

A Patch Hath Smaller Patches: Delineating Ecological Neighborhoods for Parasites

Authors: Zelmer, Derek A., and Seed, John R.

Source: Comparative Parasitology, 71(2): 93-103

Published By: The Helminthological Society of Washington

URL: https://doi.org/10.1654/4136

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

A Patch Hath Smaller Patches: Delineating Ecological Neighborhoods for Parasites

DEREK A. ZELMER^{1,3} AND JOHN R. SEED²

ABSTRACT: Use of the host individual as a boundary for parasite populations and communities provides an unambiguous spatial unit that is useful for pattern description, but this framework precludes consideration of the host landscape and within-host population dynamics. Recognizing host individuals as spatially and temporally complex landscapes requires modified concepts of parasite populations and communities. An outline of the currently accepted hierarchies of parasite populations and communities is provided on the basis of ecological neighborhoods that are delineated by discrete habitat patches or functional dynamics (or both), as opposed to host individuals. This parasite-based framework accommodates consideration of both within- and among-host dynamics and facilitates investigation into the mechanisms by which these 2 levels of investigation interact.

KEY WORDS: parasite, ecology, neighborhood, scale, population, community, dynamics.

The appropriate extent and grain (sensu Wiens, 1989) of any ecological investigation must encompass both a time scale appropriate to the process being investigated and a spatial extent determined by an organism's sphere of influence on the process during that time period (Addicott et al., 1987). This longstanding concept (von Uexküll, 1921; Hutchinson, 1953) is formalized by Addicott et al. (1987) as an "ecological neighborhood." Viewing parasitism as an interaction between an organism and its habitat (Leukhart, 1879 and Filipchenko, 1937, as cited in Dogiel, 1964; Zelmer, 1998) facilitates unique tests of current ecological theory, especially metapopulation (Levins, 1969, 1970; Hanski, 1994, 1996) and metacommunity theory (Hubbell, 2001), and affords extrapolation from host-parasite systems to more general ecological patterns and processes. Hosts represent well-defined and almost perfectly replicated habitats that harbor intrinsically hierarchical populations and communities of parasites (Esch and Fernandez, 1993), facilitating the type of multiscale, among-system contrasts of matched habitats that are exceptionally difficult in other natural systems. Unfortunately, the convenience of the unambiguous boundary of the host individual (Aho and Bush, 1993) has led to the arbitrary designation of host individuals and populations as appropriate ecological neighborhoods. Although the vagaries of adopting arbitrary temporal and spatial sampling units have been well documented in population and community investiga-

tions in free-living systems (Brown and Kodric-Brown, 1977; Connell and Sousa, 1983; Connor et al., 1983; Cunningham, 1986; Addicott et al., 1987), the potential effects of similar practices on current interpretations of the ecology of parasitic species have not been subjected to thorough examination.

Constraining the infrapopulation concept to individuals of a species in an individual host at a particular time (Margolis et al., 1982; Bush et al., 1997) rather than focusing on the actual ecological unit of an interacting parasite population is convenient for describing patterns of parasite distributions and abundance but ignores how the host landscape might affect the processes responsible for producing those ecological patterns. For parasites compartmentalized into reasonably distinct subpopulations within a host individual, either by stage (e.g., the exerythrocytic stages of Plasmodium spp.) or by physiological requirements (e.g., localization of Trypanosoma cruzi in cardiac or smooth muscle), the changes occurring within each subpopulation and the connectivity among subpopulations must be evaluated to understand the within-host dynamics of a single species and better understand the among-host dynamics.

For example, African trypanosomes infect a wide variety of mammalian hosts (Ashcroft, 1959; Ashcroft et al., 1959) and insect (*Glossina* spp.) vectors (Molyneux and Ashford, 1983; Leak, 1999), undergoing a series of dynamic population changes within the mammalian hosts (Molyneux and Ashford, 1983). Peak parasitemia is reached 5 to 7 d after inoculation, at which point all available extracellular sites and,

¹ Department of Biological Sciences, Emporia State University, Emporia, Kansas 66801, U.S.A. (e-mail: Zelmerde@esumail.emporia.edu) and

² Department of Epidemiology, CB# 7400, McGavran-Greenberg Hall, School of Public Health, The University of North Carolina, Chapel Hill, North Carolina 27599-7400, U.S.A.

³ Corresponding author.

perhaps, selected intracellular sites (e.g., brain tissue) are occupied by trypanosomes (Ormerod and Venkatesan, 1971; Jenni et al., 1983; Stoppini et al., 2000). At this point, growth of the trypanosome population is checked by density (or quorum) sensing and subsequent transformation of trypanosomes into a nondividing stage. Ultimately, the host immune response clears the bulk of the trypanosome population. What remains is a minor population of antigenically distinct trypanosomes that reproduce and repopulate the host, repeating the cycle of infection. Depending on the host involved and the species of trypanosome, these cyclic fluctuations in parasitemia can continue for weeks or years. This simplified description of African trypanosome in vivo ecology is based on monitoring of parasitemia at the level of the host individual. The actual patterns and processes of these population dynamics are far more complex.

Isolation of trypanosomes from sites other than the blood has demonstrated that both abundance and surface variant antigens (variant antigen types or VATs) differ among trypanosome subpopulations within a single host individual (Seed and Effron, 1973; Seed et al., 1984). In addition to the genotypic differences that occur among the VAT subpopulations, 2 distinct stages occur within the mammalian host. The "long slender" stage divides rapidly to populate the host but has limited infectivity for the insect vector. In contrast, the "short stumpy" stage does not divide but is infective to the insect vector. Thus, there are stage-structured, genetically disparate subpopulations (VATs) of trypanosomes, with apparently independent dynamics, occupying distinctly different foci within the host habitat at any given time. The complexity and dynamics of the mechanisms revealed when the grain of investigation is focused on ecological neighborhoods of trypanosome infrapopulations are obscured when infections are considered only at the host level.

