

High Proportion of Heterozygotes in Microsatellite DNA Loci of Wild Clonal Silver Crucian Carp, *Carassius langsdorfii*

Authors: Kenichi Ohara, Shi Dong, and Nobuhiko Taniguchi

Source: Zoological Science, 16(6) : 909-913

Published By: Zoological Society of Japan

URL: <https://doi.org/10.2108/zsj.16.909>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non-commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

High Proportion of Heterozygotes in Microsatellite DNA Loci of Wild Clonal Silver Crucian Carp, *Carassius langsdorfii*

Kenichi Ohara¹, Shi Dong² and Nobuhiko Taniguchi^{1*}

¹Laboratory of applied population genetics, Faculty of Agriculture, Tohoku University, Sendai, Miyagi 985-8555, Japan and

²Faculty of Fisheries, Agriculture College of Tianjin Xiqing, Tianjin 300381, China

ABSTRACT—The silver crucian carp, *Carassius langsdorfii* has three reproductive characteristics: gynogenesis, polyploidy (triploid or tetraploid), and genetic homogeneity within a family. In natural water, the silver crucian carp populations consist of multiple clonal lines. In the present study, three microsatellite DNA loci were used to distinguish several clonal lines of the silver crucian carp sampled from natural water. Progeny and the maternal fish had the same genotype in the three loci. In 237 fish collected from the wild, nine alleles were observed in *GF1**, sixteen alleles in *GF17**, and nine alleles in *GF29**. Ten genotypes were observed in *GF1**, seventeen in *GF17** and eight in *GF29**. The proportion of heterozygotes was very high in each locus (1.000). Sixteen clonal lines were distinguished by the combined genotypes of three microsatellite loci. Two subtypes were also detected within the clonal line KOC-011.

INTRODUCTION

The silver crucian carp, *Carassius langsdorfii* (Matsubara and Ochiai, 1965) has three reproductive characteristics: gynogenesis, polyploidy (triploid or tetraploid) (Kobayasi, 1971; Kobayasi and Ochi, 1972), and clonal nature (Dong and Taniguchi, 1996; Dong *et al.*, 1997; Umino *et al.*, 1996). Sperms of other species initiate development of their eggs (Kobayasi, 1971; Kobayasi and Ochi, 1972). However, the introduced sperm nucleus does not transform into a male pronucleus and makes no contribution to the zygotic genome (Nakakuki *et al.*, 1984; Yamashita *et al.*, 1990, 1993). A cytological study concluded that a population in the Kanto district appeared to be comprised of both triploid and tetraploid types (Kobayasi *et al.*, 1970). Thus it was suggested that the silver crucian carp is unisexual and produces unreduced polyploid eggs (Kobayasi, 1971).

Taniguchi and Sakata (1977) found that the silver crucian carp of western Japan has a specific band of muscle protein electrophoresis, which has been verified to be a creatine kinase in another study (Dong and Taniguchi, 1996). Taniguchi and Sakata (1977) also found that the silver crucian carp is independent from the gene pool of wild diploid crucian carp species. Murakami and Fujitani (1997) found that the repetitive DNA (Cal3nDr) are specific to polyploid silver crucian carp, and they suggested that this region is useful in the study of the genetic background of this species such as the origin of polyploidy and gynogenetic reproduction.

In terms of genetic homogeneity, several authors have observed that the progeny in this species are usually genetically identical to the maternal fish (Dong and Taniguchi, 1996; Dong *et al.*, 1997; Umino *et al.*, 1996). The wild silver crucian carp consist of several clonal lines (Dong *et al.*, 1996; Umino *et al.*, 1997; Ohara *et al.*, 1998). These lines have been distinguished using tissue graft, isozymes, multilocus minisatellite DNA fingerprinting and mtDNA RFLP (Murayama *et al.*, 1984; Dong *et al.*, 1996; Umino *et al.*, 1997; Ohara *et al.*, 1998). Of these methods, tissue grafts, isozymes and mtDNA RFLP are time-consuming or lack the sensitivity to identify clonal lines. By contrast, multilocus minisatellite DNA fingerprinting is more sensitive method for the identification of clonal lines. This method, however, has limited application in population genetics because it is difficult to identify the zygotic condition (Takagi *et al.*, 1997). Recently, a number of easily and highly variable genetic markers, known as microsatellite DNA, has been developed (Wright and Bentzen, 1994). Microsatellites are highly polymorphic, particularly in fish (Brocker *et al.*, 1994).

