

Illustrated Catalog of the Subtribe Helopinina Lacordaire (Coleoptera: Tenebrionidae: Blaptinae: Pedinini)

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ILLUSTRATED CATALOG OF THE SUBTRIBE HELOPININA LACORDAIRE (COLEOPTERA: TENEBRIONIDAE: BLAPTINAE: PEDININI)

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ABSTRACT

A catalog of all valid genus-group (13 genera, 10 subgenera) and species-group names (239 species and subspecies) of Helopinina darkling beetles (Coleoptera: Tenebrionidae: Blaptinae: Pedinini) and their available synonyms is presented. For each name, the author, year, and page number of the description are provided, with additional data depending on the taxon rank (e.g., type species for genus-group names, author of synonymies for invalid taxa, notes). Verified distributional records (loci typici and data acquired from revisionary publications) for all the species were gathered and illustrated. The following **new combinations** are proposed: *Amatodes angulicollis* (Gebien), *Amatodes canaliculata* (Fabricius), *Amatodes cordofana* (Haag-Rutenberg), *Amatodes delahayei* (Robiche), *Amatodes delicatula* (Fairmaire), *Amatodes demeyeri* (Robiche), *Amatodes drumonti* (Robiche), *Amatodes fasciolata* (Gebien), *Amatodes gebieni* (Robiche), *Amatodes girardi* (Robiche, Le Gall, and Goergen), *Amatodes granata* (Gebien), *Amatodes haroldi* (Haag-Rutenberg), *Amatodes hirsuta* (Solier), *Amatodes hirsutula* (Solier), *Amatodes jocquei* (Robiche), *Amatodes kochi* (Robiche), *Amatodes legalli* (Robiche), *Amatodes mucorea* (Fairmaire), *Amatodes muellerae* (Robiche), *Amatodes planicollis* (Haag-Rutenberg), *Amatodes planipennis* (Gebien), *Amatodes congoensis* (Robiche), *Amatodes limbourgi* (Robiche), *Amatodes setulosa* (Quedenfeldt), *Amatodes spinosa* (Robiche), *Amatodes claudegirardi* (Robiche), *Amatodes ertli* (Gebien), *Amatodes ferreri* (Robiche), *Amatodes jubae* (Gridelli), *Amatodes merkli* (Robiche).

Amatodes multicostata (Robiche), *Amatodes wernerii* (Robiche), *Psectes criberrimus* (Koch), and *Drosochrus costulatus* (Brancsik). Finally, a neotype is designated for *Nicandra okahandja* (Gebien) in order to fix the taxonomic status of the species.

Keywords: Africa, nomenclature, darkling beetles, Tenebrionoidea

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INTRODUCTION

Helopinina Lacordaire are a morphologically diverse subtribe of darkling beetles (Tenebrionidae Latreille) distributed across the whole African continent and parts of the Middle East (Gebien 1943; present findings). For the majority of their taxonomic history, Helopinina have been interpreted as a separate tribe within Tenebrioninae (Gebien 1943),

morphologically resembling Pedinini Eschscholtz (Doyen and Tschinkel 1982). As such, representatives of this taxon were included in a recently conducted phylogeny of pedinoid beetles as potential outgroups (Kamiński *et al.* 2019a). Surprisingly, all members of Helopinina were recovered deeply within Pedinini among representatives of subtribes Pedinina and Leichenina Mulsant (Fig. 1A). Some topologies suggested the paraphyly of Helopinini

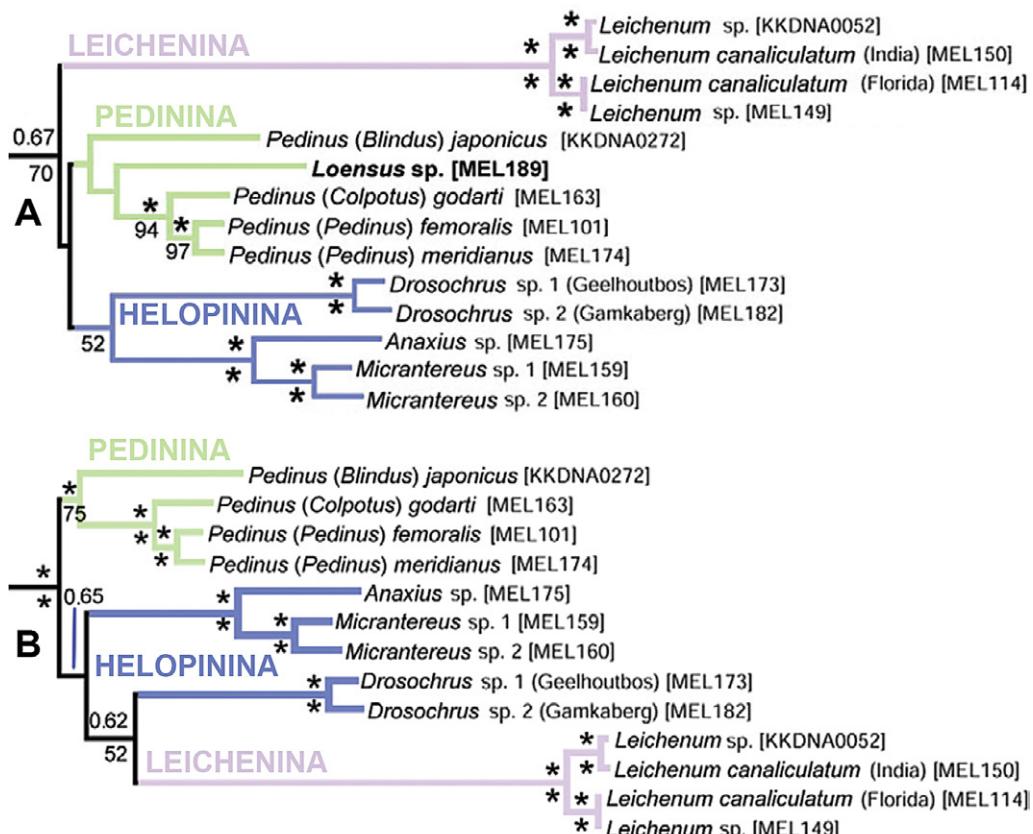


Fig. 1. Conflicting phylogenetic hypotheses of the tribe Pedinini (for details see Kamiński *et al.* 2019a). A) Preferred topology obtained with inclusion of sequences originating from a museum specimen of *Loensus* Lucas (Pedinina), B) Rejected topology rendering Helopinina paraphyletic with regard to Leichenina. An asterisk indicates either Bayesian posterior probability of 1.0 (above) or maximum likelihood bootstrap percentage of 100% boot support (below) of a particular clade.

with regard to Leichenina (Fig. 1B). However, the recovered branch support values for relations within Pedinini were low; therefore, based on morphological traits (basal margin of mentum wide, shiny; epistomal margin near maxillary fossa fold-like; metanepisternum triangular; abdominal process equal to or wider than hind coxae), the taxonomic concept of Helopinini was maintained but the taxonomic rank was lowered to subtribe (Kamiński *et al.* 2019a). Furthermore, no larval characters were found to link Helopinina and Leichenina (Kamiński *et al.* 2019b). Helopinina are currently considered one of three subtribes within Pedinini alongside Pedinina (4 genera, 134 species and subspecies; morphological phylogeny available in Kamiński and Iwan 2017) and Leichenina (2 genera, ~13 species; never fully revised). The recently revised classification places Pedinini as one of seven tribes of the darkling beetle subfamily Blaptinae Leach (Kamiński *et al.* 2021).

Although some members of Helopinina are commonly collected in the field, taxonomic contributions to the subtribe are quite scant compared to other groups of darkling beetles. This is likely due to the lack of modern revisions for most genera, and the difficulty of identifying specimens using the original descriptions and existing taxonomic publications. As a result, species-level identifications and generic assignments within Helopinina are often problematic, or impossible, without referring to type material (MJK, personal observation). The most comprehensive morphological study of beetles currently classified within Helopinina was conducted by Koch (1958), who proposed new diagnoses for some generic components, and identified “nearly 300 new species”, most of which he planned to describe in subsequent papers. However, he stopped working on this group after 1958, explicitly leaving several unsolved taxonomic problems, such as delimitation between *Blastarnodes* Koch, *Diestecopus* Solier, *Drosochrus* Erichson, *Nicandra* Fairmaire, and *Psectes* Hesse. Together these taxa represent nearly half of helopinine diversity (Fig. 2), while their definitions mainly rely on highly ambiguous features, such as the number and visibility of elytral rows (Koch 1958). Since 1958, few authors have published taxonomic contributions on Helopinina, e.g., Schulze (1968), Kaszab (1971), Ferrer (1999), and Robiche (2004a, 2008a, b, c, 2012a, b, 2013), with the first generic revision concerning a single genus (*Anaxius* Fähræus) performed only recently (Kamiński and Schoeman 2018).

Despite not being the most speciose subtribe within Pedinini, Helopinina is one of the most morphologically diverse groups not just within Pedinini but in the subfamily Blaptinae. Throughout the literature it has been noted that representatives of the subtribe resemble many distantly related tenebrionid groups

(Fähræus 1870; Haag-Rutenberg 1871; Koch 1958, 1962) such as Adesmini Lacordaire (representing subfamily Pimeliinae Latreille), Amarygmini Gistel, or Dendarini Mulsant and Rey (both Tenebrioninae) (Fig. 3). Sets of morphologically convergent species were found to be both co-occurring and allopatric, with the latter case adequately illustrated by the xerophilous species *Micrantereus scortercci physosternooides* Koch (Somalia) representing Helopinina and *Physosterna cribripes* (Haag-Rutenberg) (Angola, Namibia) of Adesmini (Koch 1962). This phenomenon of resemblance may be due to alternate defensive strategies: predator deterrence (mimicry) or predator avoidance (crypsis), as seen in the tribe Asidini (Smith *et al.* 2014). Helopinina and morphologically similar species within Tenebrioninae have paired caudal gland and reservoir systems for chemical defense, raising the possibility that their similarities are due to Müllerian mimicry. However, all Pimeliinae lack defensive glands, suggesting a potential Batesian mimicry relationship with Helopinina as the model.

Some Helopinina seem to avoid predation as adults through a combination of coloration and encrustation of the dorsal surface with surrounding substrate (Koch 1958; Robiche 2003, 2004a, 2008b, 2012a). Judging solely on general morphology, this defensive strategy is expected to afford protection to species in *Amatodes* Dejean, *Ametrocera* Fähræus, *Aptila* Fähræus, *Asidodema* Koch, *Oncopteryx* Gebien, and *Piscicula* Robiche. Furthermore, many of these cryptic taxa display a great degree of morphological resemblance to selected groups of Pimeliinae darkling beetles, particularly those of tribes Asidini Fleming and Sepidiini Eschscholtz (Fig. 3). The resemblance is commonly manifested in the similar pronotal shape (angular sides), presence of carinae or tubercles on the pronotum and elytra, and dense setation of the body. It can be hypothesized that at least some of those characters enable individuals to blend and coat themselves with particles from the substrate (Smith *et al.* 2014). However, a comprehensive overview of cryptic strategies within darkling beetles is needed to test this assumption.

Due to the interest in Helopinina taken by Carl Koch and Lieselotte Schulze, a large, identified collection of material concerning this subtribe is preserved at the Ditsong Museum (Pretoria, South Africa). Material identified to species level in this collection currently includes 45 drawers. Furthermore, due to collecting efforts made by Ditsong Museum personnel, new specimens of Helopinina are available for study in the collection (~10 unsorted drawers). Many of the aforementioned “species” designated by Koch (1958) have not been formally described, but bear labels indicating their proposed name-bearing status and specific names (Fig. 4). The specific set of

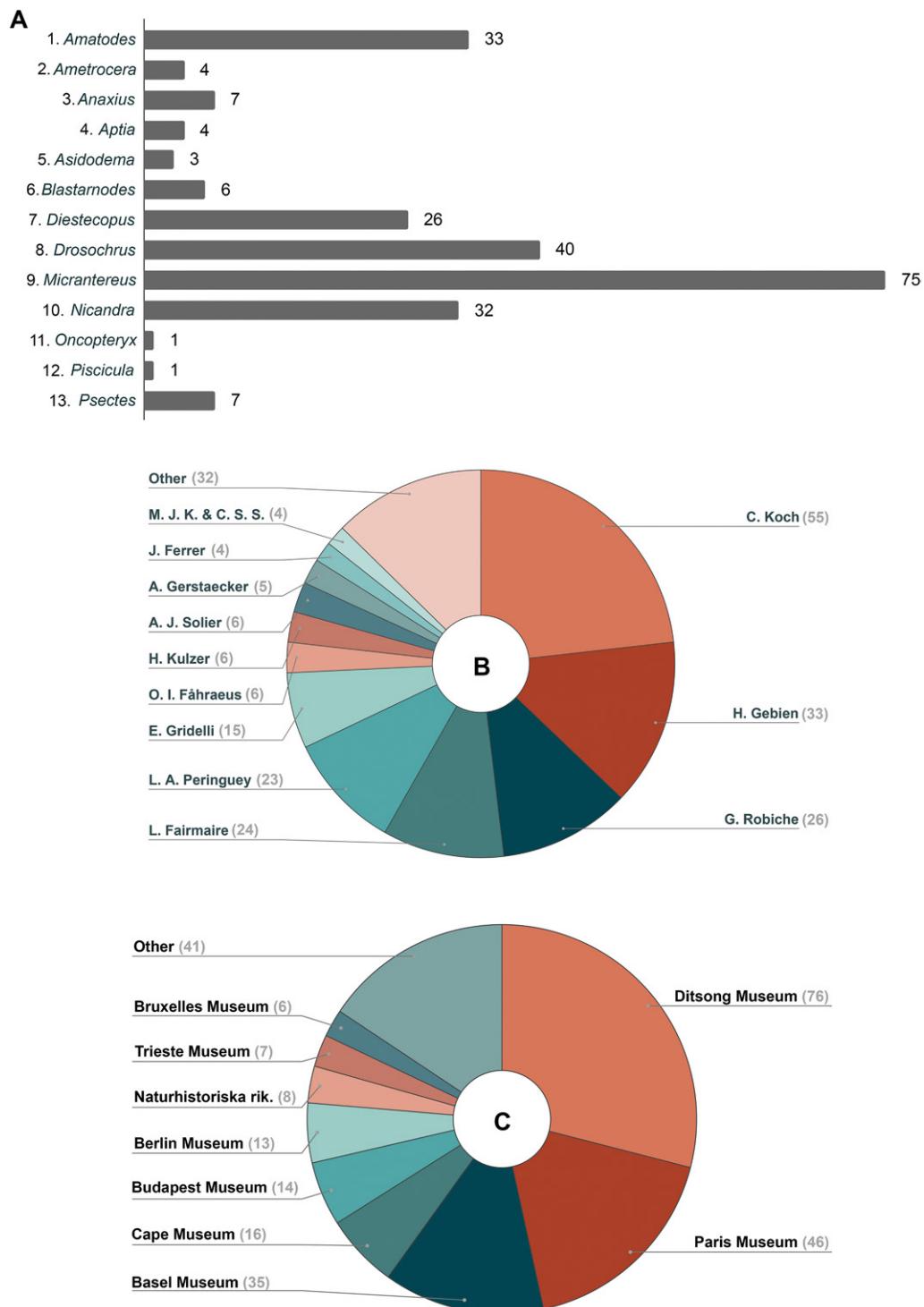


Fig. 2. Taxonomic diversity of the subtribe Helopinina and summary of collections and descriptors. A) Species and subspecies diversity of the genera, B) Authors of species and subspecies descriptions, C) Entomological collections hosting the primary types of the species and subspecies representing Helopinina.

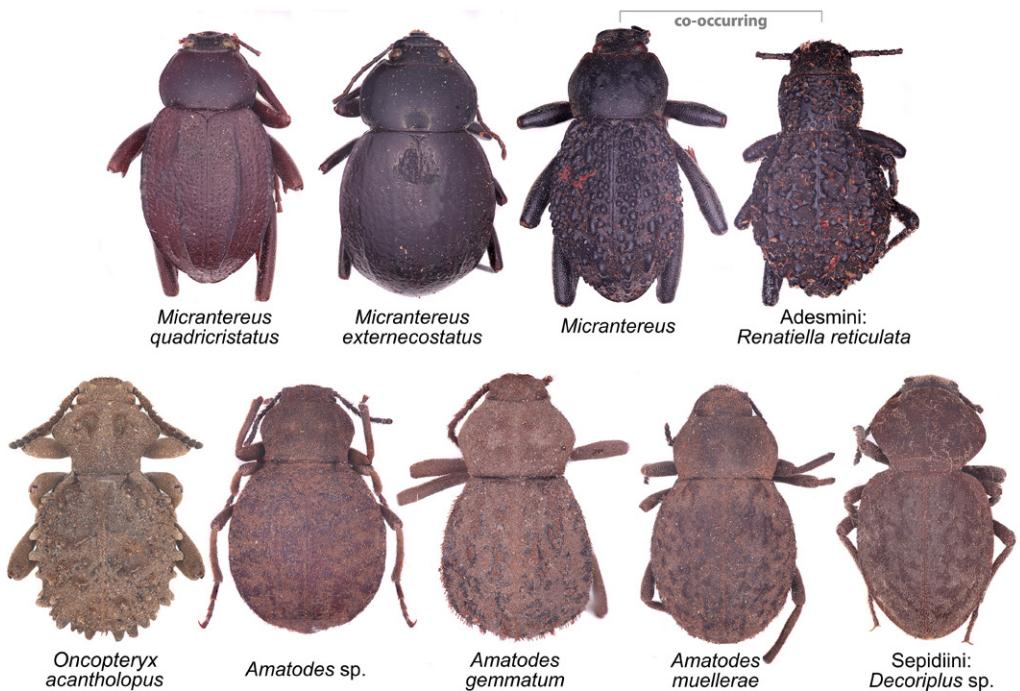


Fig. 3. Morphological diversity of Helopinina and their resemblance to some distantly related darkling beetle tribes.

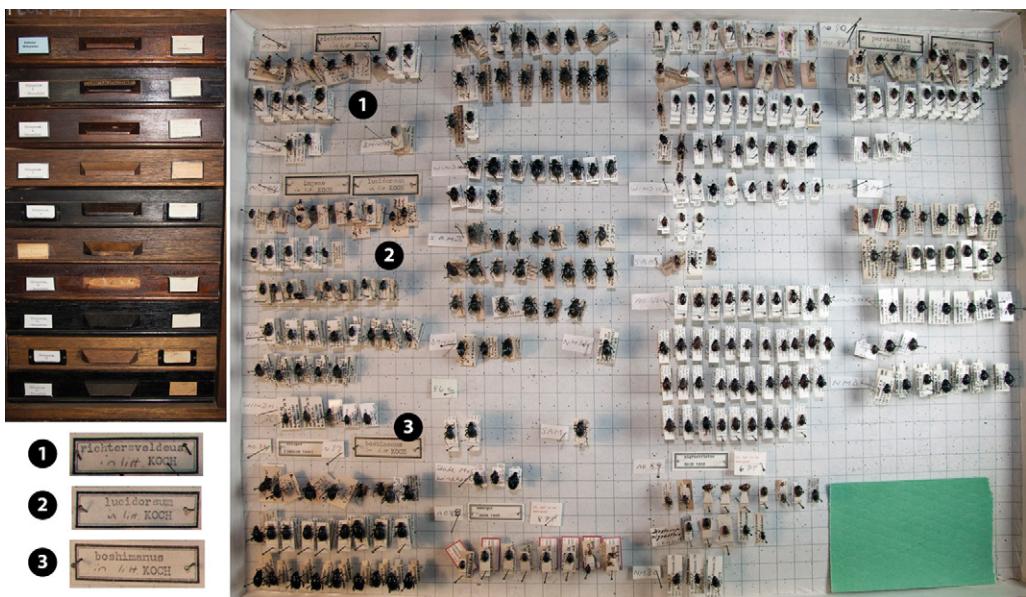


Fig. 4. Overview of the Helopinina material deposited at Ditsong Museum (Pretoria, South Africa). Selected drawers with identified material (top left), exemplar drawer with undescribed yet designated as new ("in litt.") species of helopinine beetles.

circumstances created by the collection in the Ditsong museum creates a unique environment for taxonomic work, which can be efficiently used to address unresolved questions within the group such as the actual number of species currently described, the validity and accuracy of descriptions, generic and species concepts, and the phylogenetic placement of taxa.

The main purpose of this publication is to generate a foundation for future work in Helopinina by compiling the available nomenclatural, taxonomic and distributional knowledge for the subtribe in the form of a catalog, as the last estimate of species diversity was done over 80 years ago. At that time Gebien (1943) listed 121 species in 12 genera. Furthermore, as no reliable identification tools exist for Helopinina, habitus and label photos of select type specimens preserved at the Ditsong Museum are presented.

MATERIAL AND METHODS

Nomenclatural Data. All nomenclaturally available genus- and species-group names are listed. The author, year, and page on which the generic or species name is described, type data, and distributional records are provided for each name. If an original description was not provided in the publication that made a name available, the page on which the name is made available is presented. For each valid genus-group name, gender and type fixation are included. The reference in which a given generic or specific name is first placed in synonymy with the current valid name is listed, e.g., [syn. by Gebien 1911: 563]. For every species-group name the original genus-group name is provided. Type deposition data and the status of the name-bearing types was taken from the original publication, unless specified differently.

The following format for deposition information was used through the catalog:

Basel Museum – Naturhistorisches Museum Basel, Basel, Switzerland

Berlin Museum – Museum für Naturkunde, Berlin, Germany

British Museum – The Natural History Museum, London, United Kingdom

Brussels Museum – Muséum des sciences naturelles de Belgique, Brussels, Belgium

Budapest Museum – Magyar Természettudományi Múzeum, Budapest, Hungary

California Academy – Museum of the California Academy of Sciences, San Francisco, CA, USA

Cape Museum – Iziko South African Museum, Cape Town, South Africa

Carmagnola Museum – Museo Civico di Storia Naturale di Carmagnola, Turin, Italy

Companhia Diamantes – Companhia de Diamantes de Angola, Luanda, Angola

Copenhagen Museum – Statens Naturhistoriske Museum, University of Copenhagen, Copenhagen, Denmark

Coryndon Museum – Natural History Museum of Kenya, Nairobi, Kenya

Dahlem Museum – Dahlem Museum, Berlin, Germany

Ditsong Museum – Ditsong National Museum of Natural History, Pretoria, South Africa

Dundo Museum – Dundo Museum, Dundo, Angola

Florence Museum – Museo di Storia Naturale di Firenze, Florence, Italy

Geneva Museum – Muséum d'Histoire Naturelle, Geneva, Switzerland

Genoa Museum – Museo Civico di Storia Naturale “G. Doria”, Genoa, Italy

Hamburg University – Universität Hamburg, Hamburg, Germany

Harvard Museum – Harvard Museum of Comparative Zoology, Cambridge, MA, USA

Humboldt University – Humboldt-Universität, Berlin, Germany

Leiden Museum – Naturalis Museum, Leiden, Holland

Lund University – Zoological Museum, Lund University, Lund, Sweden

Milan Museum – Museo Civico di Storia Naturale, Milan, Italy

Munich Museum – Bayerisches Nationalmuseum, Munich, Germany

Naturhistoriska riksmuseet – Naturhistoriska riksmuseet, Stockholm, Sweden

Oxford University – Oxford University Museum of Natural History, Oxford, United Kingdom

Paris Museum – Muséum national d'Histoire naturelle, Paris, France

Robiche coll. – private collection of author GR

Tervuren Museum – Musée royal de l'Afrique centrale, Tervuren, Belgium

Torino Museum – Museo Regionale di Scienze Naturali di Torino, Turin, Italy

Trieste Museum – Museo Civico di Storia Naturale, Trieste, Italy

Vernouillet coll. – private collection of Vernouillet (France); see Robiche (2004a)

Vienna Museum – Naturhistorisches Museum Wien, Wien, Austria

Warsaw Museum – Muzeum i Instytut Zoologii, Polska Akademia Nauk, Warsaw, Poland

Distributional Data. The distribution of all genera was illustrated using Quantum GIS (QGIS) v. 2.4. All vector layers were downloaded from the Natural Earth webpage (www.naturalearthdata.com). The list of localities was built by consulting available literature and is available free of charge on Harvard Dataverse repository (Kamiński

2020). Geographic data with low degrees of accuracy (e.g., countries or regions) were not georeferenced, and therefore are absent on the maps and are marked with an asterisk in the distributional sections of particular species.

