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Article

Effects of host plant species on the development and reproduction of *Neoseiulus bicaudus* (Phytoseiidae) feeding on *Tetranychus turkestanii* (Tetranychidae)

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Abstract

The leaf structure of the host plant can affect the development and reproduction of the predatory enemies of spider mites. The objective of this study was to compare the effects of five host plant species [cucumber (*Cucumis sativus* L.), cotton (*Gossypium hirsutum* L.), eggplant (*Solanum melongena* L.), tomato (*Lycopersicon esculentum* Mill.) and green bean (*Phaseolus vulgaris* L.)] on the development and reproduction of *Neoseiulus bicaudus* feeding on *Tetranychus turkestanii*. There were significant differences in the characteristics of trichomes on the surfaces of the host plant species. Tomato leaves had the greatest trichome density (782 trichomes/cm²) whereas green bean leaves had the least density (58 trichomes/cm²). Cucumber had the longest trichomes (1.58 mm) and green bean had the shortest (0.14 mm). *N. bicaudus* developed into adulthood and completed its development on all five plant species. The total immature period of *N. bicaudus* feeding on *T. turkestanii* was shortest (5.07 d) on tomato. *N. bicaudus* on cucumber had the longest previposition period (2.78 d) and the shortest oviposition period (11.96 days). Total fecundity was highest on green bean (42.4 eggs/day/female) and lowest on cucumber (20.81 eggs/day/female). The female/male sex ratio was highest on cucumber (0.76) and lowest on tomato (0.65). The net reproductive rate (R_0) was 14.55 on cucumber, 23.54 on cotton, 21.79 on eggplant, 24.05 on tomato, and 34.61 on green bean. In conclusion, among the five host plant species in this study, green bean was best and cucumber was worst for the development and reproduction of *N. bicaudus* preying on *T. turkestanii*.

Key words: host plant, predatory mite, spider mite, predation, life parameters

Introduction

The host plant species can affect the population establishment and performance of predator mites (Walter, 1996; Romero & Benson, 2004; Tanga & Mohamed, 2013). The microenvironment of plant surfaces seems uniform to the human eye, but quite complex to arthropods (Buitenhuis *et al.*, 2015; Roda & English, 2003). The characteristics of host plant can modify interaction between herbivore insects and their enemies (Grostal & O'Dowd, 1994; Norton *et al.*, 2001; Momen & Hussein, 2011). The host plant species affects the oviposition, growth and development of predatory mites in the following ways: (i) by influencing the activity of predatory mites, (ii) by providing shelter for the mites from natural enemies, (iii) by altering the microenvironment, and (iv) by trapping and retaining food sources such as pollen and fungal spores (Romero *et al.*, 2011; Rebecca, 2014; Skirvin & Fenlon, 2001; Buitenhuis *et al.*, 2014).

Neoseiulus bicaudus (Wainstein) is a species of predatory mites, while belong to phytoseiidae. It was recently observed in the Xinjiang Uygur Autonomous Region of northwest China (Wang *et*

al., 2015a). It was previously recorded from Iran, Russia, Greece, Cyprus, and the United States (De Leon, 1962; Wainstein, 1962; Schuster & Pritchard, 1963; Faraji *et al.*, 2007; Palevsky *et al.*, 2009). Wang *et al.*, (2015a) redescribed this species and discussed the morphological differences between the Chinese population of this species with *Cydnodromus comitatus* (De Leon, 1962), *Amblyseius scyphus* (Schuster & Pritchard, 1963), *Amblyseius micmac* (Chant and Hansell, 1971) and *Amblyseius hirotae* (Ehara, 1985), which were considered as the synonyms of *N. bicaudus*. Wang *et al.* (2015b) described the different stage morphology and predatory behavior of *N. bicaudus*. Li *et al.* (2015) studied the effects of temperature on the development and reproduction of *N. bicaudus*.

The spider mite *Tetranychus turkestanii* is mainly distributed in Russia, Kazakhstan, America, the Middle East, and Xinjiang, China (Li *et al.*, 2014; Lu & Zhang 2000). It is the most important pest mite in Xinjiang, causing damage to many crops including cotton (*Gossypium hirsutum* L.), eggplant (*Solanum melongena* L.), corn (*Zea mays* L.), sorghum (*Sorghum bicolor* L.), medlar (*Achras Sapota* L.), tomato (*Lycopersicon esculentum* Mill.) and beans (*Phaseolus vulgaris* L.). *T. turkestanii* directly damages plant leaves. Under severe infestation, plant leaves become yellow and then dry or drop off the plant (Sohrabi & Shishehbor, 2008; Guo *et al.*, 2013).