Empirical evidence suggests that the phenomenon of multiple neighborhoods within a single host can be applied across a broad range of parasitic taxa. It has long been recognized that high rates of mutation can produce genetically distinct subpopulations of viruses and bacteria within host individuals (Eggers and Tamm, 1965; Holland et al., 1982; Sedivy et al., 1987; Dominingo and Holland, 1988; Eigen and Biebricher, 1988; Smith and Palese, 1988; Steinhauer et al., 1989; de la Torre and Holland, 1990; de la Torre et al., 1990) and that significant ecological and immunological interactions can occur among these coexisting strains (Berchieri and Barrow, 1990; Sernicola et al., 1999; Barrow and Page, 2000; Lipsitch et al., 2000).

The potential for similar metapopulation dynamics among the protozoa is not restricted to the trypanosomes (Read and Taylor, 2001). Species of Plasmodium also demonstrate variable antigenic types (Brown and Brown, 1965; McLean et al., 1982; Hommel et al., 1983; Handunetti et al., 1987; Fandeur et al., 1995) in addition to the within-host genetic diversity that arises as a product of sexual reproduction and continual colonization (Daubersies et al., 1996; Mercereua-Puijanon, 1996; Bruce and Day, 2003) with dramatic effects on the outcome of infection (Hargreaves et al., 1975; Snounou et al., 1989). It is likely that similar dynamics also occur in metazoan parasite systems where stages are separated spatially within a single host (e.g., schistosomes and ascarids) or reproduce within a host (e.g., digenean parthenitae and taeniid metacestodes).

The importance of genetically variable (Saag et al., 1988; Parry et al., 1990; Wolfs et al., 1990; Nowak et al., 1991; de Boer and Boerlijst, 1994; Mittler et al., 1995), spatially structured (Embretson, Zupancic, Ribas et al., 1993; Pantaleo et al., 1993) subpopulations and refugia (Embretson, Zupancic, Beneke et al., 1993; Antia et al., 1996; Pilyugin et al., 1997) in promoting the persistence of viral and bacterial infections also has been demonstrated on theoretical grounds. Although most theoretical considerations of within-host dynamics explicitly (Mittler et al., 1996; Pilyugin et al., 1997; Austin et al., 1998) or implicitly (Bonhoeffer and Nowak, 1994; Antia et al., 1996; Antia and Lipsitch, 1997; Hoshen et al., 2000) assume the host environment to be a "well-mixed bucket" (Mittler et al., 1996), there are significant limitations on the explanatory power of models that do not incorporate a compartmentalized host model (Antia and Halloran, 1996). The host organism represents a complex landscape of habitats (Smith and Holt, 1996) and demands a framework that considers populations at a scale smaller than the host individual. Such a framework requires a neighborhood approach to the concept of an infrapopulation (sensu Margolis et al., 1982; Bush et al., 1997) to accommodate investigation of the dynamics of parasitic infections.

Populations and communities can be delineated spatially and functionally depending on whether one is concerned with patterns or processes. Based solely on the description of pattern, a population can be delineated as a temporally and spatially contiguous set of individuals of a particular species. However, understanding the mechanisms that govern the abundance of a species at a particular locality requires consideration of a population as a functioning, interacting unit. Unlike pattern description, the ecological neighbor-

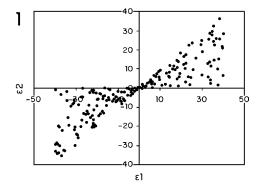
hood of a population is not simply measured but rather is determined by the extent of gene flow and the potential for positive and negative interactions. Recalling the earlier African trypanosome example, the observation that there are genetically and ecologically asynchronous dynamics in different sites within the same host individual suggests that there are discrete populations within a single host.

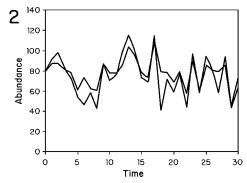
One obvious solution is to delineate infrapopulations and therefore infracommunities according to the habitat patches occupied by reasonably distinct subpopulations of parasites. The primary goal of the work presented here is to establish just what constitutes an ecological neighborhood for parasitic organisms. Although the work focuses on within-host population and community dynamics, among-host dynamics are addressed, where appropriate, both to clarify ecological concepts and to demonstrate that a functional delineation of populations and communities crosses theoretical boundaries between different levels of ecological investigation.

ECOLOGICAL NEIGHBORHOODS OF PARASITE POPULATIONS

Evaluating the host individual as a relevant scale for parasite infrapopulations requires quantification of infrapopulation dynamics at a grain finer than that circumscribed by the host individual to determine whether spatially disjunct parasitic individuals function as a panmictic infrapopulation or as a set of relatively discrete populations or demes. The connectivity among infrapopulations within a single host individual (i.e., the spatial component of the ecological neighborhood) can be evaluated in a discrete fashion by monitoring genotypes (as described for the VATs of the African trypanosomes). Alternatively, emergent properties of infrapopulations can be used to assess the spatial extent of ecological neighborhoods by evaluating the degree to which the dynamics of the infrapopulations are synchronous.

Moran (1953) postulated that the correlation between the changes in abundance of spatially separated populations with similar population growth rates is equivalent to the correlation between the stochastic variations in their environments. Thus, spatially separated groups of individuals of a single species of parasite demonstrating synchronous within-host dynamics (Figs. 1, 2) either are responding to the same scale of environmental changes (Moran, 1953; Royama, 1992; Ranta et al., 1995; Haydon and Steen, 1997) or are interconnected by sufficient dispersal to allow the sum of the infrapopulations to

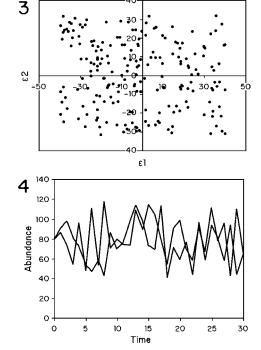




Figures 1, 2. Synchronous population dynamics for 2 populations produced by a discrete logistic growth model with an intrinsic rate of increase of 0.5 individuals per unit time and a carrying capacity of 80 individuals. At each time step, a stochastic event (£1) alters the abundance of the first population by -40 to 40 individuals. This random change is multiplied by a random number between 0 and 1 to determine the change in the second population (£2). **1.** Correlation between stochastic events affecting the 2 populations. **2.** Resultant synchronous population dynamics.

