species-specific markers. This study evaluated the utility of COX-I for the quick and accurate species diagnosis of eggs, larvae, pupae and adults of B. zonata Saunders, B. tau Walker, and B. dorsalis Hendel. Furthermore the utility of species-specific markers in differentiating B. zonata (500bp) and B. tau (220bp) was shown. Phylogenetic relationships among five subgenera, viz., Austrodacus, Bactrocera, Daculus, Notodacus and Zeugodacus have been resolved employing the 5’ region of COX-I (1490-2198); where COX-I sequences for B. dorsalis Hendel, B. tau Walker, B. correcta Bezzi and B. zonata Saunders from India were compared with other NCBI-GenBank accessions. Phylogenetic analysis employing Maximum Parsimony (MP) and Bayesian phylogenetic approach (BP) showed that the subgenus Bactrocera is monophyletic.

Key Words: Bactrocera, COX-I, species-specific marker, Bayesian phylogeny, monophyletic

RESUMEN

Las moscas de la fruta que pertenecen al género Bactrocera (Diptera: Tephritidae) son las principales plagas invasoras de los cultivos agrícolas en Asia y Australia. El aumento en el movimiento transfronterizo de los productos agrícolas ha resultado en la posibilidad de la introducción de muchas especies invasoras que incluyen Bactrocera principalmente por los estados inmaduros. Por lo tanto, el diagnóstico de especies con rapidez y precisión es importante en el puerto de entrada, donde la identificación morfológica tiene un papel limitado, ya que requiere la presencia de ejemplares adultos, la disponibilidad de especialistas, y donde la escasez de datos en la taxonomía de los estados inmaduros de las especies impiden el diagnóstico exacto. En esta situación, el diagnóstico molecular de especies basado en el gene COX-I ha llegado a ser útil, ya que el diagnóstico de especies no está limitado por el estadio de desarrollo. Sin embargo, otro método de diagnóstico rápido y exacto de especies de Bactrocera spp. es basado en el desarrollo de marcadores específicos de la especie. En el presente estudio, se evaluó la utilidad de usar el COX-I para el diagnóstico rápido y preciso de especies usando los huevos, larvas, pupas y adultos de B. zonata Saunders, B. tau Walker y B. dorsalis Hendel. También demosladamos la utilidad de usar marcadores específicos de las especies para diferenciar B. zonata (500bp) y B. tau (220bp). Relaciones filogenéticas entre los cinco subgéneros, es decir, Austrodacus, Bactrocera, Daculus, Notodacus y Zeugodacus se ha resuelto por primera vez empleando la región 5’ de la COX-I (1490 a 2198) donde se compararon las secuencias de COX-I de B. dorsalis Hendel, B. tau Walker, B. correcta Bezzi y B. zonata Saunders, de la India con las accesiones del NCBI-GenBank. El análisis filogenético utilizando tanto el programa Union de Vecinos (NJ, Neighbor-Joining) y la máxima parsimonia (MP) demostró que los subgéneros del Bactrocera son monofílicos.
Fruit flies are among the world's most serious pests causing enormous yield losses. Tephritid flies of the genus *Bactrocera* are of particular concern throughout Asia and Australia (Kim et al. 1999). There are about 500 described species of *Bactrocera* that are grouped into 28 subgenera (Drew and Hancock 2000). Accidental introduction of alien invasive species, such as *Bactrocera* species, poses a serious and mounting threat to crop production with increasing long distance transportation of agricultural produce. Therefore it is necessary to identify the species quickly and accurately at the port of entry. In this regard identification using morphological characters falls short because it requires adult specimens and the availability of expertise in taxonomy; this is further complicated by the lack of identification keys for identification of immature stages (eggs, larvae and pupae). The problematic immature stages are most often encountered in the import consignments. At this juncture, molecular species diagnostics based on COX-I become handy because they are not limited by developmental stage or sexual gender. Yet another method of resolving identification of closely related species is achievable by the development of species-specific primers that produce a specific amplicons. Such species-specific markers could be used effectively for species diagnosis even by a non-specialist.