The objective of this study was to develop life tables for *N. bicaudus* preying on *T. turkestanii* on five host plant species. Information from this study can be used (i) to understand the effects of different host plant species on *N. bicaudus* populations and to determine which host plant species is the best habitat for *N. bicaudus*, (ii) to increase understanding the interactions among host-pest-predator and (iii) to provide a scientific basis for the efficient release of predatory mites to control field pests.

Materials and methods

Mite colony

Tetranychus turkestanii was from a laboratory colony. The colony was initiated in 2010 with individuals collected from a cotton field near Huayuan, Shihezi City, Xinjiang. The colony was maintained on green bean. The laboratory colony of *Neoseiulus bicaudus* was originally collected from Ili, Xinjiang and established in 2013. The colony had been acclimated and propagated on *T. turkestanii* for two years. Both colonies were kept in a growth chamber (FLI-2000H, Eyela, Japan) at 26±1 °C, 60% relative humidity, and a 16:8 h (light : dark) photoperiod.

Plant material

Cucumber (SG), cotton (HG-49), eggplant (ZCQ), tomato (HMN-140) and green bean (SJD) were cultivated in a field near Shihezi University, Xinjiang. When the plants reached their maturity, three plants were selected in each plot. Three leaves were collected from each of the upper, middle, and lower parts of the plants (i.e., nine leaves per plant). This allowed us to compare both trichome density (number of trichomes per cm²) and trichome length among plant species. The number and length of trichomes on the surface of each leaf were determined under a stereo microscope (Zeiss Discovery V20). The values were then averaged.

Experiment set-up

In a preliminary experiment, two generations of *N. bicaudus* and *T. turkestanii* were raised on the five host plant species. Small chambers were constructed by drilling a hole (1 cm diameter) through solid pieces of plexiglass (3 cm long × 2 cm wide × 0.3 cm high) (Xu *et al.*, 2013). A section of a plant leaf was placed on a strip of filter paper and then positioned over the hole at one end of the chamber. One gravid adult female of *N. bicaudus* female and ten adult *T. turkestanii* females were

transferred into the chambers. Both ends of the chamber were then covered with thin pieces of glass to keep the mites from escaping. The chambers were then placed in growth chambers at 26 °C, 60% R.H. and 16:8 h (L:D). This humidity is typical of the arid/semi-arid climate of Xinjiang Province. There were 120 replications for each plant species.

After the *N. bicaudus* eggs were laid, the adult female and all the eggs but one was removed from each chamber. The development of the immature *N. bicaudus* was observed at 12 h intervals. The duration of each developmental stage was noted until all of the *N. bicaudus* reached adulthood. The leaf was not changed until the nymph stage. Afterwards, the leaf was changed daily. After reaching adulthood, the mites were sexed and males and females from the same host plant species were paired inside the chambers. Ten adult *T. turkestanii* females were provided as food (Wang *et al.*, 2015b). The number of eggs laid by each female was recorded at 24 h intervals until it died. The eggs were counted and then transferred to clean chambers. The hatchlings were reared to adulthood and their sex-ratio was determined (Xia *et al.* 2012; Gotoh *et al.* 2004; McMurtry & Scriven 1964).

Life history parameters and statistical analysis

Life tables were constructed from the survival and fecundity rates of the *N. bicaudus*. The parameters for each host plant species were calculated using the methods of Birch (1948). Population parameters were calculated according to the following equations. The net reproductive rate, R_0 , is given by $R_0 = \sum l_x m_x$; the intrinsic rate of increase, r_m is given by $r_m = (L_n R_0)/T$; the finite rate of increase, λ , is given by $\lambda = e^{r_m}$; the mean generation time, T , is given by $T = \sum x l_x m_x / \sum l_x m_x$; and the doubling time, Dt , is given by $Dt = (L_n 2)/r_m$.

Analysis of variance was conducted using SPSS version 17.0 to determine the effect of host plant species on *N. bicaudus* at different life stages. The means were compared using Duncan's multiple range tests. Differences were considered significant at $P < 0.05$.

Results

Surface traits of the five host plant species

The type and number of trichomes differed among the five host plant species (Table 1). The trichomes of cotton and eggplant were in clusters, whereas those of cucumber, tomato and green bean were single. Cucumber and eggplant trichomes were acicular and hard, whereas trichomes in the other host plants were villiform and soft (Fig. 1). Tomato and eggplant had significantly more trichomes than others ($P < 0.05$). There was no significant difference in the number of trichomes among cotton, cucumber and green bean. Cucumber had the longest trichomes, averaging 1.58 mm in length. Cotton and eggplant had the second longest trichomes (1.30 and 1.10 mm long, respectively) and the bean had the shortest trichomes (0.14mm).

TABLE 1. Description of the trichomes of five host plants.