experience the "average" host environment (Holmes et al., 1994; Molofsky, 1994; Bascompte and Solé, 1998; Kendall et al., 2000). In either case, the biology of the situation suggests that delineating the infrapopulation at the level of the host individual is justified. In contrast, asynchronous dynamics (Figs. 3, 4) are indicative of either disparate population growth rates or a response to stochastic environmental changes at a grain finer than that delineated by the host individual. One could reasonably conclude that such asynchronous, spatially disjunct infrapopulations were behaving as functionally distinct infrapopulations, but patterns require cautious interpretation because the interaction between host immune responses and parasite density dependence can produce complete asynchrony among serotypes within a single host (Bruce and Day, 2003).

The host individual might not be the smallest



Figures 3, 4. Asynchronous population dynamics for 2 populations produced by a discrete logistic growth model with an intrinsic rate of increase of 0.5 individuals per unit time and a carrying capacity of 80 individuals. At each time step, a stochastic event (£1) alters the abundance of the first population by -40 to 40 individuals, and an independent stochastic event (£2) alters the abundance of the second population by -40 to 40 individuals. **3.** Uncorrelated stochastic events affecting the 2 populations. **4.** Resultant asynchronous population dynamics.

relevant habitat unit of a parasite infrapopulation; nonetheless, it does provide an unambiguous boundary (Bush et al., 1997). For parasites exhibiting disjunct within-host dynamics, the host individual constitutes an ecological neighborhood for investigations of metapopulation dynamics. Essentially, a metapopulation of free-living organisms can be considered in 2-dimensional space as a distribution of organisms within suitable habitat patches that are surrounded by unsuitable habitat but interconnected by dispersal (Levins, 1969, 1970; Hanski, 1994, 1996). This model is appropriate for the within-host dynamics of parasites that reproduce in stage-independent, disjunct habitats within a host individual and for parasites with direct transmission, where host individuals comprise the patches of suitable habitat (Fig. 5).

For parasite species with complex life cycles, the model is more complex. Instead of a 2-dimensional distribution of host patches, host distributions are

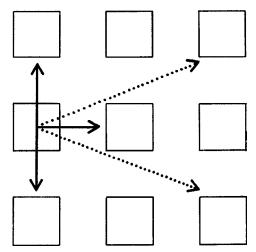


Figure 5. Two-dimensional representation of direct transmission of a parasite between individuals (squares) in a host population. Solid arrows represent transmission probabilities that would be high in hosts with low vagility. Broken arrows represent transmission probabilities that only would be high in hosts with high vagility.

envisioned as a vertical series of horizontal planes (i.e., host distributions are "stacked"; upon one another [Fig. 6]). Each horizontal plane represents a host species or group of species in which the individuals of a given parasite species share a particular ontological sequence determined by the nature of the habitat (i.e., they are individuals of the same stage or set of stages). Thus, the individual parasites inhabiting a single horizontal plane constitute a component population (sensu Bush et al., 1997) of parasites. The same model, with each horizontal plane representing a specific habitat type (e.g., organs or tissues), applies to within-host dynamics when spatial structuring of a parasite species is stage-dependent.

Conventional 2-dimensional metapopulation models assume horizontal dispersal between patches. In a 3-dimensional parasite metapopulation model, among-patch transmission occurs only in the vertical plane (i.e., from one component population to the next). Spatial structuring of the model becomes a question of which hosts on a horizontal plane can be infected by propagules generated on the plane immediately below. Given the focal nature of parasite transmission (Audrey, 1958; Pavlovsky, 1966), spatial structuring of this model is determined by the vagility of the hosts on either of the 2 planes and the infective stages of the parasites. If the motility of both host species and the infective stage of the parasite are restricted, transmission probabilities will be high only for transmission of propagules from a host to the

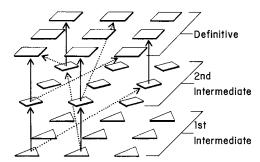


Figure 6. Three-dimensional representation of transmission of a parasite with a 3-host life cycle. Each shape represents a host individual, and each horizontal plane of shapes represents a host population. Solid arrows represent transmission probabilities that would be high when both species of host on planes connected by transmission and the dispersing stages of the parasites exhibit low vagility. Broken arrows represent transmission probabilities that only would be high when hosts on 1 or both connected planes, or the dispersing stages of the parasites, exhibit high vagility.

neighboring host on the horizontal plane directly above (solid transmission arrows of Fig. 6). When host vagility on either of the 2 interacting planes is high or the motility of the dispersing stage of the parasite connecting the 2 planes is high, the spatial structure decreases and probabilities of infecting any host in the next higher plane increase as a function of the increased probability of spatial overlap between the 2 hosts (broken transmission arrows of Fig. 6).

The means of continuing the spatial model through subsequent parasite generations requires transmission from the definitive host component population to the first intermediate host component population and depends on the life span of the hosts and parasites. If the parasite propagules from definitive hosts are likely to encounter a naive first intermediate host component population, the model continues in a noncyclic fashion by adding another series of horizontal planes (component populations), as shown in Figure 7. If there is substantial overlap in parasite generations, a cyclic model of transmission is appropriate (Fig. 8).

The same 3-dimensional models are applied to the within-host dynamics of parasites with stage-dependent spatial structuring if the planes are viewed as specific tissues or organs and the shapes as individuals within an infrapopulation. The same criteria for continuation of generations applies, with parasites whose dynamics allow for minimal overlap of generations at a site within a host following the noncyclic "stack model" (Fig. 7) and parasites with substantial overlap of generations within a host following the cyclic model (Fig. 8).