In previous studies mitochondrial DNA has been utilized to resolve the phylogeny of *Bactrocera* (Muraji & Nakahara 2001; Smith et al. 2003; Zhang et al. 2010) and host associated genetic differences in thrips (Brunner et al. 2004). In this regard, there are contradicting views on whether the phylogeny of *Bactrocera* is monophyletic or paraphyletic (Zhang et al. 2010). A monophyletic theory for *Bactrocera* was proposed by Drew (1989) based on 5 morphological characters. On the other hand White (2000) proposed the employment of DNA sequencing to resolve the phylogeny of *Bactrocera*. Subsequently Muraji & Nakahara (2001) employed 1.6 kb mitochondrial DNA sequences and concluded that *Bactrocera* is paraphyletic.

In the present study the utility of various developmental stages namely, egg, larva, pupa and adult (male, female) of 4 economically important species of *Bactrocera* (*B. zonata* Saunders, *B. correcta* Bezzi, *B. tau* Walker, and *B. dorsalis* Hendel) were analyzed for accurate species diagnosis by using COX-I sequencing. The utility of species-specific markers in discriminating *B. zonata* from *B. tau* were also evaluated. Phylogenetic analyses carried out employing COX-I (1490-2198) for these species, which were compared to the NCBI-Genbank accessions to understand their phylogeny. Bayesian methods of phylogeny that calculate the posterior probabilities of the clades based nucleotide substitution models were employed to assess the monophyly of the group.

### Materials and Methods

**Stock Culture Maintenance**

Adults of *B. dorsalis*, *B. correcta*, *B. tau* and *B. zonata* were obtained from Delhi using bait traps and net sweeps in 2008 and 2009. These insects were mass-reared on pumpkin *Cucurbita maxima*, kept in plastic cages (24.5 cm × 20.5 cm × 20.5 cm) and provided with water, yeast powder bacto applied on sugar cubes and a piece of pumpkin as a food source and site of oviposition. Individual pairs of newly emerged adults were separated from this stock and maintained separately. These were observed for oviposition, and as soon as oviposition started pieces of pumpkin were removed to separate plastic jars (14.0 cm × 10.5 cm) each provided with a layer of sand (10-15 cm) to facilitate pupation; the mouths of these plastic jars were covered with muslin cloth and secured by rubber bands. Observations were made on the larva and puparia daily, and puparia were removed to separate containers before adult emergence to record the sex ratio and other details. Morphological identification of all 4 species of *Bactrocera*, viz., *B. tau*, *B. zonata*, *B. correcta* and *B. dorsalis*, was carried out according to White (1992) and Drew (1994) prior to molecular studies. The pure cultures were maintained in insect proof cages (24.5 cm × 20.5 cm × 20.5 cm) at room temperature (30-32 °C) and RH 70-90% for supplying samples of the immature stages from time to time. *B. zonata*, *B. tau*, *B. correcta* and *B. dorsalis* were used for developmental stage non-limiting studies, in which the egg, larva, pupa and adult were drawn from the insect cages from time to time and molecular identifications were carried out.

**DNA isolation and Polymerase Chain Reaction**

Total DNA was extracted from individual fruit fly adults using the CTAB method (Saghai Maroof et al. 1984). Thoracic tissue was homogenized with liquid nitrogen using a sterile micropestle in 1.5 mL microcentrifuge tube filled with 100 μL STE buffer (100 mM NaCl, 10 mM Tris HCl (pH 8.0), and 1 mM EDTA (pH 8.0)). The homogenate was heated at 65 °C for 60 min before being centrifuged at 6000 rpm for 15 min at room temperature followed by precipitation of DNA and disolution in molecular biology grade water (DNAsafe-free and RNAase free) (Eppendorf, Germany). Two μL was used as the template for Polymerase Chain Reaction (PCR).