	Description	Number per cm ²	Length (mm)
Cucumber	Single, erect, acicular, hard	113±25.4 c	1.58±0.13 a
Cotton	Clustered, horizontal, villiform, soft	42±3.5 c	1.30±0.10 b
Eggplant	Clustered, erect, acicular, hard	250±7.1 b	1.10±0.06 b
Tomato	Single, erect, villiform, soft	782±86.1 a	0.62±0.03 c
Green bean	Single, erect, villiform, soft	58±7.3 c	0.14±0.02d

Note: Values are means ± SE. Values within a column followed by different letters are significantly different at $P < 0.05$.

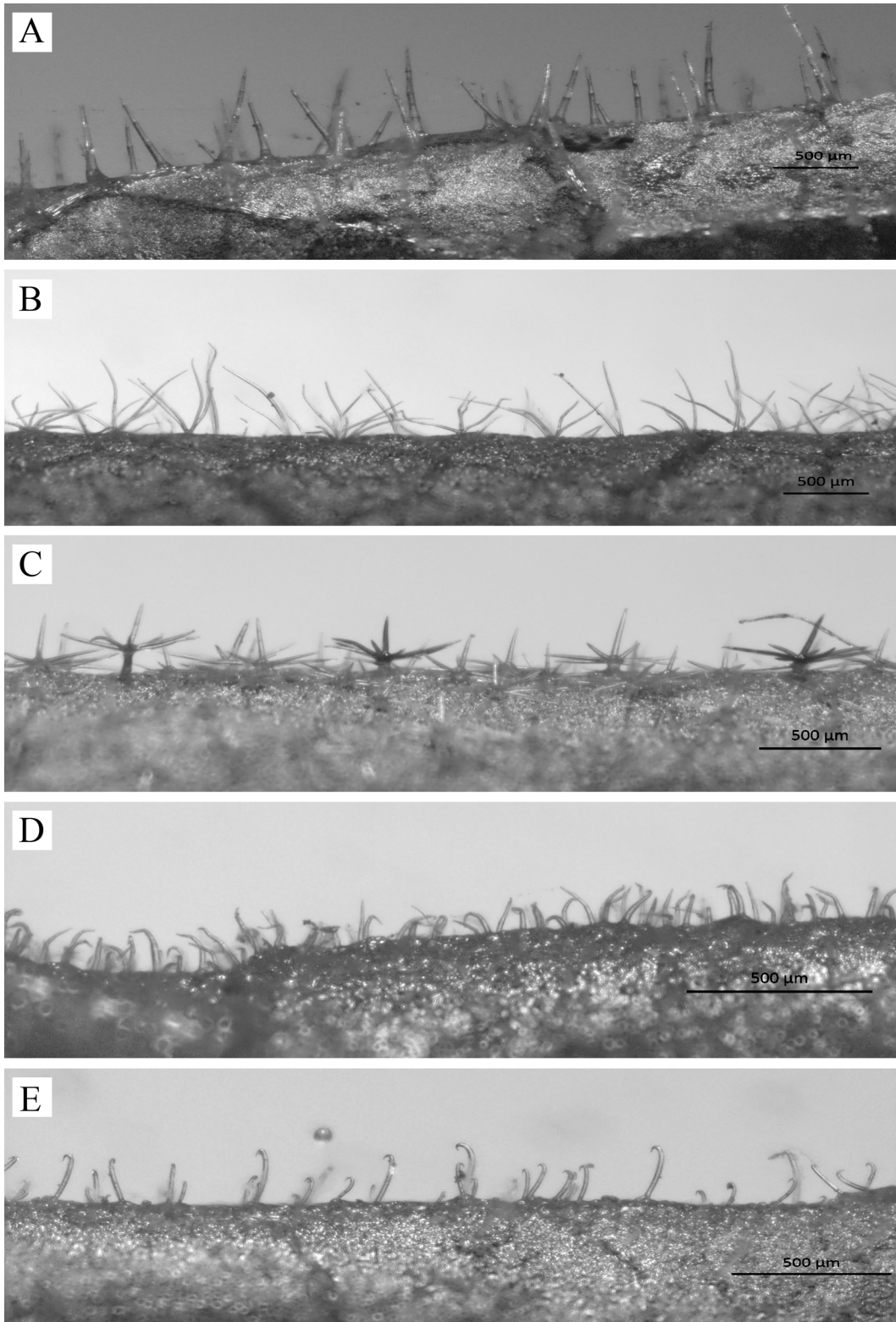


FIGURE 1. Trichomes on leaves of (a) cucumber, (b) cotton, (c) eggplant, (d) tomato, and (e) green bean.