The concept of a host-delineated parasite neighbor-

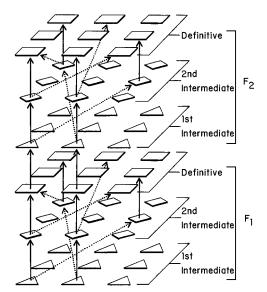


Figure 7. Three-dimensional noncyclic model of transmission of a parasite with a 3-host life cycle and non-overlapping generations. Each shape represents a host individual, and each horizontal plane of shapes represents a host population. Solid arrows represent transmission probabilities that would be high when both species of host on planes connected by transmission and the dispersing stages of the parasites exhibit low vagility. Broken arrows represent transmission probabilities that only would be high when hosts on 1 or both connected planes, or the dispersing stages of the parasites, exhibit high vagility.

hood pools the infective stages of parasites that are vectored or transmitted through trophic interactions with the reproductive stages of the parasite. Thus, the link among host individuals between component populations is not given separate consideration. Identifying all transmissible (infective) infrapopulations within a single host at 1 point in time emphasizes the fact that different infrapopulations within a host individual have very different biological properties by treating infective, dispersing stages at each point in the life cycle as distinct from vegetative and reproductive stages within a host individual. Investigations conducted at the suprapopulation level (Margolis et al., 1982) also might necessitate separate consideration of the dispersing infective stages in which case identification of all of the transmissible infrapopulations within a given component population is recommended as a theoretically useful delineation (Fig. 9).

ECOLOGICAL NEIGHBORHOODS OF PARASITE COMMUNITIES

Communities also can be considered in terms of pattern and process. Determining why a certain

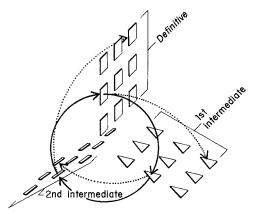


Figure 8. Three-dimensional cyclic model of transmission of a parasite with a 3-host life cycle and overlapping generations. Each shape represents a host individual, and each plane of shapes represents a host population. Solid arrows represent transmission probabilities that would be high when both species of host on planes connected by transmission and the dispersing stages of the parasites exhibit low vagility. Broken arrows represent transmission probabilities that only would be high when hosts on one or both connected planes, or the dispersing stages of the parasites, exhibit high vagility.

assemblage of organisms occurs at a particular locality is fundamentally different from a description of the emergent properties of that community. Thus, the question of whether communities should be delineated spatially (Klopfer, 1969) or in terms of energetic relations (Odum, 1959) is irrelevant: the grain of the investigation (Wiens, 1989) must be dictated by the question. At either level of investigation, the "relevant environment" of the parasites (the *Umwelt* of von Uexküll [1921]) must be given full consideration. It is clear that in many cases the smallest relevant ecological neighborhood for parasitic organisms is not the host individual.

In a biogeographical sense, a community consists of populations of different species occupying a common space at a particular time (Begon et al., 1996; Fauth et al., 1996), often restricted to specific phylogenetic groups for practical considerations (the assemblages of Fauth et al. [1996]). However, community ecology also is concerned with the emergent properties of communities. To focus attention on what cannot be explained by the sum of the parts, it has been suggested that groups of organisms with similar functional roles (the guilds of Root [1967, 1973] or the local guilds and ensembles of Fauth et al. [1996] [or both]) be the focus of community ecologists (Colwell, 1979; Tudge, 1991; Ricklefs and Miller, 1999). For certain parasitic organisms, such a resource-

based neighborhood for community processes will entail delineation of assemblages within the host landscape as opposed to among host individuals.

The ecological neighborhood for investigations into the emergent properties of parasitic communities would include species that have a direct positive or negative effect on spatially and temporally congruent species in equilibrium communities (MacArthur, 1960) (the interactive communities of Holmes and Price [1986]) or those coexisting species that are tracking shared resources in nonequilibrium communities (Wiens, 1984) (a functional subset of the isolationist communities of Holmes and Price [1986]). Thus, a community delineated by emergent properties, theoretically, can contain fewer species than a spatially delineated community within the same geographical boundaries. In equilibrium communities, one expects compensatory changes in the abundances of species with negative interactions, whereas in nonequilibrium communities, species tracking fluctuations in a shared resource should exhibit positive covariation (Schluter, 1984). These expectations facilitate the functional delineation of neighborhoods for community processes by examination of patterns of covariance among candidate species.

Patterns of covariance can be examined using a matrix of abundances (species represented by rows and census times represented by columns) to compare the sum of individual species variances to the variance of their sums (Pielou, 1972; Robson, 1972; Schluter, 1984). The variance of the sums includes the average covariation between pairs of species and is a measure of compensatory fluctuations. The ratio of this variance to the sum of the species variances (V) is a means of quantifying whether species fluctuations are independent (V = 1), congruent (V > 1), or compensatory (V < 1) (Schluter, 1984; Gotelli and Graves, 1996). Monitoring change in V as species are added to the analysis or as the scale of the neighborhood is increased should facilitate a quantitative functional delineation of both equilibrium (compensatory) and nonequilibrium (congruent) communities. It is important for this approach that species or assemblages to be excluded from the covariance analysis be predetermined based on biologically meaningful justifications (Pielou, 1972) and not as the result of "data dredging" (Selvin and Stuart, 1966).

Delineating infracommunities based on assemblages of parasite infrapopulations occupying a specific habitat patch at a particular time facilitates scaling from the level of a single cell to an organ, to an organ system, to the entire host. This delineation encompasses both spatial and functional community

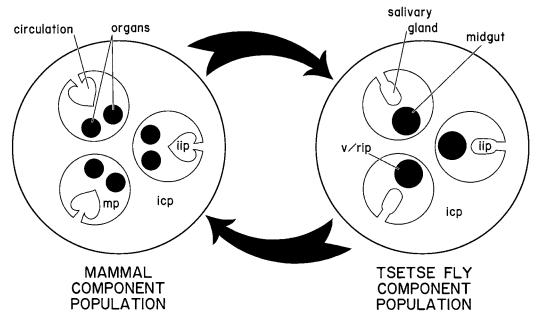


Figure 9. A diagrammatic representation of an African trypanosome suprapopulation. The largest circles delineate the mammal and tsetse component populations. The 3 circles circumscribed by each component population represent host individuals, each of which contains a metapopulation of trypanosomes (mp) comprising the trypanosome infrapopulations in the various organ systems. The heart represents the parasites occupying the host mammal's circulatory system, which constitutes the infective infrapopulation (iip). The space within the large circles representing the component populations connects the infective infrapopulations within each host, thereby representing the infective component population (icp). The midgut of the tsetse fly host represents the vegetative—reproductive infrapopulation (v/rip) within that host.

definitions, necessitating a clear and biologically meaningful demarcation of the ecological neighborhood of the community under investigation. The unambiguous host boundary of Bush et al. (1997) is maintained, but free-living stages within a particular subset of the abiotic environment (i.e., a habitat patch within a locality) also are described as an infracommunity rather than as a component community as suggested by Bush et al. (1997).