PCR was carried out in a thermal cycler (AB- Applied Biosystems, Veriti 96 wells) with the following cycles: 94 °C for 4 min as initial denaturation followed by 35 cycles of 94 °C 40 s, 47 °C for 45 s and 72 °C for 45 s and 72 °C for 20 min as final extension. Use of primers specific to mitochondrial cytochrome oxidase I (COX-I), (LCO-1490-5′...
GGT CAA CAA ATC ATA AAG ATA TTG G -3';
HCO-2198- 5´- TAA ACT TCA GGG TGA CCA
AAA AAT CA -3'; Hebert et al. 2003) resulted in
the amplification of an approximately 700bp frag-
ment. PCR was performed in 25 μL total reaction
volume containing 20 pmol of each primer, 10
mM Tris HCl (pH 8.3), 50 mM KCl, 2.5 mM
MgCl2, 0.25 mM of each dNTP and 0.5 U of Taq
DNA polymerase (Thermo Scientific, Fermentas).
The amplified products were resolved in 1.0%
agarose gel, stained with ethidium bromide (10
μg/ml) and visualized in a gel documentation sys-
tem (UVP). For the species-specific primers men-
tioned in Table 1, PCR mix and PCR cycle param-
eters were the same except for annealing temper-
ature, viz., 62 °C for 45 s for both B. tau and B. zo-
nata.

**Molecular Cloning and Sequencing**

The PCR amplified fragments were eluted using
Nucleospin® Extract II according to the manu-
facturer's protocol (Macherey-Nagel [MN], Ger-
many) and ligated into the general purpose clon-
ing vector, InsT/Aclone (Fermentas GmbH, Ger-
many) according to the manufacturer's protocol.
Five μL of the ligated vector was cloned into 200
μL of competent *Escherichia coli* (DH5?) cells by
heat treatment at 42 °C for 45 s and the whole
content was transferred into a tube containing
800 μL of SOC (tryptone - 2% w/v, yeast extract -
0.5% w/v, NaCl - 8.6 mM, KCl - 2.5 mM, MgSO4 -
2.0 mM, glucose - 20 mM in 1000 mL water, pH
7.0) and rotated at 150 rpm, 37 °C for 1 h; 200 μL
of the above culture was spread on Luria Bertani
agar (LBA) (tryptone - 10 g, yeast extract - 5 g,

