Flora and factors affecting species diversity in the islet groups of the protected “Natura 2000” sites of the Amvrakikos Gulf and Mesolongi Lagoon (Ionian area, Greece)

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Abstract

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The vascular plant species diversity of the islet groups in the Amvrakikos Gulf and Mesolongi Lagoon, hitherto unexplored, is documented by an inventory of vascular plants, filling gaps in the floristic information available for the two National Parks to which these islet groups belong, as well as for the Ionian land-bridge island system for which detailed studies concerning floristic and phytogeographical relationships were available before. The total vascular flora of the two islet groups comprises 226 plant taxa. Spectra of life-form and chorological elements are presented and discussed. Floristic cross-correlations (beta-diversity) show that the individual islet floras differ from each other from 39 % to 91 %. Concerning the role of eco-geographical variables as influencing plant species diversity in the area, regression analyses indicate island area as the major predictor variable of species richness.

Additional key words: island biogeography, land-bridge islands, phytogeography, Mediterranean region, vascular plants, beta-diversity

Introduction
Plant species richness and distribution patterns on oceanic versus chersogenous islands have been extensively studied in island biogeography. An important point is to determine factors affecting species richness and repartition on islands. This becomes critical when the islands concerned are protected by law and conservation efforts are claimed.

The ecological importance of the Ionian area from a nature conservation point of view was mentioned by Iliadou & al. (2014). The designation of two national parks and Ramsar Convention sites of international importance for birds on the border of W mainland Greece and the Ionian archipelago makes it important to elucidate the biogeographical relationship of the Ionian area to the adjacent parts of the Greek mainland.

Plant species diversity of some larger Ionian islands and islet groups has been studied already, but there is still a large number of small islands and islets that have not been investigated at all and have remained completely unexplored floristically, such as the islet groups in the marginal seas of the Amvrakikos Gulf and the Mesolongi Lagoon (Fig. 1).

The islet group in the Amvrakikos Gulf belongs to the Natura 2000 Site of Community Importance GR2110001, “Amvrakikos Gulf, Louros and Arachthos Delta”, which also includes the Special Protection Area GR2110004, “Amvrakikos Gulf, Katakourko lagoon and Korakonisia”. Both sites belong to the “Amvrakikos Wetlands National Park”. The islet group in the Mesolongi Lagoon is part of two overlapping Natura 2000 sites, a Special Protection Area (GR2310015) and a Site of European
Community Importance (GR2310001), which compose the “National Park of Mesolongi – Aitoliko lagoons, estuaries of Acheloos and Evinos River and Echinades islet group”. These two national parks are of the most diverse and complex sites of the Natura 2000 ecological network in Greece and also represent two of Greece’s 11 Ramsar Convention wetlands, which are of international importance for the conservation of their avifauna (Dimopoulos & al. 2005).

The floristic composition of the two islet groups under discussion here (Fig. 1) was hitherto unknown, and for this reason the main objectives of the present study are: (1) to provide for the first time a floristic inventory of the protected islet groups, thus continuing current studies of the plant species diversity of the Ionian islets (Iliadou & al. 2014); (2) to investigate the floristic similarities among the individual islets; and (3) to explore the main factors affecting plant species richness in the two islet groups in order to highlight the role of geographical variables in shaping their flora. In the given conservation context, factors determining plant species richness and habitat composition may also directly or indirectly influence animal and especially bird species richness (Kissling & al. 2008).

Material and methods

1. Study area

The survey of the two investigated islet groups covers six islets of the Amvrakikos Gulf and four islets of the Mesolongi Lagoon with areas ranging from 0.41 ha to 65.9 ha, very low elevation ranging from 1 m to 8 m a.s.l. and distance from the nearest mainland coastline ranging from 10 m to 5.7 km (Table 1). The low hilly and coastal slopes are generally gentle. The names of the constituting islets and their geographical data are provided in Fig. 1 and Table 1. The islets are uninhabited, but there are one or two small houses for people working in the fish farms that are the main human activity in the surrounding sea. Grazing has been observed on the larger islets. The Amvrakikos Gulf islets belong to the prefecture of Preveza, except the islet of Kefalos, which belongs to the prefecture of Aitolioakarnania as also do the islets of the Mesolongi Lagoon. All the islets are owned by the Greek state.

