Atlas Florae Europaeae notes, 19–22. Nomenclatural changes and taxonomic adjustments in some native and introduced species of Malinae (Rosaceae) in Europe

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Abstract

Compilation of the nomenclatural checklist of Rosaceae for volumes 16 and 17 of Atlas Florae Europaeae resulted in nomenclatural and taxonomic adjustments presented here. The synonymy of Cotoneaster bullatus is verified; the name C. tomentellus is shown to have been validly published in 2009 instead of 1961. Cotoneaster marianus, an alleged endemic to Macedonia, is a synonym of C. pannasicus from Greece. The new combination Sorbaronia mitschurinii is proposed to reflect the intergeneric hybrid origin of Aronia mitschurinii, an apomictic species of culligenous origin (Aronia melanocarpa x Sorbus aucuparia). The name Crataegus chrysocarpa var. rotundifolia J. B. Phipps & Sennikov is proposed to replace the illegitimate C. chrysocarpa var. phoenicea E. J. Palmer, and the illegitimate C. horrida Medik. ex Cinovskis 1971 is added to its synonymy. The new name Crataegus chrysocarpa var. phoeniceoides J. B. Phipps & Sennikov is proposed for C. chrysocarpa var. phoenicea J. B. Phipps, nom. illeg., non E. J. Palmer. The name Sorbus tauricola Zaik. ex Sennikov is validly published with a new description and type designation. The lectotype of Cotoneaster bullatus and the neotypes of C. moupinensis f. floribundus and Mespilus rotundifolia are designated.

Additional key words: apomictic taxa, Aronia, Cotoneaster, Crataegus, Ehrhart, Maloideae, new names, Pyrinae, Sorbaronia, Sorbus, synonymy, typifications

Introduction
Production of the forthcoming volumes 16 (Rosaceae: Cydonia to Prunus in the sequence of Flora Europaea, excluding Sorbus) and 17 (Rosaceae: Sorbus) of Atlas Florae Europaeae revealed the need for corrections in the nomenclature and synonymy of some names and in the taxonomic position of one species in subtribe Malinae (formerly subfamily Maloideae or subtribe Pyrinae). References to the International Code of Nomenclature (ICN) are according to the Melbourne edition (McNeill & al. 2012).

19. Nomenclature and synonymy in some native and alien species of Cotoneaster
The genus Cotoneaster Medik. is especially rich in problems because numerous minor variants were described as species on the basis of their presumably obligate apomictic mode of reproduction; the most comprehensive survey of the genus can be found in Fryer & Hyldmø (2009). This monograph, although written in the full taxonomic format and including reportedly all the species described...
up to date, is peculiar in its extreme splitter’s approach and numerous inaccuracies in nomenclature, descriptions, distributions, and historical background (Sennikov 2010). A series of corrections to the nomenclature, taxonomy and distribution of cotoneasters native to or alien in East Europe and the Caucasus has been published earlier (Sennikov & Somlyay 2011; Sennikov 2011); the present contribution adds nomenclatural corrections for one native species that is endemic to the Balkans and two taxa of Chinese origin that are cultivated and locally naturalized in Europe.


**Cotoneaster bullatus** Bois is a species of *C*. *ser*. *Bullati* Flinck & B. Hylmö that is common in ornamental cultivation in Europe (Fryer & Hylmö 2009), frequently escapes from cultivation, and is found as casual or naturalized alien in several countries of Europe (Sennikov 2009; Dickoré & Kasperek 2010).

The lectotypification of *Cotoneaster bullatus* was attempted by Fryer & Hylmö (2009) who, however, missed the word “here” in the expression of intention “designated here” as required by Art. 7.10. This typification is confirmed and formally effected in the present note.