**Table 1. List of taxa examined with geographic origin and NCBI-GenBank accession numbers.**

<table>
<thead>
<tr>
<th>SPECIES (Bactrocera)</th>
<th>ORIGIN</th>
<th>COI</th>
</tr>
</thead>
<tbody>
<tr>
<td>B. correcta</td>
<td>India</td>
<td>GU323782</td>
</tr>
<tr>
<td>B. zonata</td>
<td>India</td>
<td>GU323777</td>
</tr>
<tr>
<td>B. verbascifoliae</td>
<td>India</td>
<td>DQ116350</td>
</tr>
<tr>
<td>B. umbrosa</td>
<td>Indonesia</td>
<td>DQ116348</td>
</tr>
<tr>
<td>B. trivialis</td>
<td>Papua New Guinea</td>
<td>DQ116342</td>
</tr>
<tr>
<td>B. trilineola</td>
<td>Vanuatu</td>
<td>DQ116339</td>
</tr>
<tr>
<td>B. tryoni</td>
<td>New Caledonia</td>
<td>DQ116337</td>
</tr>
<tr>
<td>B. redunca</td>
<td>Vanuatu</td>
<td>DQ116330</td>
</tr>
<tr>
<td>B. quadrisetosa</td>
<td>Vanuatu</td>
<td>DQ116328</td>
</tr>
<tr>
<td>B. papayae</td>
<td>Thailand</td>
<td>DQ116326</td>
</tr>
<tr>
<td>B. philippinensis</td>
<td>Philippines</td>
<td>DQ116320</td>
</tr>
<tr>
<td>B. psidii</td>
<td>New Caledonia</td>
<td>DQ116313</td>
</tr>
<tr>
<td>B. occipitalis</td>
<td>Indonesia</td>
<td>DQ116311</td>
</tr>
<tr>
<td>B. melanotus</td>
<td>Cook Islands</td>
<td>DQ116299</td>
</tr>
<tr>
<td>B. latifrons</td>
<td>USA</td>
<td>DQ116297</td>
</tr>
<tr>
<td>B. kandiensis</td>
<td>Sri Lanka</td>
<td>DQ116295</td>
</tr>
<tr>
<td>B. kirki</td>
<td>Tonga</td>
<td>DQ116294</td>
</tr>
<tr>
<td>B. jarvisi</td>
<td>Australia</td>
<td>DQ116288</td>
</tr>
<tr>
<td>B. frauenfeldi</td>
<td>Solomon Islands</td>
<td>DQ116287</td>
</tr>
<tr>
<td>B. facialis</td>
<td>Tonga</td>
<td>DQ116285</td>
</tr>
<tr>
<td>B. endriandrae</td>
<td>Australia</td>
<td>DQ116284</td>
</tr>
<tr>
<td>B. distincta</td>
<td>Tonga</td>
<td>DQ116282</td>
</tr>
<tr>
<td>B. caryae</td>
<td>India</td>
<td>DQ116266</td>
</tr>
<tr>
<td>B. curvipesinis</td>
<td>New Caledonia</td>
<td>DQ116261</td>
</tr>
<tr>
<td>B. carambolaee</td>
<td>Malaysia</td>
<td>DQ116259</td>
</tr>
<tr>
<td>B. cognata</td>
<td>Philippines</td>
<td>DQ116252</td>
</tr>
<tr>
<td>B. arecae</td>
<td>Thailand</td>
<td>DQ116236</td>
</tr>
<tr>
<td>B. aquilonis</td>
<td>Australia</td>
<td>DQ116235</td>
</tr>
<tr>
<td>B. cacuminata</td>
<td>Unknown</td>
<td>DQ116241</td>
</tr>
<tr>
<td>B. dorsalis</td>
<td>India</td>
<td>H056093</td>
</tr>
<tr>
<td>B. tau</td>
<td>India</td>
<td>GU323773</td>
</tr>
<tr>
<td>B. cucurbitae</td>
<td>Philippines</td>
<td>DQ116248</td>
</tr>
<tr>
<td>B. xanthodes</td>
<td>Vanuatu</td>
<td>DQ116356</td>
</tr>
<tr>
<td>B. cucumis</td>
<td>Australia</td>
<td>DQ116237</td>
</tr>
<tr>
<td>B. oleae</td>
<td>Italy</td>
<td>DQ116307</td>
</tr>
<tr>
<td>C. capitata</td>
<td>USA</td>
<td>DQ116371</td>
</tr>
<tr>
<td>A. ludens</td>
<td>Mexico</td>
<td>DQ116207</td>
</tr>
</tbody>
</table>
Asokan et al.: Molecular Identification and Phylogeny of Bactrocera Species

NaCl - 5 g, agar - 15 g in 1000 mL of water, pH 7.0) containing ampicillin (100 mg/mL), IPTG (4 mg/mL) and X-gal (40 mg/mL) and were incubated at 37 °C for 16 h. Blue/white selection was carried out and all the white colonies (colonies with insert) were maintained on LBA containing ampicillin (100 mg/mL), incubated at 37 °C overnight and stored at 4 °C. Plasmids were prepared from the overnight culture of the positive colonies cultured in LB broth (enzymatic casein - 10 g, yeast extract - 5 g, NaCl - 5 g in 1000 ml of water, pH-7.0) using modified alkali lysis method (Birnboim & Dolly 1979). Plasmids were resolved in 1.0% agarose gel, stained with ethidium bromide (10 μg/mL) and visualized by gel electrophoresis. Clones that had 2.5 kb as compared to control plasmid (1.8 kb) were selected for sequencing. For the purpose of sequencing, plasmids were isolated using GeneJET™ Plasmid Miniprep Kit (Fermentas, Germany) according to manufacturer's protocol from overnight cultures of the 5 randomly selected clones multiplied in LB broth. Sequencing was carried out on an automated sequencer (ABI Prism® 3730 XL DNA Analyzer; Applied Biosystems, USA) using M13 universal primers both in forward and reverse directions.