The Amvrakikos Gulf is also a semi-closed sea, watered by plenty of fresh water with sediments. Vött & al. (2007) showed geomorphological, sedimentological and geoarchaeological evidence of multiple tsunami impacts on the Aktio headland (the entrance of the Amvrakikos Gulf). May & al. (2012), studying the Lefkadi–Preveza...
coastal zone, which is characterized by an active barrier system and related extensive beachrock sequences protecting the entrance to the gulf, concluded that tsunami events in particular have induced sudden impulses of coastal evolution in the study area contributing to coastal and environmental changes involving a recurrent reorganization of the coastal system by gradual, long-term, steadily acting processes such as longshore drift, tending to the establishment of a state of renewed coastal balance.

The islets in the Amvrakikos Gulf are composed of Triassic/Lower Jurassic or Jurassic limestones (IGME 1989). The Mesolongi Lagoon is connected to the Gulf of Patras, which forms a part of the Ionian Sea. The lagoon lies behind a fragmented sandy coastal ridge that separates it from the open sea. It is semi-closed and shallow (the depth ranges from a few cm in the interior regions up to roughly 2.5 m, with a mean depth of approximately 0.5 m) and the circulation is dominated by the tide entering through the Gulf of Patras, at least for ordinary wind conditions (Leftheriotis & al. 2013). The islets in the Mesolongi Lagoon are composed of post-tectonic and late-tectonic sediments of the Holocene (alluvium) in valleys, plains and coastal deposits (IGME 1989). They form a kind of barrier islets (Fig. 1).

Based on the climatic diagram of Emberger (1952), the study area belongs to a subhumid bioclimatic characterized by mild winters. The xerothermic index (Bagnouls & Gaussen 1953) and the ombrothermic diagram (UNESCO-FAO 1963) characterize the study area as thermo-Mediterranean with a dry period from early May to mid-October.

Three habitat types of Annex I of the Directive 92/43/EU have been recognized in the study area of the Amvrakikos Gulf islets: vegetated sea cliffs of the Mediterranean coasts (habitat type code 1240), Mediterranean and thermo-Atlantic halophilous scrub (Sarcocornietea fruticosi) (1420) and Olea europaea – Pistacia lentiscus scrub (9320). Five habitat types of Annex I have been recognized on the islets of the Mesolongi Lagoon: habitat type 1420, as in the Amvrakikos Gulf islets, Salicornia and other annuals colonizing mud and sand (1310), Mediterranean salt meadows (Juncetalia maritimi) (1410), embryonic shifting dunes (2110) and coastal dunes with Juniperus species (2250*); plus one more habitat type not included in Annex I: reedbeds (72A0).

2. Annotated floristic inventory

The floristic inventory presented here (see Appendix) includes species and subspecies, with their life-form and chorological categories, of the flora of the individual islets of the two groups. Repeated field trips were carried out between 2007 and 2012 in different seasons of the year, in order to acquire an integrated knowledge of the flora of the islets.

Families, genera, species and subspecies are given in an alphabetic order within the higher groups gymnosperms and angiosperms. For simplicity, “taxa” is statistically used hereafter for both species and subspecies (the latter in case a species has more than one subspecies in the area). The plant collections are deposited in Agrinio (W Greece) in the herbarium of the Department of Environmental and Natural Resources Management, Laboratory of Ecology and Nature Conservation.

Nomenclature, life-form and chorological categories follow Dimopoulos & al. (2013) except for Spergularia marina, the correct name for S. salina (see Buttler & Thieme 2013, the latter designation erroneously accepted in Dimopoulos & al. 2013: 83). For the determination of the plant material, Tutin & al. (1968–1980, 1993), Pignatti (1982) and Strid & Tan (1997, 2002) were used.

3. Floristic similarity analysis

Floristic similarity of the islets per pair (beta-diversity) was calculated using the Sørensen similarity coefficient (Sørensen 1948): Cs=2j/(a+b)/C, where j = the number of taxa common to both islets compared, a = the number of taxa recorded from islet 1, and b = the number of taxa recorded from islet 2. In addition, the proportion of islets on which a taxon occurs was calculated. The dendrogram presenting floristic relations between the islets studied (Fig. 2) is based on the hierarchical cluster analysis using Statistica 6 (StatSoft, Inc. 2001).

4. Statistical analysis of the factors affecting plant species richness

Geographical data concerning surface area (A), highest elevation (E) and maximum distance to the nearest mainland (Dm) were determined by open-access digital topographic maps (1:50 000 scale) of the Hellenic Military Geographical Service.

The relationships between plant species richness and the geographical variables affecting it were tested using simple and multiple linear regression analyses and the Spearman correlation coefficient. All regressions and the estimations of parameters were carried out with Statistica 6 (StatSoft, Inc. 2001).