The specimen *Vilmorin* 2123 (4483) (K 000442346), taken from a plant cultivated in the Arboretum Vilmorin at Les Barres, France, preserved on 3 Jul 1902 and received by Kew on 14 Jul 1902, was indicated as the holotype of *Cotoneaster moupinensis* f. *floribundus* Stapf by Lu & Brach (2002). However, this is not an element on which the original description of this taxon was based (Art. 9.3); neither is this type designation correctable to an effectively published neotypification under Art. 7.10 and Art. 9.23. As explained by Prain (1909) in the comment to Plate 8284 alongside which Stapf’s *Cotoneaster* name was published, the original description (Stapf 1909) was based on the material taken from a bush cultivated at the Royal Botanic Gardens, Kew, and originated from living material given by Maurice de Vilmorin in 1905. As evident from identifications, Stapf examined the Kew specimen and treated it as *C. moupinensis* Franch., but at the later date of 10 Jan 1910. Since this specimen is of the same origin as the plant that was cultivated at Kew, its designation as neotype is reasonable and is formally effectuated here.

Klotz (1972) described *Cotoneaster boisianus* G. Klotz as different from *C. bullatus* s.str. in its smaller leaves 2–5 cm long (vs. 5–8 cm long), a smaller inflorescence with 10–20 flowers (vs. 20–30 flowers), and smaller fruits 5–8 mm in diam. (vs. 8–10 mm in diam.). This taxon was recognized among the plants that were cultivated in the Arboretum Vilmorin under the same accession number that was the basis of *C. boisianus* (Klotz 1972), and its type was part of the material subsequently used for lectotypification of the latter name (Fryer & Hylmö 2009). The complexity of the type citation of *C. boisianus* that included two mentions of the collection date and accession number, one for the original seed material (no. 2123) and the other for the preserved specimen (no. 4483), misled Lu & Brach (2002) to conclude that this name was not validly published because of two types having been simultaneously designated.

The present typifications fix the current recognition of two variants of *Cotoneaster bullatus*, one with smaller dimensions of plant parts (Klotz 1972) and the other with larger ones (Bois 1906; Stapf 1909; Rehder & Wilson 1912; Lu & Brach 2003). As this difference reflects the variability in a limited area from which the species was originally collected, even though the plants are tetraploid and presumably apomictic because of producing uniform progeny from seeds (Fryer & Hylmö 2009), we refrain from accepting these morphotypes as separate taxa at any rank. The species rank is preferred for the third variant included by Lu & Brach (2003), *C. bullatus* var. *macrophyllus* Rehder & E. H. Wilson = *C. rehderi* Pojark., because of much larger leaves and the pubescent hypanthium (Rehder & Wilson 1912).

After many years of oblivion, *Cotoneaster parnassicus* Boiss. was accepted as a member of *C. sect. Racemiflori* (Pojark.) G. Klotz by Hylmö & Fryer (1999). This species was described from Greece (Boissier 1856), and also erroneously reported from Crete in place of *C. nummularius* (Hylmö & Fryer 1999).

Andonoski & Andonovski (1996) reported a new species of *Cotoneaster*, *C. marianus* Andonoski & Andonovski from Mt Galičica in Macedonia. When originally described, the new species was compared only with the two species previously known from Galičica, *C. integerrimus* Medik. and *C. tomentosus* (Aiton) Lindl. Micevski (1999) took no notice of this report. Recently Teofilovski & al. (2012) accepted *C. marianus* and invented its distribution in Macedonia, with one of the records at the border with Greece.

The original description of *Cotoneaster marianus* is very incomplete, stating that the hypanthium of the species is hairy, the fruits have two nutlets, the flowers are solitary or in pairs on very short pedicels, and the leaves are small and slightly hairy above. These characters are at odds with *C. integerrimus* and *C. tomentosus* but fit those of *C. parnassicus*. Good photographs of the Macedonian plants, kindly offered by Aco Teofilovski, provide clear evidence that *C. marianus* is indistinguishable from *C. parnassicus*, which is very widely distributed in Greece. The new synonymy established here makes a new country record of *C. parnassicus*.

Fryer & Hylmö (2009: 75) attempted to lectotypify *Cotoneaster parnassicus* by Orphanides 420, but that gathering is not part of the original material of the name. Two syntypes cited in the protologue are available for lectotypification.