RESULTS

A total of 239 valid species and subspecies divided into 13 valid genera are listed in this catalog (Fig. 2A). The genus *Micrantereus* Solier is the most diverse with 75 valid species-group taxa, followed by *Drosochrus* (40), *Amatodes* (33), *Nicandra* (32), *Diestecopus* (26), *Anaxius* (7), *Psectes* (7), *Blastarnodes* (6), *Ametrocera* (4), *Aptia* (4), *Asidodema* (3), and the two monotypic genera *Oncopteryx* and *Piscicula*. Nearly 75% of the total helopinine species diversity was described by the following six researchers (Fig. 2B): C. Koch (55 species and subspecies), H. Gebien (33), G. Robiche (26), L. Fairmaire (24), L. A. Péringuey (23) and E. Gridelli (15). However, the majority of the type species of different generic-level taxa (15 out of 20 genera and subgenera) were described in older contributions authored by other entomologists, i.e., Fabricius (1801), Guérin-Méneville (1834), Erichson (1843), Solier (1848), Fähræus (1870), Quedenfeldt (1885), Fairmaire (1887, 1888a), Gebien (1910a, 1920), and Hesse (1935).

The name-bearing types of the species representing Helopinina are preserved mainly in European and African collections (Fig. 2C), although syntypes of three species are deposited in USA institutions (*Nicandra costatula* Koch, *Nicandra namaquensis* Koch, both in California Academy, and *Psectes kokoanus* Koch in Harvard Museum). The Ditsong Museum and Paris Museum host over 45% of the total name-bearing types of Helopinina.

The majority of the new combinations introduced in the catalog below are a result of a synonymy noticed by Bousquet and Bouchard (2013), which concerns precedence of *Amatodes* Dejean over *Oncosoma* Westwood. Additionally, the type digitization effort conducted here reveals inconsistencies concerning the definitions of some currently accepted genera. Especially apparent is the case of *Diestecopus* and *Psectes* (Figs. 8, 17); however, several other genera need taxonomic revision as well (e.g., *Nicandra*, personal observation).

A database containing 440 distributional records (64 not georeferenced) was created during this study (Figs. 5, 6; Kamiński 2020). According to the acquired data, Helopinina species are widely distributed throughout the eastern Afrotropical Realm (Fig. 5). Only the representatives of *Drosochrus* and *Micrantereus* have distribution ranges extending into the Palearctic Realm (Fig. 6). However, due to

the shortage of available contributions, most species are known only from the type series, which precludes any detailed biogeographic analysis at the species level. Furthermore, this phenomenon probably contributes to the seemingly disjunct ranges of many genera (Figs. 5, 6). One of the oddest distributional patterns revealed here concerns *Amatodes* and *Micrantereus*, of which the majority of species were collected from the eastern part of the African continent, while a few species of these genera seem to be restricted to West Africa (Fig. 5). Namely, *Amatodes girardi* (Robiche, Le Gall, and Goergen) was reported from Benin, Burkina Faso, Ghana, and Ivory Coast, *Amatodes delicatula* (Fairmaire) and *Amatodes mucorea* (Fairmaire) from Mali, *Amatodes canaliculata* (Fabricius) from Guinea, *Amatodes gemmata* (Fabricius) from Gambia, Guinea-Bissau, and Senegal, *Amatodes hirsutula* Solier from Guinea and Senegal, and *Amatodes hirsuta* Solier and *Micrantereus anomalus* (Guérin-Méneville) from Senegal.

CATALOG OF THE HELOPININA

Subtribe Helopinina Lacordaire, 1859

Hélopinides Lacordaire 1859: 457.

- = *Aptilina* Koch 1958: 139 [syn. by Kamiński *et al.* 2019a]. Type genus: *Aptila* Fähræus, 1870.
- = *Drosochrini* Koch 1958: 133 [unnecessary replacement name for *Helopinini* Lacordaire, 1859]. Type genus: *Drosochrus* Erichson, 1843.
- = *Micrantereimi* Reitter 1917: 60 [syn. by Kamiński *et al.* 2019a]. Type genus: *Micrantereus* Solier, 1848.
- = *Oncosomina* Koch 1958: 134 [syn. by Kamiński *et al.* 2019a]. Type genus: *Oncosoma* Westwood, 1843.

Type Genus. *Helopinus* Solier, 1848.

Diagnosis. Helopinina can be distinguished from other subtribes of Pedinini by the following characteristics (Kamiński *et al.* 2019a, 2021): supraorbital crest flat (carinated in Leichenina); antennae elongate (short in Leichenina); eyes not covered with scales (covered with scales in Leichenina); the basal margin of mentum wide and shiny (narrow and dull in Pedinina); epistomal margin near maxillary fossa fold-like (flat in Pedinina); metanepisternum triangular (rectangular in Pedinina); abdominal process equal to or wider than hind coxae (narrower than hind coxae in Pedinina).

From the remaining tribes of the opatrionoid clade (Kamiński *et al.* 2019a, 2021), Helopinina differs by having gula smooth or covered with irregular rugosities (with stridulatory surface in Platynotini), mentum with lateral wings visible (mentum flat covering the wings in Dendarini), and protrochanter without elongate base (with elongate base in Opatrini).

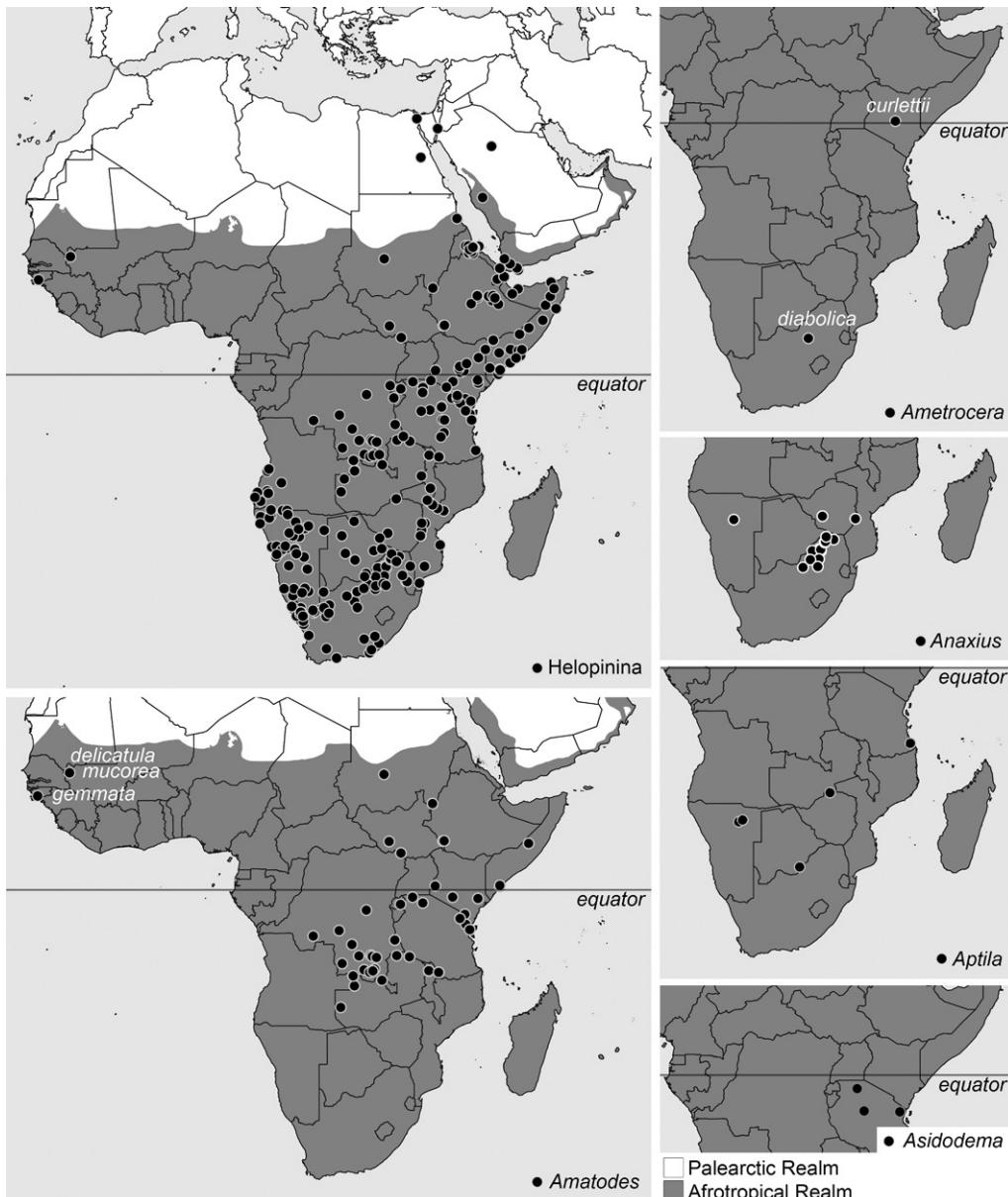


Fig. 5. Distribution of the subtribe Helopinina and specific genera.

TAXONOMIC DIVERSITY (13 genera, 239 species). *Amatodes* (33 spp.), *Ametrocera* (4), *Anaxius* (7), *Aptila* (4), *Asidodema* (3), *Blastarnodes* (6), *Diestecopus* (26), *Drosochrus* (40), *Micrantereus* (75), *Nicandra* (32), *Oncopteryx* (1), *Piscicula* (1), *Psectes* (7).

DISTRIBUTION. Africa and Arabian Peninsula (Figs. 5, 6).

Genus *Amatodes* Dejean, 1834

Subgenus *Amatodes* Dejean, 1834

Amatodes Dejean 1834: 189 [feminine].
= *Ogcosoma* Westwood 1843: 121 [syn. by Gemminger in Gemminger and Harold (1870: 1897)]. Type species: *Ogcosoma granulare* Westwood, 1843; by monotypy.

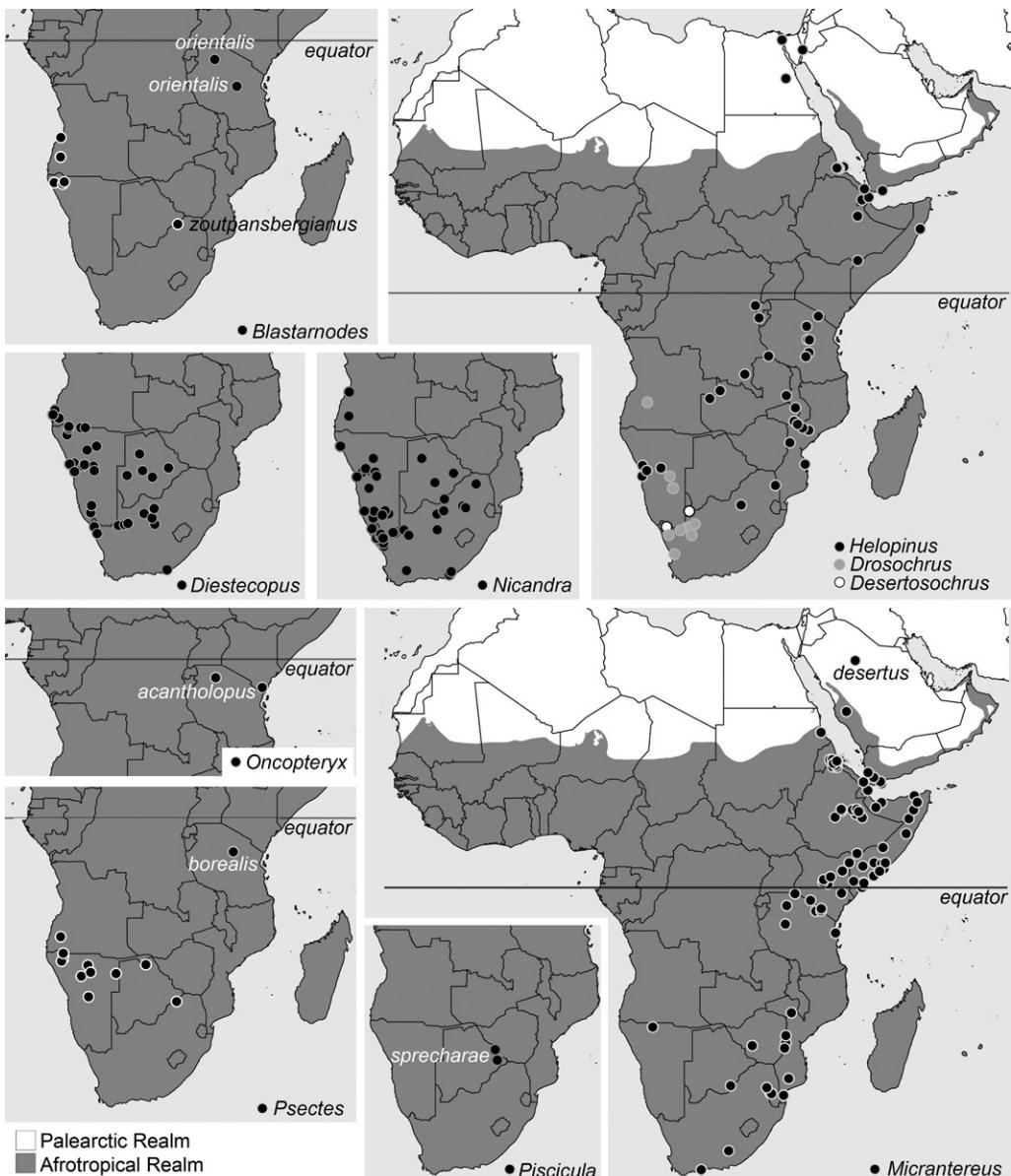


Fig. 6. Distribution of specific genera of the subtribe Helopinina.

= *Oncosoma* Gebien 1911: 563 [unjustified emendation of *Oncosoma* (as “*Ogcosoma*”), not in prevailing usage].

Note. *Oncosoma* is an unjustified emendation from *Ogcosoma* by Agassiz (1846: 259), in prevailing usage, treated as justified emendation (ICZN 1999, Art. 33.2.3.1) (see Bouchard *et al.* 2005).

Type Species. *Pimelia gemmata* Fabricius, 1801; by monotypy.

1. *Amatodes angulicollis* (Gebien, 1910), new combination

Oncosoma angulicolle Gebien 1910a: 144.

Type Data. Syntypes, six specimens (Basel Museum), single specimen (Budapest Museum): “Bunkeya, Kapema-Kipaila, Kamfua”.

Distribution. DEMOCRATIC REPUBLIC OF THE CONGO: Bunkeya, Kapema-Kapaila, Kamfua (Gebien 1910a).

2. *Amatodes canaliculata* (Fabricius, 1801), new combination

Pimelia canaliculata Fabricius 1801: 132.
= *Ogcosoma guineense* Haag-Rutenberg 1871: 30
[syn. by Gebien 1911: 563].

Type Data. *Pimelia canaliculata* Fabricius, 1801, **syntypes**, not specified (Copenhagen Museum): “Guinea”.

Oncosoma guineense Haag-Rutenberg, 1871, **syn-types**, not specified (Munich Museum): “Guinea”.

Distribution. GUINEA* (Haag-Rutenberg 1871).

3. *Amatodes cordofana* (Haag-Rutenberg, 1871), new combination

Ogcosoma cordofanum Haag-Rutenberg 1871: 30.

Type Data. Holotype, male (Vienna Museum): “Kordofan”.

Distribution. SUDAN: Kordofan (Haag-Rutenberg 1871).

4. *Amatodes delahayei* (Robiche, 2009), new combination

Oncosoma delahayei Robiche 2009: 4.

Type Data. Holotype, male (Paris Museum): “Zambie, prov. N. O., 80 km sud Mwinilunga, 19.XI.2003, K. Wemer & SMRZ leg.”. **Paratype**, female (Robiche coll.): same data as holotype.

Distribution. ZAMBIA: Mwinilunga (Robiche 2009).

5. *Amatodes delicatula* (Fairmaire, 1893), new combination

Oncosoma delicatulum Fairmaire 1893a: 149.

Type Data. Holotype, not specified (Paris Museum): “Kayes”.

Distribution. MALI: Kayes (Fairmaire 1893a).

6. *Amatodes demeyeri* (Robiche, 2001), new combination

Oncosoma demeyeri Robiche 2001: 78.

Type Data. Holotype, male (Tervuren Museum): “Kasai, Lula, terr. Luisa. VIII.1956 (Dr. M Poll leg.)”. **Paratypes**, two specimens (Tervuren Museum): same data as holotype; two specimens (Tervuren Museum): “Kasai, Lula, 1958 (A.J. Jobaert leg.)”; nine specimens (Tervuren Museum): “Sandoa, XI.1931 (F.G. Overlaet leg.)”; three specimens (Paris Museum): “Sandoa, XI.1931 (A.J. Jobaert leg.)”; two specimens (Robiche coll.): “idem, (F.G. Overlaet leg.)”; three specimens (Paris Museum): “idem, (F.G. Overlaet leg.)”; seven specimens (Tervuren Museum): “Lulua, Muteba, XI. 1931 (F.G. Overlaet leg.)”; single specimen (Paris Museum): “Lulua, Muteba (F.G. Overlaet leg.)”; single specimen (Paris Museum): “Kwango, Kianza, terr. De Feshl, IX.1959 (H. Daems leg.)”.

Distribution. DEMOCRATIC REPUBLIC OF THE CONGO: Kaaasi*, Lula*, Sandoa, Muteba, Kianza (Robiche 2001).

7. *Amatodes drumonti* (Robiche, 2009), new combination

Oncosoma drumonti Robiche 2009: 2.

Type Data. Holotype, male (Ditsong Museum): “République Démocratique du Congo, Lualaba, Kamina, 1950, Dr. Buis leg.”. **Paratypes**, female (Ditsong Museum): same data as holotype; female (Robiche coll.): “République Démocratique du Congo, Lualaba, Kamina, XI.1954, R. Ledieu leg.”.

Distribution. DEMOCRATIC REPUBLIC OF THE CONGO: Kamina, Lualaba (Robiche 2009).

8. *Amatodes fasciolata* (Gebien, 1910), new combination

Oncosoma fasciolatum Gebien 1910a: 144.

Type Data. Syntypes, five specimens (Basel Museum): “Madona, Kambove-Bunkeya, Sunba, Kapema-Kipaila”.

Distribution. DEMOCRATIC REPUBLIC OF THE CONGO: Bunkeya, Kambove, Kapema-Kipaila. Madona*, Sunba (Gebien 1910a).

9. *Amatodes gebieni* (Robiche, 2008), new combination

Oncosoma gebieni Robiche 2008b: 529.

Type Data. Holotype, male (Ditsong Museum): “République Démocratique du Congo, Kasenga, 02.II.1912, Dr Bequaert leg.”.

Distribution. DEMOCRATIC REPUBLIC OF THE CONGO: Kasenga (Robiche 2008b).

10. *Amatodes gemmata* (Fabricius, 1801)

Pimelia gemmata Fabricius 1801: 132.

Amatodes gemmata: Dejean 1834: 189.

= *Ogcosoma granulare* Westwood 1843: 121 [syn. by Gebien 1911: 563].

Type Data. Pimelia gemmata Fabricius, 1801, **syn-types**, not specified (Copenhagen Museum): “Guinea”.

Ogcosoma granulare Westwood, 1843, **holotype**, not specified (Oxford University): “Gambia”.

Distribution. SENEGAL* (Dejean 1834); GAMBIA* (Westwood 1843); GUINEA-BISSAU: Bolama (Gebien 1921).

11. *Amatodes girardi* (Robiche, Le Gall, and Goergen, 2002), new combination

Oncosoma girardi Robiche et al. 2002: 409.

Type Data. Holotype, male (Paris Museum): “Haute Volta, près de Ouagadougou, Pabré, VIII.1974, R. P. Fernandez leg.”. **Paratypes**, female (Paris Museum): same data as holotype; six specimens (Paris Museum): same data as holotype; one specimen

(Paris Museum): "Haute Volta, Léo, VII.1960, R.P. Nicolas leg.;" one specimen (Paris Museum): "Haute Volta, Bobo Dioulasso, VIII.1960, P. Jolivet leg.;" twenty specimens (Paris museum and Robiche coll.): "Côte d'Ivoire, sud du parc de la Comoé, IV.1998, P. Moretto leg.;" one specimen (Paris Museum): "northern region, Tamale, 15.VII.1970, Dr S. Endrödy-Younga leg.;" two specimens (Paris Museum): "Upper region, Tumu, N.10°.08'-W.02°.00', 12.VIII.1971. Dr S. Endrödy-Younga leg.;" six specimens (Robiche coll.): "Bénin, Parakou, IV.VI.1996, P. Le Gall and G. Robiche leg.;" five specimens (Paris Museum and Robiche coll.): "Bénin, Djougou, IX. 1996, P. Le Gall and G. Robiche leg.;" one specimen (Robiche coll.): "Bénin, Djougou, IX. 1996, P. Le Gall leg.;" one specimen (Genova Museum): "Haut Uélé, Dongou, IV.1927, F.S. Patrizi leg.."

Distribution. BURKINA FASO: Bobo-Dioulasso (Robiche *et al.* 2002); IVORY COAST: Comoé National Park (Robiche *et al.* 2002); GHANA: Tamale, Tumu (Robiche *et al.* 2002); BENIN: Djougou, Parakou Pabré (Robiche *et al.* 2002).

12. *Amatodes granata* (Gebien, 1921), new combination

Oncosoma granatum Gebien 1921: 124.

Type Data. Syntypes, two specimens (Brussels Museum): "Niam-Niam (Bohdorff); single specimen (Kongo Museum): "Sassa (Colmant)".

Distribution. SOUTH SUDAN: Niam-Niam*, Sassa (Gebien 1921).

13. *Amatodes haroldi* (Haag-Rutenberg, 1875), new combination

Oncosoma haroldi Haag-Rutenberg 1875: 67.

Type Data. Syntypes, not specified (Munich Museum): "Fassoglu, Bogos-Ländern".

Distribution. SUDAN: Fassoglu, Bogos-Ländern* (Haag-Rutenberg 1875); SOMALIA: Surroundings of Dusa-Mareb (Ferrer 1995).