Results

1. Floristic inventory and analysis

There is no previous literature information at all concerning plant species richness of the islets studied. Our pioneer field investigations on all of the ten islets revealed the occurrence of 226 plant taxa.

The vascular plant diversity of the six Amvrakikos Gulf islets comprises 142 plant taxa, belonging to 45 families, 117 genera, 130 species and 12 subspecies, while the vascular plant diversity of the four Mesolongi Lagoon islets comprises 164 plant taxa, belonging to 46 families, 126 genera, 155 species and nine subspe-
cies (Table 2). Except for two gymnosperms, all of the taxa are angiosperms. Vascular cryptogams have not yet been found.

In both island groups a high percentage (c. 43%) of the taxa belongs to three families: Poaceae (23 taxa), Asteraceae and Fabaceae (19 taxa each). Therophytes predominate (52.1% – 53%) and Mediterranean plant taxa represent 75.2% of the Amvrakikos Gulf islets flora and 66.7% of the Mesolongi Lagoon islets flora. For the life-form spectrum see Table 3; for the chorological spectrum see Table 4.

Four alien taxa, according to Arianoutsou & al. (2010), are present in the flora of the islets studied, viz.: Erigeron bonariensis, Phalaris canariensis, Symphyotrichum squamatum and Xanthium orientale subsp. italicum.

Eleven of the taxa recorded are included in the IUCN Red List of Threatened Species (IUCN 2013) and three are included in the European Red List of Vascular Plants (Bilz & al. 2011), all characterized as Least Concern (LC); 27 taxa are included in the list of UNEP (2014) (see Table 5).

2. Floristic similarity among the studied islets

Floristic cross-correlations (beta-diversity) between the studied islets by means of the Sørensen similarity coefficient revealed values (0.09–0.61) showing that the floras of each of the studied islets differ from each other from 39% to 91% (Fig. 2). The hierarchical cluster analysis showed that the island of Vouvalos (AG1), highest and largest of the Amvrakikos Gulf islets group, has a very different floristic composition as also does the medium-sized islet of Kefalos (AG5), when compared to all the other islets (Fig. 2). The other four, smaller Amvrakikos Gulf islets group together, as also do the four Mesolongi Lagoon islets (Fig. 2).

Concerning the number of islets on which each taxon occurs, 33.2% of the taxa occur on only one of the ten studied islets, 23.5% on only two of them and 15.9% on only three of them. Only 1.3% of the taxa were found to be common to all ten islets, and 0.4% common to nine of them. The most widespread plant taxa recorded on the studied islets are Asparagus acutifolius, Limbarda crithmoides and Pistacia lentiscus (on all ten studied islets), Smilax aspera (on nine islets) and Asphodelus ramosus, Elytrigia juncea and Rubia peregrina (on eight islets).

Except the above-mentioned taxa, there are others found on five or all six islets of the Amvrakikos Gulf, such as Lonicera impexa, Myrtus communis and Olea euro-

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Table 2. Numbers of vascular plant taxa in the flora of the two studied islet groups. – AG = Amvrakikos Gulf islets; ML = Mesolongi Lagoon islets.

<table>
<thead>
<tr>
<th>Area</th>
<th>Families</th>
<th>Genera</th>
<th>Species</th>
<th>Subspecies</th>
<th>Taxa</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gymnosperms AG</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>0</td>
<td>2</td>
<td>1.4</td>
</tr>
<tr>
<td>ML</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0.6</td>
</tr>
<tr>
<td>Angiosperms AG</td>
<td>43</td>
<td>115</td>
<td>128</td>
<td>12</td>
<td>140</td>
<td>98.6</td>
</tr>
<tr>
<td>ML</td>
<td>45</td>
<td>125</td>
<td>154</td>
<td>9</td>
<td>163</td>
<td>99.4</td>
</tr>
<tr>
<td>Total AG</td>
<td>45</td>
<td>117</td>
<td>130</td>
<td>12</td>
<td>142</td>
<td>100</td>
</tr>
<tr>
<td>ML</td>
<td>46</td>
<td>126</td>
<td>155</td>
<td>9</td>
<td>164</td>
<td>100</td>
</tr>
</tbody>
</table>

Table 3. Life-form spectrum of the flora of the two studied islet groups.