In Europe *Cotoneaster tomentellus* Pojark. is reported as possibly a naturalized alien in the United Kingdom (Stace 2010) and as possibly a casual alien in Sweden (Karlsson 2002). This species is native to China (Sichuan) and belongs to a complex group of *C. racemiflorus* (Desf.) Bosse s.l. (Fryer & Hylmö 2009).

Pojarkova (1961) explicitly cited two gatherings (“Jun 1908, fl., Oct 1908, fr.”), both under *Wilson 1317* as types of *Cotoneaster tomentellus*, consequently making it impossible for the name to be validly published as a name in its own right (Art. 6.9). Moreover, her citation of “*C. racemiflora* var. *soongoricus* Rehd. et Wils. in Sarg., Pl. Wilson, 1 (1912) 168, quoad descript. sed excl. basion. et syn.” in synonymy does not qualify as a replaced synonym because it refers to a misapplication, not to a name (Art. 41, Note 3). The species name was consequently not validly published in 1961 (Art. 40.1 & 40.2).

Buzunova (2005) attempted to lectotypify the name *Cotoneaster tomentellus* with *Wilson 1317* in flower at LE, but she failed to fulfill the conditions of valid publication because certain provisions of the ICN were not met: Art. 36.1(c), i.e. the name was not accepted by her but treated as synonym of *C. soongoricus* (Regel & Herder) Popov, and Art. 40.6, i.e. the word “lectotypus” was used. These conditions were first fulfilled by Fryer & Hylmö (2009), who accepted the name, provided a full and direct reference to the Latin description in Pojarkova (1961), and indicated *Wilson 1317* in fruit at A as holotype.

Lu & Brach (2003) returned *Cotoneaster tomentellus* to the synonymy of *C. soongoricus*. The latter is a Central Asian species described from Kazakhstan that readily differs in its larger and broader leaves, which are totally glabrous above (vs. rather densely hairy in *C. tomentellus*). *Cotoneaster tomentellus* occurs in Central China, whereas the distribution area of *C. soongoricus* embraces mountainous areas of E Central Asia.

20. A new species transfer in *Sorbaronia*


Skvortsov & Maitulina (1982) compared the cultivar of chokeberries that was very common in Russian orchards with the wild species *Aronia melanocarpa* (Michx.) Elliott native to North America. The cultivated plants were demonstrably monomorphic and obviously differed in their fruit, which is at least twice as large, fleshy, depressed-globose, and dull black because of pubescence (vs. pyriform, shiny black, and completely glabrous), and in their leaves, which are elliptic (vs. lanceolate) with a subrounded (vs. acute) apex. The cultivated plants were also much hardier (Rehder’s hardiness zone II vs. IV), and their fruits had a different taste and were much more juicy. The change of fruit dimensions may be ascribed to artificial selection, whereas the fruit shape and hairiness and the leaf shape are obviously novelties of the cultivar.

In its native area the black-fruited *Aronia melanocarpa* is predominantly diploid in New England and tetraploid outside (Brand 2010), reproducing sexually with production of highly heterogeneous offspring, whereas the cultivated plants are tetraploid, with the offspring being identical, thus indicating apomixis (Skvortsov & al. 1983; Persson-Hovmalm & al. 2004; Leonard 2011). Most of researchers who discussed the origin of cultivated chokeberries (Skvortsov & al. 1983; Persson-Hovmalm & al. 2004) inferred that this cultivar is most
likely an infraspecific hybrid derived from *A. melanocarpa*.

The early proof of hybrid origin of the cultivated chokeberries came from Ivan Michurin, who created the cultivar and introduced it to the Russian market. Michurin (1948) stated that in 1905, in order to produce sweet-fruited cultivars of “rowans” (*Sorbus* s.l.) that may be suitable to the Russian North and Siberia, he pollinated the native Russian *S. aucuparia* L. with the introduced *Aronia melanocarpa*. The resulting hybrid was named “Likernaya” (Liquor Rowan); after Michurin’s death in the 1940s it was distributed nameless to the Soviet market (Skvortsov & al. 1983).