14. *Amatodes hirsuta* Solier, 1843

Amatodes hirsutum Solier 1843: 268.

Type Data. Syntypes, not specified (Paris Museum): "Du Sénégal, Collection de Mr Dupont".

Distribution. SENEGAL* (Solier 1843).

15. *Amatodes hirsutula* Solier, 1843

Amatodes hirsutulum Solier 1843: 267.

Type Data. Syntypes, not specified (Paris Museum): "Du Sénégal".

Distribution. GUINEA* (Haag-Rutenberg 1871); SENEGAL* (Solier 1843).

16. *Amatodes jocquei* (Robiche, 2001), new combination

Oncosoma jocquei Robiche 2001: 77.

Type Data. Holotype, male (Tervuren Museum): "Malawi North Reg., Nyika plateau, Chelinda 2600 m, XI.1981 (R. Jocqué leg.)". **Paratypes**, six specimens (Tervuren Museum): same data as holotype; single specimen (Tervuren Museum): "Malawi, entre Chelinda et Rumphi, 1700 m, XI.1981 (R. Jocqué leg.)"; three specimens (Paris Museum): "Malawi North Re., Nyika, Chelinda 2300m, XI.1981 (R. Jocqué leg.)"; two specimens (Robiche coll.): "idem, (R. Jocqué leg.)".

Distribution. MALAWI: Chelinda, Nyika plateau (Robiche 2001).

17. *Amatodes kochi* (Robiche, 2008), new combination

Oncosoma kochi Robiche 2008b: 525.

Type Data. Holotype, male (Ditsong Museum): "Congo Belge, PNU, Kasenga, Mabwe (585 m), 24-31.XII.1948, Miss. G.F. de Witte". **Paratypes**, female (Ditsong Museum): same data as holotype; eight specimens (Ditsong Museum and Robiche coll.): "19.XI.1948, Mis. G.F. de Witte"; five specimens (Ditsong Museum): same data except "12-15. XII.1948"; four specimens: same data except "17-27.XII.1948"; two specimens: same data except "12-15.XII.1948"; single specimen (Ditsong Museum): "Congo Belge, Kinda, XII.1926".

Distribution. DEMOCRATIC REPUBLIC OF THE CONGO: Kindu, Mabwe (Robiche 2008b).

18. *Amatodes legalli* (Robiche, 2008), new combination

Oncosoma legalli Robiche 2008b: 530.

Type Data. Holotype, male (Ditsong Museum): "Congo Belge, PNU., Kaswabilenga (700 m), 23-24.X.1947, Miss. G. F. de Witte". **Paratypes**, female (Ditsong Museum): same data as holotype; two specimens (Robiche coll.): same data as holotype; two specimens (Ditsong Museum): same data except "XI.1947"; 19 specimens (Ditsong Museum and Robiche coll.): same data except "Kankunda (1300 m), 19-24.XI.1947"; 17 specimens (Ditsong Museum and Robiche coll.): same data except "r. Kateke-s/afl. Lufira (950 m), 23.XI.1947"; three specimens (Ditsong Museum): same data except "Kankunda affl. G Lupiala (1300 m), 22-26.X.1947"; single specimen (Ditsong Museum): same data except "piste Lupiala (900-1200 m), 23.X.1947"; single specimen (Ditsong Museum): same data except "Lupiala (850 m), 24.X.1947"; single specimen (Ditsong Museum): same data except "Lukawe (700 m), 28.X.1947"; single specimen (Ditsong Museum): same data except "Lusinga (1760 m), 1-8.XII.1947".

Distribution. DEMOCRATIC REPUBLIC OF THE CONGO: Kankunda*, Kaswabilenga*, Lukawe*, Lupiala*, Lusinga (Robiche 2008b).

19. *Amatodes mucorea* (Fairmaire, 1893), new combination

Oncosoma mucoreum Fairmaire 1893a: 148.

Type Data. Holotype, not specified (Paris Museum): “Kayes”.

Distribution. MALI: Kayes (Fairmaire 1893a).

20. *Amatodes muellerae* (Robiche, 2008), new combination

Oncosoma muellerae Robiche 2008b: 526.

Type Data. Holotype, male (Ditsong Museum): “Rhodésie du Nord (Zambie), Abercorn, H-J. Bredo leg., X.1943 et 15.XII.1943”. **Paratypes**, female (Ditsong Museum): same data as holotype; five specimens (Ditsong Museum and Robiche coll.): same data except “11.XI.1942” and “30.X.1943”; four specimens (Ditsong Museum and Robiche coll.): same data except “Mweru, Wantipa, Buleya”; five specimens (Ditsong Museum and Robiche coll.): same data except “XII.1943”; single specimen (Ditsong Museum): same data except “H.J. Bredo leg.”.

Distribution. ZAMBIA: Abercorn, Buleya, Mweru Wantipa (Robiche 2008b).

21. *Amatodes planicollis* (Haag-Rutenberg, 1879), new combination

Ogcosoma planicolle Haag-Rutenberg 1879: 289.

Type Data. Syntypes, not specified (Munich Museum): “Nyassa Coll. Bates”.

Distribution. Nyassa*.

22. *Amatodes planipennis* (Gebien, 1910), new combination

Oncosoma planipenne Gebien 1910a: 147.

Type Data. Syntypes, male (Basel Museum): “Ukerewe, Deutsch-Ostafrika von Herrn Schulinspektor Ertl”; two males (Basel Museum): “Herr von Kigonsera”.

Distribution. TANZANIA: Kigonsera, Ukerewe island (Gebien 1910a).

Subgenus *Conophthalmus* Quedenfeldt, 1885

Conophthalmus Quedenfeldt 1885: 13 [masculine].

Type Species. *Conophthalmus setulosus* Quedenfeldt, 1885; by monotypy.

23. *Amatodes congoensis* (Robiche, 2012), new combination

Oncosoma congoensis Robiche 2012a: 19.

Type Data. Holotype, male (Brussels Museum): “Congo Belge, Mpala, VII.1948, leg. R. P. Lefèvre”.

Distribution. DEMOCRATIC REPUBLIC OF THE CONGO: Mpala (Robiche 2012a).

24. *Amatodes limbourgi* (Robiche, 2012), new combination

Oncosoma limbourgi Robiche 2012a: 17.

Type Data. Holotype, male (Brussels Museum):

“Rhodésie du nord (Zambie), Abercorn, 15.XII.1943, I.G. N°15.333, leg. H. J. Bredo”.

Paratypes, male and unsexed specimen (Brussels Museum): same data as holotype.

Distribution. ZAMBIA: Abercorn (Robiche 2012a).

25. *Amatodes setulosa* (Quedenfeldt, 1885), new combination

Conophthalmus setulosum Quedenfeldt 1885: 14.

Type Data. Lectotype, designated by Robiche (2013), unsexed (Paris Museum): “*Conophthalmus* n. sp. *Setulosus* n. sp. Qdf”; “Lectotype *Conophthalmus setulosus* Qued. Robiche G. Dés. 2013”.

Distribution. ANGOLA* (Quedenfeldt 1885).

26. *Amatodes spinosa* (Robiche, 2008), new combination

Oncosoma spinosum Robiche 2008c: 525.

Type Data. Holotype, male (Paris Museum): “Zambie, prov. N.O, Nchila-Ikelenge, 13.XI.2003, K. Werner & SMRZ/leg.”.

Paratype, female (Robiche coll.): same data as holotype.

Distribution. ZAMBIA: Nchila-Ikelenge (Robiche 2008c).

Subgenus *Strophiamixa* Robiche, 2005

Strophiamixa Robiche 2005: 358 [feminine], replacement name.

= *Strophia* Robiche 2004a: 130 [junior homonym of *Strophia* Meigen, 1832 (Insecta: Lepidoptera)].

Type Species. *Oncosoma ertli* Gebien, 1910; by original designation.

27. *Amatodes claudegirardi* (Robiche 2004), new combination

Oncosoma claudegirardi Robiche 2004a: 134.

Type Data. Holotype, male (Paris Museum): “Kenya, région du Parc de Tsavo, Boura Wa-Taita, 1904 (Ch. Alluaud leg.)”. **Paratypes**, female (Paris Museum): “Kenya, Makuro, Rift Valley, 1904 (Ch. Alluaud leg.)”; male and three females (Paris Museum and Vernouillet coll.): “Kenya, Kibwesi, Wa Kamba, 1904 (Ch. Alluaud leg.)”.

Distribution. KENYA: Boura Wa-Taita (Tsavo National Park), Wa Kamba*, Makuro* (Rift Valley) (Robiche 2004a).

28. *Amatodes ertli* (Gebien, 1910), new combination

Oncosoma ertli Gebien 1910a: 148.

Type Data. Holotype, not specified (Basel Museum): “Kigonsera am Nyassa-See, 1906”.

Distribution. TANZANIA: Kigonsera (Gebien 1910a); RWANDA: Gashora; TANZANIA: Katoma, Kibaro*, Kigonsera, Uasi*, Upa*; KENYA: Nairobi (Robiche 2004a), surroundings of Malindi (Ferrer 1996).

29. *Amatodes ferreri* (Robiche, 2004), new combination
Oncosoma ferreri Robiche 2004a: 138.

Type Data. Holotype, male (Tervuren Museum): “Ethiopie, Kaffa prov., Magi airfield (Washa Wuha, 650m, 11. IV. 1972”). Paratypes, female (Tervuren Museum): same data as holotype; female (Vernouillet coll.): “Mui game reserve, 700 m, 10.IV.1972”; two males and three females (Tervuren Museum and Vernouillet coll.): “Mui, 08-10.IV. 1972”.

Distribution. ETHIOPIA: Mui, Washa Wuha (Robiche 2004a).

30. *Amatodes jubae* (Gridelli, 1939), new combination

Micrantereus jubae Gridelli 1939b: 47.

Type Data. Syntypes, male (Geneva Museum): “Somalie, Bulo Kero, 1934 (F. Patrizi leg.)”, male (Trieste Museum): “Somalie, Belet Amin, 1934 (F. Patrizi leg.)”, male (Basel Museum): “Somalie, Bulo Kero, dét. Gridelli Micrantereus jubae”, male (Basel Museum): “Somalie italienne, Belet Amin, VIII. 1934, (F. Patrizi leg.)”.

Distribution. SOMALIA: Belet Amin, Bulo Kero (Gridelli 1939b).

31. *Amatodes merkli* (Robiche, 2004), new combination

Oncosoma merkli Robiche 2004a: 136.

Type Data. Holotype, male (Paris Museum): “Tanzanie, West Usambara”. Paratype, male: same data as holotype.

Distribution. TANZANIA: Usambara Mountains (Robiche 2004a).

32. *Amatodes multicostata* (Robiche, 2004), new combination

Oncosoma multicostatum Robiche 2004a: 136.

Type Data. Holotype, male (Paris Museum): “Tanzanie, Pangani”. Paratype, male (Paris Museum): “Tanzanie, Pare Gebirge”.

Distribution. TANZANIA: Pangani, Pare Mountain (Robiche 2004a).

Incertae Sedis

33. *Amatodes werneri* (Robiche, 2003), new combination

Oncosoma werneri Robiche 2003: 15.

Type Data. Holotype, female (Paris Museum): Kenya, Hola (0°12'28"S 40°01'E), 10. V. 2000, (K. Werner leg.). Paratype, female (Robiche coll.): same data as holotype.

Distribution. KENYA: Hola (Robiche 2003).

Genus *Ametrocera* Fähræus, 1870

Ametrocera Fähræus 1870: 260 [feminine].

= *Idricus* Fairmaire 1888b: 199 [syn. by Péringuay 1904b: 296]. Type species: *Idricus diabolicus* Fairmaire, 1888; by monotypy.

Type Species. *Ametrocera aurita* Fähræus, 1870; subsequent designation by Lucas (1920: 88).

1. *Ametrocera aurita* Fähræus, 1870

Ametrocera aurita Fähræus 1870: 261.

= *Ametrocera turrita* Fähræus 1870: 261 [syn. by Gebien 1910b: 149].

= *Idricus pacificus* Péringuay 1892b: 122 [syn. by Péringuay 1904b: 296].

Type Data. *Ametrocera aurita* Fähræus, 1870, holotype, female (Naturhistoriska riksmuseet): no data.

Ametrocera turrita Fähræus, 1870, holotype, male (Naturhistoriska riksmuseet): no data.

Idricus pacificus Péringuay, 1892, holotype, unsexed (Cape Museum): “Bechuanaland”.

Distribution. SOUTH AFRICA: Bechuanaland* (Fähræus 1870; Péringuay 1892b).

2. *Ametrocera curlettii* Ferrer, 1996

Ametrocera curlettii Ferrer 1996: 95.

Type Data. Holotype, male (Carmagnola Museum): “Meru Dist.: Materi Mitunguu, 5.XI.1988, Dr. D. Dianasso leg.”.

Distribution. KENYA: Mitunguu (Ferrer 1996).

3. *Ametrocera diabolica* (Fairmaire, 1888)

Idricus diabolicus Fairmaire 1888b: 199.

Ametrocera diabolica: Gebien 1910b: 149.

Type Data. Holotype, not specified (Paris Museum): “Potchefstroom, Transvaal”.

Distribution. SOUTH AFRICA: Potchefstroom (Fairmaire 1888b).

4. *Ametrocera tribulus* (Fairmaire, 1894)

Idricus tribulus Fairmaire 1894b: 658.

Ametrocera tribulus: Gebien 1910b: 149.

Type Data. Holotype, not specified (Paris Museum): “Lagoa”.

Distribution. Lagoa* (Fairmaire 1894b).

Genus *Anaxius* Fähræus, 1870

Anaxius Fähræus 1870: 307 [masculine].

Type Species. *Anaxius obesus* Fähræus, 1870; by monotypy.

1. *Anaxius bloubergensis* Kamiński and Schoeman, 2018

Anaxius bloubergensis Kamiński and Schoeman 2018: 473.

Type Data. Holotype, male (Ditsong Museum): “S. Afr.; Limpopo Prov./Blouberg Nat. Res./22.59 S–29.08 E”, “25–27.11.2016E-Y:3987/day, sandy bushveld/leg. Ruth Müller”. Paratypes, 3 males (Ditsong Museum): same data as holotype; male

(Ditsong Museum) and male (MIZ PAN): “S. Afr.; Limpopo Prov. Blouberg NR North/dd22 98S, dd29.12E”, “6.12.2012; 866m, BLN1/Roodeberg Bushveld/leg. Colin Schoeman”; male (Warsaw Museum): “Blouberg NR/Rodeberg Bushveld/-22.98; 29.12”, “leg. Colin Schoeman/BLN7c/I”.

Distribution. SOUTH AFRICA: Blouberg Nature Reserve (Kamiński and Schoeman 2018).

2. *Anaxius campbellae* Koch, 1958

Anaxius campbellae Koch 1958: 213.

= *Anaxius montiscaerulei* Koch 1958: 214 [syn. by Kamiński and Schoeman 2018: 475].

Type Data. *Anaxius campbellae* Koch, 1958, **holotype**, male (Ditsong Museum): “Pienaar River, 1898./v. Jutrzenka”, “182”, “HOLOTYPE/Anaxius/CAMPBELLAE”. **Paratype**, male (Ditsong Museum): “Plat River.I, II; 1903/ (Jutrzenka)”.

Anaxius montiscaerulei Koch, 1958, **holotype**, male (Ditsong Museum): “BLOUBERG, Tvl./Leipzig Miss. Stat./3–5.I.1955/Transv. Mus. Exp.”.

Distribution. SOUTH AFRICA: Blouberg, Blouberg Nature Reserve, Gondeni, Pienaar River, Plat River, Pretoria; ZIMBABWE: Bulawayo (Kamiński and Schoeman 2018; Koch 1958).

3. *Anaxius limpopoensis* Kamiński and Schoeman, 2018

Anaxius limpopoensis Kamiński and Schoeman 2018: 477.

Type Data. Holotype, male (Ditsong Museum): “S. Afr.; Limpopo Prov./Lindani Nat. Res 1336m/24.02 S–28.23 E”, “8.12.2005; E-Y:3687/ single, bushveld/leg. Gusmann, Müller”. **Paratypes**, male (Ditsong Museum) and male (Warsaw Museum): same data as holotype; 2 males (Ditsong Museum): “S. Afr. Limpopo Prov./Mabote farm/24.07S 28.39 E”, “14.11.2009/Leg. Ruth Müller”; male (Ditsong Museum): “S. Afr.; Limpopo Prov./25 km N Mookgophong/24.25 S–28.41 E”, “12–15.1.2014 E-Y:3951/mixed woodland 1185m/leg. Ruth Müller”; male (Ditsong Museum): “S. Afr.; Limpopo Prov./Waterberg Game Res./24.11 S–28.20 E”, “4–7.11.2002/leg. B. Dombrowsky”.

Distribution. SOUTH AFRICA: Lindani Nature Reserve, Mabote farm, surroundings of Mookgophong, Waterberg Game Reserve (Kamiński and Schoeman 2018).

4. *Anaxius meletsensis* Kamiński and Schoeman, 2018

Anaxius meletsensis Kamiński and Schoeman 2018: 478.

Type Data. Holotype, male (Ditsong Museum): “S. Afr.; Limpopo Prov./Meletse Reserve 1003m/24.36

S–27.39 E”, “27.11.2014; E-Y:3954/on ground, bushveld/leg. Ruth Müller”. **Paratypes**, 2 males and female (Ditsong Museum) and male (Warsaw Museum): same data as holotype.

Distribution. SOUTH AFRICA: Geelhoutbush farm, Meletse Reserve (Kamiński and Schoeman 2018).

5. *Anaxius obesus* Fåhraeus, 1870

Anaxius obesus Fåhraeus 1870: 307.

Type Data. Lectotype, designated by Kamiński and Schoeman (2018), male (Ditsong Museum): “Caffra-ria”. **Paralectotype**, male (Ditsong Museum): same data as holotype.

Distribution. SOUTH AFRICA: Amathole Municipality (Kamiński and Schoeman 2018).

6. *Anaxius prozeskyi* Koch, 1958

Anaxius prozeskyi Koch 1958: 209.

Type Data. Holotype, male (Ditsong Museum): “Anaxius/magnificus/Koch/C. Koch det. 195”, “Blouberg, Tvl./N. side, Glenfernness/16–21.I.1955/Transva. Mus. Exp.”, “HOLOTYPE/Anaxius/PROZESKYI”. **Paratype**, male (Ditsong Museum): “Helopininae/anaxius/?” , “186”, “3811”, “3427”, “Makgaberg/- 2.03./2457”, “PARATYPE/Anaxius/PROZESKYI”.

Distribution. NAMIBIA: Okahandja Dis.; SOUTH AFRICA: Blouberg, Blouberg Nature Reserve, Makgabeng (Kamiński and Schoeman 2018).

7. *Anaxius pseudoloensis* Kamiński and Schoeman, 2018

Anaxius pseudoloensis Kamiński and Schoeman 2018: 481.

Type Data. Holotype, male (Ditsong Museum): “Espungabera/80.K.Jutha 1954”.

Distribution. MOZAMBIQUE: Espungabera (Kamiński and Schoeman 2018).

Genus *Aptila* Fåhraeus, 1870

Aptila Fåhraeus 1870: 258 [feminine].

Type Species. *Aptila costata* Fåhraeus, 1870; subsequent designation by Lucas (1920: 115).

1. *Aptila costata* Fåhraeus, 1870

Aptila costata Fåhraeus 1870: 259.

= *Aptila tuberculata* Fåhraeus 1870: 258 [syn. by Haag-Rutenberg 1871: 32].

= *Micrantereus litigiosus* Péringuey 1899: 308 [syn. by Gebien 1943: 905].

Type Data. *Aptila costata* Fåhraeus, 1870, **holotype**, unsexed (Naturhistoriska riksmuseet): no data.

Aptila tuberculata Fåhraeus, 1870, **holotype**, female (Naturhistoriska riksmuseet): no data.

Micrantereus litigiosus Péringuey, 1899, **holotype**, unsexed (Cape Museum): “Bechuanaland (Ramotsa). Miss M. Good”.

Distribution. BOTSWANA: Ramotsa (Péringuey 1899).

2. *Aptila debilis* Fähræus, 1870

Aptila debilis Fähræus 1870: 260.

= *Aptila noxia* Fähræus 1870: 259 [syn. by Haag-Rutenberg 1871: 32].

= *Micrantereus parvulus* Péringuey 1899: 307 [syn. by Gebien 1943: 905].

Type Data. *Aptila debilis* Fähræus, 1870, **holotype**, unsexed (Naturhistoriska riksmuseet): no data.

Aptila noxia Fähræus, 1870, **holotype**, female (Naturhistoriska riksmuseet): no data.

Micrantereus parvulus Péringuey, 1899, **holotype**, male (Cape Museum): “Transvaal”.

Distribution. SOUTH AFRICA: Transvaal* (Péringuey 1899); NAMIBIA: Neitsas (Gebien 1920).

3. *Aptila micranteroides* Koch, 1958

Aptila micranteroides Koch 1958: 204.

Type Data. Holotype, male (Ditsong Museum): “Southern Rhodesia, Mashonaland: Lower Sabi River, D. Townley”. **Paratype**: same data as holotype.

Distribution. ZIMBABWE: Mashonaland (Lower Sabi River) (Koch 1958).

4. *Aptila tropicalis* Koch, 1958

Aptila tropicalis Koch 1958: 207.

Type Data. Holotype, male (Tervuren Museum): “Southern Tanganyika Territory: Lindi, III.1903, ex. coll. Staudinger”. **Paratypes**, two females: same data as holotype.

Distribution. TANZANIA: Lindi (Koch 1958).

Genus *Asidodema* Koch, 1958

Asidodema Koch 1958: 139 [feminine].

Type Species. *Oncosoma alternicostis* Gebien, 1910; by original designation.

1. *Asidodema alternicostis* (Gebien, 1910)

Oncosoma alternicostis Gebien 1910a: 144.
Asidodema alternicostis: Koch 1958: 139.

Type Data. Syntypes, male and female (Basel Museum): “Wembere-Steppe”.

Distribution. TANZANIA: Wembere-Steppe (Gebien 1910a).

2. *Asidodema collaris* (Gebien, 1910)

Oncosoma collare Gebien 1910a: 145.
Asidodema collaris: Koch 1958: 139.

Type Data. Holotype (Basel Museum): “Wugiri, Deutsch-Ostafrika”.

Distribution. TANZANIA: Wugiri (Gebien 1910a).

3. *Asidodema suturalis* (Gebien, 1910)

Oncosoma suturale Gebien 1910a: 146.

Asidodema suturalis: Koch 1958: 139.

Type Data. Syntypes, male and female (Basel Museum): “Ukerewe”.

Distribution. TAZNANIA: Ukerewe (Gebien 1910a).

Genus *Blastarnodes* Koch, 1958

Blastarnodes Koch 1958: 154 [masculine].

Type Species. *Blastarnodes herero* Koch, 1958; by original designation.

1. *Blastarnodes carpi* Koch, 1958 (Fig. 7B)

Blastarnodes carpi Koch 1958: 202.

Type Data. Syntypes, nine specimens (Ditsong Museum): “Kaokoveld: Kaoko Otavi, VII.1951, B. Carp Exped.”.

Distribution. NAMIBIA: Kaoko Otavi, Oropembe (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 7B), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 7B should still be considered a syntype.