Homology searches carried out using BLAST (http://www.ncbi.nlm.nih.gov), and the differences in COX-1 sequences of B. tau, B. correcta, B. zonata, and B. dorsalis were determined using the sequence alignment editor BioEdit version 7.0.5.3 (Hall 1999). Sequences for B. tau, B. correcta, B. zonata, and B. dorsalis were deposited with the NCBI database, and the accession numbers were GU323773 - GU323782 and HQ658090 - HQ658093. For the development of species-specific markers for B. tau and B. zonata, 7 sets of forward and reverse primers were synthesized based on the variable regions in the aligned sequences of GU323774 - B. tau and GU323777 - B. zonata. The primers thus designed were tested on different identified Bactrocera species collected on mango and guava from the experimental farm of Indian Agricultural Research Institute (IARI), New Delhi, India. The PCR amplified fragments resulting from species-specific markers for B. tau and B. zonata were further cloned, sequenced and analyzed as above. Multiple sequences of all B. tau, B. zonata, B. correcta, and B. dorsalis from India and Bactrocera species from other geographical areas (acquired from GenBank) (Table 1) were aligned using the multiple alignment programme BioEdit v.7.0.

Phylogenetic Analysis

Phylogeny was assessed using maximum parsimony (MP), and Bayesian phylogeny (BP) using PAUP v4b10 (Swofford 1998) and MrBayes v3.1.2 (Ronquist & Heulsenbeck 2003). MP analysis was performed with 1000 replicates with random sequence addition, TBR (tree bisection reconnection) swapping, and multtrees option in effect. Parsimony Bootstrap support (BS) values were calculated based on 10,000 simple stepwise addition replicates with TBR branch swapping and 10 trees saved per replicate. For Bayesian analysis, the best-fit model of nucleotide evolution was evaluated using Modeltest v3.7 (Posada & Crandall 1998) based on the Akaike information criterion (AIC). MrBayes calculates the posterior probability of the phylogenetic tree based on the Bayesian theorem and nucleotide substitution models used in maximum likelihood can be imple-

Fig. 1. Sequence comparison for B. tau and B. zonata from the mitochondrial Cytochrome Oxidase I (COX-I) gene showing differences. Analyses performed using BioEdit v.7.0.
ment. The program uses Monte Carlo Markov Chain (MCMC) approach to approximate the posterior probability of the trees based on the best-fit model for that particular sequence. (Huelsnsebeck & Ronquist 2001). The posterior probabilities (PP) of the clades were obtained by 2 independent runs with 4 MCMC each. The run was continued for 5 million generations and trees were sampled every hundredth generation resulting in 50,000 trees. The first 15% were considered as the burn in phase and the remaining 42,000 trees were summarized. The trees obtained were visualized using Figtree v1.3.1 (Rambaut 2009).

RESULTS AND DISCUSSION

The PCR amplicon of the same size (approx. 700bp) was amplified for both B. tau and B. zonata. Sequencing results showed that the total nucleotide length obtained was 658 bp for both B. tau and B. zonata. The BLAST search for the eight sequences viz. GU323773 - B. tau (egg), GU323774 - B. tau (adult), GU323775 - B. tau (larva), GU323776 - B. tau (pupa), GU323777 - B. zonata (adult), GU323778 - B. zonata (larva), GU323779 - B. zonata (pupa), GU323780 - B. zonata (egg) shows that both of the sequences had the maximum hits for the respective species. Alignment of the sequences for various developmental stages, viz., egg, larva, pupa and adult, did not show any differences in the COX-I nucleotide sequences, which clearly proved that molecular identification is not limited by stage of development, polymorphism and sex of the target species. Hence, it also proved that the species-specific marker developed can be extremely useful for molecular identification in any given developmental stage, in particular for the B. zonata and B. tau, the 2 species which occur on mango (Mangifera indica L.), is significant.