<table>
<thead>
<tr>
<th>Life-form category</th>
<th>Amvrakikos Gulf islets</th>
<th>Mesolongi Lagoon islets</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of taxa</td>
<td>%</td>
<td>Number of taxa</td>
</tr>
<tr>
<td>Phanerophytes</td>
<td>15</td>
<td>10.6</td>
</tr>
<tr>
<td>Chamaephytes</td>
<td>12</td>
<td>8.4</td>
</tr>
<tr>
<td>Hemicryptophytes</td>
<td>24</td>
<td>16.9</td>
</tr>
<tr>
<td>Geophytes</td>
<td>17</td>
<td>12</td>
</tr>
<tr>
<td>Therophytes</td>
<td>74</td>
<td>52.1</td>
</tr>
<tr>
<td>Total</td>
<td>142</td>
<td>100</td>
</tr>
</tbody>
</table>

Table 4. Chorological spectrum of the flora of the two studied islet groups.

<table>
<thead>
<tr>
<th>Chorological category</th>
<th>Amvrakikos Gulf islets</th>
<th>Mesolongi Lagoon islets</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of taxa</td>
<td>%</td>
<td>Number of taxa</td>
</tr>
<tr>
<td>1. Widespread taxa</td>
<td>34</td>
<td>24.1</td>
</tr>
<tr>
<td>Cosmopolitan</td>
<td>7</td>
<td>5</td>
</tr>
<tr>
<td>Subtropical-tropical</td>
<td>3</td>
<td>2.1</td>
</tr>
<tr>
<td>Euro-Siberian</td>
<td>2</td>
<td>1.4</td>
</tr>
<tr>
<td>European-SW Asian</td>
<td>13</td>
<td>9.2</td>
</tr>
<tr>
<td>Paleotemperate</td>
<td>9</td>
<td>6.4</td>
</tr>
<tr>
<td>2. Mediterranean taxa</td>
<td>106</td>
<td>75.2</td>
</tr>
<tr>
<td>East Mediterranean</td>
<td>5</td>
<td>3.5</td>
</tr>
<tr>
<td>Mediterranean</td>
<td>69</td>
<td>49</td>
</tr>
<tr>
<td>Mediterranean-Atlantic</td>
<td>7</td>
<td>5</td>
</tr>
<tr>
<td>Mediterranean-European</td>
<td>15</td>
<td>10.6</td>
</tr>
<tr>
<td>Mediterranean-SW Asian</td>
<td>10</td>
<td>7.1</td>
</tr>
<tr>
<td>3. Alien taxa</td>
<td>1</td>
<td>0.7</td>
</tr>
<tr>
<td>Total</td>
<td>141</td>
<td>100</td>
</tr>
</tbody>
</table>
3. Statistical analysis of the factors affecting plant species richness

The Spearman correlation coefficient was calculated among biogeographical factors to evaluate multi-collinearity of these predictor variables. Surface area (A), elevation (E) and distance to the mainland (Dm) are not correlated to each other.

The results of simple regression analyses (log-log) of plant species richness indicated island surface area as a predictor variable of species richness since they are strongly correlated ($R^2 = 0.741$). Elevation and distance to the mainland do not show a significant effect on species richness. Fig. 3 presents the logarithmic species-area relationship (SAR) for the islets studied with the equation $\log S = 1.507 + 0.343 \log A$. Stepwise multiple regression analyses were conducted among the number of plant taxa and the set of the studied biogeographical factors. Only the surface area (A) is a strong predictor variable of plant species richness since plant species richness is not significantly correlated with elevation (E) and distance to the adjacent species pool (Dm).

Discussion

There has been a complete gap so far concerning plant species richness of the islets studied, which are included in the protected areas of the “Amvrakikos Wetlands National Park” and the “National Park of Mesolongi – Aitoliko lagoons, estuaries of Acheloos and Evinos River and Echinades islet group”. During our field investigations on the six islets in the Amvrakikos Gulf and the four islets in the Mesolongi Lagoon, 226 plant taxa were registered representing 11.1% of the total flora of the Ionian area (2027 taxa) as recorded by Dimopoulos & al. (2013). A large proportion of these taxa belong to the families Asteraceae, Fabaceae and Poaceae, which are among the families adapted best to the ecological conditions of the Mediterranean area as is also confirmed by many floristic studies of Greek insular areas (Panitsa & Tzanoudakis 2001, 2010; Kougioumoutzis & al. 2012; etc.). The Mediterranean character of the flora of the studied islet groups is reflected also by the high proportions of the Mediterranean plant taxa, in conjunction with the high percentage of therophytes.