2. *Blastarnodes gebieni borgesii* Koch, 1958 (Fig. 7C)

Blastarnodes gebieni borgesii Koch 1958: 201.

Type Data. Syntypes, 15 specimens (Ditsong Museum): “South-western Angola: E. of Humpata, VI.1954, Vernay-Transv. Mus. Exped.”.

Distribution. ANGOLA: surroundings of Humpata (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 7C), this designation was never formalized in publication. Therefore, the specimen presented in Fig. 7C should still be considered a syntype.

3. *Blastarnodes gebieni gebieni* (Kulzer, 1951) (Fig. 7D)

Nicandra gebieni Kulzer 1951: 571.

Blastarnodes gebieni gebieni: Koch 1958: 154.

Type Data. Holotype, female (Basel Museum): “Benguela, leg. Dr. Wellmann”.

Distribution. ANGOLA: Benguela (Kulzer 1951).

4. *Blastarnodes herero* Koch, 1958 (Fig. 7A)

Blastarnodes herero Koch 1958: 201.

Type Data. Syntypes, 14 specimens (Ditsong Museum), single specimen (Budapest Museum): “Kaokoveld; Ohopoho, VII.1951, H. Hall”.

Distribution. NAMIBIA: Kaoko Otavi, Opuwo, Oropembe (Koch 1958).

5. *Blastarnodes orientalis* (Gebien, 1937)

Helopinus orientalis Gebien 1937: 54.

Blastarnodes orientalis: Koch 1958: 154.

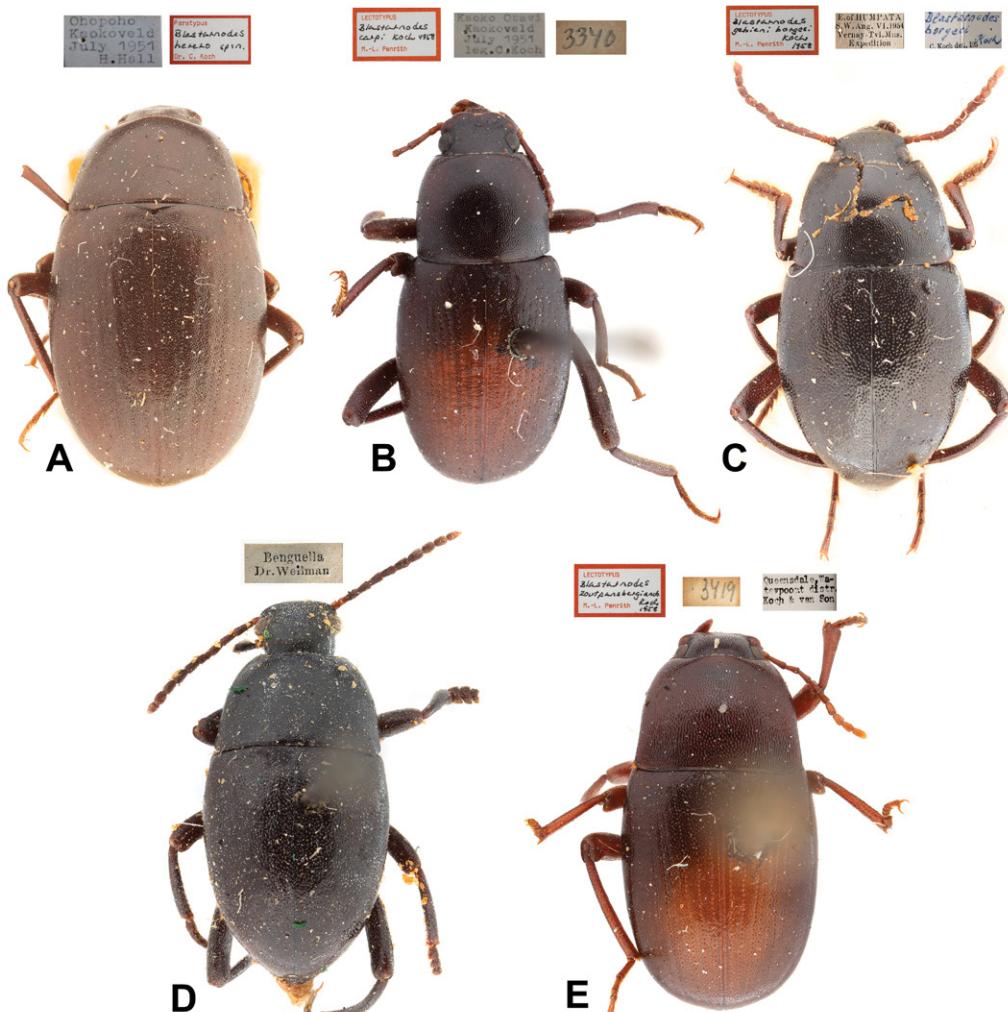


Fig. 7. Type specimens of species representing the genus *Blastarnodes*. A) *B. herero*, B) *B. carpi*, C) *B. gebieni borgesii*, D) *B. gebieni gebieni*, E) *B. zoutpansbergianus*.

Type Data. Syntypes, two males and one female (Basel Museum, Trieste Museum): “Ugogo: Dodoma & N.W. Usagara”.

Distribution. TANZANIA: Dodoma, Usagara (Gebien 1937).

6. *Blastarnodes zoutpansbergianus* Koch, 1958 (Fig. 7E)

Blastarnodes zoutpansbergianus Koch 1958: 203.

Type Data. Syntypes, three specimens (Ditsong Museum): “Queensdale, Zoutpansberg District, VIII.1949, C. Koch & G. van Son”.

Distribution. SOUTH AFRICA: Queensdale (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 7E), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 7E should still be considered a syntype.

Genus *Diestecopus* Solier, 1848

Diestecopus Solier 1848: 194 [masculine].
= *Blastarnus* Fairmaire 1897: 132 [syn. by Koch 1958: 153]. Type species: *Blastarnus grallator* Fairmaire, 1897; subsequent designation by Koch (1958: 153).

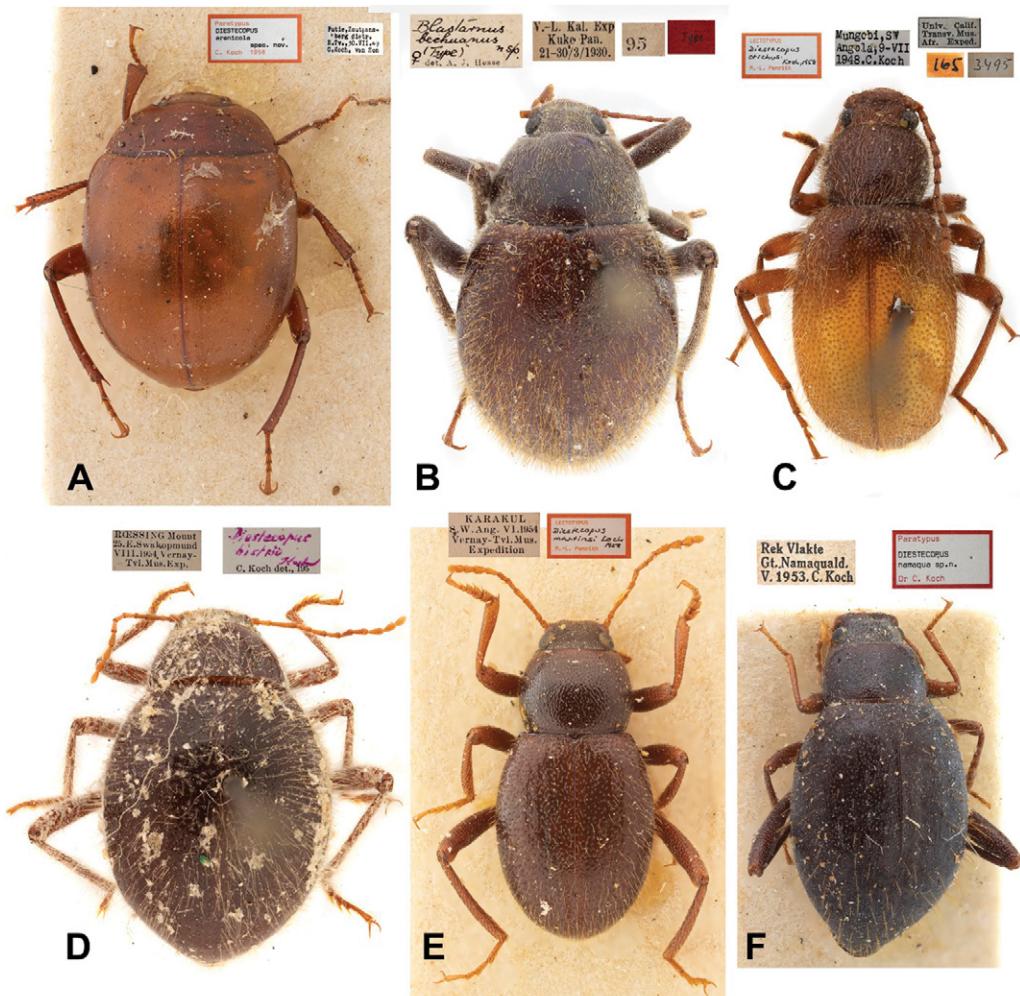


Fig. 8. Type specimens of species representing the genus *Diestecopus*. A) *D. arenicola*, B) *D. bechuanus*, C) *D. citicus*, D) *D. histrio*, E) *D. martinsi*, F) *D. namaqua*.

Type Species. *Diestecopus erodiooides* Solier, 1848; by monotypy.

1. *Diestecopus arenicola* Koch, 1958 (Fig. 8A)
Diestecopus arenicola Koch 1958: 195.

Type Data. Syntypes. 25 specimens (Ditsong Museum), single specimen (Budapest Museum); “Northern Transvaal: Futie, Zoutpansberg District, VII.1950, C. Koch & G. van Son”.

Distribution. SOUTHAFRICA: Futie (Koch 1958).

2. *Diestecopus bechuanus* (Hesse, 1935) (Fig. 8B)
Blastarnus bechuanus Hesse 1935: 567.
Diestecopus bechuanus: Koch 1958: 153.

Type Data. Holotype, female (Ditsong Museum); “Kuke Pan, 21-30.iii.30”. **Paratypes:** male: same data as holotype; female (Ditsong Museum); “Kaotwe, 8-12.iv.30”; male (Ditsong Museum); “Kopjes, Mabeapudi, 7-9.iv.30”.

Distribution. BOTSWANA: Kaotwe, Kuke Pan, Mabeapudi (Hesse 1935).

3. *Diestecopus conspersus* (Müller, 1887)
Drosochrus conspersus Müller 1887: 304.
Diestecopus conspersus: Koch 1958: 153.

Type Data. Syntypes, not specified (Berlin Museum); no data.

Distribution. No data.

4. *Diestecopus ctichus* Koch, 1958 (Fig. 8C)

Diestecopus ctichus Koch 1958: 185.

Type Data. Syntypes, six specimens (Ditsong Museum): “Angola: Mungobi, VII.1948, Univ. California-Transv. Mus. Exped.”.

Distribution. ANGOLA: Mungobi; NAMIBIA: Oshikango, Kaoko Otavi (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 8C), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 8C should still be considered a syntype.

5. *Diestecopus erodioides* Solier, 1848

Diestecopus erodioides Solier 1848: 196.

Type Data. Holotype, unsexed (Torino Museum): “Cap de Bonne Espérance”.

Distribution. SOUTH AFRICA: Cap de Bonne Espérance* (Solier 1848).

6. *Diestecopus gracilipes* Koch, 1958

Diestecopus gracilipes Koch 1958: 193.

Type Data. Holotype, male (Ditsong Museum): “Western Damaraland: Rossing Mountains, VIII.1954, Vernay-Transv. Ms. Exped.”.

Distribution. NAMIBIA: Rossing Mountains (Koch 1958).

7. *Diestecopus grallator* (Fairmaire, 1897)

Blastarnus grallator Fairmaire 1897: 132.

Diestecopus grallator: Koch 1958: 153.

Type Data. Syntypes, not specified (Paris Museum): “Kimberley (Transvaal.”).

Distribution. SOUTH AFRICA: Kimberley (Fairmaire 1897).

8. *Diestecopus gynandromorphus* (Gebien, 1920)

Blastarnus gynandromorphus Gebien 1920: 159.

Diestecopus gynandromorphus: Koch 1958: 153.

Type Data. Syntypes (Basel Museum): “Kuibis 15.VII.1911 (Michaelsen)”.

Distribution. NAMIBIA: Kuibis (Gebien 1920).

9. *Diestecopus hirtulus* (Fairmaire, 1897)

Blastarnus hirtulus Fairmaire 1897: 133.

Diestecopus hirtulus: Koch 1958: 153.

Type Data. Holotype, not specified (Paris Museum): “Kuisip”.

Distribution. SOUTH AFRICA: Kuisip* (Fairmaire 1897).

10. *Diestecopus histrio* Koch, 1958 (Fig. 8D)

Diestecopus histrio Koch 1958: 189.

Type Data. Syntypes, two males and female (Ditsong Museum): “Western Damaraland: Rossing Mountains, VIII.1954, Vernay-Transv. Ms. Exped.”.

Distribution. NAMIBIA: Rossing Mountains, Swakopmund (Koch 1958).

11. *Diestecopus laminiger* (Gebien, 1920)

Blastarnus laminiger Gebien 1920: 158.

Diestecopus laminiger: Koch 1958: 153.

Type Data. Holotype, male (Basel Museum): “Otavifontein, 5 km ostlich Otavi 6. VI. 1911 (Michaelsen)”.

Distribution. NAMIBIA: Otavifontein (Gebien 1920).

12. *Diestecopus latipalpis* (Kulzer, 1951)

Nicandra latipalpis Kulzer 1951: 567.

Diestecopus latipalpis: Koch 1958: 153.

Type Data. Syntypes, 2 males and 2 females (Basel Museum): “D.S.W. Afr. Oranje”.

Distribution. SOUTH AFRICA: Orange State* (Kulzer 1951).

13. *Diestecopus longulus* (Gebien, 1920)

Blastarnus longulus Gebien 1920: 158.

Diestecopus longulus: Koch 1958: 153.

Type Data. Holotype, male (Basel Museum): “Usakos, 22. IV. —22. VI. 1911 (Michaelsen)”.

Distribution. NAMIBIA: Usakos (Gebien 1920).

14. *Diestecopus marginicollis* (Gebien, 1920)

Blastarnus marginicollis Gebien 1920: 156.

Diestecopus marginicollis: Koch 1958: 153.

Type Data. Syntypes, male (Basel Museum): “Karibib, 23. —26. IV. 1911 (Michaelsen)”; female (Basel Museum): “Brakwater, 20 km nördlich Windhuk 23. V. 1911 (Michaelsen)”.

Distribution. NAMIBIA: Karibib, surroundings of Windhoek (Gebien 1920).

15. *Diestecopus martinsi* Koch, 1958 (Fig. 8E)

Diestecopus martinsi Koch 1958: 188.

Type Data. Syntypes, eight specimens (Ditsong Museum): “Angola: Karakul, VI.1954, Vernay-Transv. Mus. Exped.”.

Distribution. ANGOLA: Pediba, Porto Alexandre, Mocamedes (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 8E), this designation was never formalized in publication. Therefore, the specimen presented in Fig. 8E should still be considered a syntype.

16. *Diestecopus namaqua* Koch, 1958 (Fig. 8F)

Diestecopus namaqua Koch 1958: 192.

Type Data. Syntypes, eight specimens (Ditsong Museum): “South-western Great Namaqualand: Rek Vlakte, V.1953, C. Koch & W. Graaf”.

Distribution. SOUTH AFRICA: Richtersveld, Witputs; NAMIBIA: Rek Vlakte (Koch 1958).

17. *Diestecopus nigrostriatus* Koch, 1958 (Fig. 9)

Diestecopus nigrostriatus Koch 1958: 191.

Type Data. Syntypes, six specimens (Ditsong Museum): “North-eastern Cape Province: Kuruman, V.1948, Univ.California-Transv. Mus. Exped.”.

Kuruman, Kal.
V-48. C.Koch



Fig. 9. Syntype specimen of *Diestecopus nigrostriatus*.

Distribution. SOUTH AFRICA: Kuruman, Niekerk's Hope, Vryburg (Koch 1958).

18. *Diestecopus nudus* Koch, 1958

Diestecopus nudus Koch 1958: 187.

Type Data. Syntypes, male and nine females (Ditsong Museum): "Kaokoveld: Swartboois Drift, VII.1951, B. Carp Exped.".

Distribution. ANGOLA: Ehomba Mountains, Swartbooisdrift (Koch 1958).

19. *Diestecopus ovalis* (Kulzer, 1951)

Nicandra ovalis Kulzer 1951: 570.

Diestecopus ovalis: Koch 1958: 153.

Type Data. Holotype, male (Basel Museum): "Ontjo, D.S. W. Afr.".

Distribution. NAMIBIA: Ontjo, Windhoek (Kulzer 1951).

20. *Diestecopus physopterus* (Gebien, 1920)

Blastarnus physopterus Gebien 1920: 159.

Diestecopus physopterus: Koch 1958: 153.

Type Data. Holotype, female (Basel Museum): "Okahandja VI. 1901".

Distribution. NAMIBIA: Okahandja (Gebien 1920).

21. *Diestecopus piliger* (Gebien, 1920: 153)

Blastarnus piliger Gebien 1920: 153.

Diestecopus piliger: Koch 1958: 153.

Type Data. Holotype, female (Basel Museum): "Brit. Südwestafrika: Kl. Xamaland, Kamaggas".

Distribution. SOUTH AFRICA: Kamaggas (Gebien 1920).

22. *Diestecopus pruinosis* (Fairmaire, 1897)

Blastarnus pruinosis Fairmaire 1897: 132.

Diestecopus pruinosis: Koch 1958: 153.

Type Data. Holotype, unsexed (Paris Museum): "Lac N'gami".

Distribution. BOTSWANA: Ngami (Fairmaire 1897).

23. *Diestecopus subpellucens* (Gebien, 1920)

Blastarnus subpellucens Gebien 1920: 154.

Diestecopus subpellucens: Koch 1958: 153.

Type Data. Syntypes, two males and one female (Basel Museum): "Deutsch- Südwestafrika"; female (Basel Museum): "Kuibis 15. VII. 1911 (Michaelsen)".

Distribution. NAMIBIA: Kuibis (Gebien 1920).

24. *Diestecopus subplanatus* (Gebien, 1920)

Blastarnus subplanatus Gebien 1920: 151.

Diestecopus subplanatus: Koch 1958: 153.

Type Data. Syntypes, two males (Basel Museum): "Tsumeb 13.—19. VI. 1911 (Michaelsen)".

Distribution. NAMIBIA: Tsumeb (Gebien 1920).

25. *Diestecopus subseriatus* (Gebien, 1920)

Blastarnus subseriatus Gebien 1920: 155.

Diestecopus subseriatus: Koch 1958: 153.

Type Data. Syntypes, two males and three females (Basel Museum, Dahlem Museum): "Windhoek" and "Cap Croß".

Distribution. NAMIBIA: Cape Cross, Windhoek (Gebien 1920).

26. *Diestecopus tibidens* Koch, 1958

Diestecopus tibidens Koch 1958: 194.

Type Data. Holotype, male (Ditsong Museum): "Bushmanland: Nabies, 30m. E of Kakamas, XII.1948, Univ. California-Transv. Mus. Exped.".

Paratype: same data as holotype.

Distribution. SOUTH AFRICA: Nabies (Koch 1958).

Genus *Drosochrus* Erichson, 1843

Subgenus *Desertosochrus* Koch, 1958

Desertosochrus Koch 1958: 149 [masculine].

Type Species. *Drosochrus piligaster* Koch, 1958; by original designation.

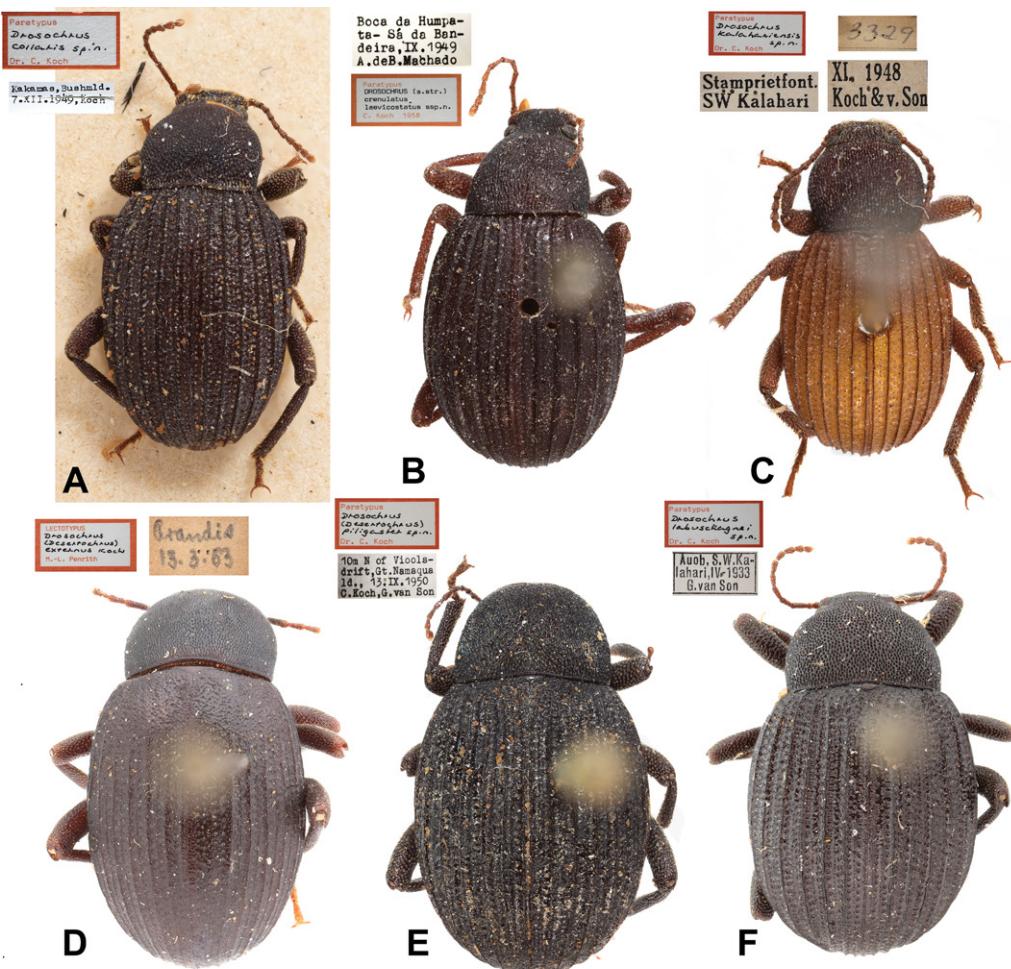


Fig. 10. Type specimens of species representing the genus *Drosochrus*. A–C) Subgenus *Drosochrus*: A) *D. collaris*, B) *D. crenulatus laevicostatus*, C) *D. kalahariensis*. D–F) Subgenus *Desertosochrus*: D) *D. externus*, E) *D. piligaster*, F) *D. labuschagnei*.