In the present study, we used microsatellite DNA to examine the clonal nature of the artificially propagated silver crucian carp, and examined the zygotic condition of the loci of several clonal lines in this species collected from natural water in Kochi, Japan. As for the scientific name, we tentatively used *C. langsdorfii* for the triploid silver crucian carp while several other scientific names have been given to the silver crucian carp (Ginbuna), such as, *C. auratus langsdorfii* (Hosoya, 1993; Nakamura, 1969), *C. gibelio langsdorfii* (Miyadi *et al.*, 1976) and *C. langsdorfii* (Matsubara and Ochiai, 1965).

* Corresponding author: Tel. 022-717-8740;
FAX. 022-717-8743.

MATERIALS AND METHODS

Sampling and DNA extraction

One silver crucian carp, *C. langsdorfii*, caught from the Monobe River in Kochi Prefecture, Japan, was used as the maternal fish. Sperm from several goldfishes were used to activate egg development without the effect of genetic contribution to the offspring (Kobayasi, 1971; Dong and Taniguchi, 1996). The fry were reared until 150 days after hatching, and 40 fish were used for the analysis.

Samples of silver crucian carp from natural waters were collected from two locations, the Niyodo and Monobe Rivers in Kochi Prefecture, Japan. The triploidy of these samples was determined by analyzing the red blood cell size using the methods detailed in Sezaki *et al.* (1977). In this study, we used the samples of triploid silver crucian carp that had previously been identified as 17 clones (designated as KOC-001 to 015 and 017) by minisatellite DNA fingerprinting in a previous report (Ohara *et al.*, 1998). The samples of KOC-016 were not used due to denaturation of DNA. Total DNA was extracted from the blood of each fish by standard SDS-phenol/chloroform procedures and stored at 4°C prior to PCR analysis (Takagi *et al.*, 1997).

Microsatellite analysis

Three microsatellite loci, *GF1**, *GF17**, and *GF29**, developed by Zheng *et al.* (1995), were used in this study. Nomenclature of loci and alleles follow Shaklee *et al.* (1990). Radiolabeling and chemiluminescence methods were used to visualize microsatellite DNA. The radiolabeling method was performed using the methods reported by Takagi *et al.* (1997). The reverse primer was 5' end-labeled with [γ^{32} P]ATP. The PCR was programmed for 7 cycles of 1 min at 94°C, 30 sec at 53°C, 30 sec at 72°C, 33 cycles of 30 sec at 90°C, 30 sec at 53°C, and 30 sec at 72°C, respectively. Following amplification, PCR products were mixed with denaturing stop dye, heated at 95°C for 15 min, and electrophoresed on 6% polyacrylamide gel. Alleles were sized according to an M13 sequence ladder.

The chemiluminescence detection method was performed according to the methods given in Perez-Enriquez *et al.* (1998). The reverse primer was 5' end-labeled with biotin. The PCR and electrophoresis conditions were the same as in the radiolabeling method. After electrophoresis, DNA was transferred to a nylon membrane by blotting, then the membrane was dried and UV crosslinked. DNA on the membrane was detected by using the Phototope™-Star Detection Kit (New England Biolabs). A sequence ladder obtained from the pUC19 plasmid was used as a size marker and was prepared with the CircumVent™ Phototope™ Kit (New England Biolabs).

Estimation of genetic distance

We estimated the genetic distance (*D*) among the clonal lines based on the band sharing index (*BSI*) at the three microsatellite loci in order to determine the genetic relationship among the clonal lines (Wetton *et al.*, 1987; Gillbert *et al.*, 1990). The *BSI* between clonal lines was calculated with the formula $BSI = 2Nab / (Na + Nb)$, where *Na* and *Nb* are the number of bands (alleles) present in each clone, and *Nab* is the number of bands shared. The genetic distance (*D*) calculated with the formula $D = 1 - BSI$. The value varies from zero, when the two individuals are identical, to 1, when there are no bands in common. A dendrogram was drawn by the UPGMA method (Sneath and Sokal, 1973) based on *D* values, using a program in the PHYLIP Ver 3.5 software package (Felsenstein, 1994).