Effects of host plant species on immature stages of N. bicaudus feeding on T. turkestanii

The egg stage of *N. bicaudus* feeding on *T. turkestanii* was significantly longer on cucumber (2.02 d) than on the other host plant species (Table 2). Differences among the other host plant species were not significant. The larval stage was significantly longer on green bean than on cucumber ($P<0.05$). The protonymph stage was significantly longest on green bean followed by cucumber. The deutonymph stage was longest on cucumber and green bean. The deutonymph stage was shortest on tomato. For *N. bicaudus* females feeding on *T. turkestanii*, the immature stage was significantly longer on cucumber and green bean than on eggplant, cotton and tomato. The differences among the latter three plant species were not significant. Overall, the total immature period of *N. bicaudus* feeding on *T. turkestanii* was shortest on tomato.

TABLE 2. Effect of host plant species on the development time (days) of immature *N. bicaudus* feeding on *T. turkestanii*

Plant species	Egg (d)	Larval (d)	Protonymph(d)	Deutonymph(d)	Total immature(d)
Cucumber	2.02±0.09 a	0.54±0.02 b	1.51±0.06 b	1.74±0.07 a	5.82±0.13 a
Cotton	1.59±0.05 b	0.56±0.03 ab	1.46±0.05 bc	1.53±0.05 bc	5.13±0.07 c
Eggplant	1.63±0.03 b	0.59±0.02 ab	1.37±0.04 c	1.60±0.04 ab	5.19±0.06 c
Tomato	1.69±0.03 b	0.59±0.02 ab	1.35±0.04 c	1.44±0.06c	5.07±0.08 c
Green bean	1.65±0.03 b	0.62±0.02 a	1.66±0.03 a	1.71±0.03 a	5.65±0.06 b

Note: Values are means ± SE. Values within a column followed by different letters are significantly different at $P<0.05$.

Effects of host plants on adult development of N. bicaudus feeding on T. turkestanii

The pre-oviposition period of *N. bicaudus* feeding on *T. turkestanii* was significantly longer on cucumber (2.78 d) than on the other host plant species. Conversely, the oviposition period of *N. bicaudus* was significantly shorter on cucumber (11.96 d) than on the other host plant species. The post-oviposition period and total life span were both longest on green bean (11.93 and 31.05 d, respectively) and shortest on cucumber (4.37 and 19.11 d, respectively). There was no significant difference in either the post-oviposition period or the life span of *N. bicaudus* among cotton, eggplant and tomato. The female/male sex ratio was highest on cucumber (0.76) and lowest on tomato (0.65) (Table 3).

TABLE 3. Effect of host plant species on the development time (days) of *N. bicaudus* adults feeding on *T. turkestanii*

Plant species	Pre-oviposition (d)	Oviposition (d)	Post- oviposition (d)	Life span (d)	Female : male
Cucumber	2.78±0.11 a	11.96±0.77 b	4.37±0.66 c	19.11±1.12 c	0.76
Cotton	2.30±0.09 b	15.48±0.65 a	9.44±0.83 ab	27.22±1.08 b	0.73
Eggplant	2.31±0.083 b	16.03±0.74 a	8.63±1.06 b	26.97±1.45 b	0.70
Tomato	1.70±0.13 c	16.27±0.64 a	7.03±1.18 b	25.00±1.41 b	0.65
Green bean	2.52±0.44 b	16.60±0.77 a	11.93±1.47 a	31.05±1.66 a	0.66

Note: Values are means ± SE. Values within a column followed by different letters are significantly different at $P<0.05$.

Effects of different host species on the fecundity of N. bicaudus feeding on T. turkestanii

Daily fecundity and daily female fecundity were both highest on green bean (2.55 eggs/day/female and 1.69 female eggs/day/female, respectively) and lowest on cucumber (1.71 eggs/day/female and 1.29 female eggs/day/female, respectively). The daily fecundity of *N. bicaudus* was significantly greater on green bean than on the other host plants. Total fecundity was greatest on

green bean (42.2 eggs /female) and least on cucumber (20.81 eggs/female). There was no significant difference in total fecundity on cotton and on eggplant (Table 4).

TABLE 4. Effect of host plant species on the fecundity of *N. bicaudus* feeding on *T. turkestanii*

Plant species	Daily fecundity (Eggs/day/female)	Daily female fecundity (Female eggs/day/female)	Total fecundity (Eggs/female)
Cucumber	1.71±0.08 d	1.29±0.069 d	20.81±1.86 d
Cotton	2.10±0.07 bc	1.54±0.05 b	32.37±1.78 c
Eggplant	1.98±0.07 c	1.39±0.05 cd	32.03±2.06 c
Tomato	2.25±0.08 b	1.47±0.05 bc	37.66±1.77 b
Green bean	2.55±0.10 a	1.69±0.06 a	42.40±2.36 a

Note: Values are means ± SE. Values within a column followed by different letters are significantly different at $P < 0.05$.

