

SERUM CHEMISTRY VALUES OF THE ENDANGERED SAN JOAQUIN KIT FOX (*VULPES MACROTIS MUTICA*)

Authors: McCue, Patrick M., and O'Farrell, Thomas P.

Source: Journal of Wildlife Diseases, 28(3) : 414-418

Published By: Wildlife Disease Association

URL: <https://doi.org/10.7589/0090-3558-28.3.414>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

SERUM CHEMISTRY VALUES OF THE ENDANGERED SAN JOAQUIN KIT FOX (*VULPES MACROTIS MUTICA*)

Patrick M. McCue^{1,2} and Thomas P. O'Farrell¹

¹ EG&G Energy Measurements, Inc., Santa Barbara Operations, 130 Robin Hill Road, Goleta, California 93117, USA

² Present address: Department of Reproduction, School of Veterinary Medicine, University of California, Davis, California 95616, USA

ABSTRACT: Serum chemistry values were obtained from 64 adult San Joaquin kit foxes (*Vulpes macrotis mutica*) in western Kern County, California (USA). The goal of the study was to establish normal chemistry values for this endangered species. No significant differences were found for mean values of alanine aminotransferase (217.1 IU/l), alkaline phosphatase (44.2 IU/l), cholesterol (145.6 mg/dl), total protein (5.8 g/dl), creatinine (0.63 mg/dl), calcium (8.2 mg/dl), albumin (3.0 g/dl), glucose (129.2 mg/dl), amylase (196.8 IU/l), sodium (153.7 mEq/l) and phosphorus (5.42 mg/dl) between sexes or seasons. Significant differences were noted for aspartate aminotransferase, blood urea nitrogen and potassium between seasons. Possible disturbances in normal hepatic and renal functions were noted.

Key words: *Vulpes macrotis mutica*, San Joaquin kit fox, endangered species, serum chemistry.

INTRODUCTION

Measurement of hematologic and serum chemistry values are commonly used as diagnostic techniques for the assessment of health status of individual animals. These parameters may also be useful in assessing health status of populations and may indirectly serve as indicators of habitat condition and changing habitat quality (Franzmann, 1972; Seal et al., 1975; Seal, 1977). In order to correctly evaluate physiological parameters, normal values must be established for that species. This paper presents normal ranges of serum chemistry values for the endangered San Joaquin kit fox (*Vulpes macrotis mutica*), a small canid inhabiting the arid central valley of California. Hematologic values have been previously reported (McCue and O'Farrell, 1987). The distribution, life history and status of this and other subspecies has also been published (O'Farrell, 1987).

MATERIALS AND METHODS

Kit foxes were trapped on the Elk Hills Naval Petroleum Reserve which is located approximately 48 km southwest of Bakersfield, Kern County, California (35°08'N, 119°28'W). An ecological description of Elk Hills was presented by McCue and O'Farrell (1987). Animals were captured in live-traps (National Live Trap Corporation, Tomahawk, Wisconsin 54487, USA) and handled using methods designed to mini-

mize capture stress and excitement. Blood samples (6 to 9 ml) were taken from the jugular vein in 10 ml syringes and transferred immediately into sterile Vacutainers (Becton Dickinson Vacutainer Systems, Rutherford, New Jersey 07070, USA) containing potassium oxalate and sodium fluoride for blood glucose analysis or no anticoagulant for analysis of other biochemical parameters. Blood samples were refrigerated before being shipped by mail to Veterinary Reference Laboratory, Inc. (P.O. Box 25978, Santa Ana, California 92799, USA) on the same day they were collected.

Aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase, blood urea nitrogen (BUN), cholesterol, total protein, creatinine, phosphorus, calcium, albumin, and glucose were analyzed using a SMA-1260 (Technicon, Inc., Tarrytown, New York 10591, USA), amylase was measured by the iodometric method (Van Loon et al., 1951), and potassium and sodium were measured on a Beckman System E-2 (Beckman Instruments, Inc., Fullerton, California 92634, USA).