RESULTS

Microsatellite DNA patterns of artificial propagated offspring

Microsatellite DNA markers of 40 offspring from one maternal fish were examined. Since both the offsprings and

the maternal fish had the same genotype at the three polymorphic loci, they are of the same clonal line. The genotypes of the clonal line were determined as *307/ (*307 or *311) /

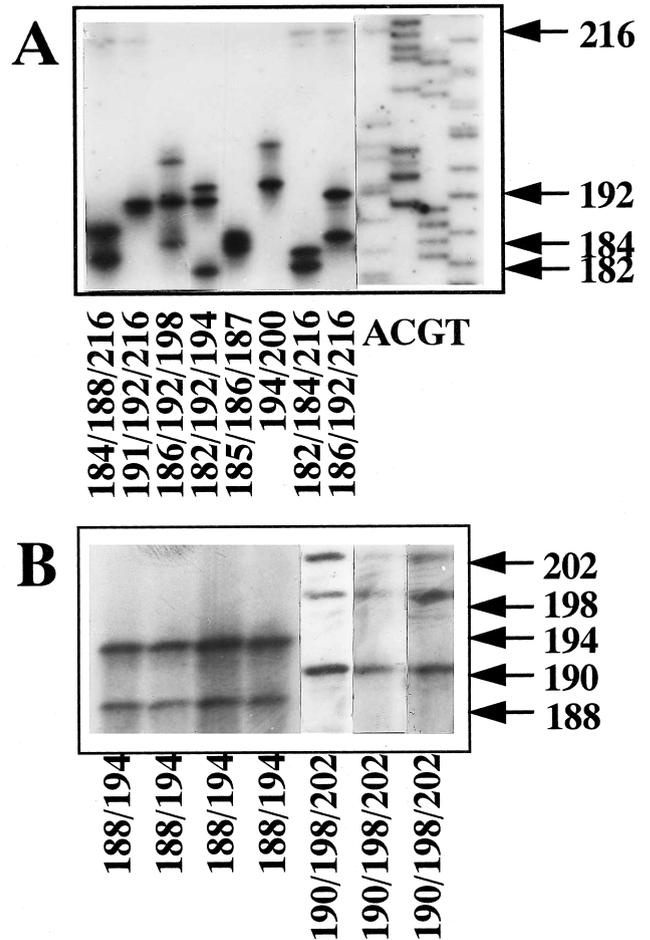


Fig. 1. Microsatellite banding pattern of Silver crucian carp, *Carassius langsdorfii*. (A) Microsatellite banding pattern of eight clones (left to right: KOC-010, 006, 002, 011-1, 012, 013, 014, 015) in *GF17** obtained with the radiolabeling method. The size standard is a sequencing of M13 mp18. (B) Microsatellite banding pattern of two clones (KOC-002, 003) in *GF29** obtained with the chemiluminescence detection method.

Table 1. Numbers of alleles, genotypes and fragment size as well as proportion of heterozygotic individuals and allelic combinations of 17 clonal lines of silver crucian carp, *Carassius langsdorfii*

	<i>GF1*</i>	<i>GF17*</i>	<i>GF29*</i>
No. of samples	237	237	237
No. of alleles	9	16	9
No. of genotypes	10	17	8
Fragment size(bp)	293–335	182–216	186–202
Proportion of heterozygotes*	1.000	1.000	0.992
Allelic combination			
Triallelic	0.253	0.734	0.376
Diallelic	0.747	0.266	0.624
Monoallelic	0	0	0

* Proportion of heterozygotic individuals

Table 2. Genotypes of three microsatellite DNA loci in 17 clonal lines of silver crucian carp, *Carassius langsdorfii*