Effects of host plant species on life table parameters of N. bicaudus feeding on T. turkestanii

The net reproductive rate (R_0) of *N. bicaudus* feeding on *T. turkestanii* ranged from 14.55 on cucumber to 34.61 on green bean (Table 5). The longest mean generation time was 14.47 d on green bean followed by 9.45 d on eggplant, 9.29 d on cotton, 8.84 d on tomato, and 8.20 d on cucumber. The maximum intrinsic rate of natural increase (r_m) was 0.36 on tomato; the minimum was 0.25 on green bean. However, the maximum doubling time was on green bean (2.83) whereas the minimum was on tomato (1.93). The finite rate of increase (λ) ranged from 1.28 on green bean to 1.44 on tomato (Table 5).

TABLE 5. Life table parameters on *N. bicaudus* feeding on *T. turkestanii* on five host plants

Plant species	Net reproductive rate (R_0)	Mean generation time (T , in days)	Intrinsic rate of natural increase (r_m)	Population doubling time (Dt)	Finite rate of increase (λ)
Cucumber	14.55	8.20	0.33	2.12	1.39
Cotton	23.55	9.29	0.34	2.04	1.41
Eggplant	21.79	9.45	0.33	2.13	1.39
Tomato	24.05	8.84	0.36	1.93	1.44
Green bean	34.61	14.47	0.25	2.83	1.28

Note: Values are means ± SE. Values within a column followed by different letters are significantly different at $P < 0.05$.

Survival rate and daily female fecundity of N. bicaudus feeding on T. turkestanii

The survival curves of *N. bicaudus* on different host species were all type I (Fig. 2). Furthermore, there were differences among the survival rates. The survival rate (l_x) decreased significantly in the later developmental stages, especially on cucumber. On cucumber, *N. bicaudus* females began dying on d 11. All of the females died by d 35. On cotton, the female mites began dying on d 16. All of the females died by d 40. The survival was longest on eggplant (49 d).

Most *N. bicaudus* individuals completed their life span on these five plant species. The curves of age-specific fecundity (m_x) peaked soon after the onset of reproduction and varied considerably among the different host plant species. The age-specific fecundity of *N. bicaudus* peaked on d 3 on cucumber and cotton, d 8 on eggplant, d 6 on tomato, and d 11 on green bean. The life span of *N. bicaudus* was longest on green bean and cotton and shortest on cucumber.

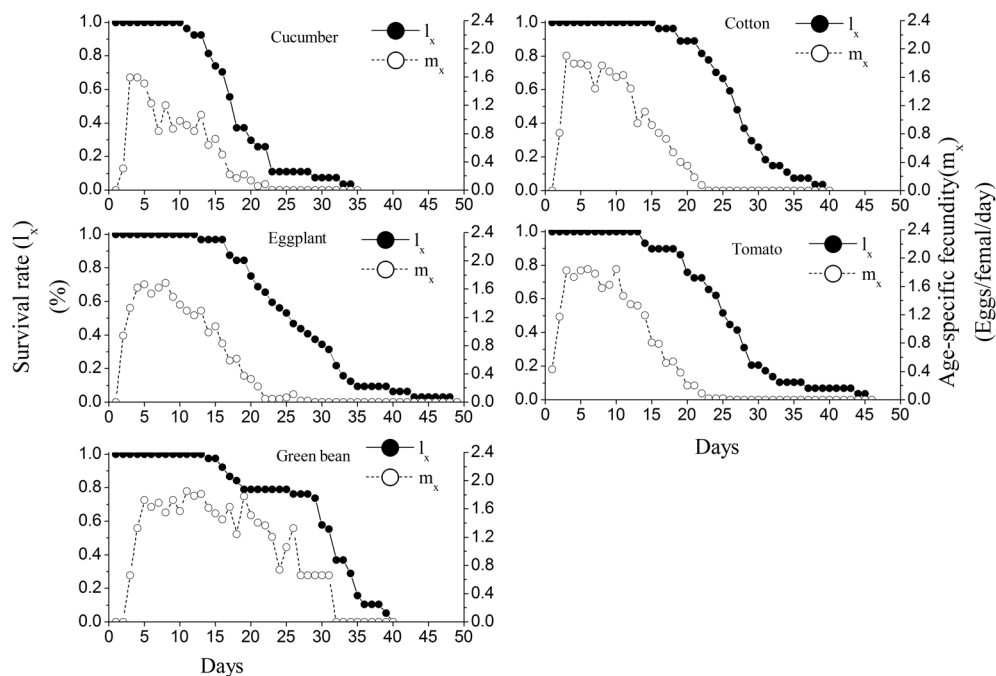


FIGURE 2. Effects of five host plant species on the survival and daily fecundity of *N. bicaudus* females feeding on *T. turkestani*. Abbreviations: l_x , age-specific survival rate; m_x , age-specific fecundity.