Alignment of the sequences for GU323774 - B. tau (adult) and GU323777 - B. zonata (adult) in BioEdit V. 7.0 shows that there were variations in 103 nucleotides out of 658 bp; amounting to a 15.65% difference between B. tau and B. zonata (Fig. 1). The rapid and accurate identification of invasive insect pest species at any given time and for all the developmental stages is important in the aspect of biosecurity and quarantine purpose. The species-specific markers require only conventional PCR (with degenerate primer sequences) which is readily available, rapid and inexpensive for molecular identification of species in question. Out of 12 primer sets identified each for B. tau and B. zonata. One primer set, viz. Rebi(BT)- F & Rebi(BT)- R and Rebi(BZ)- F & Rebi(BZ)- R, could successfully identify B. tau and B. zonata, respectively,(Table 2; Fig. 2). These species-specific markers amplified an expected fragment size of 280 bp and 498 bp for B. tau and B. zonata, respectively. Cloning and sequencing of these products and BLAST search of these sequences shows a maximum hit for the respective species only, which is already sequenced and submitted to NCBI-GenBank. Similarly, same size bands were obtained from test fruit fly specimens collected from mango and guava and was corroborated with morphological markers.

The validation of these species-specific markers, which were developed in the present study, were carried out by employing PCR using both these primers and genomic DNA isolated from the following Bactrocera species, viz., B. correcta, B. caryae, B. dorsalis, and B. cucurbitae, which

![Fig. 2. Validation of species-specific markers for B. zonata and B. tau (M - 100bp DNA ladder, (Thermo Scientific- Fermentas) 1 & 2 - PCR amplified product (COX-1) B. zonata and B. tau respectively, 3 - B. zonata- specific marker, 4 - B. tau- specific marker.](https://bioone.org/journals/Florida-Entomologist/2011/94/December/1030/images/Fig2.jpg)
were morphologically identified and collected from IARI, New Delhi. None of the PCR reactions produced any amplification, including non-specific amplicons, where the PCR mix and PCR cycling conditions were kept same, including the annealing temperature, viz., 62 °C for 45 s, for all the reactions. However the applicability of these primers on other populations of *B. tau* & *B. zonata* within and outside India depend on the variation in the nucleotide sequences both in forward (274-298 and 58-83 for *B. tau* and *B. zonata*, respectively) and reverse primer binding regions (529-550 and 529-556 for *B. tau* and *B. zonata*, respectively). Comparison of forward primer binding regions for *B. tau* (GU323776) with only 1 existing COX-I (658 bp) deposit showed that there were variations at 2 positions viz., at the 276th position 1 accession had T instead of C, and at the 278th position 1 accession had T instead of C. Similarly examination of the reverse primer binding region showed that there was no variation in any of the nucleotide positions. A comparison of the forward primer binding region for *B. zonata* (GU323780) with 4 other NCBI accessions showed that there were variations at 8 positions, viz., at the 59th position 1 accession had G instead of T; at the 60th and 67th positions 1 accession had A instead T; at the 64th and 76th position 1 accession had C instead of T & A, respectively; at the 74th and 79th position 1 accession had T instead of C & A, respectively; at the 73rd position 1 accession had G and another having T instead of A. Similarly in the reverse primer binding region, variations were found at the 529th, 530th and 539th positions, where 2 accessions had T instead of C. In this regard, development of degenerate primers would be a valuable tool in identifying the other populations of *B. tau* and *B. zonata*. Other criteria to be taken into consideration in developing a species-specific marker for *Bactrocera* species are intra- and inter-specific variations (Bayar et al. 2002; Brunner et al. 2004). Color morphs and size variations in *Bactrocera* species are influenced by many factors, which could be impediments for species identification. Hence molecular identification using species-specific markers is an advantage where there is polymorphism in the target species. Development of species-specific markers for *B. tau* and *B. zonata* would be of immense value to identify these invasive pests at any developmental stages like egg, larva, pupa or adult. Hence, the species-specific markers developed in this study could be used for the identification of *B. tau* and *B. zonata* in any stage of development without even sequencing. The species-specific marker is also available tool when there is a problem of polymorphism as it has been shown in the case of *Ceratitis capitata* (Wiedemann) and *Anastrepha fraterculus* (Wiedemann) (Sonvico et al. 1996). In addition to the above, molecular identification could reveal multiple infestations as against the morphological identification, which could be used to separate the species (Barbosa et al. 2005).