Clonal lines	Microsatellite locus			Number of individuals		
	<i>GF1*</i>	<i>GF17*</i>	<i>GF29*</i>	Niyodo River	Monobe River	Total
KOC-001	307/(307 or 311)/311	185/192/193	188/(188 or 194)/194	7	3	10
KOC-002	299/303/335	186/192/198	190/198/202	10	6	16
KOC-003	303/307/311	188/(188 or 192)/192	188/(188 or 194)/194	2	3	5
KOC-004	303/(303 or 305)/305	185/187/216	188/(188 or 194)/194	25	1	26
KOC-005	299/(299 or 303)/303	188/189/198	186/188/194	0	4	4
KOC-006	303/307/311	191/192/216	186/188/194	10	10	20
KOC-007	303/(303 or 311)/311	194/(194 or 204)/204	188/(188 or 194)/194	2	2	4
KOC-008	293/(293 or 303)/303	196/(196 or 204)/204	192/200/202	36	0	36
KOC-009	303/(303 or 311)/311	184/(184 or 216)/216	188/(188 or 194)/194	6	0	6
KOC-010	293/309/311	184/188/216	194/(194 or 196)/196	10	0	10
KOC-011-1	307/(307 or 311)/311	182/192/194	188/(188 or 194)/194	65	0	65
KOC-011-2	307/(307 or 311)/311	182/(182 or 194)/194	188/(188 or 194)/194	1	0	1
KOC-012	303/(303 or 305)/305	185/186/187	188/(188 or 194)/194	3	0	3
KOC-013	303/(303 or 305)/305	194/(194 or 200)/200	196/(196 or 202)/202	11	0	11
KOC-014	303/(303 or 311)/311	182/184/216	188/192/196	11	0	11
KOC-015	303/313/335	186/192/216	190/194/196	2	0	2
KOC-017	299/303/311	188/192/216	194/(194 or 196)/196	7	0	7

*311 in *GF1**, *185 / *192, / *193 in *GF17**, and *188 / (*188 or *194) / *194 in *GF29**. This clonal line was identified as the clone KOC-001.

Microsatellite DNA polymorphism of wild crucian carp

Microsatellite polymorphism is shown in Fig. 1 as examples of the radiolabeling and chemiluminescence methods. A total of 237 silver crucian carp from natural water were analyzed. The number of alleles per locus, amplified product size range, proportion of heterozygotes, allelic combination and the number of genotypes for the three loci are summarized in Table 1. In the two bands types, the one allele was recognized to be duplicated to the one of two bands. The assumed genotypes of the silver crucian carp are given in Table 2. In *GF1**, *303 and *311 were predominant and appeared in 13 and 10 clonal lines, respectively. In *GF17**, *192 and *216 appeared in seven clonal lines. In *GF29**, *188 and *194 appeared in 11 and 13 clonal lines, respectively.

The observed number of alleles and genotypes were quite large in the three loci comparing with isozyme analysis (Simizu *et al.*, 1993; Don *et al.*, 1996). The proportion of heterozygotes in the wild population was very high over the three microsatellite loci comparing with gold fish (Zheng *et al.*, 1995). The observed number of genotypes was quite small although a large number of alleles was observed. In the results of allelic combinations, the ratio of triallelic types were high in *GF17** (0.734). In *GF1** and *GF29**, the ratio of diallelic types were relatively high with values of 0.747 and 0.626, respectively, compared with isozyme analysis (Simizu *et al.*, 1993; Dong *et al.*, 1996). Yet, monoallelic type was not observed in this population.

The 16 clonal lines, which were identified by minisatellite DNA fingerprinting, are clearly distinguished based on the

genotype in *GF17**. Based on the analysis of each locus, all individuals within each clonal lines except KOC-011 had the same genotype. However, among the 66 individuals of KOC-011, one fish had two alleles (*182 and *194) while the others had three alleles (*182, *192, and *194) in *GF17**. This fish had the same with the others in genotype of *GF1** and *GF29**.