1. *Drosochrus externus* Koch, 1958 (Fig. 10D)
Drosochrus externus Koch 1958: 162.

Type Data. Syntypes, five specimens (Ditsong Museum): “Damaland: Arandis, III.1953, F. Gaerdes”.

Distribution. NAMIBIA: Arandis (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 10D), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 10D should still be considered a syntype.

2. *Drosochrus labuschagnei* Koch, 1958 (Fig. 10F)
Drosochrus labuschagnei Koch 1958: 161.

Type Data. Syntypes, six specimens (Ditsong Museum): “Aoub River, IV.1933, G. van SON”.

Distribution. NAMIBIA: Aoub River* (Koch 1958); SOUTH AFRICA: Inkbosch Pan (Koch 1958).

3. *Drosochrus piligaster* Koch, 1958 (Fig. 10E)
Drosochrus piligaster Koch 1958: 161.

Type Data. Syntypes, six specimens (Ditsong Museum): “10 m. N. of Vioolsdrift, IX.1950, C. Koch & G. van Son”.

Distribution. SOUTH AFRICA: surroundings of Vioolsdrift (Koch 1958).

Subgenus *Drosochrus* Erichson, 1843

Drosochrus Erichson 1843: 243 [masculine].

Type Species. *Drosochrus brunnipes* Erichson, 1843; subsequent designation by Gebien (1943: 910).

4. *Drosochrus brunnipes* Erichson, 1843
Drosochrus brunnipes Erichson 1843: 244.

Type Data. Holotype, unsexed (Humboldt University): “Cap. Von Hrn. Buquet”.

Distribution. No data.

5. *Drosochrus collaris* Koch, 1958 (Fig. 10A)

Drosochrus collaris Koch 1958: 158.

Type Data. Syntypes, four specimens (Ditsong Museum): “Pofadder, VIII.1950. C. Koch & G. van Son”.

Distribution. SOUTH AFRICA: Between Kenhardt and Onderstedoorn, Kakamas and its surroundings, Klaver, Nabies, Pofadder, Springbok, Upington (Koch 1958).

6. *Drosochrus crenulatus crenulatus* Erichson, 1843

Drosochrus crenulatus Erichson 1843: 244.

Type Data. Holotype, unsexed (Humboldt University): “Angola”.

Distribution. ANGOLA* (Erichson 1843).

7. *Drosochrus crenulatus laevicostatus* Koch, 1958 (Fig. 10B)

Drosochrus crenulatus laevicostatus Koch 1958: 157.

Type Data. Syntypes, 18 specimens (Dundo Museum): “Angola: Boca da Humpata - Sá de Bandeira, IX.1949, A. de Barros Machado”.

Distribution. ANGOLA: Bandeira (Koch 1958).

8. *Drosochrus kalahariensis* Koch, 1958 (Fig. 10C)

Drosochrus kalahariensis Koch 1958: 160.

Type Data. Syntypes, seven specimens (Ditsong Museum): “Stamprietfontein. IX.1951. C. Koch & G. van Son”.

Distribution. NAMIBIA: Neuhof-Kowas, Stampriet, surroundings of Stampriet (Koch 1958).

9. *Drosochrus tristis* (Fåhraeus, 1870)

Emyon tristis Fåhraeus 1870: 305.

Drosochrus tristis: Koch 1958: 148.

Type Data. Syntypes, not specified (Naturhistoriska riksmuseet).

Distribution. SOUTH AFRICA: Bechuanaland* (Péringuéy 1892b).

Subgenus *Helopinus* Solier, 1848

Helopinus Solier 1848: 152 [masculine].

= *Pteraulus* Solier 1848: 200 [syn. by Koch 1958: 149]. Type species: *Pteraulus sulcatipennis* Solier, 1848; see Bouchard et al. (2021: 327).
= *Emyon* Gerstaecker 1854: 532 [syn. by Koch 1958: 149]. Type species: *Emyon caelatus* Gerstaecker, 1854; by monotypy.

Type Species. *Helopinus costatus* Solier, 1848; by monotypy.

10. *Drosochrus abyssinicus* (Gebien, 1937)

Helopinus abyssinicus Gebien 1937: 51.

Drosochrus abyssinicus: Koch 1958: 150.

Type Data. Syntypes, two specimens (Basel Museum): “Dire Daona”.

Distribution. ETHIOPIA: Dire Daona (Gebien 1937).

11. *Drosochrus acanthocnemis* (Gebien, 1937)

Helopinus acanthocnemis Gebien 1937: 53.

Drosochrus acanthocnemis: Koch 1958: 150.

Type Data. Holotype, male (Berlin Museum): “Somalia: Handodu: Karo Lola”.

Distribution. KENYA: Karo-Lola (Gebien 1937).

12. *Drosochrus caelatus* (Gerstaecker, 1854)

Emyon caelatus Gerstaecker 1854: 532.

Drosochrus caelatus: Koch 1958: 150.

Type Data. Lectotype, designated by Ferrer (1999), unsexed (Berlin Museum): “15722”, “Tete, Peters”, “Emyon caelatus Gerstaecker”, “Hist. Coll. Coleoptera n° 15722 1 ex. Emyon caelatus Gerstaecker, Tete, Peters. Zool. Mus. Berlin”.

Paraleotypes, two specimens (Berlin Museum): same data as lectotype.

Distribution. MOZAMBIQUE: Boroma (Ferrer 1999); ZAMBIA: Zambesi (Ferrer 1999).

13. *Drosochrus costatus aegyptiacus* (Gridelli, 1939) (Fig. 11A)

Helopinus costatus aegyptiacus Gridelli 1939a: 198.

Drosochrus costatus aegyptiacus: Koch 1958: 150.

Type Data. Holotype, male (Basel Museum):

“Cairo”. **Paratypes**, female (Basel Museum): “Tourah, W. Dougla”; male (Basel Museum): “di Wadi Digla (Cairo)”; female (Basel Museum): “Cairo”.

Distribution. EGYPT: Cairo, Ismailia, Tabah Valley, Qena Valley (Gridelli 1939a).

14. *Drosochrus costatus costatus* (Solier, 1848)

Helopinus costatus Solier 1848: 199.

Drosochrus costatus costatus: Koch 1958: 150.

= *Helopinus misolampoides* Lacordaire 1876: 20 and pl. 58 [syn. by Gebien 1943: 909].

Type Data. *Helopinus costatus* Solier, 1848, **syntypes**, not specified (Torino Museum – Spinola coll.): “De l’Arabie”.

Helopinus misolampoides Lacordaire, 1876, **syntypes**, not specified (Paris Museum): “De Breme Arabie”.

Distribution. “Arabia”* (Solier 1848).

Note. Lacordaire (1876) did not provide a description for *Helopinus misolampoides*, but the name was made available in the explanation of plates on page 20.

15. *Drosochrus costatus elegans* (Baudi, 1881)

Helopinus elegans Baudi 1881: 287.

Drosochrus costatus elegans: Koch 1958: 150.

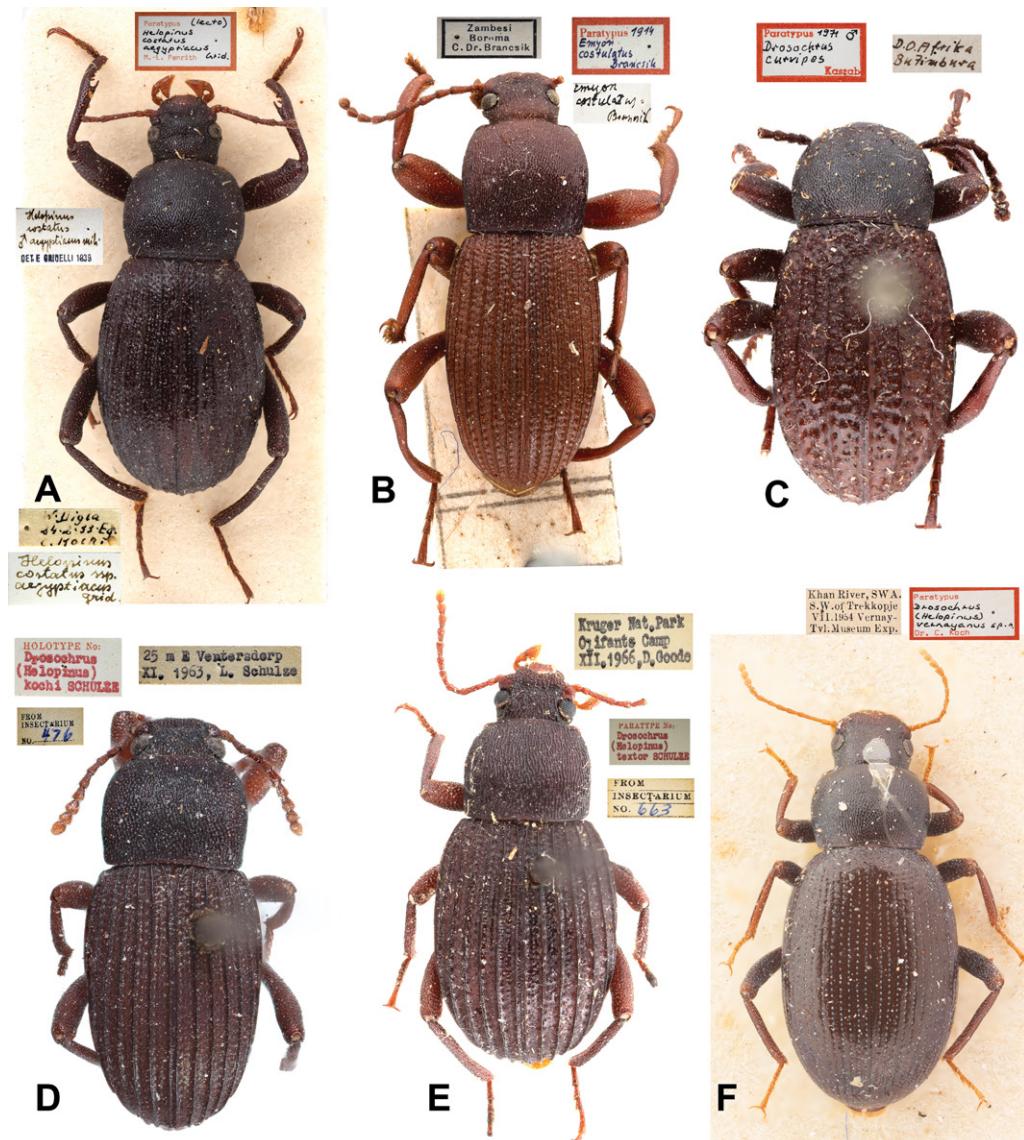


Fig. 11. Type specimens of species representing the genus *Drosochrus* subgenus *Helopinus*. A) *D. costatus aegyptiacus*, B) *D. costulatus*, C) *D. curvipes*, D) *D. kochi*, E) *D. textor*, F) *D. vernayanus*.

Type Data. Syntypes, not specified (Paris Museum): “Ad Maris Arabici abyssinica litora prope Assab D. Doria”.

Distribution. DJIBOUTI: 30 km from lake Assal (Ardoïn 1979); ERITREA: Assab, Dahlak Kebir Island, Emberemi, Massawa, Otumlo, Ras Ghedem, Saati (Gridelli 1939a); DJIBOUTI: Obock (Gridelli 1939a).

16. *Drosochrus costulatus* (Brancsik, 1914), new combination (Fig. 11B)

Emyon costulatus Brancsik 1914: 65.

Type Data. Syntypes, not specified (Budapest Museum): “Zambesi, Boroma, C. Dr. Brancsik”.

Distribution. MOZAMBIQUE: Boroma (Brancsik 1914).

Note. Koch did not discuss this species in his 1958 paper, which includes a revision of subgeneric concepts for *Drosochrus*, but examination of types clearly places it within the subgenus *Helopinus*.

17. *Drosochrus cristatus* (Solier, 1848)

Pteraulus cristatus Solier 1848: 201.

Drosochrus cristatus: Gebien 1911: 566.

Type Data. Holotype, unsexed (Torino Museum): “Cap de Bonne Espérance”.

Distribution. SOUTH AFRICA: “Cap de Bonne Espérance”* (Solier 1848).

Note. Gebien (1911) originally synonymized *Pteraulus* with *Drosochrus*. Koch (1958) in establishing a subgeneric classification for *Drosochrus*, synonymized *Pteraulus* with the subgenus *Helopinus*.

18. *Drosochrus curlettii* Ferrer, 1999

Drosochrus curlettii Ferrer 1999: 59.

Type Data. Holotype, male (Carmagnola Museum): “Tanzania, Iringa, Mafinga, 10-15.I.1994, G. Curletti leg.”. **Paratypes** (Carmagnola Museum), two females: same data as holotype; three males and three females: “Mafinga, 10-20.XII.1997, G. Curletti leg.”.

Distribution. TANZANIA: Iringa, Mafinga (Ferrer 1999).

19. *Drosochrus curvipes* Kaszab, 1971 (Fig. 11C)

Drosochrus curvipes Kaszab 1971: 237.

Type Data. Holotype, male (Budapest Museum): “Tanganyika: Katesh, S of Mt. Hanang, 5900 feet, 26. VI.—1. VII 1965, leg. Dr. J. SZUNYOGHY”.

Paratype, female (Budapest Museum): “Butumbura”.

Distribution. BURUNDI: Bujumbura (Kaszab 1971); TANZANIA: Mountain Hanang (Kaszab 1971).

Note. Kaszab did not directly provide the subgeneric affiliation of this species. However, according to his diagnosis this species is closely related to *D. caelatus*. This statement was used here to assign *D. curvipes* to the subgenus *Helopinus*.

20. *Drosochrus desolatus* Koch, 1958

Drosochrus desolatus Koch 1958: 164.

Type Data. Holotype, male (Ditsong Museum): “Khan River, SW. of Trekkopje. VII.1954. Vernay Transv. Mus. Exped.”.

Distribution. NAMIBIA: Surroundings of Trekkopje (Koch 1958).

21. *Drosochrus drumonti* Robiche, 2010

Drosochrus drumonti Robiche 2010: 371.

Type Data. Holotype, male (Brussels Museum): “R. D. Congo, Jadotville: Numbi, V.1957, R. P. Th. De Carters leg.”. **Paratype**, male (Robiche coll.): “R. D. Congo, Katanga, Lukafu, 18.X-15.XI.2003, T. Bouyer leg.”.

Distribution. DEMOCRATIC REPUBLIC OF THE CONGO: Numbi, Lukafu (Robiche 2010).

22. *Drosochrus duvivieri* (Gebien, 1921)

Emyon duvivieri Gebien 1921: 123.

Drosochrus duvivieri: Koch 1958: 150.

Type Data. Syntypes, 13 syntypes (Basel Museum): “Moliro, Tanganyika III.-V. 1895 (Duvivier)”.

Distribution. DEMOCRATIC REPUBLIC OF THE CONGO: Moliro (Gebien 1921).

23. *Drosochrus excavatus* Robiche, 2010

Drosochrus excavatus Robiche 2010: 367.

Type Data. Holotype, male (Paris Museum): “Mozambique, Prov. Tete, Chipembéré, 21-25. XII.2009, G. Robiche leg.”. Paratypes, female and two females (Paris Museum, Robiche coll.): same data as holotype; male (Paris Museum): “Zambèze, amont de Tambara, Alfiate (Garé), 1929, P. Lesne leg.”.

Distribution. MOZAMBIQUE: Chipembéré, Zambèze (Robiche 2010).

24. *Drosochrus exsculptus* (Gebien, 1920)

Emyon exsculptus Gebien 1920: 144.

Drosochrus exsculptus: Koch 1958: 150.

Type Data. Syntypes, male and female (Basel Museum): “Okahandja”; male: “Osona bei Okahandja VI.1911 (Michaelsen)”; two females: “Okahandja 27.—28.IV., 1911 (Michaelsen)”.

Distribution. NAMIBIA: Okahandja (Gebien 1920).

25. *Drosochrus girardi* Robiche, 2008

Drosochrus girardi Robiche 2008a: 392.

Type Data. Holotype, male (Paris Museum): “Mozambique, province de Tete, Chipembéré, 23.II.2008, Robiche G. leg.”. **Paratypes**, 44 specimens (Paris Museum and Robiche coll.): same data as holotype.

Distribution. MOZAMBIQUE: Chipembéré (Robiche 2008a).

26. *Drosochrus gridellii* Ferrer, 1995

Drosochrus gridellii Ferrer 1995: 51.

Type Data. Holotype, male (Florence Museum): “Gardo, Eil, 8.IV.1980”.

Distribution. SOMALIA: Eil (Ferrer 1995).

27. *Drosochrus kochi* Schulze, 1968 (Fig. 11D)

Drosochrus kochi Schulze 1968: 177.

Type Data. Holotype, male (Ditsong Museum): “25 miles East of Ventersdorp (Tvl.), 26°18'S, 27°05'E, November 1963, L. Schulze”. **Paratypes**, seven specimens: same data as holotype.

Distribution. SOUTH AFRICA: Ventersdorp (26°18'S, 27°05'E) (Schulze 1968).

28. *Drosochrus meruensis* (Gebien, 1910)

Helopinus meruensis Gebien 1910c: 388.

Drosochrus meruensis: Koch 1958: 150.

Type Data. Syntypes, five males and three females (Naturhistoriska riksmuseet): “Meru-Niederung 22.-23. Nov.” and “Ngare na Nyuki, Jan”.

Distribution. TANZANIA: Ngare Nanyuki (Gebien 1910c).

29. *Drosochrus minor* (Fairmaire, 1882)*Helopinus minor* Fairmaire 1882a: 77.*Drosochrus minor*: Koch 1958: 150.**Type Data. Holotype**, female (Paris Museum): no data.**Distribution.** SOMALIA* (Fairmaire 1882a).**30. *Drosochrus muellerae lesnei* Robiche, 2008***Drosochrus muellerae lesnei* Robiche 2008a: 392.**Type Data. Holotype**, male (Paris Museum): "Zambèze, Nova Choupenga, près Chemba, 1929, P. Lesne leg.". **Paratypes**, 23 specimens (Paris Museum and Robiche coll.): same data as holotype; three specimens (Paris Museum and Robiche coll.): "Zambèze, Nova Choupenga, 1928, J. Surcouf leg."; nine specimens (Paris Museum and Robiche coll.): "Zambèze, env. de Chemba, Inhacoro, 1928, P. Lesne leg."; five specimens (Paris Museum and Robiche coll.): "Zambèze, Tambara, 1929, P. Lesne leg.".**Distribution.** MOZAMBIQUE: Chemba, Inhassoro, Tambara (Robiche 2008a).**31. *Drosochrus muellerae muellerae* Robiche, 2008***Drosochrus muellerae muellerae* Robiche 2008a: 389.**Type Data. Holotype**, male (Paris Museum): "Mozambique, province de Manica, Vanduzi, 500 m, 04.I.2004, Robiche G. & Camiade D. leg.". **Paratypes**, 34 specimens: same data as holotype.**Distribution.** MOZAMBIQUE: Vanduzi (Robiche 2008a).**32. *Drosochrus pilosus* (Gebien, 1921)***Helopinus pilosus* Gebien 1921: 122.*Drosochrus pilosus*: Koch 1958: 150.**Type Data. Syntypes**, three males and female (Basel Museum): "Aden, Arabien".**Distribution.** YEMEN: Aden (Gebien 1921).**33. *Drosochrus psalidiformis* (Ancey, 1881)***Helopinus psalidiformis* Ancey 1881: 397.*Drosochrus psalidiformis*: Koch 1958: 150.**Type Data. Syntypes**, two specimens (Paris Museum): "Aden (Yemen)".**Distribution.** YEMEN: Aden (Ancey 1881).**34. *Drosochrus simplicipes* (Gebien, 1937)***Helopinus simplicipes* Gebien 1937: 50.*Drosochrus simplicipes*: Koch 1958: 150.**Type Data. Holotype**, unsexed (Basel Museum): "Mpangwe". **Paratypes**, two specimens (Trieste Museum): "Ugogo: Dodoma, U. de Cieuta. II.1935".**Distribution.** TANZANIA: Dodoma, Ugogo (Gebien 1937); ZAMBIA: Mpangwe (Gebien 1937).**35. *Drosochrus sulcatipennis* (Solier, 1848)***Pteraulus sulcatipennis* Solier 1848: 202.*Drosochrus sulcatipennis*: Gebien 1910: 566.**Type Data. Syntypes**, not specified (Torino Museum): "Cap de Bonne Espérance".**Distribution.** SOUTH AFRICA: "Cap de Bonne Espérance" (Solier 1848).**36. *Drosochrus swierstrae* (Müller, 1887)***Emyon swierstrae* Müller 1887: 303.*Drosochrus swierstrae*: Koch 1958: 150.**Type Data. Syntypes**, not specified (Leiden Museum, Munich Museum): no data.**Distribution.** No data.**37. *Drosochrus textor* Schulze, 1968 (Fig. 11E)***Drosochrus textor* Schulze 1968: 178.**Type Data. Holotype**, male (Ditsong Museum): "Olifants Camp (Kruger National Park), 24.03°S, 31.44°E, December 1966, D. Goode". **Paratypes**, nine specimens: same data as holotype.**Distribution.** SOUTH AFRICA: Kruger National Park (Schulze 1968).**38. *Drosochrus tichyi* Robiche, 2010***Drosochrus tichyi* Robiche 2010: 369.**Type Data. Holotype**, male (Robiche coll.): "Zambie, NO Prov., 185 km S Mwinilunga, 06.XII.2004, Snyzek & Tichy leg". **Paratype**, female (Robiche coll.): same data as holotype.**Distribution.** ZAMBIA: 185 km south of Mwinilunga (Robiche 2010).**39. *Drosochrus vernayanus* Koch, 1958 (Fig. 11F)***Drosochrus vernayanus* Koch 1958: 163.**Type Data. Syntypes**, four specimens (Ditsong Museum): "Khan River, SW. of Trekkopje".**Distribution.** NAMIBIA: surroundings of Trekkopje, Rossing Mountains; Walvis Bay (Koch 1958).***Incertae Sedis*****40. *Drosochrus depressus* Erichson, 1843***Drosochrus depressus* Erichson 1843: 244.**Type Data. Holotype**, unsexed (probably Humboldt University): "Cap. Von Krebs".**Distribution.** No data.**Genus *Micrantereus* Solier, 1848***Micrantereus* Solier 1848: 175 [masculine].= *Solenomerus* Fåhraeus 1870: 306 [syn. by Fairmaire 1897: 131]. Type species: *Solenomerus longipes* Fåhraeus, 1870; by monotypy.**Type Species.** *Acanthomerus anomalus* Guérin-Méneville, 1834; by original designation**1. *Micrantereus algoensis* Péringuey, 1896***Micrantereus algoensis* Péringuey 1896: 176.**Type Data. Holotype**, male (Cape Museum): "Mozambique (Rikatla)".**Distribution.** MOZAMBIQUE: Elias (Péringuey 1896).

2. *Micrantereus ambiguus* Péringuey, 1899
Micrantereus ambiguus Péringuey 1899: 306.

Type Data. Holotype, male (Ditsong Museum): “Zambesia (Buluwayo). F. C. Selous”.

Distribution. ZIMBABWE: Bulawayo (Péringuey 1899).

3. *Micrantereus angolensis* Koch, 1958
Micrantereus angolensis Koch 1958: 214.