The ecological neighborhood of a component community must encompass the interactions of component populations of different species that overlap in habitat use at 1 point in time. This differentiates the free-living stages in a patch (an infracommunity) from the sum of those patches (a component community). The source community (sensu Bush et al., 1993) for a host individual, whether the infective stages are free-living or within the confines of an intermediate host, is included within the infracommunity neighborhood, whereas a host population recruits from a neighborhood that encompasses the component community. The sum of these component communities at a particular locality makes up the neighborhood of the parasite supracommunity, which

can be more restrictively defined as a collection of the suprapopulations of different species occupying a particular locality at a particular time.

CONCLUDING REMARKS

As suggested by Bush et al. (1997), the most important aspect of an ecological investigation is the unambiguous definition of the parameters of that investigation. Rather than attempt to subdivide parasite populations and communities into their smallest discernable units or depend on units of sampling convenience and geometry, herein we have described a biologically meaningful framework to delineate parasite populations and communities that is broadly applicable to the empirical population biology and community ecology of both micro- and macroparasites.

Infrapopulation and infracommunity dynamics are poorly understood and until recently have not been the focus of a great deal of research. Such investigations must proceed with the understanding that hosts do not represent a single, stable habitat. Studies of infrapopulation and infracommunity dynamics

must elucidate the degree to which the spatially and temporally complex host landscape affects infrapopulation—infracommunity and metapopulation—metacommunity dynamics within host individuals. Host-based delineation of the ecological neighborhoods encompassing parasite population and community processes cannot be applied to investigations of in vivo dynamics and can bias investigations of parasite populations and communities at larger scales. Modification of current ecological concepts to accommodate both within-host and among-host dynamics is necessary not only for the formulation of clear questions regarding in vivo dynamics but also to provide a parasite-based framework for parasite ecology in general.

ACKNOWLEDGMENTS

The ideas presented herein originated from discussions with numerous students and colleagues and were culled from a much broader pool of vague notions with the helpful suggestions of Mary Everhart, Mike Barger, John Janovy, Jr., Dennis J. Richardson, and especially Rich Clopton.

LITERATURE CITED

- Addicott, J. F., J. M. Aho, M. F. Antolin, D. K. Padilla, J. S. Richardson, and D. A. Soluk. 1987. Ecological neighbourhoods: scaling environmental patterns. Oikos 49:340–346.
- Aho, J. M., and A. O. Bush. 1993. Community richness of parasites of some freshwater fishes from North America. Pages 185–193 in R. E. Ricklefs and D. Schluter, eds. Species Diversity in Ecological Communities: Historical and Geographic Perspectives. University of Chicago Press, Chicago, Illinois.
- Antia, R., and M. E. Halloran. 1996. Recent developments in theories of pathogenesis of AIDS. Trends in Microbiology 4:282–285.
- Antia, R., J. C. Koella, and V. Perrot. 1996. Models of within-host dynamics of persistent mycobacterial populations. Proceedings of the Royal Society of London Series B 263:257–263.
- Antia, R., and M. Lipsitch. 1997. Mathematical models of parasite responses to host immune defenses. Parasitology 115:S155–S167.
- Ashcroft, M. J. 1959. The importance of wild animals as reservoirs for trypanosomiasis. East African Medical Journal 36:289–297.
- Ashcroft, M. J., E. Burtt, and H. Fairbairn. 1959. The experimental infection of some African wild animals with *Trypanosoma rhodesiense*, *T. brucei*, and *T. congolense*. Annals of Tropical Medicine and Parasitology 53:147–161.
- Audrey, J. R. 1958. The localization of disease with special reference to the zoonoses. Transactions of the Royal Society of Tropical Medicine and Hygiene 52:308– 334.