In recent years mitochondrial DNA has become the common molecular marker in phylogenetic analysis and population genetic studies in animals (Langor & Sperling 1997), because it has the advantage that rare mutations create new haplotypes. Therefore, 2 individuals that share the same haplotypes are likely to have a common ancestor (Li 1997). Mitochondrial DNA has also been used in analyze phylogenetic relationships among tephritid fruit fly species (Smith et al. 2003, 2005; Zhang et al. 2010), because the COX-I sequences provide slightly better resolution, and also quantitative support in terms of bootstrap and divergence values for species-level identification than was previously possible (Armstrong 1997). Based on the mitochondrial DNA sequence data, the phylogenetic relationship of some tephritid taxa has been resolved (Han 2000) especially at the generic level (Smith & Bush 1997). In this connection, we used a nucleotide sequence of 658 bp variable region of the mitochondrial cytochrome oxidase I (COX-I), for constructing the phylogenetic relationship among the five subgenera of *Bactrocera*. The aligned dataset consisted of 600 nucleotides with 193 variable sites which were parsimoniously informative. The heuristic MP search resulted in 4 equally parsimonious trees with 995 steps (CI = 0.360, RI = 0.511, HI = 0.639). All of the parsimonious trees were similar in topology and one of the trees is shown in Figure 3. The best model of nucleotide substitution according the AIC was GTR + I + G. The Bayesian phylogenetic analysis implementing GTR + I + G resulted in a consensus tree similar in topology to the parsimony analysis except for minor changes in not well supported nodes (figure not shown). The 30 ingroup taxa belonging to *Bactrocera* subgenus. *Bactrocera* were retrieved as a monophyletic group with good statistical support at the nodes (BS = 89% and PP = 1.0: Fig. 3). COX-I sequences of the same region for *Ceratitis capitata* and *Anastrepha ludens* were used in the analysis as out groups.

Researchers have proposed a phylogenetic analysis of the *Bactrocera* subgenus groupings based on morphological characters (Drew & Hancock 2000; White 2000). Muraji & Nakahara (2001) used mitochondrial DNA sequences of 18 *Bactro- cera* species in 4 subgenera, indicated that *B. (Bactrocera)* was paraphyletic and *B. (Zeugodac- cus)* was monophyletic. Drew (1989) divided the subgenera of *Bactrocera* into 4 groups, viz. *Bactrocera* group, *Queenslandacus* group, *Zeugodac- cus* group and *Melanodacus* group. There are many views on the phylogeny of the subgenus *Bactrocera* whether it is monophyletic or para- phyletic (Zhang et al. 2010). In this regard, White (2000) reported that *Bactrocera* is paraphyletic in
both weighted and unweighted analysis, which was also supported by the work of Muraji & Nakahara (2001) and Zhang et al. (2010). On the contrary, Smith et al. (2003) reported that the subgenus Bactrocera is monophyletic based on the analyses of mitochondrial DNA sequences. In the present study, by employing both maximum parsimony and likelihood analyses (as implemented in MrBayes) we have shown that the subgenus Bactrocera is monophyletic (Fig. 3).