Type Data. Holotype, female (Ditsong Museum): “South-western Angola, Huila Province and District: Onguera, VI.1954, Vernay-Transv. Mus. Exped.”.

Distribution. ANGOLA: Onguera (Koch 1958).

4. *Micrantereus anomalus* (Guérin-Méneville, 1834)

Acanthomerus anomalus Guérin-Méneville 1834: 24.
Micrantereus anomalus: Solier 1848: 177.

Type Data. Holotype, not specified (Paris Museum): “Du Senegal”.

Distribution. SENEGAL* (Guérin-Méneville 1834).

5. *Micrantereus arabicus* Blair, 1933

Micrantereus arabicus Blair 1933: 6.

Type Data. Holotype, male (British Museum): “Hijaz, Taif foothills, Sahluj, 4.iv.1932”.

Distribution. SAUDI ARABIA: Sahluj (Blair 1933).

6. *Micrantereus armipes* Fairmaire, 1897

Micrantereus armipes Fairmaire 1897: 131.

Type Data. Syntypes, not specified (Paris Museum): “Ikouta, Afrique orientale”.

Distribution. Ikouta* (Fairmaire 1897); SOMALIA: Bur Akaba (Ferrer 1995).

7. *Micrantereus assimilis* Ancey, 1882

Micrantereus assimilis Ancey 1882: 54.

Type Data. Holotype, not specified (Paris Museum): “Uzagara”.

Distribution. TANZANIA: Usagara (Ancey 1882).

8. *Micrantereus capicola* Péringuey, 1904

Micrantereus capicola Péringuey 1904a: 245.

Type Data. Syntypes, not specified (Cape Museum): “Cape Colony (Mossel Bay; Cradock). Dr. H. Martin.”.

Distribution. SOUTH AFRICA: Mossel Bay, Cradock (Péringuey 1904a).

9. *Micrantereus carinatus* Péringuey, 1896

Micrantereus carinatus Péringuey 1896: 173.

Type Data. Syntypes, not specified (Cape Museum): “Betschuanaland (Ramoutsaa), Upper Limpopo”.

Distribution. BOTSWANA: Ramutosa, Upper Limpopo* (Péringuey 1896).

10. *Micrantereus citernii* Gridelli, 1939

Micrantereus citernii Gridelli 1939b: 45.

Type Data. Holotype, male (Geneva Museum): “Bohotlch e Berbera da Citerni nel 1903”.

Distribution. SOMALIA: surroundings of Berbera (Gridelli 1939b).

11. *Micrantereus costatus* Gerstaecker, 1854

Micrantereus costatus Gerstaecker 1854: 532.

Type Data. Holotype, not specified (Berlin Museum): no data.

Distribution. MOZAMBIQUE: Tete (Gestro 1873).

12. *Micrantereus desertus* Koch, 1965

Micrantereus desertus Koch 1965: 146.

Type Data. Holotype, male (Milan Museum): “Nefud Gebiet.- Hail, III.G. Popov”. **Paratypes**, two males and two females (British, Ditsong, Milan Museums): same data as holotype; male (Ditsong Museum): “70 km ostnordostlich von Hail, III., G. Popov”.

Distribution. SAUDI ARABIA: Ha'il, surroundings of Ha'il (Koch 1965).

13. *Micrantereus devexus* Péringuey, 1896

Micrantereus devexus Péringuey 1896: 176.

Type Data. Syntypes, not specified (Cape Museum): “Mozambique (Rikatla)”.

Distribution. MOZAMBIQUE: Elias (Péringuey 1896).

14. *Micrantereus drosochrooides* Ferrer, 1995

Micrantereus drosochrooides Ferrer 1995: 48.

Type Data. Holotype, male (Florence Museum): “Jessoma, 15.VIII.1968”. **Paratypes**, female: same data as holotype; female: “Mogadisco, at S. of Belet, 1.X.1973”.

Distribution. SOMALIA: Jessoma*, Mogadishu, surroundings of Beledweyne (Ferrer 1995).

15. *Micrantereus externecostatus* Fairmaire, 1884

Micrantereus externecostatus Fairmaire 1884: LXXVI.

Type Data. Syntypes, not specified (Paris Museum): “Makdischu”.

Distribution. SOMALIA: Balad, Kismayo (Ferrer 1995); Mogadishu (Fairmaire 1884).

Note. This taxon was redescribed as new by Fairmaire (1887: 295).

16. *Micrantereus externus* Fairmaire, 1899

Micrantereus externus Fairmaire 1899: 182.

Type Data. Syntypes, not specified (Paris Museum): “Delagoa”.

Distribution. MOZAMBIQUE: Maputo Bay (Fairmaire 1899).

17. *Micrantereus fallax* Péringuey, 1896

Micrantereus fallax Péringuey 1896: 175.

Type Data. Syntypes, not specified (Cape Museum): “Zambezia (Upper Limpopo)”.

Distribution. MOZAMBIQUE: “Upper Limpopo”* (Péringuey 1896).

18. *Micrantereus femoratus femoratus* Gerstaecker, 1871

Micrantereus femoratus Gerstaecker 1871: 64.
 = *Micrantereus luteopubens* Fairmaire 1882b: LII [syn. by Fairmaire 1887: 294].
 = *Micrantereus lacrymosus* Gerstaecker 1884: 56 [syn. by Gridelli 1939b: 14].

Type Data. *Micrantereus femoratus* Gerstaecker, 1871, **Syntypes**, male and female (Berlin Museum): “Von Mbaram und Endara”.

Micrantereus luteopubens Fairmaire, 1882, **Syn-types**, not specified (Paris Museum): no data.

Micrantereus lacrymosus Gerstaecker, 1884, **Syntypes**, not specified (Berlin Museum): “Massai”.

Distribution. ZANZIBAR: Mbaram*, Endara (Gerstaecker 1871); Massai* (Gerstaecker 1884).

Note. Gridelli (1939b) noted that many subspecies may represent variation or geographic forms. Rather than separating them into different species or combining them under a single name, he preserved the forms within different subspecies to organize them until more data could be gathered.

19. *Micrantereus femoratus septentrionalis* Gridelli, 1939

Micrantereus femoratus septentrionalis Gridelli 1939b: 16.

Type Data. Holotype, male (Trieste Museum): “Harar”. **Paratypes**, four males and two females (Basel Museum): same data as holotype; several specimens (Berlin Museum): “Erlanger-Neumann, Harr”; ten males and eight females (Berlin Museum): “7 ed il 30 aprile 1900, Argobba”; five males and six female (Berlin Museum): “11 maggio 1900 Abd el Kadr”; single specimen (Berlin Museum): “16 maggio 1900; Valle dell’ Errer”; three males (Berlin Museum): “sud di Harar”; four specimens (Budapest Museum): “Lafto”.

Distribution. ETHIOPIA: Harar, Lafto (Gridelli 1939b).

20. *Micrantereus femoratus sjoestedti* Gebien, 1910

Micrantereus femoratus sjoestedti Gebien 1910c: 390.

Type Data. Syntypes, 10 males and six females (Naturhistoriska riksmuseet): “Meru-Niederung 23.-25. XI”.

Distribution. KENYA: Meru National Park (Gebien 1910c), Shompole Conservancy (Gridelli 1939b).

Note. Gridelli (1939b), who moved Gebien’s species to a subspecies of *M. femoratus*, created an unjustified emendation of the name *sjöstedti*.

21. *Micrantereus femoratus variolosus* Gerstaecker, 1871

Micrantereus variolosus Gerstaecker 1871: 64.

Type Data. Holotype, female (Berlin Museum): “Von Endara”.

Distribution. ZANZIBAR: Endara (Gerstaecker 1871); TANZANIA: Arusha, Kilimandjaro, Moshi, Shirati, Tabora (Gridelli 1939b); KENYA: Meru National Park (Gebien 1910c); SOMALIA*; UGANDA* (Gridelli 1939b).

22. *Micrantereus gasanus* Péringuey, 1904

Micrantereus gasanus Péringuey 1904a: 247.

Type Data. Syntypes, not specified (Ditsong Museum): “Southern Rhodesia (Melseter, Gazaland). G. A. K. Marshall”.

Distribution. ZIMBABWE: Melsetter (Péringuey 1904a).

23. *Micrantereus gerstaeckeri andreinii* Gridelli, 1939

Micrantereus gerstaeckeri andreinii Gridelli 1939b: 29.

Type Data. Holotype, male (Trieste Museum): “Acrur”.

Distribution. ERITREA: Mendefera, Keyh, Segheneyti; ETHIOPIA: Fare, Kerteta, Tembien* (Gridelli 1939b).

24. *Micrantereus gerstaeckeri crenatus* Gridelli, 1939

Micrantereus gerstaeckeri crenatus Gridelli 1939b: 31.

Type Data. Syntypes, male (Berlin Museum or Trieste Museum): “Mojo!, 1 June 1900”; male (Berlin Museum or Trieste Museum): “da Mojo ad Atschabo!, 2 June 1900”; male (Berlin Museum or Trieste Museum): “Oda!, 13 June 1900”.

Distribution. ETHIOPIA: Alto Uebi Scebeli*, Atschabo*, Mojo, Oda* (Gridelli 1939b).

25. *Micrantereus gerstaeckeri fimbritibius* Fairmaire, 1882

Micrantereus fimbritibius Fairmaire 1882c: 68.

Type Data. Holotype, not specified (Paris Museum): no data.

Distribution. No data.

Note. Gridelli (1939b) notes that this subspecies (originally described as a species) is a synonym of either the nominative form or *M. gerstaeckeri andreinii*. However, the female holotype lacks the characters needed to further clarify the synonymy, so until more evidence is examined, this subspecies is retained as valid.

26. *Micrantereus gerstaeckeri gerstaeckeri* Gestro, 1873

Micrantereus gerstaeckeri Gestro 1873: 354.

Type Data. Holotype, male (Genoa Museum): “Keren dal Dott. O. Beccari nel 1870”.

Note. The species name was originally *Micrantereus gerstäckeri*, but was subsequently corrected to *M. gerstaeckeri*.

Distribution. ERITREA: Asmara, Gheleb (Gridelli 1939b), Keren (Gestro 1873), Massawa (Gridelli 1939b); SUDAN: Suakin (Gridelli 1939b).

27. *Micrantereus gerstaecheri laevior* Fairmaire, 1892

Micrantereus laevior Fairmaire 1892: 114.

Type Data. Syntypes, not specified (Paris Museum): no data.

Distribution. DJIBOUTI: Obock, Maro in Danakil Plain*; ETHIOPIA: Auasc Valley*, Dire Dawa, Gota, Harar (Gridelli 1939b).

28. *Micrantereus gerstaecheri laevissimus* Gridelli, 1939

Micrantereus gerstaecheri laevissimus Gridelli 1939b: 32.

Type Data. Syntypes, two males (Berlin Museum and Trieste Museum): “da Gorgora a Goloda, 17 giugno 1900”.

Distribution. SOMALIA: Surroundings of Goloda (Gridelli 1939b).

29. *Micrantereus gratiosus* Péringuey, 1896

Micrantereus gratiosus Péringuey 1896: 174.

Type Data. Syntypes, males (Cape Museum): “Zambezia (between the Zambeze and Limpopo Rivers)”.

Distribution. MOZAMBIQUE: “between the Zambeze and Limpopo Rivers”* (Péringuey 1896).

30. *Micrantereus gridelli* Koch, 1963

Micrantereus gridelli Koch 1963: 165.

Type Data. Holotype, male (Ditsong Museum): “Jesomina, VIII.1959, C. Koch leg.”. **Paratypes**, male and female, same data as holotype; female (Trieste Museum): “Hiran-Provinz: Belet Uen, IV. 1936, C. Lomi leg.”.

Distribution. SOMALIA: Belet Uen (Koch 1963).

31. *Micrantereus hirsutus* Péringuey, 1904

Micrantereus hirsutus Péringuey 1904a: 248.

Type Data. Holotype, female (Ditsong Museum): “Transvaal (Lydenburg)”.

Distribution. SOUTH AFRICA: Lydenburg (Péringuey 1904a).

32. *Micrantereus kuntzeni* Gridelli, 1939

Micrantereus kuntzeni Gridelli 1939b: 17.

Type Data. Syntypes, male (Trieste Museum): “S. Galla. - 21 April 1901, Tarre Fada Gumhi”; female (Berlin Museum): “S. Galla. - 2 May 1901, Malka Re-Handodu”; male (Berlin Museum): “S. Somali - 3 May 1901, Handodu-Haro Lola”; single male and single female (Berlin Museum): “S. Somali. - 6 May 1901, Haro Lola”; female (Berlin Museum): “S. Somali. - May 1901, Haro Lola”.

Distribution. SOMALIA: Haro Lola*, Malke Re (Gridelli 1939b).

33. *Micrantereus laevigatus* Gridelli, 1939

Micrantereus laevigatus Gridelli 1939b: 26.

Type Data. Syntypes, single male (Berlin Museum or Trieste Museum): “Djilanden (Somalia mer.), 20 may 1901”; single male (Berlin Museum or Trieste Museum): “Bardera-Makka Gele Gedid (Somalia mer.), 2 June 1901”; single male (Berlin Museum or Trieste Museum): “S. Galla, Hasa (Mauc?), 31 marzo 1901”.

Distribution. SOMALIA: Bardera, Makka Gele Gedid*, Merca (Gridelli 1939b).

34. *Micrantereus lequeuxi* Robiche, 2019

Micrantereus lequeuxi Robiche 2019: 357.

Type Data. Holotype, male (Paris Museum): “République du Somaliland, région de Woqooyi Galbeed, Abudla Hill, Arabaiyo, 1300 m, 6.VI.2011, 9°40'38"N-43°45'46"E, J-P. Lequeux leg.” Paratypes. Two males, single female (Robiche coll.) same data as holotype.

Distribution. SOMALIA: Abudla Hill, Arabaiyo (Robiche 2019).

35. *Micrantereus limpopoanus* Péringuey, 1896

Micrantereus limpopoanus Péringuey 1896: 172.

Type Data. Syntypes, not specified (Cape Museum): “Upper Limpopo River”.

Distribution. SOUTH AFRICA: “Upper Limpopo River” (Péringuey 1896).

36. *Micrantereus longipes* (Fåhraeus, 1870)

Solenomerus longipes Fåhraeus 1870: 306.

Micrantereus longipes: Péringuey 1896: 173.

Type Data. Syntypes, not specified (Naturhistoriska riksmuseet): no data.

Distribution. SOUTH AFRICA: Transvaal*, Limpopo River* (Péringuey 1892a).

Note. Péringuey (1896) treated *Solenomerus* as a subgenus of *Micrantereus*.

37. *Micrantereus luctuosus* Péringuey, 1896

Micrantereus luctuosus Péringuey 1896: 173.

Type Data. Holotype, male (Cape Museum): “Mozambique (Rikatla)”.

Distribution. MOZAMBIQUE: Elias (Péringuey 1896).

38. *Micrantereus lydenburgiensis* Péringuey, 1904

Micrantereus lydenburgiensis Péringuey 1904a: 247.

Type Data. Syntypes, not specified (Ditsong Museum): “Transvaal (Lydenburg)”.

Distribution. SOUTH AFRICA: Lydenburg (Péringuey 1904a).

39. *Micrantereus machadoi* Koch, 1958

Micrantereus machadoi Koch 1958: 215.

Type Data. Holotype, male (Companhia Diamantes): “North-eastern Angola, Province of Malange, Lunda District: Dundo, XII.1949. A. de Barros Machado”.

Distribution. ANGOLA: Dundo (Koch 1958).

40. *Micrantereus mantillerii* Robiche, 2012*Micrantereus mantillerii* Robiche 2012b: 220.

Type Data. Holotype, male (Paris Museum): “Kenya, Isiolo à Wajir, 01. V. 2001, K. Werner leg.”. **Paratypes**, 10 males and four females (Paris Museum and Robiche coll.): same data as holotype; two females (Robiche coll.): “Kenya, N.E. prov. 20.IV.2001, K. Werner leg.”; female (Paris Museum): “Kenya, N.E. prov., El Wak, 3.V.2001, K. Werner leg.”; female (Robiche coll.): “Kenya, El Wak, 26.IV.2001, K. Werner leg.”; male (Brussels Museum): “Kenya, coast Garissa, N of Bura, 5.XII.2007, Snižek leg.”.

Distribution. KENYA: Isiolo, El Wak, Wajir, surroundings of Bura (Robiche 2012b).

41. *Micrantereus marginipennis* Fairmaire, 1884*Micrantereus marginipennis* Fairmaire 1884:

LXXVI.

Type Data. Holotype, female (Paris Museum): “Makdischu”.

Distribution. SOMALIA: Mogadishu (Fairmaire 1884).

Note. This taxon was redescribed as new by Fairmaire (1887: 295).

42. *Micrantereus nitidus nitidus* Gahan, 1896*Micrantereus nitidus* Gahan 1896: 455.

Type Data. Syntypes, not specified (British Museum): “Aden and Lahej (Yerhury)”.

Distribution. YEMEN: Aden, Lahej (Gahan 1896).

43. *Micrantereus nitidus yemensis* Koch, 1965*Micrantereus nitidus yemensis* Koch 1965: 145.

Type Data. Holotype, male (Milan Museum): “Yemen Taiz, 1963, G. Benardelli”. **Paratypes**, two males (Milan and Ditsong Museums): same data as holotype.

Distribution. YEMEN: Taiz (Koch 1965); Delemi* (Kaszab 1972).

44. *Micrantereus ovampoanus* Péringuey, 1896*Micrantereus ovampoanus* Péringuey 1896: 175.

Type Data. Syntypes, not specified (Cape Museum): “Ovampoland”.

Distribution. NAMIBIA: Ovampoland (Péringuey 1896).

45. *Micrantereus parvidens* Gebien, 1910*Micrantereus parvidens* Gebien 1910c: 389.

Type Data. Syntypes, 30 specimens (Naturhistoriska riksmuseet and Basel Museum), single specimen (Budapest Museum): “Kilimandjaro: 1.16.XI”; “Kibonoto 1,300-1,900 m. 15.XI.-16.XII”, “Obstgartensteppe 16. XI”, “Natronsee 28.II”, “Nieder Kibonoto 28.II”, “Mwika XI.1907”.

Distribution. TANZANIA: Kilimandjaro (Gebien 1910c).

46. *Micrantereus patrizii* Gridelli, 1939*Micrantereus patrizii* Gridelli 1939b: 20.

Type Data. Holotype, unsexed (Geneva Museum): “Archer’s Post, Gennaio 1920”. **Paratype**, unsexed (Basel Museum): “Bulessa, Gennaio 1920”.

Distribution. KENYA: Archer’s Post, Bulessa (Gridelli 1939b), between Mararal and Turkana* (Ferrer 1996).

47. *Micrantereus paulschulzei* Schulze, 1968*Micrantereus paulschulzei* Schulze 1968: 180.

Type Data. Holotype, male (Ditsong Museum): “5 miles East of Jambila (Tvl.), 25°45’S, 30°53’E, December 1965, L. Schulze”. **Paratypes**, 12 females: same data as holotype.

Distribution. SOUTH AFRICA: Umjindi (25°45’S, 30°53’E) (Schulze 1968).

48. *Micrantereus procursus* Péringuey, 1904*Micrantereus procursus* Péringuey 1904a: 245.

Type Data. Syntypes, not specified (Cape Museum): “Southern Rhodesia (Melsetter, Gazaland). G. A. K. Marshall.”.

Distribution. ZIMBABWE: Chimanimani (Péringuey 1904a).

49. *Micrantereus pronus* Péringuey, 1896*Micrantereus pronus* Péringuey 1896: 176.

Type Data. Syntypes, not specified (Cape Museum): “Transvaal (Barberton)”.

Distribution. SOUTH AFRICA: Barberton (Péringuey 1896).

50. *Micrantereus propinquus* Péringuey, 1899*Micrantereus propinquus* Péringuey 1899: 307.

Type Data. Syntypes, not specified (Cape Museum): “Zambesia (Buluwayo). F. C. Selous”.

Distribution. ZIMBABWE: Bulawayo (Péringuey 1899).

51. *Micrantereus quadricristatus* Fairmaire, 1884*Micrantereus quadricristatus* Fairmaire 1884: LXXVI.

Type Data. Holotype, female (Paris Museum): “Makdischu”.

Distribution. SOMALIA: Jezeera (Ferrer 1995); Mogadishu (Fairmaire 1884).

Note. This taxon was redescribed as new by Fairmaire (1887: 296).

52. *Micrantereus recticostatus* Fairmaire, 1882*Micrantereus recticostatus* Fairmaire 1882a: 79.

Type Data. Holotype, female (Paris Museum): no data.

Distribution. SOMALIA* (Gridelli 1939b).

53. *Micrantereus rugulosus berberanus* Gridelli, 1939*Micrantereus rugulosus berberanus* Gridelli 1939b: 36.

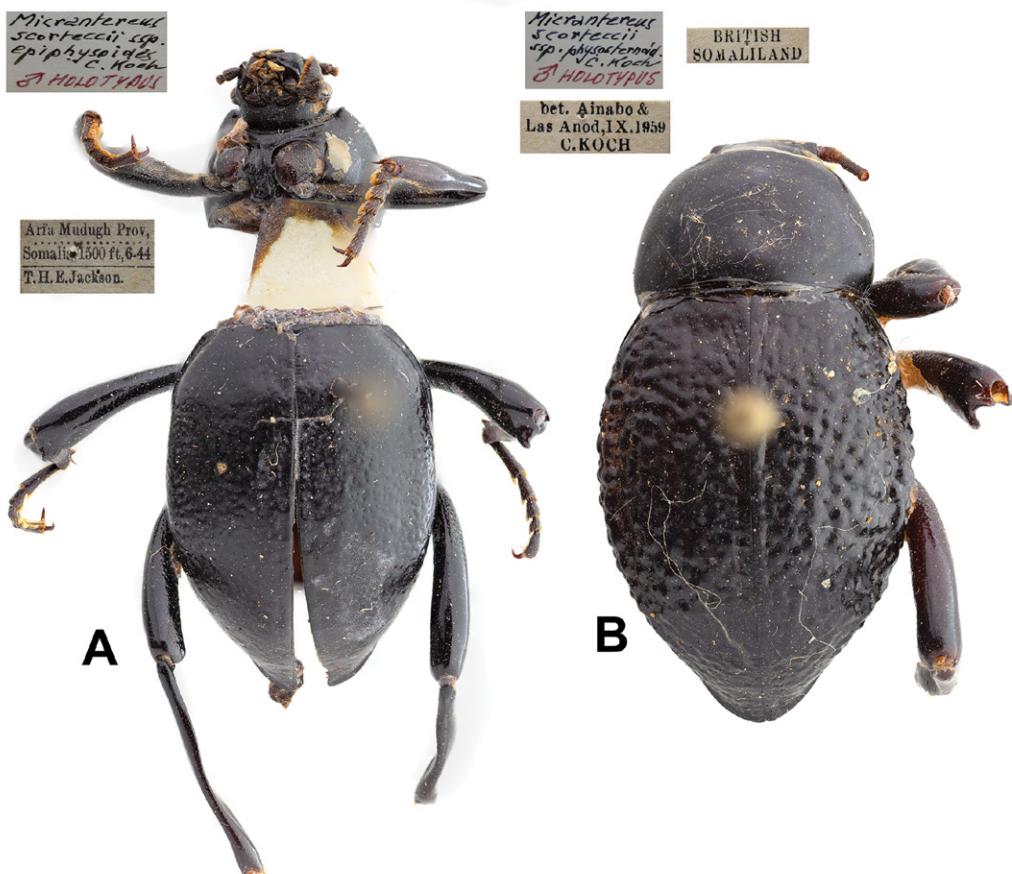


Fig. 12. Type specimens of species representing the genus *Micrantereus*. A) *M. scortecii epiphysoides*, B) *M. scortercci physosternoides*.