Leonard (2011) studied the relationships of *Aronia mitschurinii* to the wild *Aronia* species and several other genera of *Malinae* Reveal (*Pyrinae* Dumort., formerly *Maloidae* C. Weber) using amplified fragment length polymorphism (AFLP) analysis. In this study Leonard confirmed that *A. mitschurinii* was clearly distinct from the wild species of *Aronia* and its first parent was most likely *A. melanocarpa* (less likely *A. prunifolia* (Marshall) Rehder, which is a hybrid between the black-fruited species and the red-fruited *A. arbutifolia* (L.) Pers.). The second parent was confirmed to be *Sorbus aucuparia*, and the genetic proximity to the *Aronia* parent indicated one or more acts of backcrossing with it. Again, Michurin’s memoirs agree with these genetic studies: Michurin (1948) reported that he selected the new cultivar from three generations of crosses.

The intergeneric hybrids between *Aronia Medik.* and *Sorbus* L. are known under the name *Sorbaronia* C. K. Schneid. *Sorbaronia* embraces highly variable morphotypes that are mostly intermediate between the parents. The hybrids of *A. melanocarpa* and *S. aucuparia* are called *Sorbaronia fallax* (C. K. Schneid.) C. K. Schneid.; they are small trees with compound leaves and purple-black fruit (Rehder 1940; Mezhensky 2005) that have never been considered taxonomically identical with *A. mitschurinii*. Because of its distinctive morphology and stable characters caused by predominantly apomictic reproduction, *A. mitschurinii* should be treated as an apomictic microspecies of cultigenous origin that is taxonomically distinct from *S. fallax*.

In spite of the intergeneric origin, Leonard (2011) decided to maintain *Aronia mitschurinii* in *Aronia* because of its greater similarity to the species of the latter. However, an analogous case when variable intergeneric hybrids with *Sorbus* as one of the parents are accommodated in a single hybridogenous genus is *Sorbocotoneaster* Pojark. The natural hybrid between *Sorbus aucuparia* L. and *Cotoneaster laxiflorus* Lindl. (syn. *C. melanocarpus* (Bunge) Loudon), *Sorbocotoneaster pozdnjakovii* Pojark., was studied morphologically (Pojarkova 1953) and cytologically (Krügel 1992). Three distinct, yet sympatric, morphotypes were discovered: the primary hybrid (triploid) closely approaching *S. aucuparia*, the putative backcross with *S. aucuparia* (tetraploid) intermediate between the parents, and the putative backcross with *C. laxiflorus* (pentaploid) closely approaching *C. laxiflorus*. If it were not for the apomictic reproduction, *A. mitschurinii* would have been taxonomically included in *S. fallax*; as far as its taxonomic separation is maintained, its transfer to *Sorbaronia* is justified and is consequently effected here.
Fig. 3. Neotype of *Mespilus rotundifolia* – Ehrhart (1792), Plantae selectae hortuli proprii 3: No. 26 (M 0213350).
When established in the wild, in East Europe Sorb-ronia mitschurinii shares the ecological preferences of its Aronia parent. In their native area the species of Aronia occur in bogs, swamps and wet woods, but can be found also on dry soils (Gleason & Cronquist 1991). In European Russia a few instances of naturalization of S. mitschurinii were noted, e.g. in pine forests with Polytrichum spp. around peat bogs or in secondary dry pine forests in the Vladimir Region (Seregin 2010; A. Seregin pers. comm.), in moist pine forests along margins of bogs in the Leningrad Region (pers. obs.), and along margins of peat bogs in Bashkiria (A. Muldashev pers. comm.). All these records are no less than 20 years old (e.g. Seregin 2010); although the active and very rapid migration of this species into natural habitats has just started, it seems to be the beginning of a large future expansion because the species is very common in crop and ornamental cultivation and is very much liked by birds who actively disperse it. In some areas of the Vladimir Region the occurrence of this species in natural habitats is already regular (A. Seregin pers. comm.).