From the above phylogenetic analysis, our result supports the morphological group classifica-

![Fig. 3. One of the four equally parsimonious trees obtained from MP analysis of the COX-I dataset. Bayesian posterior probabilities (PP) > 0.9 are indicated above the branches and bootstrap > 50% are indicated below the branches. Ceratitis capitata (DQ116371) and Anastrepha ludens (DQ116207) were used as out groups.](image-url)
tion: Bactrocera group (Bactrocera) and Zeugodacus group (Zeugodacus + Austrodacus + Notodacus) (Drew & Hancock 2000). This study showed that the subgenus Zeugodacus is not paraphyletic, since Austrodacus is not located within the Zeugodacus clade because there is a 100% bootstrap separating them. Here the result was against Smith et al. (2003), who proposed that the subgenus Zeugodacus is paraphyletic. Whereas Muraji & Nakahara (2001) reported that the subgenus Zeugodacus is a monophyletic clade. The subgenus Daculus represented by B. (Daculus) oleae, shows different classification positions with maximum parsimony and likelihood analyses (as implemented in MrBayes). Drew (1989) suggested B. (Daculus) oleae belongs to the Melanodacus group and did not fall into the Bactrocera group or Zeugodacus group. Whereas Smith et al. (2005) indicated that B. (Daculus) oleae is a sister group to B. (Bactrocera) group. Our study indicated subgenera Daculus and Notodacus are a single lineage by itself, and the subgenus Austrodacus is closely related to the subgenus Zeugodacus (Fig. 3).

Considering the pest status, it is necessary to analyze the molecular diversity in all 4 important Indian Bactrocera species viz. B. tau, B. correcta, B. zonata and B. dorsalis. For molecular diversity analysis, all the corresponding species sequences were acquired from NCBI-GenBank. The phylogram for the 19 NCBI accessions of B. dorsalis revealed that there were 2 major groups, namely group I, which represents the B. dorsalis population from India, Taiwan, China, New Zealand, Tanzania and Vietnam, while the Group II is the population from USA (Fig. 4); whereas analysis of the 7 available NCBI accessions of B. zonata revealed that Group I represents India, Russia, Africa and Egypt, while group II is the population from Pakistan (Fig. 5). For B. correcta detailed information is lacking, nevertheless the phylogram for the available accessions is shown in Fig. 6.

In the present study we showed the utility of species-specific markers which can be used for the species identification of B. tau and B. zonata in any of the developmental stages. Thus it will help in timely, accurate and stage non-limiting identification of these 2 species of fruit flies, which will in turn help for better quarantine purpose. Our results indicated that the COX-I gene is useful for inferring the phylogeny of Bactrocera taxa included in the present study. Clearly more taxa must be analyzed and more data, particularly from genes that show sufficient information vari-

Fig. 4. Maximum- Parsimony (MP) tree (MEGA 4.0) with bootstrap support (1000 replicates) showing clustering of B. dorsalis species for COX-1 sequences. Two distinct clades can be seen in which, the Indian populations of B. dorsalis clustered with the Taiwan, Vietnam, Tanzania, China and New Zealand population and USA populations stand as a separate clade with 100% bootstrap support. Ceratitis capitata (DQ116371) and Anastrepha ludens (DQ116207) were used as out groups.
ation to resolve some of the internal nodes, are needed (Smith et al. 2003). However our maximum parsimony and Bayesian analysis suggested that the subgenus *Bactrocera* is monophyletic.

**ACKNOWLEDGMENTS**

Authors gratefully acknowledge the financial support received from the Indian Council of Agricultural Research (ICAR), New Delhi through the XIth Plan Network Project on Insect Biosystematics (NPIB). Our sincere thanks are due to the anonymous reviewers, whose critical comments helped in shaping up of this article. Technical and administrative support received from the Indian Institute of Horticultural Research (IIHR), Bangalore is also acknowledged.

**REFERENCES CITED**


---

**Fig. 5.** Maximum Parsimony (MP) tree (MEGA. 4.0) with bootstrap support (1000 replicates) showing clustering of *B. zonata* species for COX-I sequences from various countries. *Ceratitis capitata* (DQ116371) and *Anastrepha ludens* (DQ116207) were used as out groups.

**Fig. 6.** Maximum Parsimony (MP) tree (MEGA. 4.0) with bootstrap support (1000 replicates) showing clustering of *B. correcta* species for COX-I sequences from India and Thailand. *Ceratitis capitata* (DQ116371) and *Anastrepha ludens* (DQ116207) were used as out groups.