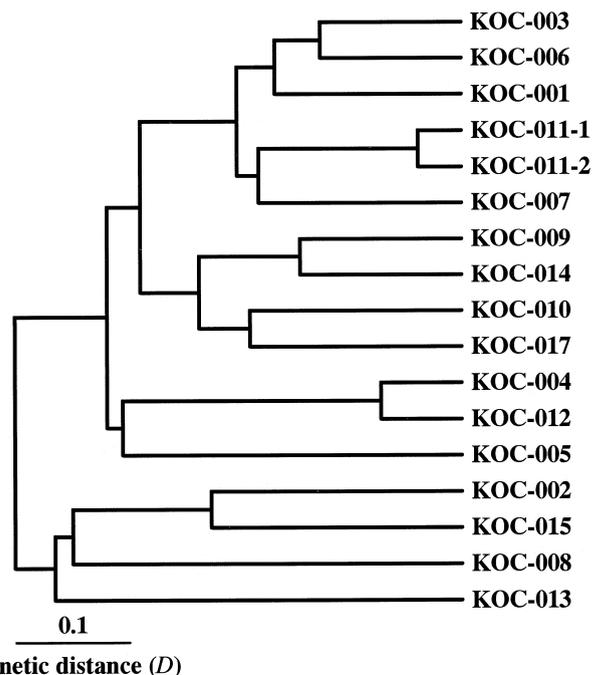


Fig. 2. UPGMA dendrogram drawn on the basis of the genetic distance of three microsatellite loci between each pair of 17 clonal lines in the silver crucian carp, *Carassius langsdorfii*, using PHYLIP ver 3.5.

Thus KOC-011 was subdivided into two clonal lines: one with three alleles (*182, *192, and *194), and the other with two alleles (*182 and *194), which were designated KOC-011-1 and KOC-011-2, respectively.

The average genetic distance (D) between clonal lines was calculated as 0.637 ± 0.191 . The BSI within clonal lines, except for KOC-011, was equal to 1.000, indicating genetic uniformity. The dendrogram drawn by the UPGMA method based on D values is given in Fig. 2. KOC-011-1 and KOC-011-2 had a very close genetic relationship compared to other clones.

DISCUSSION

The analysis of microsatellite DNA loci can be used as a fingerprinting method to distinguish clonal lines. In the loci, information of the zygotic condition of each locus can also be obtained. The maximum allele number of an individual at each microsatellite locus was three as observed in this study. This suggests that the silver crucian carp *C. langsdorfii* is a true triploid. Since the genotypes can be recorded by each locus, in the future, microsatellite DNA loci will be applied to establish a registration system of clonal lines of silver crucian carp.

The proportion of heterozygotes was very high over the three microsatellite loci. Similar results were observed in the electrophoretic analysis (Simizu *et al.*, 1993; Dong *et al.*, 1996; Ohara *et al.*, 1998). The fixed multilocus heterozygosity observed in this study is a characteristic of unisexual-hybrid vertebrates (Vrijenhoek, 1990). The unisexual vertebrates were generally originated from hybridization between congeneric species (Dawley, 1989). The silver crucian carp may be originated from hybridization (Simizu *et al.*, 1993). One explanation for the high proportion of heterozygotes is that during the oogenesis of silver crucian carp, the first meiotic division does not occur, but rather the oocyte undergoes a homotypic nuclear division as in somatic cells, which are called apomixis (Kobayasi, 1976; Yamasita *et al.*, 1993; Arai, 1997). Therefore, they can maintain an almost heterozygous state for each generation within a clonal population. The multilocus heterozygous condition may lead to an increase of fitness in the clonal line of silver crucian carp.

In this study, we detected two subtypes within the clone KOC-011 collected from natural water. Kojima *et al.* (1984) observed the partial pairing of homologue-like chromosomes at the zygote stage in silver crucian carp. Zhang *et al.* (1992) observed the putative recombinant in offspring from a female of the silver crucian carp based on the electrophoretic pattern of isozyme. Two different processes can be considered to explain the origin of the subtype: 1) Synapsis and recombination occurred between homologous chromosomes within the microsatellite locus existed. 2) The microsatellite DNA mutated to a new allele (size changed or null allele). Information on this mechanism is important to understand how genetic diversity is maintained within silver crucian carp populations. Further studies are needed to determine the developing processes of these subtypes.

In this study, we used three microsatellite loci, yet, it is not enough to fully discuss the genetic relationship of the silver crucian carp. It is necessary to investigate many other microsatellite loci. Nonetheless, we found that the microsatellite loci used in this study are very valuable due to their application not only to identify clonal lines of this species, but also to study the origin of the silver crucian carp.