Floristic cross-correlations of the studied islets (beta-diversity) according to the Sørensen similarity coefficient revealed values from 0.09, indicating high floristic heterogeneity, to 0.61, showing significant floristic similarity. Hierarchical cluster analyses divided the islets studied into three subgroups mainly based on surface area and the geographical position.

paea subsp. europaea, and also on all four islets of the Mesolongi Lagoon, such as Juncus maritimus, Limonium narbonense, L. virgatum, Salsola tragus, Scirpoidea holoschoenus and Tamarix hampeana.
Furthermore, the proportion of islets on which a taxon occurs showed that about 20% of the taxa are frequent on more than half of the islets, while 72.6% of the taxa are rare or not common in the islet group, being found only on one to three of the islets. Island similarities concerning all native taxa show a significant geographical component, reflecting the spatial autocorrelation of floras before the fragmentation of the mainland – or the effects of intensive post-fragmentation dispersal (Triantis & al. 2008b).

The high occurrence on the studied islets of taxa such as *Myrtus communis*, *Olea europaea* subsp. *europaea* and *Pistacia lentiscus* is a nice example of one of the most outstanding ecological plant-avian frugivore interactions. Panitsa & Tzanoudakis (2010) also mentioned that the high incidence of *Olea europaea* and *Pistacia lentiscus* represent characteristic examples of plant species introduction via sea birds to islets around the E Aegean island of Leros.

As the islet groups of the Amvrakikos Gulf and Mesolongi Lagoon belong to national parks that are nature reserves under the International Ramsar Convention and designated as Important Bird Areas, it is critical to mention here that the islets studied, as a part of warm lowlands in the Mediterranean, are also important wintering quarters for European frugivorous birds. González-Varo (2010) suggested that, in a conservation context, it may be possible to sustain frugivore animal numbers by promoting succession within extant patches of vegetation.

### Table 5. Plant taxa on the two studied islet groups that are registered under a protection status.

<table>
<thead>
<tr>
<th>Taxon</th>
<th>AG</th>
<th>ML</th>
<th>Presidential Decree 67/1981</th>
<th>CITES</th>
<th>IUCN</th>
<th>European Red List</th>
<th>UNEP WCMC</th>
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<tbody>
<tr>
<td>Achillea maritima</td>
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<tr>
<td>Aegilops biuncialis</td>
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<td>Aeluropus littoralis</td>
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<td>Alkanna tinctoria</td>
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<tr>
<td>Arthrocnemum macrostachyum</td>
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<td>Calystegia soldanella</td>
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<tr>
<td>Centaurium erythraea</td>
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<td>Cyclamen hederifolium</td>
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<td>Ephedra foeminea</td>
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<td>Euphorbia dendroides</td>
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<td>Euphorbia paralias</td>
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<td>Euphorbia peplis</td>
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<td>Geranium purpureum</td>
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<td>Glaucium flavum</td>
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<td>Juncus inflexus</td>
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<td>Juniperus phoenicea</td>
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<td>Medicago marina</td>
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<td>Parapholis incurva</td>
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<td>Ranunculus chius</td>
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instead of increasing woodland cover (reforestation) at landscape level.

In addition, the role of the Mesolongi Lagoon islets as barrier islets should be noted. Looney & Gibson (1995) noticed that on coastal barrier islands the seed bank of low disturbance, late-succession vegetation types was well developed in contrast to a poorly developed and transient seed bank occurring in more frequently disturbed, early successional dredge-spoil unvegetated areas and strand. Stallins & Parker (2003) outlined the complex interplay between disturbance and environmental gradients (plant species abundances, landform-mediated physical gradients, sediment mobility) that influence biogeographical patterns. This is in accordance with relatively high plant species richness and the absence of endemic plant taxa on the investigated islets that could additionally be attributed to their low elevation and their relatively recent isolation from the mainland.

The investigation of factors affecting plant species richness showed that, for the islet groups studied here, the total species richness is strongly correlated to island surface area. There is a strong species-area relationship \( R^2 = 0.74 \), as was also found for the islands of the S Aegean \( R^2 = 0.73 \) by Kagiampaki & al. (2011). This corroborates that, in general, surface area is considered the most influential explanatory variable of species richness in island biogeography (Whittaker & Fernández-Palacios 2007; Triantis & al. 2008a, 2008b).

The logarithmic approach of the power function \( S = cA^z \) of Arrhenius's model (1921) explains best the variation in species richness (Willerslev & al. 2002) and shows higher slopes for “island”-like species-area relationships than “mainland” relationships according to Rosenzweig (1995). The slope as expressed by the \( z \)-value of the species-area relationship for the studied islet groups is 0.343. This is not close to the “canonical” value of 0.263 set up by Preston (1962) and MacArthur & Wilson (1967), but it is within the range of 0.2–0.5 proposed by Rosenzweig (1995) for island groups indicating the above-mentioned “mainland” species-area relationship. It should be mentioned that for the neighbouring islet group of Echinades situated in the Ionian open sea, the \( z \)-value is 0.236 (Iliadou & al. 2014). The slope \( (z-value) \) shows the increase rate of species richness with area and it can vary according to the examined geographical unit and the taxonomic group analysed (Duarte & al. 2008; MacArthur & Wilson 1967).