- Austin, D. J., N. J. White, and R. M. Anderson. 1998. The dynamics of drug action on the within-host population growth of infectious agents: melding pharmokinetics with pathogen population dynamics. Journal of Theoretical Biology 194:313–339.
- Barrow, P. A., and K. Page. 2000. Inhibition of colonisation of the alimentary tract in young chickens with *Campylobacter jejuni* by pre-colonisation with strains of *C. jejuni*. FEMS Microbiology Letters 182:87–91.
- Bascompte, J., and R. V. Solé. 1998. Monitoring Spatiotemporal Dynamics in Ecology. Springer, Berlin, Germany. 230 pp.
- Begon, M., J. L. Harper, and C. R. Townsend. 1996. Ecology, 3rd ed. Blackwell Science, Oxford, U.K. 1092 pp.
- Berchieri, A., Jr., and P. A. Barrow. 1990. Further studies on the inhibition of colonization of the chicken alimentary tract with *Salmonella typhimurium* by precolonization with an avirulent mutant. Epidemiology and Infection 104:427–441.
- Bonhoeffer, S., and M. A. Nowak. 1994. Mutation and the evolution of virulence. Proceedings of the Royal Society of London Series B 258:133–140.
- Brown, K. N., and I. N. Brown. 1965. Immunity to malaria: antigenic variations in chronic infections of *Plasmodium knowlesi*. Nature 208:1286–1288.
- **Brown, J. H., and A. Kodric-Brown.** 1977. Turnover rates in insular biogeography: effect of immigration on extinction. Ecology 58:445–449.
- Bruce, M. C., and K. P. Day. 2003. Cross-species regulation of *Plasmodium* parasitemia in semi-immune children from Papua New Guinea. Trends in Parasitology 19:271–277.
- Bush, A. O., R. W. Heard, Jr., and R. M. Overstreet. 1993. Intermediate hosts as source communities. Canadian Journal of Zoology 71:1358–1363.
- Bush, A. O., K. D. Lafferty, J. M. Lotz, and A. W. Shostak. 1997. Parasitology meets ecology on its own terms: Margolis et al. revisited. Journal of Parasitology 83:575–583.
- Colwell, R. K. 1979. Toward a unified approach to the studies of species diversity. Pages 75–91 in G. P. Patil, W. Smith, and C. Taillie, eds. Ecological Diversity in Theory and Practice. International Cooperative Publishing House, Fairland, Maryland. 365 pp.
- Connell, J. H., and W. P. Sousa. 1983. On the evidence needed to judge ecological stability or persistence. American Naturalist 121:789–824.
- Connor, E. F., S. H. Faeth, and D. Simberloff. 1983. Leaf-miners on oak: the role of immigration and in situ reproductive requirement. Ecology 64:191–204.
- Cunningham, M. A. 1986. Dispersal in white-crowned sparrows: a computer simulation of the effect of studyarea size on estimates of local recruitment. Auk 103: 79–85.
- Daubersies, P., S. Sallenave-Sales, S. Magne, J. F. Trape,
 H. Contamin, T. Fandeur, C. Rogier, O. Mercereau-Puijalon, and P. Druilhe. 1996. Rapid turnover of Plasmodium falciparum populations in asymptomatic individuals living in a high transmission area. American Journal of Tropical Medicine and Hygiene 54:
- de Boer, R. J., and M. C. Boerlijst. 1994. Diversity and virulence thresholds in AIDS. Proceedings of the

- National Academy of Sciences of the United States of America 91:544–548.
- de la Torre, J. C., and J. J. Holland. 1990. RNA virus quasispecies populations can suppress vastly superior mutant progeny. Journal of Virology 64:6278–6281.
- de la Torre, J. C., E. Wimmer, and J. J. Holland. 1990. Very high frequency of reversion to guanidine resistance in clonal pools of guanidine-dependent type 1 poliovirus. Journal of Virology 64:664–671.
- Dogiel, V. A. 1964. General Parasitology. Oliver and Boyd, London, U.K. 516 pp.
- Dominingo, E., and J. J. Holland. 1988. High error rates, population equilibrium, and evolution of RNA replication systems. Pages 3–36 in E. Dominingo, J. J. Holland, and P. Ahlquist, eds. RNA Genetics. Vol. III. Variability of RNA Genomes. CRC Press, Boca Raton, Florida.
- Eggers, H. J., and I. Tamm. 1965. Coxsackie A9 virus: mutation from drug dependence to drug independence. Science 148:97–98.
- Eigen, M., and C. K. Biebricher. 1988. Sequence space and quasispecies distribution. Pages 211–245 *in* E. Dominingo, J. J. Holland, and P. Ahlquist, eds. RNA Genetics. Vol. III. Variability of RNA Genomes. CRC Press, Boca Raton, Florida.
- Embretson, J., M. Zupancic, J. Beneke, M. Till, S. Wolinsky, J. L. Ribas, A. Burke, and A. T. Haase. 1993. Analysis of human immunodeficiency virus-infected tissues and in situ hybridization reveals latent and permissive infections at single-cell resolution. Proceedings of the National Academy of Sciences of the United States of America 90:357–361.
- Embretson, J., M. Zupancic, J. L. Ribas, A. Burke, P. Racz, K. Tenner-Racz, and A. T. Haase. 1993. Massive covert infection of helper T lymphocytes and macrophages by HIV during the incubation period of AIDS. Nature 362:359–362.
- **Esch, G. W., and J. C. Fernandez.** 1993. A functional Biology of Parasitism. Chapman and Hall, London, U.K. 337 pp.
- Fandeur, T., C. Le Scanf, B. Bonnemains, C. Slomianny, and O. Mercereau-Puijalon. 1995. Immune pressure selects for *Plasmodium falciparum* parasites presenting distinct red blood cell surface antigens and inducing strain-specific protection in *Saimiri* monkeys. Journal of Experimental Medicine 181:283–295.
- Fauth, J. E., J. Bernardo, M. Camara, W. J. Resetarits, Jr., J. Van Buskirk, and S. A. McCollum. 1996. Simplifying the jargon of community ecology: a conceptual approach. American Naturalist 147:282–286.
- Gotelli, N. J., and G. R. Graves. 1996. Null Models in Ecology. Smithsonian Institution Press, Washington, D.C. 368 p.
- Handunetti, S. H., K. N. Mendis, and P. H. David. 1987. Antigenic variation of cloned *Plasmodium fragile* in its natural host *Macaca sinica*. Journal of Experimental Medicine 165:1269–1283.
- Hanski, I. 1994. Patch occupancy dynamics in fragmented landscapes. Trends in Ecology and Evolution 9: 131–135.
- Hanski, I. 1996. Metapopulation ecology. Pages 13–43 in O. E. Rhodes, Jr., R. K. Cheeser, and M. H. Smith, eds. Population Dynamics in Ecological Space and Time. University of Chicago Press, Chicago, Illinois. 388 pp.