The mean and standard deviation (SD) were calculated for each data set. Aberrant values that differed from the mean by greater than three standard deviations were classified as "outliers" and excluded (Lumsden and Mullen, 1978; Lumsden et al., 1979). Statistics for data sets were recalculated subsequently with the "outliers" removed. Ranges of normal chemistry values were calculated based on two standard deviations above and below the mean. This statistical method also was used for calculating normal hematological ranges of kit fox (McCue and O'Farrell, 1987). Means of chemical variables were examined for differences between sexes and seasons (summer, August–September;

TABLE 1. Blood chemistry values of adult San Joaquin kit foxes sampled in western Kern County, California, between 1981–82 that did not differ between sexes or seasons.

Value	Units	Sample size (n)	Mean (\bar{x})	SD	Normal range ($\bar{x} \pm 2$ SD)
ALT	(IU/l)	63	217.1	112.9	0–443
Alkaline phosphatase	(IU/l)	62	44.2	32.6	0–109
Cholesterol	(mg/dl)	64	145.6	43.1	59–232
Total protein	(g/dl)	64	5.8	0.6	4.6–7.0
Creatinine	(mg/dl)	63	0.63	0.29	0.1–1.2
Calcium	(mg/dl)	62	8.2	0.96	6.3–10.1
Albumin	(g/dl)	64	3.0	0.36	2.3–3.7
Glucose	(mg/dl)	61	129.2	28.8	72–187
Amylase	(IU/l)	58	196.8	68.7	59–335
Sodium	(mEq/l)	58	153.7	2.5	149–159
Phosphorus	(mg/dl)	62	5.42	1.7	2.0–8.9

winter, November–January) using analysis of variance techniques (SAS Institute Inc., 1985). Differences were considered to be significant when $P < 0.05$.

RESULTS

Blood samples were collected from 64 (34 males, 30 females) adult (>6-mo-old) kit foxes. Mean values of ALT, alkaline phosphorus, cholesterol, total protein, creatinine, calcium, albumin, glucose, amylase, sodium and phosphorus were not significantly different between sexes or seasons (Table 1). AST levels were significantly higher in winter than summer, while BUN and potassium levels were significantly higher in summer than winter (Table 2).

DISCUSSION

Alanine aminotransferase (ALT) is an enzyme found within hepatocytes and is

elevated in serum following hepatocyte plasma membrane damage or hepatocellular degeneration (Meyer, 1982). Plasma half-life for ALT is approximately 2.5 hr (Zinkl et al., 1971) and persistence of elevated serum enzyme levels may reflect continued enzyme leakage (Meyer, 1982). Ninety-five percent of the kit foxes had ALT levels greater than the normal range for domestic dogs (Wolford et al., 1986), coyotes (*Canis latrans*) (Rich and Gates, 1979), red foxes or silver foxes (*V. vulpes*) (Benn et al., 1986). Factors that may be associated with the elevated liver enzyme levels in San Joaquin kit foxes include infectious canine hepatitis virus, which is known to be present in the fox population (McCue and O'Farrell, 1988), parasites, toxins and heavy metals.

Aspartate aminotransferase (AST) is an enzyme that is not liver specific because it

TABLE 2. Blood chemistry values of adult San Joaquin kit foxes sampled in western Kern County, California, between 1981–82 that differed between sexes or seasons.

Value	Units	Variables*	Sample size (n)	Mean (\bar{x})	SD	Normal range ($\bar{x} \pm 2$ SD)
AST	(IU/l)	S	30	158.7	79.0	1–317
		W	29	306.4	164.0	0–634
BUN	(mg/dl)	S	31	37.2	12.1	13–61
		W	28	24.7	4.1	17–33
Potassium	(mEq/l)	S	26	5.03	0.3	4.43–5.63
		W	29	4.75	0.3	4.15–5.35

* S, summer; W, winter.

is present in both the liver and skeletal muscle. Elevations in AST in kit foxes sampled in winter, with no significant elevation in ALT, suggest extrahepatic tissue damage releasing AST. The mean serum AST level was also higher than means reported for domestic dogs (Wolford et al., 1986), coyotes (Rich and Gates, 1979), red foxes or silver foxes (Benn et al., 1986).