In conclusion, the present study shows that the islet groups of the Amvrakikos Gulf and the Mesolongi Lagoon host a significant plant species richness and high overall heterogeneity. The role of geographical position, geological history, palaeogeography and of variables such as island surface area determines the individual islet florae. The inventory of the plant species diversity of the islets in the Amvrakikos Gulf and the Mesolongi Lagoon, the exploration of plant species-richness patterns and of the effects of factors affecting them, fills one more gap in the floristic information available for both the “Amvrakikos Wetlands National Park” and the “National Park of Mesolongi – Aitoliko lagoons, estuaries of Achelous and Evinos River and Echinades islet group”, thus adding to and complementing the case of the Echinades islets group (Iliadou & al. 2014). In order to propose and implement proper measures for the conservation management of species and habitats in the given protected areas, knowledge of the species diversity basically presented here is of crucial importance.

Acknowledgements

This research has been co-financed by the European Union (European Social Fund – ESF) and Greek national funds through the Operational Program “Education and Lifelong Learning” of the National Strategic Reference Framework (NSRF) – Research Funding Program: Heraclitus II. Investing in knowledge society through the European Social Fund. In addition, the authors would like to thank two anonymous reviewers for their valuable comments on an earlier draft of this paper.

References


Arrhenius O. 1921: Species and area. – [J. Ecol. 9]: 95–99


Appendix: Vascular plant catalogue

Symbols and abbreviations:

Life-form categories (following Dimopoulos & al. 2013): 
C = chamaephyte; 
H = hemikryptophyte; 
G = geophyte; 
P = phanerophyte; 
T = therophyte.

Chorological categories (following Dimopoulos & al. 2013): 
Co = Cosmopolitan; 
EA = European-SW Asian; 
EM = East Mediterranean; 
ES = Euro-Siberian; 
MA = Mediterranean-Atlantic; 
Me = Mediterranean; 
ME = Mediterranean-European; 
MS = Mediterranean-SW Asian; 
Pt = Paleotemperate; 
ST = Subtropical-tropical.

AG1 – AG6, ML1 – ML4 = Amvrakikos Gulf and Meso-longi Lagoon islet codes; see Table 1.

[ ] = Alien taxa according to Arianoutsou & al. (2010).

Gymnosperms

Cupressaceae

Juniperus phoenicea L. – P, Me; AG1, AG6, ML1, ML2, ML3, ML4.

Ephedraceae

Ephedra foeminea Forssk. – P, Me; AG5.

Angiosperms

Alliaceae

Allium ampeloprasum L. – G, Me; AG1, AG3, AG5, AG6.

Allium subhirsutum L. – G, Me; AG1, AG2, AG3.

Amaryllidaceae

Narcissus tazetta L. – G, MS; AG1, AG2, AG5.

Pancratium maritimum L. – G, Me; ML3.

Anacardiaceae

Pistacia lentiscus L. – P, Me; AG1, AG2, AG3, AG4, AG5, AG6, ML1, ML2, ML3, ML4.

Apiaceae


Asteraceae

Achillea maritima (L.) Ehrend. & Y. P. Guo – C, MA; ML1, ML2.


Andryala integrifolia L. – T, Me; ML1.

Anthemis arvensis subsp. incrassata (Loisel.) Nyman – T, Me; AG1, ML2, ML3.

Anthemis chia L. – T, Me; AG1, ML2.

Anthemis tomentosa L. – T, EM; AG1, AG5, ML1, ML2.

Bellis perennis L. – H, Me; AG1, AG2.

Carlina acaulis L. – H, Me; AG1, AG2, AG3, AG4, AG5, ML1, ML2, ML3, ML4.

Carlina glabra (L.) Less. – T, Me; ML2.

Carlina lanata L. – T, Me; AG1, ML1.

Carthamus lanatus L. – T, Me; AG1, AG5.

Crepis multiflora Sm. – T, EM; ML1.


Filago gallica L. – T, MA; ML2.

Glebionis segetum (L.) F. W. Schmidt – T, Me; AG1, AG2, AG4, AG5, ML1.

Hedypnois rhagadioides (L.) F. W. Schmidt – T, Me; AG1, AG2, AG3, AG4, AG5, ML1, ML2, ML3, ML4.

