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**SHORT COMMUNICATION**

**HNF-1 Regulates the Promoter Activity of the HP-27 Gene**

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**ABSTRACT**—The hibernation-specific HP-27 gene is expressed specifically in the liver of the chipmunk, a hibernating species of the squirrel family, and exists as a pseudogene in the tree squirrel, a nonhibernating species. In the promoter region, the chipmunk gene has a potential HNF-1 binding site, and the tree squirrel gene has two base substitutions in the corresponding sequence. In this paper, we investigated the role of HNF-1 in the HP-27 gene promoter activity. Gel retardation assays with in vitro-translated HNF-1 and super-shift assays using HepG2 nuclear extracts and an anti-HNF-1 antibody revealed that HNF-1 bound to the chipmunk gene sequence. HNF-1 also bound to the tree squirrel sequence, but with much lower affinity. In HepG2 cells, HNF-1 activated transcription from the chipmunk HP-27 gene, but not from the tree squirrel gene. In addition, the tree squirrel-type base substitutions in the HNF-1 binding site greatly reduced the promoter activity of the chipmunk HP-27 gene. These results indicate that HNF-1 is required for the promoter activity of the chipmunk HP-27 gene, and that the base substitutions in the HNF-1 binding site are involved in the lack of HP-27 gene expression in the tree squirrel.

**Key words:** hibernation, promoter, transcriptional regulation, HNF-1, HP-27

**INTRODUCTION**

Mammalian hibernation is a unique physiological adaptation that allows life to be sustained under extremely low body temperatures (Kayser 1961; Johansson et al., 1967; Swan 1972). Only certain small mammals, primarily in the orders Rodentia, Insectivora, and Chiroptera, can undergo hibernation. In the squirrel family, some species, such as the chipmunk (*Tamias asiaticus*) and the 13-lined ground squirrel (*Citellus tridecemlineatus*), hibernate; other species, such as the tree squirrel (*Callosciurus caniceps*), do not. The chipmunk hibernation-specific protein HP-27 was identified as a component of a 140-kDa complex (Kondo and Kondo 1992). The amount of this complex is drastically lowered in the blood during hibernation. In the chipmunk and the 13-lined ground squirrel, the HP-27 gene is expressed specifically in the liver and is downregulated during hibernation (Takamatsu et al., 1993). Although the tree squirrel has the HP-27 gene, its mRNA is not expressed (Takamatsu et al., 1993). In a previous paper, we showed that the 170-bp 5' flanking sequence of the chipmunk HP-27 gene contains the promoter for the liver-specific transcription, and that the transcription factors that bind to the region from nucleotides –170 to –140 play important roles in HP-27 gene transcription (Ono et al., 2003). Despite, the 69-bp 5' flanking sequence still retained about 10% of the promoter activity of the 170-bp 5' flanking sequence, which implied that there was a contribution of the HNF-1 binding site from –54 to –40 of the chipmunk HP-27 gene to the liver-specific promoter activity (Ono et al., 2003). In this paper, we investigated the role of HNF-1 in the HP-27 gene promoter activity.

**MATERIALS AND METHODS**

**Gel retardation assay**

Nuclear extracts were prepared from human hepatoma HepG2 cells as described (Kojima et al., 2000). Mouse HNF-1 was synthesized from pcDNA3/HNF-1 (Ono et al., 2001) using an in vitro transcription/translation system (Promega). HepG2 nuclear extracts or in vitro-translated HNF-1 were preincubated with 1 µg of poly (dl-D) on ice for 5 min in 20 µl of 1× gel-retardation buffer (12% glyc-
erol, 12 mM HEPES-KOH [pH 7.9], 60 mM KCl, 5 mM MgCl₂, 4 mM Tris-HCl [pH 7.9], 0.6 mM EDTA, 5 mM DTT), then mixed with 1 ng of ³²P-labeled, double-stranded oligonucleotide probe CM27G-63/-30 (5’-CTTTGTGGTTAAAAAAAAATGTCCACCTATTTG-3’), containing the chipmunk HP-27 gene sequence from nucleotides –63 to –30, or TS27G-63/-30 (5’-CTTTGTGGTTAAAAATTAGCCAACCTATTG-3’), containing the tree squirrel HP-27 gene sequence from nucleotides –63 to –30, and incubated on ice for an additional 30 min. For competition assays, a 10- or 100-fold molar excess of CM27G-63/-30 or TS27G-63/-30 was used as a competitor, and for supershift experiments, a rabbit polyclonal HNF-1 antibody (Santa Cruz Biotechnology) or normal rabbit serum was included in the binding reactions. The samples were subjected to electrophoresis through a nondenaturing 4% polyacrylamide gel at room temperature, then autoradiographed.

Promoter-reporter plasmid constructs

To construct chipmunk-type promoter-reporter plasmids, pCM27G-63/luc and pCM27G-32/luc, the 5’ flanking sequences were generated by PCR using a chipmunk HP-27 genomic clone (Ono et al., 2003) as the template, digested with HindIII at nucleotide +84, and subcloned into the Smal and HindIII sites of the promoterless firefly luciferase expression vector pGVB (Nippon Gene). The numbers –63 and –32 in the construct names indicate the 5’ end of the 5’ flanking sequence. For a tree squirrel promoter-reporter plasmid, pTS27G-63/luc, the corresponding sequence was amplified by PCR using a tree squirrel HP-27 genomic clone as the template, and was subcloned into pGVB. Three mutant constructs, pCM27G-170m-1/luc, pCM27G-170m-2/luc, and pCM27G-170m-3/luc, which carry both or either of the tree squirrel-type base substitutions at nucleotides –53 and –43, were constructed by PCR from pCM27G-170/luc (Ono et al., 2003).