Type Data. Syntypes, male and female (Paris Museum): “Berbera”.

Distribution. DJIBOUTI: Obock (Gridelli 1939b); ERITREA: Assab (Gridelli 1939b); SOMALIA: Berbera (Gridelli 1939b); ETHIOPIA: Dire Dawa (Gridelli 1939b).

54. *Micrantereus rugulosus rugulosus* Gestro, 1878
Micrantereus rugulosus Gestro 1878: 321.

Type Data. Holotype, not specified (Genoa Museum): “Mahal Uonz (Scioa)”.

Distribution. ETHIOPIA: Mahal Uonz (Gestro 1878); ERITREA: Assab (Gridelli 1939b); DJIBOUTI: Obock (Fairmaire 1893b).

55. *Micrantereus scaberrimus* Fairmaire, 1894
Micrantereus scaberrimus Fairmaire 1894a: 327.

Type Data. Holotype, female (Paris Museum): “Mozambique”.

Distribution. MOZAMBIQUE: Elias (Péringuay 1896).

56. *Micrantereus scortecii epiphysoides* Koch, 1962 (Fig. 12A)

Micrantereus scortecii epiphysoides Koch 1962: 267.

Type Data. Holotype, male (Coryndon Museum): “Arfa, 1500 ft., VI.1944, T.H.E. Jackson”. **Paratype**, female (Coryndon Museum): “Mudugh Provinz, Angabe, 800 ft. Seehöhe, VII.1945, T.H.E. Jackson”.

Distribution. SOMALIA: Mudug province (Koch 1962).

57. *Micrantereus scortercci physosternoides* Koch, 1962 (Fig. 12B)

Micrantereus scortercci physosternoides Koch 1962: 266.

Type Data. Holotype, male (Ditsong Museum): “Ainabo, Las Anod, IX.1959, C. Koch leg.”. **Paratypes**, male and three females (Genoa Museum): “Garoe, IX.1959”; female (Ditsong Museum): “Bosaso, IX.1959, C. Koch leg”.

Distribution. SOMALIA: Bosaso, Garowe (Koch 1962).

58. *Micrantereus scortercci scortercci* Koch, 1962*Micrantereus scortercci scortercci* Koch 1962: 263.

Type Data. Holotype, male (Genoa Museum): “Carear Berge, nördlich von Gardo, V.1953, Scortecci leg.”.

Distribution. SOMALIA: surroundings of Qardho (Koch 1962).

59. *Micrantereus septemcostatus* Fairmaire, 1897*Micrantereus septemcostatus* Fairmaire 1897: 131.

Type Data. Syntypes, not specified (Paris Museum): “Afrique orientale”.

Distribution. Afrique orientale* (Fairmaire 1897).

60. *Micrantereus seriegranosus* Fairmaire, 1891*Micrantereus seriegranosus* Fairmaire 1891: CCXCVI.

Type Data. Syntypes, not specified (Paris Museum): no data.

Distribution. No data.

Note. Gridelli (1939b) noted that this species was undoubtedly a form of his *femoratus* complex based on femoral morphology and elytral pubescence. Further, he mentioned the species' description matched his concept of *M. femoratus variolosus*. However, he did not explicitly state the synonymy.

61. *Micrantereus seriepunctatus* Fairmaire, 1884*Micrantereus seriepunctatus* Fairmaire 1884: LXXV.

Type Data. Holotype, male (Paris Museum): “Makdischu”.

Distribution. SOMALIA: Baraawe, Belet Amin, Bulo Kero (Gridelli 1939b), Mogadishu (Fairmaire 1884).

Note. This taxon was redescribed as new by Fairmaire (1887: 294).

62. *Micrantereus sinuatipes* Fairmaire, 1882*Micrantereus sinuatipes* Fairmaire 1882a: 78.

= *Micrantereus asidoides* Fairmaire 1893b: 152 [syn. by Gebien 1943: 908].

Type Data. *Micrantereus sinuatipes* Fairmaire, 1882, **holotype**, not specified (Paris Museum): no data.

Micrantereus asidoides Fairmaire, 1893, **holotype**, not specified (Paris Museum): “Ogadeen”.

Distribution. ETHIOPIA: “Ogadeen”* (Fairmaire 1893b); SOMALIA: Buran (Gridelli 1939b).

63. *Micrantereus somalicus* Gridelli, 1939*Micrantereus somalicus* Gridelli 1939b: 24.

Type Data. Syntypes, two males (Trieste Museum): “Merca, nella Somalia italiana costiera”.

Distribution. SOMALIA: Jawhar (Ferrer 1995), Merca (Gridelli 1939b).

64. *Micrantereus spissus* Périnquey, 1899*Micrantereus spissus* Périnquey 1899: 306.

Type Data. Holotype, male (Ditsong Museum): “Transvaal (Leydenburg)”.

Distribution. SOUTH AFRICA: Lydenburg (Périnquey 1899).

65. *Micrantereus spurius* Périnquey, 1896*Micrantereus spurius* Périnquey 1896: 174.

Type Data. Syntypes, not specified (Cape Museum): “Zambezia (between the Zambeze and Limpopo Rivers, Manica)”.

Distribution. MOZAMBIQUE: “between the Zambeze and Limpopo Rivers”*, Manica (Périnquey 1896).

66. *Micrantereus szalaymarzsoi* Kaszab, 1972*Micrantereus szalaymarzsoi* Kaszab 1972: 382.

Type Data. Holotype, male (Budapest Museum): “Yemen: Wadi Zabid, IX.1970, leg. A. Szalay-Marzso”. **Paratypes**, male (Budapest Museum): “Yemen: Wadi Zabid, 1.-24.VIII.1969, leg. A. Szalay-Marzso”; male (Budapest Museum): “Yemen: Wadi Zabid, VIII.1970, leg. A. Szalay-Marzso”.

Distribution. YEMEN: Wadi Zabid (Kaszab 1972).

Note. Type depository information after Merkl *et al.* (2015).

67. *Micrantereus szekessyi szekessyi* Gridelli, 1939*Micrantereus szekessyi szekessyi* Gridelli 1939b: 33.

Type Data. Holotype, male (Budapest Museum): “Abyssinia, Kovacs - Vallis Erer”. **Paratypes**, five specimens (Budapest Museum): same data as holotype; two specimens (Trieste Museum): same data as holotype.

Distribution. SOMALIA: Kovacs*, ETHIOPIA: Harar (Gridelli 1939b).

68. *Micrantereus szekessyi trichocnemis* Gridelli, 1939*Micrantereus szekessyi trichocnemis* Gridelli 1939b: 34.

Type Data. Syntypes, male (Berlin Museum): “G. Ali-Haririssa il 30 maggio 1900”; male (Berlin Museum): “Humbo Wabbi il 7 giugno 1900”.

Distribution. SOMALIA: Humbo Wadi; DJIBOUTI: Ali-Haririssa* (Gridelli 1939b).

69. *Micrantereus tentyrioides* Pascoe, 1882*Micrantereus tentyrioides* Pascoe 1882: 29.

Type Data. Holotype, male (British Museum): “Arabia (Yemen)”.

Distribution. YEMEN* (Pascoe 1882).

70. *Micrantereus timarchoides* Fairmaire, 1893*Micrantereus timarchoides* Fairmaire 1893b: 152.

Type Data. Holotype, not specified (Paris Museum): “Ogadeen”.

Distribution. ETHIOPIA: Ogadeen* (Fairmaire 1893b).

71. *Micrantereus transjubae* Gridelli, 1939*Micrantereus transjubae* Gridelli 1939b: 43.

Type Data. Holotype, unsexed (Trieste Museum): “Oltre Giuba, Vitaliano Bertozzo”.

Distribution. SOMALIA: Belet Amin, Oltre Giuba, Giuba (Gridelli 1939b).

72. *Micrantereus validus* Péringuey, 1899
Micrantereus validus Péringuey 1899: 305.

Type Data. Syntypes, 11 males and six females (Cape Museum): “Zambesia (Buluwayo). F. C. Selous”.

Distribution. ZIMBABWE: Bulawayo (Péringuey 1899).

73. *Micrantereus velox* (Péringuey, 1892)

Solenomerus velox Péringuey 1892a: 58.
Micrantereus velox: Gebien 1911: 565.

Type Data. Holotype, not specified (Cape Museum): no data.

Distribution. No data.

74. *Micrantereus vicarius* Péringuey, 1904

Micrantereus vicarius Péringuey 1904a: 249.

Type Data. Syntypes, not specified (Ditsong Museum): “Southern Rhodesia (Mount Shirinda, Gazaland). G. A. K. Marshall.”.

Distribution. ZIMBABWE: Mount Selinda (Péringuey 1904a).

75. *Micrantereus zoutpansbergianus* Péringuey, 1904

Micrantereus zoutpansbergianus Péringuey 1904a: 246.

Type Data. Syntypes, not specified (Ditsong Museum): “Transvaal (Zoutpansberg). Rev. H. A. Junod; A.J.T. Janse.”.

Distribution. SOUTH AFRICA: Zoutpansberg* (Péringuey 1904a).

Genus *Nicandra* Fairmaire, 1888

Subgenus *Calous* Koch, 1958

Calous Koch 1958: 151 [masculine].

Type Species. *Blastarnus michaelseni* Gebien, 1920; by original designation.

1. *Nicandra granicosta* Koch, 1958 (Fig. 13A)

Nicandra granicosta Koch 1958: 173.

Type Data. Holotype, male (Ditsong Museum): “Damaraland: Abachaus, XII.1949, F. Hobohm”.
Paratype, same data as holotype.

Distribution. NAMIBIA: Abachaus (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 13A), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 13A should still be considered a syntype.

2. *Nicandra hobohmi* Koch, 1958 (Fig. 13B)

Nicandra hobohmi Koch 1958: 172.

Type Data. Syntypes, six specimens (Ditsong Museum): “Damaraland: Abachaus, XII.1949, F. Hobohm”.

Distribution. NAMIBIA: Abachaus (Koch 1958).

3. *Nicandra michaelseni* (Gebien, 1920)

Blastarnus michaelseni Gebien 1920: 149.

Nicandra michaelseni: Koch 1958: 151.

Type Data. Syntypes, seven specimens (Basel Museum): “Windhuk, 29.IV.—8.V. 1911 (Michaelsen)”.

Distribution. NAMIBIA: Windhoek (Gebien 1920); Ebony, Erongo Mountains, Karibib, Khan River* (Koch 1958).

4. *Nicandra okahandia* (Gebien, 1920) (Fig. 13C)

Blastarnus okahandius Gebien 1920: 150.

Nicandra okahandia: Koch 1958: 151.

Type Data. Neotype, here designated, male (Ditsong Museum): “Portsmut, Ababis, VIII.1954. Vernay-Transv. Mus. Exped.”. **Holotype**, male (Hamburg Museum), destroyed: “Okahandja 27.-28. IV.1911 (Michaelsen)”.

Note. As a result of the work on this catalog, we found that the holotype of this species was destroyed during WW2 (Koch 1958). Unfortunately, in 1958 Koch incorrectly designated a neotype for this species by designating 45 specimens as “neotypes” (ICZN 1999). We hereby designate a neotype from the series investigated by Koch (1958) to provide nomenclatural stability and refer the reader to that publication for morphological details.

Distribution. NAMIBIA: Okahandja (Gebien 1920), Ababis, Svakop (Koch 1958).

5. *Nicandra subgranosa* (Gebien, 1920)

Blastarnus subgranosus Gebien 1920: 148.

Nicandra subgranosa: Koch 1958: 151.

Type Data. Syntypes, not specified (Basel Museum): “Windhuk, 29. IV.—8. V. 1911 (Michaelsen)”.

Distribution. NAMIBIA: Windhoek (Gebien 1920); SOUTH AFRICA: Kimberley (Koch 1958).

6. *Nicandra variabilis* (Hesse, 1935) (Fig. 13D)

Blastarnus variabilis Hesse 1935: 569.

Nicandra variabilis: Koch 1958: 151.

Type Data. Syntypes, 12 males and 21 females (Ditsong Museum), single specimen (Budapest Museum): “Kuke Pan, 21-30. iii. 30”; three females (Ditsong Museum): “Kopjes, Mabaleapudi, 7-9. v. 30”; two males and one female (Cape Museum): “Vryburg (Jones)”; female (Cape Museum): “Johannesburg, Transvaal (A. Ross)”.

Distribution. SOUTH AFRICA: Vryburg, Johannesburg; BOTSWANA: Kuke Pan, Mabaleapudi (Hesse 1935), Lobatsi, N’Gami (Koch 1958).

Subgenus *Heteronicandra* Koch, 1958

Heteronicandra Koch 1958: 151 [feminine].

Type Species. *Nicandra zumpti* Kulzer, 1951; by original designation

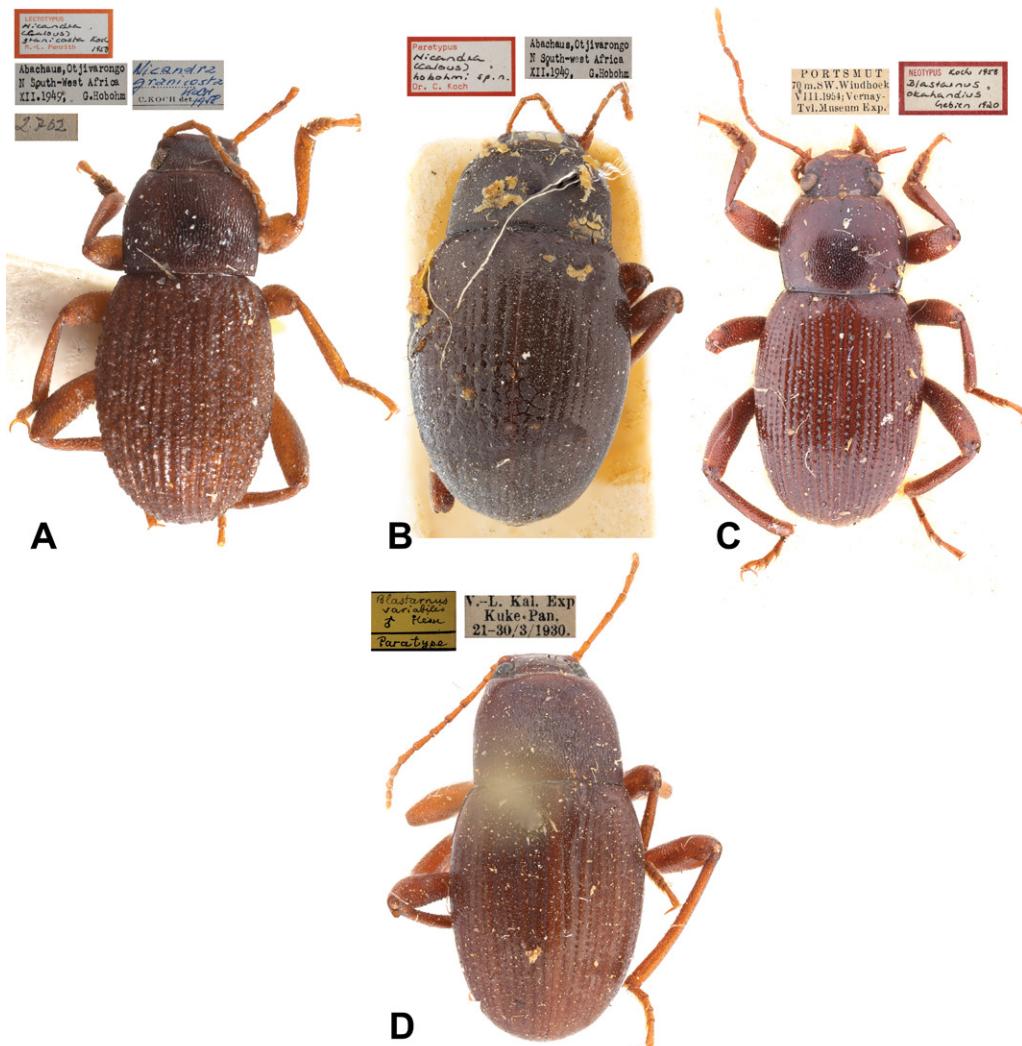


Fig. 13. Type specimens of species representing the genus *Nicandra* subgenus *Calous*. A) *N. granicosta*, B) *N. hobohmi*, C) *N. okahandia*, D) *N. variabilis*.

7. *Nicandra brincki* Koch, 1958 (Fig. 14A)
Nicandra brincki Koch 1958: 177.

Type Data. Syntypes. 10 specimens (Lund Museum), single specimen (Ditsong Museum); “North-central Cape Province: Narugas (Gordonia District). V.1951, P. Brinck & G. Rudebeck”.

Distribution. SOUTH AFRICA: Narugas (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 14A), this designation was never formalized in publication. Therefore, the specimen presented in Fig. 14A should still be considered a syntype.

8. *Nicandra dentimana* Koch, 1958 (Fig. 14B)
Nicandra dentimanus Koch 1958: 181.

Type Data. Holotype, male (Ditsong Museum); “Central Cape Province: Laingsburg, VII.1950, E.A. Pienaar”.

Distribution. SOUTH AFRICA: Laingsburg (Koch 1958).

9. *Nicandra goellnerae* Ferrer, 2004
Nicandra goellnerae Ferrer 2004: 234.

Type Data. Holotype, male (Berlin Museum); “Namibia, Kunene, 9km O de Ruacana, 24-26.

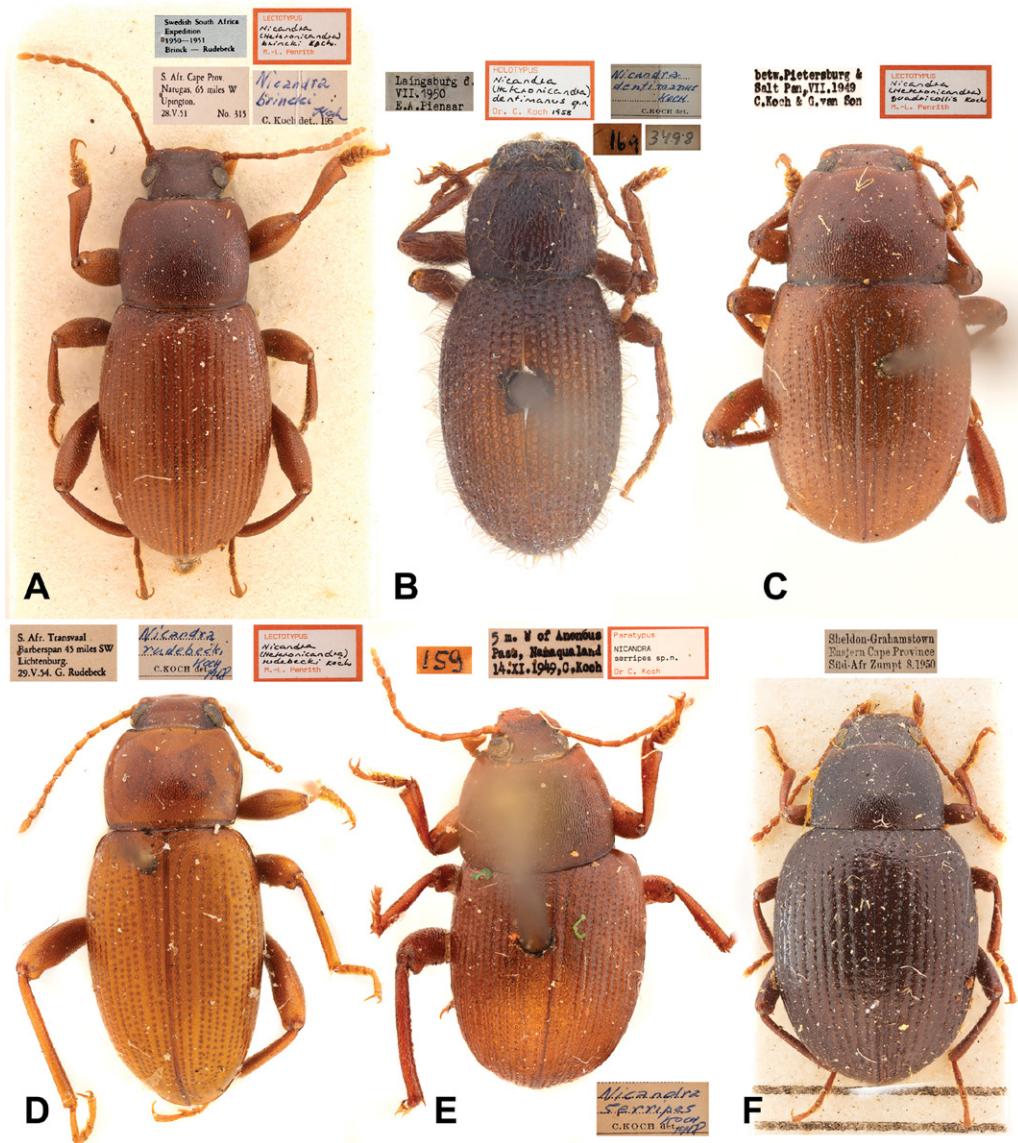


Fig. 14. Type specimens of species representing the genus *Nicandra* subgenus *Heteronicandra*. A) *N. brincki*, B) *N. dentimana*, C) *N. quadricollis*, D) *N. rudebeki*, E) *N. serripes*.

II1194, U. Göllner leg.”. **Paratypes**, three specimens (Berlin Museum and Naturhistoriska riksmuseet): same data as holotype; one specimen (Berlin Museum): “Etosha National Park, Namutoni, 27.II.1994, M. Uhlig leg.”.

Distribution. NAMIBIA: Namutoni, surroundings of Ruacana (Ferrer 2004).

10. *Nicandra piliphalla* Koch, 1958

Nicandra piliphallus Koch 1958: 180.

Type Data. Syntypes, four specimens (Ditsong Museum): “North-central Cape Province: Niekerk’s Hope, VIII.1950, C. Koch & G. van Son”.

Distribution. SOUTH AFRICA: Niekerk’s Hope, Pofadder (Koch 1958).

11. *Nicandra quadricollis* Koch, 1958 (Fig. 14C)
Nicandra quadricollis Koch 1958: 176.

Type Data. Syntypes, two males and four females (Ditsong Museum): “Northern Transvaal; btwn

Pietersburg and Salt Pan, VII.1949. C. Koch & G. van Son".

Distribution. SOUTH AFRICA: between Pietersburg and Salt Pan (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 14C), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 14C should still be considered a syntype.

12. *Nicandra rudebeki* Koch, 1958 (Fig. 14D)
Nicandra rudebeki Koch 1958: 178.

Type Data. Syntypes, seven specimens (Ditsong Museum, Lund Museum): "Westen Transvaal: Barberspan, V.1954, G. Rudebeck".

Distribution. SOUTH AFRICA: Barberspan, Brakpan (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 14D), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 14D should still be considered a syntype.