Hypochaeris achyrophora L. – T, Me; AG1, AG2, AG3, AG4, AG5, ML1, ML2, ML3, ML4.

Limonidaria crithmoides (L.) Dumort. – C, MA; AG1, AG2, AG3, AG4, AG5, ML1, ML2, ML3, ML4.

Pulicaria odora (L.) Rchb. – H, Me; AG1, AG2, AG4, AG5, ML1, ML3.

Silybum marianum (L.) Gaertn. – H, Me; AG1, AG2, AG3, AG4, AG5, ML1, ML2, ML3.

Sonchus oleraceus L. – T, Me; AG1, AG2, AG3, AG4, AG5, ML1, ML2, ML3, ML4.

Symphyotrichum sericeum (L.) F. W. Schmidt – T, Me; AG1, AG2, AG3, AG4, AG5, ML1, ML2, ML3.

Xanthium orientale subsp. italicum (Moretti) Greuter – T, [S-Eur]; ML1, ML2, ML3.

Boraginaceae

Alkanna tinctoria Tausch – H, Me; ML1, ML2.

Araceae

Arisarum vulgare Targ. Tozz. – G, Me; AG1, AG2, AG3, AG5, AG6.

Arum italicum Mill. subsp. italicum – G, Me; AG5, AG6.
**Borago officinalis** L. – T, Me; AG5.