- Hargreaves, B. J., M. Yoeli, R. S. Nussenzweig, D. Walliker, and R. Carter. 1975. Immunological studies in rodent malaria. I. Protective immunity induced in mice by mild strains of *Plasmodium berghei yoelii* against a virulent and fatal line of this plasmodium. Annals of Tropical Medicine and Parasitology 69: 289–299.
- Haydon, D., and H. Steen. 1997. The effects of largeand small-scale random events on the synchrony of metapopulation dynamics: a theoretical analysis. Proceedings of the Royal Society of London B Biological Sciences 264:1375–1381.
- Holland, J., K. Spindler, F. Horodyski, E. Grabau, S. Nichol, and S. VandePol. 1982. Rapid evolution of RNA genomes. Science 215:1577–1585.
- Holmes, E. E., M. A. Lewis, J. E. Banks, and R. R. Viet. 1994. Partial differential equations in ecology: spatial interactions and population dynamics. Ecology 75: 17–29.
- Holmes, J. C., and P. W. Price. 1986. Communities of parasites. Pages 187–213 in J. Kikkawa and D. J. Anderson, eds. Community Ecology: Patterns and Processes. Blackwell Scientific Publications, Melbourne, Australia.
- Hommel, M., P. H. David, and L. D. Oligino. 1983. Surface alterations of erythrocytes in *Plasmodium falciparum* malaria. Antigenic variation, antigenic diversity, and the role of the spleen. Journal of Experimental Medicine 137:1137–1148.
- Hoshen, M. B., R. Heinrich, W. D. Stein, and H. Ginsburg. 2000. Mathematical modelling of the within-host dynamics of *Plasmodium falciparum*. Parasitology 121:227–235.
- Hubbell, S. P. 2001. The Unified Neutral Theory of Biodiversity and Biogeography. Monographs in Population Biology 32. Princeton University Press, Princeton, New Jersey, U.S.A. 375 pp.
- Hutchinson, G. E. 1953. The concept of pattern in ecology.Proceedings of the National Academy of Sciences of the United States of America 105:1–12.
- Janovy, Jr., J., R. E. Clopton, and T. J. Percival. 1992.
 The roles of ecological and evolutionary influences in providing structure to parasite species assemblages.
 Journal of Parasitology 78:630–640.
- Jenni, L., W. Rudin, and K. F. Schell. 1983. Extravascular and vascular distribution of *Trypanosoma b. gambiense* in *Microtus montanus* after cyclical transmission. *In P.* L. Gigase and E. A. Van Marck, eds. From Parasitic Infection to Parasitic Disease. Contributions to Microbial Immunology 7:95–102.
- Kendall, B. E., O. N. Bjørnstad, J. Bascompte, T. H. Keitt, and W. F. Fagan. 2000. Dispersal, environmental correlation, and spatial synchrony in population dynamics. American Naturalist 155:628–636.
- Klopfer, P. H. 1969. Habitats and Territories: A Study of the Use of Space by Animals. Basic Topics in Comparative Psychology 465. Basic Books Inc., New York. 117 pp.
- **Leak, S. G. A.** 1999. Tsetse Biology and Ecology: Their Role in the Epidemiology and Control of Trypanosomiasis. CABI Publishing, New York. 568 pp.
- **Levins, R.** 1969. Some demographic and genetic consequences of environmental heterogeneity for biological

- control. Bulletin of the Entomological Society of America 15:237–240.
- Levins, R. 1970. Exinction. Pages 75–107 in M. Gerstenhaber, ed. Some mathematical questions in biology. Lecture notes on mathematics in the life sciences. Annals of the New York Academy of Sciences, Vol. 231.
- Lipsitch, M., J. K. Dykes, S. E. Johnson, E. W. Ades, J. King, D. E. Briles, and G. M. Carlone. 2000. Competition among *Streptococcus pneumoniae* for intranasal colonization in a mouse model. Vaccine 18:2895–2901.
- **MacArthur, R.** 1960. On the relative abundance of species. American Naturalist 94:25–36.
- Margolis, L., G. W. Esch, J. C. Holmes, A. M. Kuris, and G. A. Schad. 1982. The use of ecological terms in parasitology. Journal of Parasitology 68:131–133.
- McLean, S. A., C. D. Pearson, and R. S. Phillips. 1982.
 Plasmodium chabaudii: relationship between the occurrence of recrudescent parasitemias in mice and the effective levels of acquired immunity. Experimental Parasitology 54:213–221.
- Mercereua-Puijanon, O. 1996. Revisiting host/parasite interactions: molecular analysis of parasites collected during longitudinal and cross-sectional surveys in humans. Parasite Immunology 18:173–180.
- Mittler, J., R. Antia, and B. Levin. 1995. Population dynamics of HIV pathogenesis. Trends in Ecology and Evolution 10:224–227.
- Mittler, J., B. R. Levin, and R. Antia. 1996. T-cell homeostasis, competition, and drift: AIDS as HIV-accelerated senescence of the immune repertoire. Journal of Acquired Immune Deficiency Syndromes and Human Retrovirology 12:233–248.
- **Molofsky, J.** 1994. Population dynamics and pattern formation in theoretical populations. Ecology 75: 30–39.
- Molyneux, D. M., and R. W. Ashford. 1983. The Biology of *Trypanosoma* and *Leishmania*, Parasites of Man and Domestic Animal. Taylor and Francis, London, U.K. 294 pp.
- Moran, P. A. P. 1953. The statistical analysis of the Canadian lynx cycle. I. Structure and prediction. Australian Journal of Zoology 1:163–173.
- Nowak, M. A., R. M. Anderson, A. R. McLean, T. F. W. Wolfs, J. Goudsmit, and R. M. May. 1991. Antigenic diversity thresholds and the development of AIDS. Science 254:963–969.
- Odum, E. P. 1959. Fundamentals of Ecology, 2nd ed. W.B. Saunders Company, Philadelphia, Pennsylvania. 546 pp.
- Ormerod, W. E., and S. Venkatesan. 1971. The occult visceral phase of mammalian trypanosomes with special reference to the life cycle of *Trypanosoma* (*Trypanozoon*) brucei. Transactions of the Royal Society of Tropical Medicine and Hygiene 65: 722–735.
- Pantaleo, G., C. Graziosi, J. F. Demarest, L. Butini, M. Montroni, C. H. Fox, J. M. Orenstein, D. P. Kottler, and A. S. Fauci. 1993. HIV infection is active and progressive in lymphoid tissue during the clinically latent stage of disease. Nature 362:355–358.
- Parry, N., G. Fox, D. Rowlands, F. Brown, E. Fry, R. Acharya, D. Logan, and D. Stuart. 1990.