A majority of blood urea is synthesized in the liver from ammonia, which is formed from protein catabolism or absorbed from the gastrointestinal tract (Duncan and Prasse, 1983). The kidney is the most important route of urea excretion and BUN concentration is often used as an indicator of renal function. BUN also may increase due to decreased kidney perfusion, high protein diet, increased protein catabolism, and post-urinary obstruction or leakage. Elevated BUN levels in kit foxes, especially in summer, may have been due to a combination of factors including high protein diet, severe environmental stress, nephrotoxins and infectious disease, such as leptospirosis. However, no antibodies against five serotypes of *Leptospira interrogans* were found during a serologic survey of the Elk Hills kit foxes (McCue and O'Farrell, 1988). Captive swift foxes provided with adequate water (Mainka, 1988) had a mean BUN similar to kit foxes sampled in the winter when kit foxes were presumably experiencing less water stress. It was inferred that the increase in BUN in the summer was related to water balance, although differences in protein content of diet may also be involved. A decrease in urine volume accompanying efforts to conserve body water may have caused the elevation in BUN. However, creatinine levels which are also used to monitor renal function, were not significantly different between summer and winter.

Serum phosphorus concentration in adult kit foxes was elevated compared with values reported for adult dogs (Wolford et al., 1986) and coyotes (Smith and Rongstad, 1980). Seal et al. (1975) postulated that

high serum levels of phosphorus in wolf (*Canis lupus*) pups may have been due to high phosphorus intake from a meat diet which had a 1:20 calcium to phosphorus ratio. This probably does not explain the high phosphorus levels in kit foxes because they consume the entire carcass of their prey and probably have a more balanced calcium and phosphorus intake. Hyperphosphatemia has been associated with prerenal azotemia (Ross, 1986) and may, therefore, be related to aspects of water balance in kit foxes. Serum phosphorus elevations may also be due to methods of sample handling, because phosphorus is released from red blood cells after 12 to 24 hr, and from hemolysis (Duncan and Prasse, 1983).

Glucose elevations in captured wild animals are primarily attributed to stress (Karns and Crichton, 1978; Seal et al., 1975; Smith and Rongstad, 1980). Glucocorticoid release antagonizes the effects of insulin, leading to an elevation in blood glucose by decreasing glucose uptake and use by tissues. Methods were developed to minimize stress on captured foxes, but some individuals become hyperexcited during initial stages of handling. No attempt was made to quantify levels of stress observed; instead, every effort was made to minimize or prevent it.

Serum levels of alkaline phosphatase, cholesterol, total protein, creatine, calcium, and albumin in adult kit foxes did not differ significantly between sex or season, nor were they different from ranges reported for adult domestic dogs (Wolford et al., 1986), coyotes (Smith and Rongstad, 1980), red foxes or silver foxes (Benn et al., 1986). Levels of sodium in kit fox were at the upper range of normal or slightly higher than normal values for the domestic dog (Duncan and Prasse, 1983). Amylase levels in kit foxes were lower than those reported for dogs (Duncan and Prasse, 1983).

Interpretation of serum chemistry values must be made in relation to the physiology and ecology of the species sampled.

The kit fox is a carnivore inhabiting an extremely arid environment, where little or no free water is available. Foxes occasionally were observed drinking what free water was available, but aspects of water balance in this species are not well understood. The causes for the elevation of liver enzymes and BUN, and the electrolyte imbalances observed during this study are presently unknown. It is possible that some values in kit foxes may represent adaptations to extremes in environmental conditions and the unpredictability of food and water resources.

ACKNOWLEDGMENTS

This research was performed for the U.S. Department of Energy, Naval Petroleum Reserves in California, and Chevron USA, through the Nevada Operations Office under Contract No. DE-AC08-88NV10617 with EG&G Energy Measurements, Inc. Permission to handle this endangered species and obtain blood samples was granted by the U.S. Fish and Wildlife Service through permits PRT 2-4573 and PRT 683011, and a Memorandum of Understanding between the California Department of Fish and Game and EG&G Energy Measurements, Inc. We thank our EG&G/EM colleagues Thom Kato, Bill Berry, Brenda Evans, Dale Garner, Brad Hardenbrook, Chuck Harris, Jeff Johnson, John McManus, Gene Orth, Mike Spencer and Kris Timmerman for helping us trap foxes and obtain blood samples. JoAnne Ando and Bob Elliot helped with data storage, retrieval and compilation; Elizabeth Collins provided statistical assistance. We are most grateful for the assistance of Terry Siple, Sherre Hughes, Bryan Umstead, and Carol Rodriguez Kato of the Bakersfield Veterinary Hospital.