**Cerinthe major** L. – T, MS; AG5.

**Echium italicum** L. – H, MS; ML2.

**Echium plantagineum** L. – T, ME; ML3.

**Brassicaceae**

**Cakile maritima** Scop. – T, ME; AG1, AG5, ML2, ML3, ML4.

**Capsella bursa-pastoris** (L.) Medik. – H, Co; AG3.

**Enarthrocarpus arcuatus** Labill. – T, EM; AG5.

**Hirschfeldia incana** (L.) Lagr.-Foss. – H, EA; ML1.

**Malcolmia maritima** (L.) R. Br. – T, Me; ML3, ML4.

**Matthiola tricuspidata** (L.) R. Br. – T, Me; AG1, AG5, ML1, ML2, ML3, ML4.

**Rapistrum rugosum** (L.) All. – T, EA; AG1.

**Caryophyllaceae**

**Cerastium glutinosum** Fr. – T, EA; ML1, ML2.

**Cerastium semidecandrum** L. – T, EA; ML4.

**Petrorhagia dubia** (Raf.) G. López & Romo – T, Me; AG1, AG5, ML1.

**Sagina maritima** G. Don – T, MA; ML4.

**Spergularia marina** (L.) Besser – T, Pt; AG1, AG5, ML1.

**Caponiaceae**

**Lonicer a imp lex Aiton – P, Me; AG1, AG2, AG3, AG4, AG5, AG6, ML1.**

**Caryophyllaceae**

**Cerastium glatinosum** Fr. – T, EA; ML1, ML2.

**Cerastium semidecandrum** L. – T, EA; ML4.

**Petrophragia dubia** (Raf.) G. López & Romo – T, Me; AG1, AG5, ML1.

**Silene nocturna** L. – T, Me; AG1, AG5, ML1.

**Silene vulgaris** subsp. **macrocarpa** Turrill – H, Me; AG5.

**Spergularia marina** (L.) Besser – T, Pt; AG1, AG5, ML1.

**Chenopodiaceae**

**Arthrocnemum macrostachyum** (Moric.) K. Koch – C, Me; AG1, AG5, ML2, ML3, ML4.

**Atriplex prostrata** DC. – T, ES; AG1, AG5, ML1, ML3.

**Beta vulgaris** subsp. **maritima** (L.) Arcang. – H, EA; ML1.

**Chenopodium murale** (L.) S. Fuentes & al. – T, EA; AG1.

**Halimione portulacoides** (L.) Aellen – C, ME; AG1, AG5, AG6, ML1, ML3, ML4.

**Salicornia perennis** Willd. subsp. **perennis** – T, EA; ML1, ML3, ML4.

**Salsola soda** L. – T, Pt; AG1, AG2, AG5, ML2, ML4.

**Salsola tragus** L. – T, Pt; ML1, ML2, ML3, ML4.

**Sarcocornia fruticosa** (L.) A. J. Scott – C, MA; AG1, AG5, ML1, ML2, ML3, ML4.

**Suaeda maritima** (L.) Dumort. – T, EA; ML1, ML3, ML4.

**Cistaceae**

**Cistus salvifolius** L. – P, Me; ML1, ML2, ML4.

**Fumana thymifolia** (L.) Webb – C, Me; AG1.
Trifolium campestre Schreb. – T, EA; AG1, AG5, ML1, ML2.
Trifolium lappaceum L. – T, MS; AG1, ML1, ML2.
Trifolium nigrescens Viv. – T; Me; AG1.
Trifolium scabrum L. – T, EA; ML1, ML2, ML4.
Trifolium stellatum L. – T; Me; ML1.
Trifolium tomentosum L. – T, Me; ML1.
Trigonella corniculata subsp. balansae (Boiss. & Reut.) Lassen – T, EM; AG1, AG5.
Vicia angustifolia L. – T, Pt; AG2, ML3.
Vicia villosa subsp. microphylla (d’Urv.) P. W. Ball – T, EM; AG5.

Fumariaceae
Fumaria macrocarpa Parl. – T, Me; AG1, AG2, AG3, AG5.

Gentianaceae
Blackstonia perfoliata (L.) Huds. subsp. perfoliata – T, ME; AG1, AG5, ML1, ML2, ML3, ML4.
Centaurium erythraea Raf. – H, EA; AG1, AG5, ML2.
Centaurium tenuiflorum (Hoffmanns. & Link) Fritsch – T, ME; AG1.

Geraniaceae
Geranium purpureum Vill. – T, Me; ML1, ML3.
Geranium rotundifolium L. – T, Pt; AG5.

Hyacinthaceae
Drimia numidica (Jord. & Fourr.) J. C. Manning & Goldblatt – G, Me; AG2, ML1.
Muscari comosum (L.) Mill. – G, ME; AG1, AG2, AG5.

Hypericaceae
Hypericum tetrapterum Fr. – H, EA; AG1.

Iridaceae
Gladiolus italicus Mill. – G, MS; AG1, AG2, ML1, ML2.

Juncaceae
Juncus acutus L. – H, ES; AG1, AG5, ML1, ML2, ML3, ML4.
Juncus hybridus Brotd. – T, MA; ML1, ML2, ML4.
Juncus inflexus L. – H, Pt; AG1, ML1, ML2, ML3, ML4.
Juncus maritimus Lam. – G, ME; ML1, ML2, ML3, ML4.

Lamiaceae
Micromeria graeca (L.) Rchb. – C, Me; AG1.
Prasium majus L. – C, Me; AG1, AG2, AG3, AG5, ML1, ML2.
Thymbra capitata (L.) Cav. – C, Me; AG1, ML1, ML2.

Linaceae
Linum bienne Mill. – H, Me; ML2.
Linum corymbulosum Rchb. – T, EA; AG1, ML2.
Linum strictum L. – T, Me; ML1, ML2.

Malvaceae
Malva parviflora L. – T, MS; ML1, ML3.
Malva punctata (All.) Alef. – T, Me; AG5.

Moraceae
Ficus carica L. – P, MS; AG5, ML3.

Myrtaceae
Myrtus communis L. – P, Me; AG1, AG2, AG3, AG4, AG5, ML1, ML4.

Oleaceae
Olea europaea L. subsp. europaea – P, Me; AG1, AG2, AG4, AG5, AG6, ML1, ML4.

Orchidaceae
Ophrys scolopax subsp. cornuta (Steven) E. G. Camus – G, ME; AG1.
Serapias vomeracea (Burm. f.) Briq. - G, ME; AG1.

Orobanchaceae
Orobanche minor Sm. – T, EA; ML1, ML2.

Papaveraceae
Glaucium flavum Crantz – H, ME; ML1, ML4.

Plantaginaceae
Plantago africana L. – T, Me; AG1.
Plantago coronopus L. – T, MA; ML1, ML2.
Plantago crassifolia Forssk. – H, Me; ML1, ML2.
Plantago lagopus L. – T, Me; AG1, ML1.
Plantago lanceolata L. – H, Co; AG1, AG2, ML2.

Plumbaginaceae
Linum narbonense Mill. – H, MS; AG1, AG5, ML1, ML2, ML3, ML4.

Poaceae
Aegilops biuncialis Vis. – T, MS; ML1.
Aeluropus littoralis (Gouan) Parl. – G, MS; ML1.
Avena barbata Link – T, Me; AG1, AG2, AG3, AG4, AG5, ML1.
Brachypodium distachyon (L.) P. Beauv. – T, MS; AG1, ML1, ML2.
Brachypodium retusum (Pers.) P. Beauv. – T, Me; AG1, AG2, AG3, AG4, AG5, ML1, ML2.

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