21. The identity of “Crataegus hordida” and changes in the infraspecific taxonomy of C. chrysocarpa


A species of American origin cultivated in Europe, “Crataegus hordida Medik.” was reportedly escaped from ornamental cultivation in East Europe (Tzvelev 2001) and occasionally naturalized in the Kaluga Region of European Russia (Reshetnikova & Krylov 2006).

The name Crataegus hordida was interpreted by Cinovskis (1971), who resurrected it for C. chrysocarpa Ashe, which is the second most wide-ranging species of North American Crataegus (Phipps & O’Kennon 2004) currently classified as a member of C. sect. Coc-cineae Loudon (Tzvelev 2001) and C. ser. Rotundifolii Rehder (Phipps 1983). Nevertheless, this name has never been accepted for this species in its native distribution range; neither it is even listed in synonymy (e.g. Gleason & Cronquist 1991; Phipps & O’Kennon 2004).

In the place of its original publication, Medikus (1793) introduced Crataegus hordida without any descriptive matter but with two references: to Mespilus ro-tundifolia Ehrh. (Ehrhart 1784, 1788, 1789) as a doubtful synonym, and to C. rotundifolia Moench (Moench 1785) as a plant different from that of Medikus. Except for these citations, Medikus included no references to previously published names on which C. hordida might be based and no reference to a previously published description that was applicable to the species. Consequently, C. hordida was not validly published in 1793.

As far as we are aware, Cinovskis (1971) was the first to accept the name Crataegus hordida after Medikus. In doing so, he did not fulfill the conditions for valid publication of a new name, as he provided no validating description or reference thereto, and no type designation, but he did include full and direct references to many synonyms, of which three, C. calyciglabra Schuette, C. chrysocarpa var. phoenicea E. J. Palmer, and Mespilus rotundifolia Ehrh., are validly published names that may serve as replaced synonyms. The earliest synonym, M. rotundifolia Ehrh., is considered type-providing here. Because Cinovskis included in synonymy legitimate species names the epithets of which were not then occupied in Crataegus, the name C. hordida Moench ex Cinovskis was nomenclaturally superfluous when published and is therefore illegitimate (Art. 52.1). Since the earliest legiti-mate species name the epithet of which should have been accepted by Cinovskis, i.e. C. chrysocarpa Ashe, was included in a subordinate taxon (as C. hordida var. chrysocarpa (Ashe) Cinovskis, nom. non rite publ.), C. chrysocarpa does not automatically provide the type for C. hordida (Art. 7.5).

Mespilus rotundifolia was described by Ehrhart (1784, 1788, 1789) from cultivation in German gardens, most likely on the basis of living plants because no specimen of M. rotundifolia was preserved in Ehrhart’s main herbarium, which is currently housed at MW (Karavaev & Barsukova 1968; Balandin 2004). As is evident from his Exsiccatae (Britten 1922; Gubanov & Balandina 2000; Balandin 2006), Ehrhart cultivated the plant in his own garden in Hannover. A later gathering from this cultivation was distributed by Ehrhart (1792), and its specimen at M (best preserved with the most characteristic shape of leaves: Fig. 3) is designated here as the neotype of M. rotundifolia.

As is evident from examination of Ehrhart’s collections housed in three Herbaria (GOET, M, MW), the original Mespilus rotundifolia has nearly glabrous leaves that are hairy only along the main veins, and its inflor-escence is glabrous as correctly assumed by Sargent (1901) and Palmer (1937). Several varieties are formally recognized within the variable Crataegus chrysocarpa, the glabrous one being currently known as “C. chrysocarpa Ashe var. phoenicea E. J. Palmer” (Phipps 2012). However, the leaves of Ehrhart’s plant clearly deviate from those described in Phipps (2012) in having much longer
and acuminate lobes. A new combination at the rank of variety is proposed here to accommodate this morphotype; the new varietal name is necessarily based on *M. rotundifolia* because Sargent (1901) already legitimately used that epithet at varietal rank (Art. 11.4).

**Crataegus chrysocarpa** var. *phoeniceoides* J. B. Phipps & Sennikov, var. **nov.**


Validating description: that of *Crataegus chrysocarpa* var. *phoenicea* J. B. Phipps (see below).