- Structural and serological evidence for a novel mechanism of antigenic variation in foot-and-mouth disease virus. Nature 347:569–572.
- Pavlovsky, E. N. 1966. Natural Nidality of Transmissible Diseases. University of Illinois Press, Urbana, Illinois. 261 pp.
- **Pielou, E. C.** 1972. 2^k contingency tables in ecology. Journal of Theoretical Biology 34:337–352.
- Pilyugin, S., J. Mittler, and R. Antia. 1997. Modelling T-cell proliferation: an investigation of the consequences of the Hayflick limit. Journal of Theoretical Biology 186:117–129.
- Ranta, E., V. Kaitala, J. Lindström, and H. Lindén. 1995. Synchrony in population dynamics. Proceedings of the Royal Society of London B Biological Sciences 262:113–118.
- Read, A. F., and L. H. Taylor. 2001. The ecology of genetically diverse infections. Science 292:1099–1102.
- Ricklefs, R. E., and G. L. Miller. 1999. Ecology, 4th ed. W. H. Freeman and Company, New York. 822 pp.
- **Robson, D. S.** 1972. Appendix: statistical tests of significance. Journal of Theoretical Biology 34:350–352.
- **Root, R. B.** 1967. The niche exploitation pattern of the bluegray gnatcatcher. Ecological Monographs 37:317–350.
- **Root, R. B.** 1973. Organization of a plant-arthropod association in simple and diverse habitats: the fauna of collards (*Brassica oleracea*). Ecological Monographs 43:95–124.
- **Royama, T.** 1992. Analytical Population Dynamics. Chapman and Hall, London, U.K. 392 pp.
- Saag, M. S., B. H. Hahn, J. Gibbons, Y. Li, E. S. Parks, W. P. Parks, and G. M. Shaw. 1988. Extensive variation of human immunodeficiency virus type-1 in vivo. Nature 334:440–444.
- Schluter, D. 1984. A variance test for detecting species associations, with some example applications. Ecology 65:998–1005.
- Sedivy, J. M., J. P. Capone, U. L. RajBhandary, and P. A. Sharp. 1987. An inducible mammalian amber suppressor: propagation of a poliovirus mutant. Cell 50:379–389.
- **Seed, J. R.** 1993. Immunoecology: the origins of an idea. Journal of Parasitology 79:470–471.
- Seed, J. R., R. Edwards, and J. Sechelski. 1984. The ecology of antigenic variation. Journal of Protozoology 31:48–53.
- Seed, J. R., and H. G. Effron. 1973. Simultaneous presence of different antigenic populations of *Trypanosoma brucei gambiense* in *Microtus montanus*. Parasitology 66:269–278.
- Seed, J. R., and J. B. Sechelski. 1996. The individual host, a unique evolutionary island for rapidly dividing parasites: a theoretical approach. Journal of Parasitology 82:263–267.
- **Selvin, H. C., and A. Stuart.** 1966. Data-dredging procedures in survey analysis. American Statistician 20:20–23.
- Sernicola, L., F. Corrius, M. L. Koanga-Mogtomo, S. Baroncelli, S. DiFabio, M. T. Maggionella, R. Belli, Z. Michelini, I. Maccia, A. Cesolini, L. Cioe, P. Verani, and F. Titti. 1999. Long-lasting protection by live attenuated simian immunodeficiency virus in cynomolgus monkeys: no detection of reactivation

- after stimulation with a recall antigen. Virology 256: 291–302.
- Smith, V. H., and R. D. Holt. 1996. Resource competition and within-host disease dynamics. Trends in Ecology and Evolution 11:386–389.
- Smith, F. I., and P. Palese. 1988. Influenza viruses: High rate of mutation and evolution. Pages 123–125 in E. Dominingo, J. J. Holland, and P. Ahlquist, eds. RNA Genetics. Vol. III. Variability of RNA Genomes. CRC Press, Boca Raton, Florida.
- Snounou, G., W. Jarra, S. Viriakosol, J. C. Wood, and K. N. Brown. 1989. Use of a DNA probe to analyse the dynamics of infection with rodent malaria parasites confirms that parasite clearance during crisis is predominantly strain- and species-specific. Molecular and Biochemical Parasitology 37:37–46.
- Steinhauer, D. A., J. C. de la Torre, and J. J. Holland. 1989. High nucleotide substitution error frequencies in clonal pools of vesicular stomatitis virus. Journal of Virology 63:2063–2071.
- Stoppini, L., P.-A. Buchs, R. Brun, D. Muller, S. Duport, L. Parisi, and T. Seebeck. 2000. Infection of organotypic slice cultures from rat central nervous tissue with *Trypanosoma brucei brucei*. International Journal of Medical Microbiology 290:105–113.

- Tudge, C. 1991. Global Ecology. Oxford University Press, New York. 173 pp.
- von Uexküll, J. 1921. Umwelt und Innenwelt der Tiere, 2nd ed. Springer, Berlin, Germany. 224 pp.
- Wiens, J. A. 1984. On understanding a non-equilibrium world: myth and reality in community patterns and processes. Pages 439–457 in D. R. Strong, Jr., D. Simberloff, L. G. Abele, and A. B. Thistle, eds. Ecological Communities: Conceptual Issues and the Evidence. Princeton University Press, Princeton, New Jersey.
- Wiens, J. A. 1989. Spatial scaling in ecology. Functional Ecology 3:385–397.
- Wolfs, T. F. W., J. de Jong, H. van den Berg, J. M. G. H. Tijnagel, W. J. A. Krone, and J. Goudsmit. 1990. Evolution of sequences encoding the principle neutralization epitope of human immunodeficiency virus 1 is host dependent, rapid, and continuous. Proceedings of the National Academy of Sciences of the United States of America 87:9938–9942.
- **Zelmer, D. A.** 1998. An evolutionary definition of parasitism. International Journal for Parasitology 28: 531–533.