DNA transfection and luciferase assay

HepG2 cells were maintained in Dulbecco’s Modified Eagle’s Medium containing 10% fetal bovine serum. The cells were plated at 1.0 × 10⁵ cells per 20-mm dish, and after 20 hr were transfected with 0.75 µg of a firefly luciferase promoter-reporter plasmid and 30 ng of a Renilla luciferase internal control plasmid, pRL-TK (Promega), using FuGENE 6 (Roche Diagnostics). Where denoted, the indicated amounts of the mouse HNF-1 expression constructs pcDNA3/HNF-1 were cotransfected. After 40 hr, the luciferase activities were measured using the Dual-Luciferase Reporter Assay System (Promega).

RESULTS AND DISCUSSION

The 170-bp 5’ flanking sequence of the chipmunk HP-27 gene contains the entire promoter region necessary for its liver-specific transcription (Ono et al., 2003). Despite, the 69-bp 5’ flanking sequence still retains about 10% of the promoter activity. The sequence from nucleotides –54 to –40 of the chipmunk HP-27 gene shows an 87% identity with the consensus HNF-1 binding sequence (Fig. 1) (Tronche and Yaniv, 1992). We first examined whether HNF-1 could bind to this chipmunk sequence by gel retardation assays using a double-stranded oligonucleotide probe CM27G-63/-30 containing the sequence from nucleotides –63 to –30 of the chipmunk HP-27 gene. Incubation of in vitro-translated HNF-1 with the probe CM27G-63/-30 resulted in the formation of a protein-DNA complex (Fig. 2A, lane 3), and a similar DNA-protein complex formation was observed with the HepG2 nuclear extracts (Fig. 2A, lane 5). When an anti-HNF-1 antibody was added to the binding reaction, the protein-DNA complex was super-shifted (Fig. 2A, lanes 4 and 6). The faster migrating bands in lane 5 were considered to represent nonspecific protein binding because they were not competed out by an excess of unlabeled probe (data not shown). On the other hand, the corresponding tree squirrel HP-27 gene sequence differs from the chipmunk sequence by two nucleotides, and shows a 73% identity with the consensus HNF-1 binding sequence (Fig. 1). When incubated with in vitro-translated HNF-1, the tree squirrel-type probe TS27G-63/-30 formed a similar protein-DNA complex to that observed with CM27G-63/-30, but not as effectively as CM27G-63/-30 (Fig. 2B, lanes 2 and 4). Similarly, the complex formed between HNF-1 and the probe CM27G-63/-30 was competed by CM27G-63/-30, but not as effectively as TS27G-63/-30 (Fig. 2C, lanes 2–5). From these results, we concluded that HNF-1 binds to the 5’ flanking sequence of the chipmunk HP-27 gene from nucleotides –63 to –30, and that the two base substitutions in the corresponding tree squirrel HP-27 gene sequence disturb HNF-1 binding.

We next investigated whether HNF-1 could activate transcription from the chipmunk and tree squirrel HP-27 gene promoters in HepG2 cells. Upon cotransfection of pcDNA3/HNF-1, the luciferase activities of pCM27G-63/-30 increased in a dose-dependent manner (Fig. 3). In comparison, the promoter activities of pCM27G-32/luc, which lacks the potential HNF-1 binding site, and a tree squirrel-type promoter-reporter construct pTS27G-63/luc were hardly affected by HNF-1. These results indicate that HNF-1 can activate transcription of the chipmunk HP-27 gene, but not of the tree squirrel HP-27 gene.

Because the 63-bp 5’ flanking sequence of the chipmunk HP-27 gene retained only 10% of the promoter activity of the 170-bp 5’ flanking sequence, we then investigated the effect of HNF-1 on the promoter activity in the context of the
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entire 170-bp promoter (Ono et al., 2003). To this end, we made three mutant constructs from pCM27G-170/luc: pCM27G-170m-1/luc, pCM27G-170m-2/luc, and pCM27G-170m-3/luc. The first construct carries both the tree squirrel-type base substitutions, at nucleotide –53 and –43, in the HNF-1 binding site; the last two each carry one mutation, at nucleotide –53 or –43, respectively (Fig. 4). These mutant constructs were transfected into HepG2 cells, and those luciferase activities were compared with that of pCM27G-170/luc (Fig. 4). The promoter activity of the double mutant, pCM27G-170m-1/luc, was 70% less, and those of the single mutants, pCM27G-170m-2/luc and pCM27G-170m-3/luc, were each 50% less than the promoter activity of pCM27G-170/luc. These results indicate that the binding site for HNF-1 in the chipmunk HP-27 gene from –54 to –40 is required for effective transcriptional activation. On the other hand, in the case of the tree squirrel HP-27 gene, its low affinity for HNF-1 is likely to be insufficient for effective transactivation by HNF-1.

Like the HP-27 gene, another hibernation-specific gene, HP-25, is expressed in a liver-specific manner in the chipmunk, and exists as a pseudogene in the tree squirrel (Takamatsu et al., 1993; Kojima et al., 2000; Kojima et al., 2001). HNF-4 plays an essential role in the promoter activity of the chipmunk HP-25 gene (Kojima et al., 2000). In con-
The results presented in this paper have revealed that the binding of HNF-1 to the proximal promoter region from nucleotides –54 to –40 is required for the chipmunk HP-27 gene promoter activity. In the previous paper, we demonstrated that the transcription factors that bind to the sequence from nucleotides –170 to –140 are essential for the liver-specific promoter activity of the chipmunk HP-27 gene (Ono et al., 2003), and to further understand the transcriptional regulation of the HP-27 gene, the transcription factors that bind to this region must be identified.

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