Phipps (2012) inadvertently published a later homonym of *Crataegus chrysocarpa* var. *phoenicea* E. J. Palmer for the glabrous variant of *C. chrysocarpa* with shorter leaf lobes. This illegitimate name is changed here with a new combination based on the validating description of *C. chrysocarpa* var. *phoenicea* J. B. Phipps and the specimen that was designated as the epitype of this name. In the type designation, the flowering specimen from Rhode Island is preferred over the fruiting specimen from Vermont because it unambiguously demonstrates the glabrous inflorescence that is typical of the taxon. The fruiting specimen designated as the holotype by Phipps (2012) is inadequate as it does not unambiguously prove that the inflorescence, which was glabrous in fruit, was not hairy in flower.

22. **Validation of the name Sorbus tauricola**

*Sorbos tauricola* Zaik. ex Sennikov, sp. **nov.**

Holotype: Ukraine, Crimea, Ai-Petri Mt, NE of weather station, 960 m, gravelly slope, 22 Jun 1956, *K. Popov* (LE [flowering specimen]; isotypes: KW, SIMF) – Fig. 7.


**Illustrations** — Fig. 5 & 6; *Popov* (1959a: fig. 1, photo of a fruiting branch), *Popov* (1959a: fig. 2v, z, line drawing of a leaf and dissected flower).

**Latin description** — *Popov* (1959a: 188).

*Description* — *Trees or shrubs* up to 4–5 m high. *Bark* grey, with scattered lenticels. *Twigs* brownish grey; *young shoots* brown, loosely tomentose when young, almost glabrous at maturity, with numerous ochraceous lenticels. *Buds* narrowly ovoid, 3–4 mm long, 2–3 mm wide; *bud scales* brown, margin and apex tomentose. *Leaves* (of abbreviated fertile shoots) simple: *petiole* 1.5–2.2 mm long, tomentose; *leaf blade* slightly glossy, dark green (becoming red in autumn) above, grey-green beneath, ovate-elliptic, ± flat, regularly and prominently lobed, 7–9 cm long, 5.5–7 cm wide, widest at 53%–68% of lamina length (from apex), lower surface evenly tomentose, upper surface almost glabrous, base broadly cuneate, straight or very slightly arculate, margin ± flat, apex broadly triangular and acuminate; *veins* 6(or 7) on each side; *lobes* 4 or 5 on each side, sides slightly arculate, margin serrate with 2 or 3 teeth along shorter side and 6–10 teeth along longer side, apex acuminate; *leaf blade* slightly glossy, dark green (becoming red in autumn) above, grey-green beneath, ovate-elliptic, ± flat, regularly and prominently lobed, 7–9 cm long, 5.5–7 cm wide, widest at 53%–68% of lamina length (from apex), lower surface evenly tomentose, upper surface almost glabrous, base broadly cuneate, straight or very slightly arculate, margin ± flat, apex broadly triangular and acuminate; *veins* 6(or 7) on each side; *lobes* 4 or 5 on each side, sides slightly arculate, margin serrate with 2 or 3 teeth along shorter side and 6–10 teeth along longer side, apex acuminate; lowermost lobe slightly bent downwards, 1.2–1.5 cm wide; incision between 2 lowermost lobes 1–1.5 cm long. *Inflorescence* with 12–20 flowers, racemose-corymbose, compact, 4–6 cm in diam.; *branches* tomentose. *Flowers* 15–18 mm in diam.; *hymenium* turbinate, tomentose in flower, subglabrous in fruit; *sepals* erect, persistent, triangular, 1.3–1.8 mm long, 1.7–2 mm wide, densely tomentose on both surfaces, apex acuminate; *petals* white, elliptic to broadly elliptic, 6–7 mm long, c. 4 mm wide; *stamens* c. 20; *filaments* white; *anthers* pale lilac, 1.7–1.8 mm long; *styles* 2. *Fruits* 3–10 per inflorescence (fertility low), when ripe bright orange, ellipsoid, 10–11 × 8–9 mm, without lenticels. *Seeds* 1 or 2 per fruit, 4–5 mm long.