Discussion

The host plant is an interactive component of biological control practices and the characteristics of the host plant need to be considered when determining the predator release rate (Krips *et al.* 1999; Loughner *et al.* 2008; Madadi *et al.* 2007; Romero & Vasconcellos 2005). Life tables of both predatory mites and spider mites on different plant species provide important information that is needed for the efficient release of predatory mites to control pests in crop fields (Villanueva & Childers 2006; McMurtry *et al.* 2004; Gotoh & Kitashima 2006).

The length and number of trichomes differed among the five host plant species. Tomato and eggplant had significantly more trichomes than the other host plants. Cucumber had the longest trichomes. Cotton and eggplant had the second longest trichomes and the bean had the shortest trichomes.

In the current study, host plant species significantly affected the total immature time of *N. bicaudus* feeding on *T. turkestani*. Specifically, the generation period of *N. bicaudus* was shortest on tomato (5.0 d) and longest on cucumber (5.8 d). In contrast, Collier (2007) reported that host plant species (papaya versus snap bean) had no effect on the duration of the preimaginal stages of immature *N. idaeus* reared on *T. urticae*. The differences between our results and those of Collier may be due to differences in the characteristics of the host plant species and predatory mites in the two studies (Collier *et al.* 2007).

The fact that adult *N. bicaudus* females showed difference total fecundity in the five host plants. Total fecundity was greatest on green bean (42.2 eggs /female) and least on cucumber (20.81 eggs/female). There was no significant difference in total fecundity on cotton and on eggplant. Overall, the results indicate that the total fecundity decreases as trichome length increases. The number of trichomes have no obvious impacts on the total fecundity. However, Loughner (2008) reported that a lack of trichomes was associated with much lower predator numbers. The fecundity is an important factor that effects on the number of arthropod (Loughner *et al.* 2008).

The maximum net reproductive rate (R_0) was on green bean (34.61). The minimum was on cucumber (14.55). The total duration of the immature stages, oviposition time, total fecundity, and net reproductive rate (R_0) all indicate that *N. bicaudus* did best on green bean and tomato and worst on cucumber. This result shows the trichome length negatively affected the population growth of *N. bicaudus*. These results emphasize the importance of the host plant characteristics on the performance of natural enemies.

This study was conducted in a laboratory with constant environmental conditions, whereas, in addition to the host plants, the other environment conditions also have effect on the development and reproduction of *N. bicaudus*. So additional work must be completed before exact recommendations can be made related to the efficient release of predatory mites to control field pests.

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References

- Birch, L. (1948) The intrinsic rate of natural increase of an insect population. *The Journal of Animal Ecology*, 17, 15–26.
<http://dx.doi.org/10.2307/1605>
- Buitenhuis, R., Murphy, G., Shipp, L. & Scott-Dupree, C. (2015) *Amblyseius swirskii* in greenhouse production systems: a floricultural perspective. *Experimental and Applied Acarology*, 65(4), 451–464.
<http://dx.doi.org/10.1007/s10493-014-9869-9>
- Buitenhuis, R., Shipp, L., Scott-Dupree, C., Brommit, A. & Lee, W. (2014) Host plant effects on the behaviour and performance of *Amblyseius swirskii* (Acari: Phytoseiidae). *Experimental and Applied Acarology*, 62(2), 171–180.
<http://dx.doi.org/10.1007/s10493-013-9735-1>
- Chant, D.A. & Hansell, R.I.C. (1971) The genus *Amblyseius* (Acarina: Phytoseiidae) in Canada and Alaska. *Canadian Journal of Zoology*, 49(5), 703–758.
<http://dx.doi.org/10.1139/z71-110>
- Collier, K.F., Albuquerque, G.S., de Lima, J.O., Pallini, A. & Molina-Rugama, A. J. (2007) *Neoseiulus idaeus* (Acari: Phytoseiidae) as a potential biocontrol agent of the two-spotted spider mite, *Tetranychus urticae* (Acari: Tetranychidae) in papaya: performance on different prey stage-host plant combinations. *Experimental and Applied Acarology*, 41(1-2), 27–36.
<http://dx.doi.org/10.1007/s10493-006-9041-2>
- De Leon, D. (1962) Twenty-three new phytoseiids, mostly from southeastern United States (Acarina: Phytoseiidae). *The Florida Entomologist*, 45(1), 11–27.
<http://dx.doi.org/10.2307/3492899>
- Ehara, S. (1985) Five species of phytoseiid mites from Japan with descriptions of two new species (Acarina,