LITERATURE CITED

- BENN, D. M., D. B. McKEOWN, AND J. H. LUMSDEN. 1986. Hematology and biochemistry reference values for the ranch fox. *Canadian Journal of Veterinary Research* 50: 54-58.
- DUNCAN, J. R., AND K. W. PRASSE. 1983. *Veterinary laboratory medicine, clinical pathology*. The Iowa State University Press, Ames, Iowa, 243 pp.
- FRANZMANN, A. W. 1972. Environmental sources of variation in physiologic values of bighorn sheep. *Journal of Wildlife Management* 36: 924-932.
- KARNS, P. D., AND V. F. J. CRICHTON. 1978. Effects of handling and physical restraint on blood parameter of woodland caribou. *The Journal of Wildlife Management* 42: 904-908.
- LUMSDEN, J. H., AND K. MULLEN. 1978. On establishing reference values. *Canadian Journal of Comparative Medicine* 43: 293-301.
- , K. MULLEN, AND B. J. MCSHERRY. 1979. Canine hematology and biochemistry reference values. *Canadian Journal of Comparative Medicine* 43: 125-131.
- MAINKA, S. A. 1988. Hematology and serum biochemistry of captive swift foxes (*Vulpes velox*). *Journal of Wildlife Diseases* 24: 71-74.
- MCCUE, P. M., AND T. P. O'FARRELL. 1987. Hematologic values of the endangered San Joaquin kit fox, *Vulpes macrotis mutica*. *Journal of Wildlife Diseases* 23: 144-151.
- , AND ———. 1988. Serological survey for selected diseases in the endangered San Joaquin kit fox (*Vulpes macrotis mutica*). *Journal of Wildlife Diseases* 24: 274-281.
- MEYER, D. J. 1982. The liver. Part I. Biochemical tests for the evaluation of the hepatobiliary system. *Compendium on Continuing Education for the Practicing Veterinarian* 4: 663-674.
- O'FARRELL, T. P. 1987. Kit fox. In *Wild furbearer management and conservation in North America*, M. Novak, J. A. Baker, M. E. Obbard and B. Malloch (eds.). Ontario Trappers Association, Toronto, Ontario, Canada, pp. 423-431.
- RICH, J. E., AND N. L. GATES. 1979. Hematologic and serum chemistry values of pen-raised coyotes. *Journal of Wildlife Diseases* 15: 115-119.
- ROSS, L. A. 1986. Assessment of renal function in the dog and cat. In *Current veterinary therapy*, IX, R. D. Kirk (ed.). W. B. Saunders Company, Philadelphia, Pennsylvania, pp. 1103-1108.
- SAS INSTITUTE INC. 1985. *SAS user's guide: Basics version*, 5th ed. SAS Institute, Inc., Cary, North Carolina, 956 pp.
- SEAL, U. S. 1977. Assessment of habitat condition by measurement of biochemical and endocrine indicates of the nutritional, reproductive and disease status in free ranging animal populations. In *Classification, inventory and analysis of fish and wildlife habitat*. Proceedings of a National Symposium held in Phoenix, Arizona, U.S. Fish and Wildlife Service, Office of Biological Services Publication 78/76, Washington, D.C., pp. 305-329.
- , L. D. MECH, AND V. VAN BALLEMBERGHE. 1975. Blood analyses of wolf pups and their ecological and metabolic interpretation. *Journal of Mammology* 56: 64-75.
- SMITH, G. J., AND O. J. RONGSTAD. 1980. Serologic and hematologic values of wild coyotes in Wisconsin. *Journal of Wildlife Diseases* 16: 491-497.
- VAN LOON, E. J., M. R. LIKENS, AND A. J. SEGER. 1951. Photometric method for blood amylase by use of a starch-iodine color. *American Journal of Clinical Pathology* 22: 1134-1136.
- WOLFORD, S. T., R. A. SCHROER, F. X. GOHS, P. P. GALLO, M. BRODECK, H. B. FALK, AND R.

- RUHREN. 1986. Reference range data base for serum chemistry and hematology values in domestic animals. *Journal of Toxicology and Environmental Health* 18: 161-188.
- ZINKL, J. G., R. M. BUSH, C. E. CORNELIUS, AND R. A. FREEDLAND. 1971. Comparative studies on plasma and tissue sorbital, glutamic, lactic and hydroxybutyric dehydrogenase and transaminase activities in the dog. *Research in Veterinary Science* 12: 211-214.

Received for publication 10 April 1990.