**Chromosome number** — 2*n* = 4x = 68 (*Zaikonnikova & Kipiani* 1980).
Etymology — The species epithet is derived from Chersonesus Taurica (the Latin version of the original Greek name for the Crimean Peninsula) and the Latin word element -cola (“inhabitant”), thus meaning “inhabiting the Crimea”.

Distribution and ecology — Ukraine, endemic to the Crimea (Fig. 4). The distribution area stretches as a narrow stripe for c. 15 km along the main range of the Crimean mountains, mostly on the southern side of the range (Popov 1959a, b: fig. 3 [map]). The species grows as scattered individuals on open rocks and in underwood up to the upper limit of the forest belt, at altitudes of 700–1150 m.


Possible origin — Sorbus aria (L.) Crantz s.l. × Sorbus torminalis (L.) Crantz (Zaikonnikova & Kipiani 1980; Zaikonnikova 1985).

Other specimens examined (paratypes) — Ukraine, Crimea, Ai-Petri Mt, NE of weather station, 960 m, gravelly slope, 19 Sep 1956, K. Popov (KW, LE, SIMF); Ai-Petri Mt, beech forest, 12 Jul 1919, S. Stankov (LE, YALT). An extensive list of other specimens is provided in Popov (1959a).

The Crimean whitebeams that belong to the apomictic complex of Sorbus latifolia (Lam.) Pers. s.l. were first discovered by Zelenetzky (1906), who identified them as “S. scandica Fries”. Stankov (1927) reported the plant as S. latifolia. Popov (1959a) acknowledged the isolated occurrence of the Crimean plants and their difference from Central European whitebeams and described the Crimean populations as S. pseudolatifolia K. Popov. Popov provided an extensive description in Latin but designated two gatherings as types, one in flower and the other in fruit, making the new name not validly published under Art. 40.1. Zaikonnikova (1985) renamed the species because of homonymy but omitted the type designation, and this issue was also neglected in a recent list of Ukrainian type specimens (Fedoronchuk 2006). The name suggested by Zaikonnikova is in current use (Czerenpanov 1995; Mosyakin & Fedoronchuk 1999; Zaikonnikova 2001; Yena 2012) and is validly published here. Under Art. 46.5 and 46.10 its authorship is “Zaik. ex Sennikov”; the name may not be attributed to Zaikonnikova alone under Art. 46.2.

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Fig. 7. Holotype of *Sorbus tauricola* – Flowering (right-hand) specimen, K. Popov (LE).
graphs of Sorbaronia mitschurinii and Sorbus tauricola. Sampsø Lommi (Helsinki) produced the map of S. tauricola. Special thanks go to Mikhail Kozhin (Moscow), Marc Appelhans (Göttingen), Franz Schuhwerk and Christian Bräuchler (München) for locating and scanning specimens of the type collection of Mespilus rotundifolia. Ivan Tatanov (St Petersburg) scanned the type of S. tauricola. An image of the neotype of M. rotundifolia is reproduced with permission from the Botanische Gesellschaft bei den Naturforschern in Hannover, and Ivan Tatanov (St Petersburg) scanned the type of S. tauricola. An image of the neotype of M. rotundifolia is reproduced with permission from the Botanische Staatssammlung München; an image of the holotype of S. tauricola is reproduced with permission from the Komarov Botanical Institute. Arto Kurnto (Helsinki) is warmly thanked for continuous encouragement. The text has benefited from critical comments of Hugh McAllister and has benefited from critical comments of Hugh McAllister warmly thanked for continuous encouragement. The text has benefited from critical comments of Hugh McAllister warmly thanked for continuous encouragement. The text has benefited from critical comments of Hugh McAllister warmly thanked for continuous encouragement. The text has benefited from critical comments of Hugh McAllister warmly thanked for continuous encouragement. The text has benefited from critical comments of Hugh McAllister warmly thanked for continuous encouragement.

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