- Phytoseiidae). *Zoological Science*, 2, 115–121.
- Faraji, F., Hajizadeh, J., Ueckermann, E.A., Kamali, K. & McMurtry, J. A. (2007) Two new records for Iranian phytoseiid mites with synonymy and keys to the species of Typhloseiulus Chant and McMurtry and Phytoseiidae in Iran (Acari: Mesostigmata). *International Journal of Acarology*, 33(3), 231–239.
<http://dx.doi.org/10.1080/01647950708684527>
- Gotoh, T., Yamaguchi, K. & Mori, K. (2004) Effect of temperature on life history of the predatory mite *Amblyseius (Neoseiulus) californicus* (Acari: Phytoseiidae). *Experimental and Applied Acarology*, 32(1-2), 15–30.
- Gotoh, T., Tsuchiya, A. & Kitashima, Y. (2006) Influence of prey on developmental performance, reproduction and prey consumption of *Neoseiulus californicus* (Acari: Phytoseiidae). *Experimental and Applied Acarology*, 40(3-4), 189–204.
<http://dx.doi.org/10.1007/s10493-006-9032-3>
- Grostal, R. & O'Dowd, D.J. (1994) Plants, mites and mutualism: leaf domatia and the abundance and reproduction of mites on *Viburnum tinus* (Caprifoliaceae). *Oecologia*, 97(3), 308–315.
- Guo, Y.-L., Jiao, X.-D., Xu, J.-J., Yang, S., Duan, X.-K. & Zhang, J.-P. (2013) Growth and reproduction of *Tetranychus turkestanii* and *Tetranychus truncatus* (Acari: Tetranychidae) on cotton and corn. *Systematic and Applied Acarology*, 18(1), 89–98.
<http://dx.doi.org/10.11158/saa.18.1.10>
- Krips, O.E., Kleijn, P.W., Willems, P.E.L., Gols, G.J.Z. & Dicke, M. (1999) Leaf hairs influence searching efficiency and predation rate of the predatory mite *Phytoseiulus persimilis* (Acari: Phytoseiidae). *Experimental and Applied Acarology*, 55, 119–131.
http://dx.doi.org/10.1007/978-94-017-1343-6_29
- Li, G.-Y., Li, J.-J., Xia, W., Qu, H.-L., Yang, S. & Zhang, J.-P. (2014) Effects of Bt+CpTI transgenic cotton on the performance of *Tetranychus turkestanii* (Acari: Tetranychidae). *Systematic and Applied Acarology*, 19(2), 236–247.
<http://dx.doi.org/10.11158/saa.19.2.14>
- Li, Y.-T., JIANG, J.-Y.-Q., HUANG, Y.-Q., WANG, Z.-H. & Zhang, J.-P. (2015) Effects of temperature on development and reproduction of *Neoseiulus bicaudus* (Phytoseiidae) feeding on *Tetranychus turkestanii* (Tetranychidae). *Systematic and Applied Acarology*, 20(5), 478–490.
<http://dx.doi.org/10.11158/saa.20.5.4>
- Loughner, R., Goldman, K., Loeb, G. & Nyrop, J. (2008) Influence of leaf trichomes on predatory mite (*Typhlodromus pyri*) abundance in grape varieties. *Experimental and Applied Acarology*, 45(3-4), 111–122.
<http://dx.doi.org/10.1007/s10493-008-9183-5>
- Lu, S.-L. & Zhang, J.-P. (2000) The preliminary report of species of Phytoseiid mites in Xinjiang. *Xinjiang Agricultural Sciences*, 38(S1), 127–128. (in Chinese)
- Madadi, H., Enkegaard, A., Brodsgaard, H.F., Kharrazi-Pakdel, A., Mohaghegh, J. & Ashouri, A. (2007) Host plant effects on the functional response of *Neoseiulus cucumeris* to onion thrips larvae. *Journal of Applied Entomology*, 131(9-10), 728–733.
- McMurtry, J.A. & Scriven, G.T. (1964) Studies on the feeding, reproduction, and development of *Amblyseius hibisci* (Acarina: Phytoseiidae) on various food substances. *Annals of the Entomological Society of America*, 57(5), 649–655.
- McMurtry, J.A., Denmark, H.A. & Campos, C.B. (2004) A Revised Catalog of the Mite Family Phytoseiidae. *New Zealand: Magnolia Press*, 24(2), 108–109.
<http://www.mapress.com/zootaxa/>
- Momen, F. & Hussein, H. (2011) Influence of prey stage on survival, development and life table of the predaceous mite, *Neoseiulus barkeri* (Hughes)(Acari: Phytoseiidae). *Acta Phytopathologica et Entomologica Hungarica*, 46(2), 319–328.
<http://dx.doi.org/10.1556/APhyt.46.2011.2.16>
- Norton, A.P., English-Loeb, G. & Belden, E. (2001) Host plant manipulation of natural enemies: leaf domatia protect beneficial mites from insect predators. *Oecologia*, 126(4), 535–542.
- Palevsky, E., Gal, S. & Ueckermann, E.A. (2009) Phytoseiidae from date palms in Israel with descriptions of two new taxa and a key to the species found on date palms worldwide (Acari: Mesostigmata). *Journal of Natural History*, 43(27-28), 1715–1747.
<http://dx.doi.org/10.1080/00222930902969484>
- Rebecca, A.S. (2014) Leaf structures affect predatory mites (Acari: Phytoseiidae) and biological control: a review. *Experimental and Applied Acarology*, 62, 1–17.