ACKNOWLEDGMENT

We thank Dr. Chris Norman, lecturer of Chiba University for critical reading of the English manuscript.

REFERENCES

- Arai K (1997) Biodiversity in fish chromosomes and its applicable potential as genetic resources. *Fish Genetics Breed Sci* 24: 3–20 (In Japanese)
- Brocker AL, Cook D, Bentzen P, Wright JM, Doyle RW (1994) Organization of microsatellite differs between mammals and cold-water teleost fishes. *Can J Fish Aquat Sci* 51: 1959–1966
- Dawley RM (1989) An introduction to unisexual vertebrates. In "Evolution and Ecology Unisexual Vertebrates", Ed by R M Dawley, J P Bogart, Bull 466, New York State Museum, Albany, New York, pp 1–18
- Dong S, Taniguchi N (1996) Clonal nature of offspring of ginbuna *Carassius langsdorfii* by RAPD-PCR and isozyme patterns. *Nippon Suisan Gakkaishi* 62: 891–896 (In Japanese)
- Dong S, Taniguchi N, Tsuji S (1996) Identification of clones of ginbuna *Carassius langsdorfii* by DNA fingerprinting and isozyme pattern. *Nippon Suisan Gakkaishi* 62: 747–753 (In Japanese)
- Dong S, Ohara K, Taniguchi N (1997) Introduction of sperm of common carp *Cyprinus carpio* into eggs of ginbuna *Carassius langsdorfii* by heat shock treatment and its confirmation by DNA markers. *Nippon Suisan Gakkaishi* 63: 201–206 (In Japanese)
- Felsenstein J (1994) PHYLIP version 3.5. Department of Genetics, University of Washington, Seattle
- Gillbert D, Lehman A, O'Brien SJ, Wayne RK (1990) Genetic Fingerprinting reflects population differentiation in the California Channel Island fox. *Nature* 344: 764–767
- Hosoya K (1993) Fishes of Japan with pictorial keys to the species, Ed by T Nakabo, Tokai university press, Tokyo, pp 212–213 (In Japanese)
- Kobayasi H, Kawashima Y, Takeuchi N (1970) Comparative chromosome studies in the genus *Carassius*, especially with a finding of polyploidy in the Ginbuna *C. auratus langsdorfii*. *Japan J Ichthyol* 17: 153–160
- Kobayasi H (1971) A cytological study on gynogenesis of the triploid ginbuna (*Carassius auratus langsdorfii*). *Zool Mag* 80: 316–322 (In Japanese)
- Kobayasi H and Ochi H (1972) Chromosome studies of the hybrids, ginbuna *C. auratus langsdorfii* × kinbuna *C. auratus* subsp., and ginbuna × loach *Misgurnus anguillicaudatus*. *Zool Mag* 81: 67–71
- Kobayasi H (1976) A cytological study on the maturation division in the oogenic process of the triploid ginbuna, *Carassius auratus langsdorfii*. *Japan J Ichthyol* 22: 234–240
- Kojima K, Matsumura K, Kawashima M, Kajishima T (1984) Studies on the gametogenesis in polyploid Ginbuna *Carassius auratus langsdorfii*. *J Fac Sci Shinshu Univ* 19: 37–52
- Matsubara K, Ochiai A (1965) Ichthyology Part II. Kouseisha-kouseikaku, Tokyo, pp 530–536 (In Japanese)
- Miyadi D, H Kawanabe, N Mizuno (1976) Coloured illustrations of the freshwater fishes of Japan. Hoikusha, Osaka, pp 201–210 (In

Japanese)

- Murakami M, Fujitani H (1997) Polyploid-specific repetitive DNA sequences from triploid ginbuna (Japanese silver crucian carp, *Carassius auratus langsdorfii*). *Genes Genet Syst* 72: 107–113
- Murayama Y, Hijikata M, Nomura T, Kajishima T (1984) Analysis of histocompatibility and isozyme variation in triploid fish, *Carassius auratus langsdorfii*. *J Fac Sci Shinshu Univ* 35: 125–138
- Nakakuki M, Toya H, Sawano K, Kajishima T (1984) On the fertilization of the triploid ginbuna. *J Fac Sci Shinshu Univ* 19: 25–35
- Ohara K, Dong S, Taniguchi N (1998) Identification and distribution of clonal lines detected by DNA polymorphism in silver crucian carp, *Carassius langsdorfii* collected from the Monobe and Niyodo rivers. *Japan J Ichthyol* 45: 21–27 (In Japanese)
- Perez-Enriquez R, Takemura M, Taniguchi N (1998) Microsatellite DNA detection by chemiluminescence in red sea bream: a practical manual. *Fish Genetics Breed Sci* 26: 73–79 (In Japanese)
- Sezaki K, Kobayasi H, Nakamura M (1977) Size of erythrocytes in the diploid and triploid specimens of *Carassius auratus langsdorfii*. *Japan J Ichthyol* 24: 135–140 (In Japanese)
- Shaklee JB, Allendorf FW, Moritz DC, Whitt GS (1990) Genetic nomenclature of protein-coding loci in fish: proposed guidelines. *Trans Am Fish Soc* 119: 2–15
- Simizu Y, Oshiro T, Sakaizumi M (1993) Electrophoretic studies of diploid, triploid, and tetraploid forms of the Japanese silver crucian carp, *Carassius auratus langsdorfii*. *Japan J Ichthyol* 40: 65–75
- Sneath PHA, Sokal RR (1973) Numerical taxonomy. Freeman & Co., San Francisco, pp 114–308
- Takagi M, Taniguchi N, Cook D, Doyle RW (1997) Isolation and characterization of microsatellite loci from red sea bream *Pagrus major* and detection in closely related species. *Fisheries Sci* 63: 199–204
- Taniguchi N, Sakata K (1977) Interspecific and intraspecific variations of muscle protein in the the Japanese crucian carp-II. Starch-gel electrophoretic pattern. *Japan J Ichthyol* 24: 1–11
- Umino T, Arai K, Nakagawa H (1996) Growth performance in clonal crucian carp, *Carassius langsdorfii*. Effect of genetic difference and feeding history. *Aquaculture* 155: 271–283
- Umino T, Arai K, Maeda K, Zhang Q, Sakae K, Niwase I, Nakagawa H (1997) Natural clones detected by multilocus DNA fingerprinting in gynogenetic triploid ginbuna *Carassius langsdorfii* in Kurose River, Hiroshima. *Fisheries Sci* 63: 147–148
- Vrijenhoek RC (1990) Genetic diversity and the ecology of asexual population. In "Population Biology and Evolution", Ed by K Woehrmann S Jain, Springer-Verlag Berlin, pp 195–197
- Wetton JH, Caeter RE, Parkin DT, Walters D (1987) Demographic study of a wild house sparrow population by DNA fingerprinting. *Nature* 327: 147–149
- Wright JM, Bentzen P (1994). Microsatellite genetic markers for the future. In "Molecular Genetics in Fisheries", Ed by GR Carvalho TJ Pitcher, London, pp 117–121
- Yamashita M, Onozato H, Nakanishi T, Nagahama Y (1990) Break-down of the sperm nuclear envelope is a prerequisite for male pronucleus formation: direct evidence from the gynogenetic crucian carp *Carassius auratus langsdorfii*. *Devel Biol* 137: 155–160
- Yamashita M, Jiang J, Onozato H, Nakanishi T, Nagahama Y (1993) A tripolar spindle formed at meiosis I assures the retention of the original ploidy in the gynogenetic triploid crucian carp, ginbuna *Carassius auratus langsdorfii*. *Develop Growth and Differ* 35: 631–636
- Zhang F, Oshiro T, Takashima F (1992) Chromosome synapsis and recombination during meiotic division in gynogenetic triploid Ginbuna, *Carassius auratus langsdorfii*. *Japan J Ichthyol* 39: 151–155
- Zheng W, Stacey NE, Coffin J, Strobeck C (1995) Isolation and characterization of microsatellite loci in the goldfish *Carassius auratus*. *Mol Ecol* 4: 791–792

(Received January 18, 1999 / Accepted July 12, 1999)