- <http://dx.doi.org/10.1007/s10493-013-9730-6>
- Roda, A., Nyrop, J. & English-Loeb, G. (2003) Leaf pubescence mediates the abundance of non-prey food and the density of the predatory mite *Typhlodromus pyri*. *Experimental and Applied Acarology*, 29(3-4), 193–211.
- Romero, G.Q. & Benson, W.W. (2004) Leaf domatia mediate mutualism between mites and a tropical tree. *Oecologia*, 140(4), 609–616.
<http://dx.doi.org/10.1007/s00442-004-1626-z>
- Romero, G.Q. & Vasconcellos-Neto, J.O.Ã.O. (2005) The effects of plant structure on the spatial and microspatial distribution of a bromeliad?living jumping spider (Salticidae). *Journal of Animal Ecology*, 74(1), 12–21.
<http://dx.doi.org/10.1111/j.1365-2656.2004.00893.x>
- Romero, G.Q., Daud, R.D., Salomão, A.T., Martins, L.F., Feres, R.J.F. & Benson, W.W. (2011) Mites and leaf domatia: no evidence of mutualism in *Coffea arabica* plants. *Biota Neotropica*, 11(1), 27–34.
<http://dx.doi.org/10.1590/S1676-06032011000100002>
- Schuster, R.O. & Pritchard, A.E. (1963) Phytoseiid mites of California. *Hilgardia*, 34 (7), 191–285.
<http://dx.doi.org/10.3733/hilg.v34n07p191>
- Skirvin, D.J. & Fenlon, J.S. (2001) Plant species modifies the functional response of *Phytoseiulus persimilis* (Acari: Phytoseiidae) to *Tetranychus urticae* (Acari: Tetranychidae): implications for biological control. *Bulletin of Entomological Research*, 91(1), 61–68.
- Sohrabi, F. & Shishehbor, P. (2008) Effects of host plant and temperature on growth and reproduction of the strawberry spider mite *Tetranychus turkestanii* Ugarov & Nikolski (Acari: Tetranychidae). *Systematic and Applied Acarology*, 13, 26–32.
- Tanga, C.M., Ekesi, S., Govender, P. & Mohamed, S.A. (2013) Effect of six host plant species on the life history and population growth parameters of *Rastrococcus iceryoides* (Hemiptera: Pseudococcidae). *Florida Entomologist*, 96(3), 1030–1041.
- Villanueva, R.T. & Childers, C.C. (2006) Evidence for host plant preference by *Iphiseiodes quadripilis* (Acari: Phytoseiidae) on Citrus. *Experimental and Applied Acarology*, 39(3-4), 243–256.
<http://dx.doi.org/10.1007/s10493-006-9021-6>
- Walter, D.E. (1996) Living on leaves: mites, tomenta, and leaf domatia. *Annual Review of Entomology*, 41, 101–14.
- Wang, B.-M., Wang, Z.-H., Jiang, X.-H., Zhang, J.-P. & Xu, X.-N., (2015a) Re-description of *Neoseiulus bicaudus* (Acari: Phytoseiidae) newly recorded from Xinjiang, China. *Systematic and Applied Acarology*, 20(4), 455–461.
<http://dx.doi.org/10.11158/saa.20.4.11>
- Wang, Z.-H, Li, Y.-T, Li, T, Lu, Y.-H, Zhang, J.-P. & Xu, X.-N., (2015b) The morphology and predatory behavior of the mite *Neoseiulus bicaudus*. *Chinese Journal of Applied Entomology*, 52(3), 580–586. (in Chinese)
- Wainstein, B.A. (1962) Some new predatory mites of the family Phytoseiidae (Parasitiformes) of the USSR fauna. *Entomologicheskoe Obozrenie*, 41, 230–240.
- Xia, B., Zou, Z.-W, Li, P.-X. & Lin, P. (2012) Effect of temperature on development and reproduction of *Neoseiulus barkeri* (Acari: Phytoseiidae) fed on *Aleuroglyphus ovatus*. *Experimental and Applied Acarology*, 56(1), 33–41.
<http://dx.doi.org/10.1007/s10493-011-9481-1>
- Xu, X.-N., Wang, B.-M., Wang, E.-D. & Zhang, Z.-Q. (2013) Comments on the identity of *Neoseiulus californicus* sense lato (Acari: Phytoseiidae) with a redescription of this species from southern China. *Systematic and Applied Acarology*, 18(4), 329–344.
<http://dx.doi.org/10.11158/saa.18.4.3>

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