13. *Nicandra serripes* Koch, 1958 (Fig. 14E)
Nicandra serripes Koch 1958: 174.

Type Data. Syntypes, 33 specimens (Ditsong Museum), three specimens (Budapest Museum): "Little Namaqualand: 5 m. W. of Anenous Pass, XI.1949. C. Koch".

Distribution. SOUTH AFRICA: Anenous Pass (Koch 1958).

14. *Nicandra zumpti* Kulzer, 1951
Nicandra zumpti Kulzer 1951: 565.

Type Data. Syntypes, 35 specimens (Basel Museum), two specimens (Budapest Museum), single specimen (Ditsong Museum): "Sheldon-Grahamstown Eastern Cape Prov. 8.1.50 leg. Zumpt".

Distribution. SOUTH AFRICA: Grahamstown (Kulzer 1951).

Subgenus *Nicandra* Fairmaire, 1888

Nicandra Fairmaire 1888a: 191 [feminine].

Type Species. *Nicandra costulipennis* Fairmaire, 1888; by monotypy.

15. *Nicandra atoma* (Gebien, 1920)
Blastarnus atomus Gebien 1920: 152.
Nicandra atoma: Koch 1958: 151.

Type Data. Syntypes, not specified (Basel Museum): "Windhuk 29. IV. —8. V. 1911 (Michaelsen)"; and "Osona bei Okahandja VI. 1911 (Michaelsen)".

Distribution. NAMIBIA: Osona, Windhoek (Koch 1958).

16. *Nicandra comata* Koch, 1958

Nicandra comata Koch 1958: 165.

Type Data. Syntypes, male and female (Ditsong Museum): "Great Namaqualand, Kuibis, IX.1950, C. Koch & G. van Son".

Distribution. NAMIBIA: Kuibis, Rek Vlakte (Koch 1958).

17. *Nicandra costatula* Koch, 1958

Nicandra costatula Koch 1958: 170.

Type Data. Syntypes, five specimens (Ditsong Museum): "Little Namaqualand: Holgat, IX.1950, C. Koch & G. van Son"; five specimens (California Academy of Sciences): "15 m. S. of Alexander Bay, XII.1948, Univ. California-Transv. Mus. Exped."; two specimens (Ditsong Museum): "18 m. N. of Kleinzee, XII.1948, Univ. California-Transv. Mus. Exped.".

Distribution. SOUTH AFRICA: Holgat, surroundings of Alexander Bay, surroundings of Kleinzee (Koch 1958).

18. *Nicandra costulipennis* Fairmaire, 1888

Nicandra costulipennis Fairmaire 1888a: 191.

Type Data. Holotype, unsexed (Paris Museum): "Namaqua".

Distribution. SOUTH AFRICA: Namaqua*.

19. *Nicandra desertica* Koch, 1958 (Fig. 15A)

Nicandra desertica Koch 1958: 166.

Type Data. Holotype, male (Ditsong Museum): "Western Kaokoveld: Rocky Point, coast, VI.1951, B. Carp Exped.".

Distribution. NAMIBIA: Kaokoveld (Rocky Point) (Koch 1958).

20. *Nicandra emyonopa* Koch, 1958 (Fig. 15B)

Nicandra emyonopus Koch 1958: 170.

Type Data. Syntypes, male and female (Ditsong Museum): "Central and South-western Angola: Lobito, IV.1948, C. Koch"; male (Ditsong Museum): "Ongueria, VI.1954, Vernay-Transv. Mus. Exped.".

Distribution. ANGOLA: Lobito, Hunguéría (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 15B), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 15B should still be considered a syntype.

21. *Nicandra granipennis* (Gebien, 1920)

Blastarnus granipennis Gebien 1920: 147.

Nicandra granipennis: Koch 1958: 151.

Type Data. Syntypes, single specimen (Basel Museum): "Narubis bei Keetmanshoop, III.—IV. 1913 (H. Tsshomsen)"; single specimen (Basel Museum): "Kuibis 15. VII.1911 (Michaelsen)".

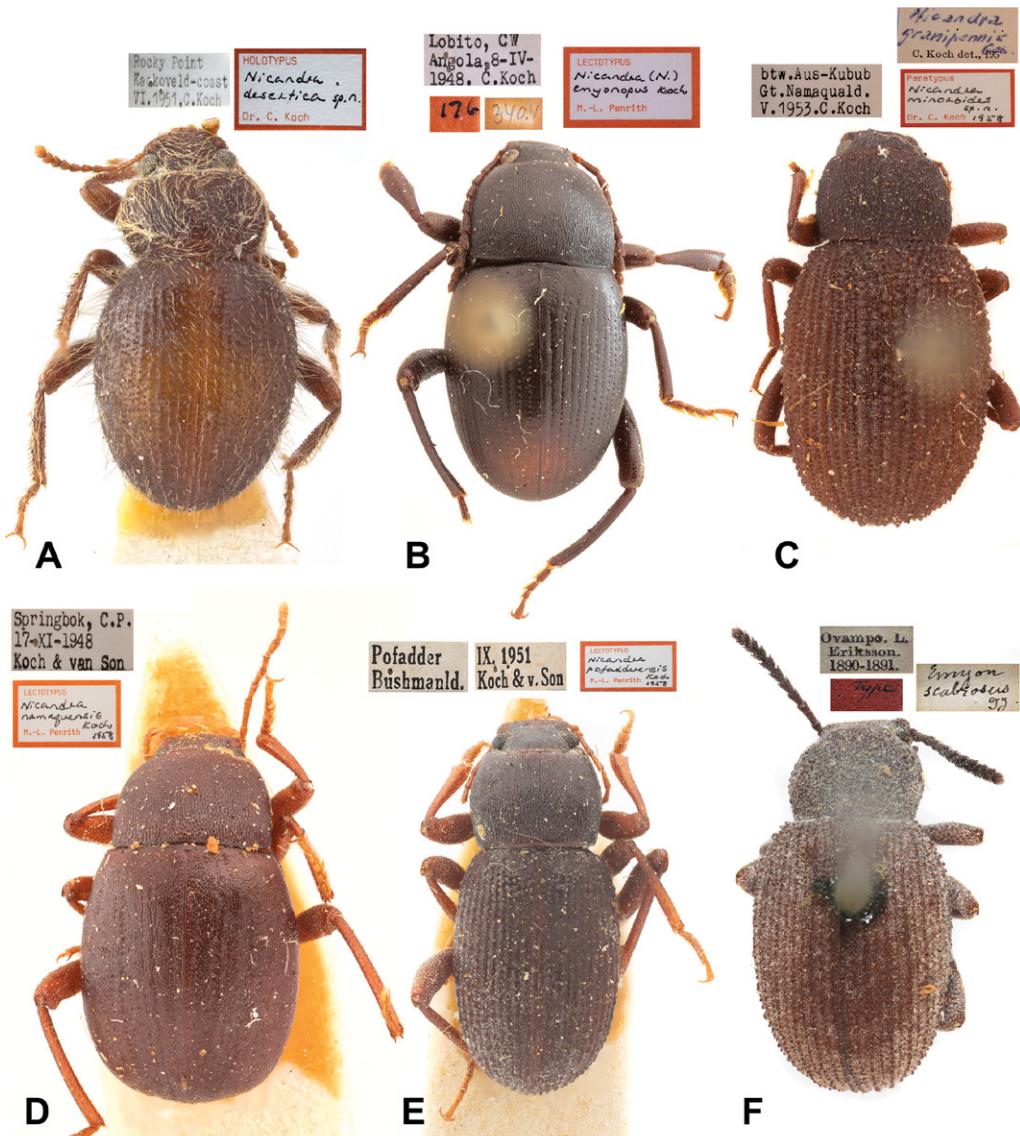


Fig. 15. Type specimens of species representing the genus *Nicandra* subgenus *Nicandra*. A) *N. desertica*, B) *N. emyonopus*, C) *N. minoroides*, D) *N. namaquensis*, E) *N. pofadderensis*, F) *N. scabrosus*.

Distribution. NAMIBIA: Kuibis, Keetmanshoop, Narubis (Gebien 1920).

22. *Nicandra lata* (Gebien, 1920)

Blastarnus latus Gebien 1920: 150.
Nicandra lata: Koch 1958: 151.

Type Data. Holotype, female (Basel Museum): “Kuibis 15.VII.1911 (Michaelsen)”.

Distribution. NAMIBIA: Kuibis (Koch 1958).

23. *Nicandra minoroides* Koch, 1958 (Fig. 15C)

Nicandra minoroides Koch 1958: 167.

Type Data. Syntypes, four specimens (Ditsong Museum): “Western Great Namaqualand: between Aus and Kubub, V.1953, C. Koch & W. Graaf”.

Distribution. NAMIBIA: between Aus and Kubub (Koch 1958).

24. *Nicandra namaquensis* Koch, 1958 (Fig. 15D)
Nicandra namaquensis Koch 1958: 168.

Type Data. Syntypes, 14 specimens (Ditsong Museum), four specimens (Budapest Museum): “Little Namaqualand: Springbok, XI.1948, Univ. California-Transv. Mus. Exped.”; 11 specimens (California Academy of Sciences): “15 m. W. of Garies, XI.1948. Univ. California-Transv. Mus. Exped.”; 28 specimens (Ditsong Museum): “Soebatsfontein. X.1949, Univ. California-Transv. Mus. Exped.”; single specimen (Ditsong Museum): “Kamieskroon, XII.1949. C. Koch”; nine specimens (Ditsong Museum): “Buffels River. near Grootm. XI.1948. Univ. California-Transv. Mus. Exped.”; two specimens (Ditsong Museum): “Steinkopf, XI.1949. C. Koch”; eight specimens (Ditsong Museum): “Anenous Pass, XII.1949, C. Koch”; single specimen (Ditsong Museum): “Wolfberg. XI.1948, Univ. California-Transv. Mus. Exped.”.

Distribution. SOUTH AFRICA: Anenous Pass, Kamieskroon, Soebatsfontein, Springbok, Steinkopf, surroundings of Garies, surroundings of Grootm. Wolfberg (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 15D), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 15D should still be considered a syntype.

25. *Nicandra pofadderensis* Koch, 1958 (Fig. 15E)
Nicandra pofadderensis Koch 1958: 168.

Type Data. Syntypes, 34 specimens (Ditsong Museum): “Bushmanland: Pofadder. IX.1951. C. Koch & G. van Son”; six specimens (Ditsong Museum): “Pella, VIII.1950. C. Koch & G. van Son”.

Distribution. SOUTH AFRICA: Pella, Pofadder (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 15E), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 15E should still be considered a syntype.

26. *Nicandra scabrosa* (Péringuey, 1892) (Fig. 15F)
Emyon scabrosus Péringuey 1892a: 58.
Nicandra scabrosa: Koch 1958: 151.

Type Data. Holotype, unsexed (Ditsong Museum): “Ovampo, Eriksson”.

Distribution. NAMIBIA: Ovampo* (Koch 1958).

27. *Nicandra sinuatipes* Koch, 1958 (Fig. 16A)
Nicandra sinuatipes Koch 1958: 169.

Type Data. Syntypes, 56 specimens (Ditsong Museum): “Little Namaqualand: 15 m. N. of Springbok, XII.1948, Univ. California-Tansv. Mus. Exped”.

Distribution. SOUTH AFRICA: Surroundings of Springbok (Koch 1958).

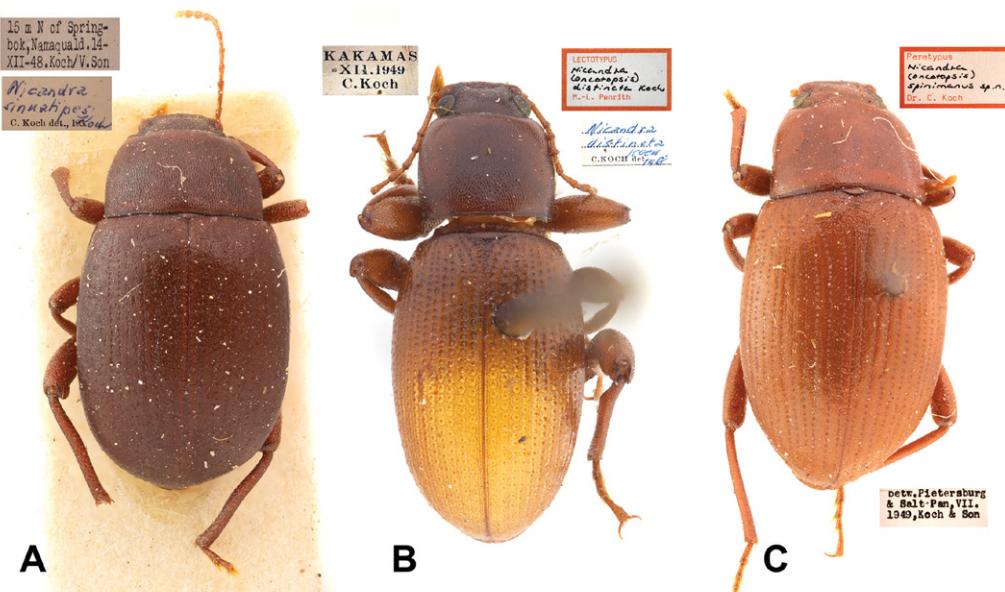


Fig. 16. Type specimens of species representing the genus *Nicandra*. A) Subgenus *Nicandra*: A) *N. sinuatipes*. B–C) Subgenus *Oncotopsis*: B) *N. distincta*, C) *N. spinimanus*.

28. *Nicandra spinulosa* Koch, 1958
Nicandra spinulosa Koch 1958: 167.

Type Data. Holotype, female (Ditsong Museum): “Western Great Namaqualand: 10 m. W. of Haalenberg, IX.1950, C. Koch & G. van Son”.

Distribution. NAMIBIA: Surroundings of Haalenberg (Koch 1958).

Subgenus *Oncotopsis* Koch, 1958

Oncotopsis Koch 1958: 152 [feminine].

Type Species. *Nicandra bicolor* Kulzer, 1951; by original designation.

29. *Nicandra bicolor* Kulzer, 1951

Nicandra bicolor Kulzer 1951: 566.

Type Data. Syntypes, not specified (Basel Museum), two specimens (Budapest Museum): “Sheldon-Grahamstown Eastern Cape Prov. 8.1.50 leg. Zumpt”.

Distribution. SOUTH AFRICA: Grahamstown (Kulzer 1951).

30. *Nicandra distincta* Koch, 1958 (Fig. 16B)
Nicandra distincta Koch 1958: 182.

Type Data. Syntypes, five specimens (Ditsong Museum): “Bushmanland: Kakamas, XII.1949, C. Koch”.

Distribution. SOUTH AFRICA: Kakamas, Kenhardt (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 16B), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 16B should still be considered a syntype.

31. *Nicandra rufipennis* Kulzer, 1951
Nicandra rufipennis Kulzer 1951: 569.

Type Data. Syntypes, four males and female (Munich Museum): “Cap (Kraatz)”.

Distribution. SOUTH AFRICA* (Kulzer 1951).

32. *Nicandra spinimana* Koch, 1958 (Fig. 16C)
Nicandra spinimanus Koch 1958: 185.

Type Data. Syntypes, male and two females (Ditsong Museum): “btwn. Pietersburg and Salt Pan, Zoutpansberg District, VII.1949, C. Koch & G. van Son”.

Distribution. SOUTH AFRICA: between Pietersburg and Salt Pan (Koch 1958).

Genus *Oncopteryx* Gebien, 1943

Oncopteryx Gebien 1943: 905 [masculine], replacement name.

= *Oncopterus* Fairmaire 1887: 178 [junior homonym of *Oncopterus* Steindachner, 1875 (Vertebrata: Pisces)].

Type Species. *Oncopterus acantholopus* Fairmaire, 1887; by monotypy.

1. *Oncopteryx acantholopus* (Fairmaire, 1887)

Oncopterus acantholopus Fairmaire 1887: 178.
Oncopteryx acantholopus: Gebien 1943: 905.

Type Data. Holotype, not specified (Paris Museum): “Uzagara”.

Distribution. KENYA: between Voi and Mombasa (Ferrer 1996); TANZANIA: Usagara (Fairmaire 1887).

Genus *Piscicula* Robiche, 2004

Piscicula Robiche 2004b: 736 [feminine].

Type Species. *Piscicula sprechareae* Robiche, 2004; by monotypy.

1. *Piscicula sprechareae* Robiche, 2004

Piscicula sprechareae Robiche 2004b: 738.

Type Data. Holotype, male (Robiche coll.): “Botswana, 50 km N.O. Selebi Phikwe, 15.XII.2001 (K. Werner leg.)”. **Paratype**, female (Basel Museum): “Botswana, Bechuanaland, Tsessebe, XII.1955 (Zumpt leg.)”.

Distribution. BOTSWANA: Surroundings of Selebi Phikwe, Tshesebe (Robiche 2004b).

Genus *Psectes* Hesse, 1935

Psectes Hesse 1935: 572 [masculine].

Type Species. *Psectes bechuanus* Hesse, 1935; by original designation.

1. *Psectes bechuanus* Hesse, 1935 (Fig. 17A)

Psectes bechuanus Hesse 1935: 573.

Type Data. Syntypes, two males and female (Ditsong Museum): “Tsotsoroga Pan, 17. vi.-9. vii. 30”.

Distribution. BOTSWANA: Tsotsoroga Pan (Hesse 1935).

2. *Psectes borealis* Kaszab, 1971

Psectes borealis Kaszab 1971: 236.

Type Data. Holotype, male (Budapest Museum): “Katesh, S of Mt. Hanang, 5900 feet, 26. VI.— 1. VII. 1965”.

Distribution. TANZANIA: Katesh (Kaszab 1971).

3. *Psectes criberrimus* (Koch, 1958), new combination (Fig. 17F)

Diestecopus criberrimus Koch 1958: 186.

Type Data. Syntypes, male and female (Ditsong Museum): “Angola: Ongueria, VI.1954, Vernay-Transv. Mus. Exped.”.

Distribution. ANGOLA: Ongueria (Hungúeria), Vanavelombe Mountain* (Koch 1958).

Note. Based on investigation of type material this species is here transferred to *Psectes*.

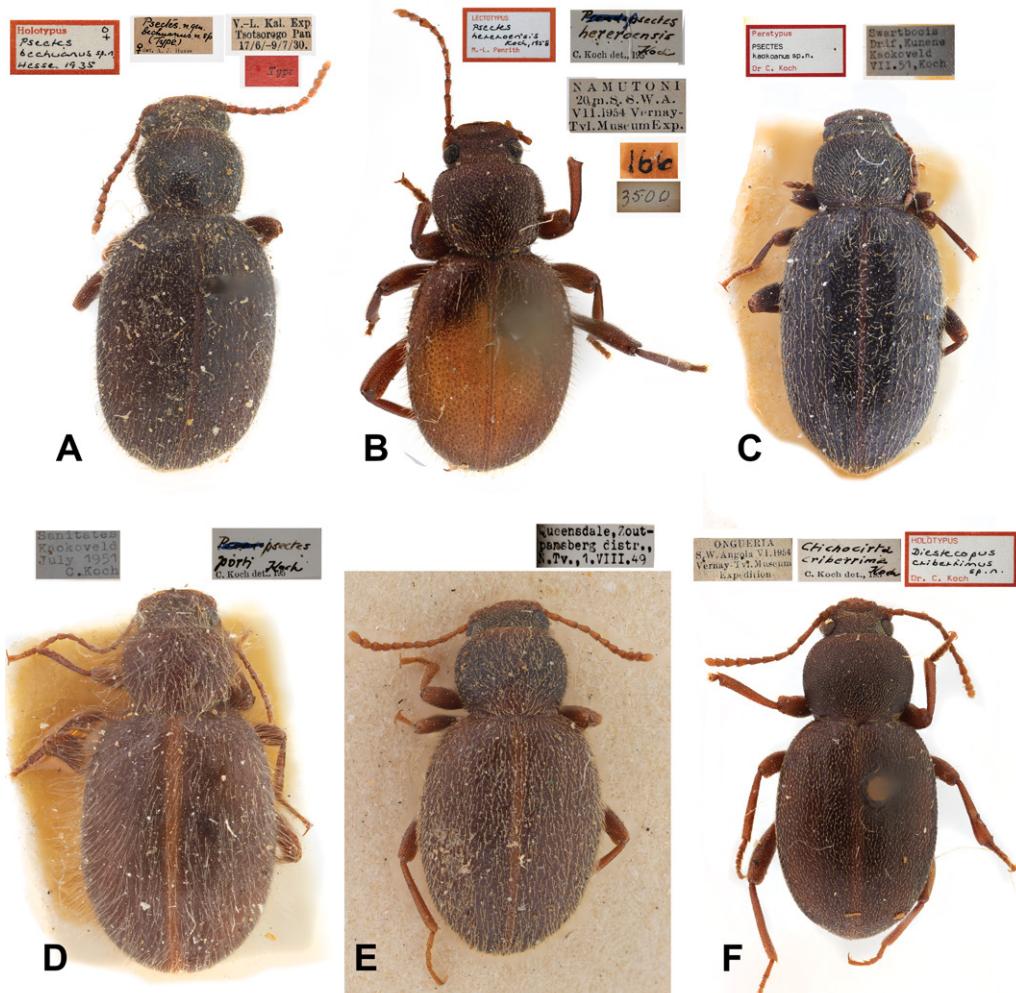


Fig. 17. Type specimens of species representing the genus *Psectes*. A) *P. bechuanus*, B) *P. hereroensis*, C) *P. kaokoanus*, D) *P. porti*, E) *P. transvaalensis*, F) *P. criberrimus*.

4. *Psectes hereroensis* Koch, 1958 (Fig. 17B)

Psectes hereroensis Koch 1958: 198.

Type Data. Syntypes, male and female (Ditsong Museum): “Damaraland: 20 m. S. of Narnutoni, VII.1954, Vernay-Transv. Mus. Exped.”; six specimens (Lund Museum): “Outjo, VI.1951, P. Brink & G. Rudebeck”; two females (Ditsong Museum): “Otavi, VII.1954, Vernay-Tranv. Mus. Exped.”.

Distribution. NAMIBIA: surroundings of Namutoni, Otavi, Outjo (Koch 1958).

Note. Although the studied type specimen is labeled as a lectotype (Fig. 17B), this designation was never formalized in publication. Therefore, the specimen presented on Fig. 17B should still be considered a syntype.

5. *Psectes kaokoanus* Koch, 1958 (Fig. 17C)

Psectes kaokoanus Koch 1958: 197.

Type Data. Syntypes, 81 specimens (Ditsong Museum), five specimens (Budapest Museum): “Kaokoveld: Swartboois Drift. VII.1951, B. Carp Exped.”; 11 specimens (Ditsong Museum): “Ehomba Mountains. VII.1951, B. Carp Exped.”; 17 specimens (Ditsong Museum): “Ohopoho. VII.1951, B. Carp Exped.”; male (Ditsong Museum): “Kaoko-Otavi, VII.1951. B. Carp Exped.”; male (Harvard Museum): “Kaukau - Kungveld: Gautscha Pan, IX.1951, Harvard-Transv. Mus. Exped.”; male (Ditsong Museum): “Damaraland: Windhoek, E. Scherz”.

Distribution. ANGOLA: Ehomba Mountains, Swartbooisdrift (Koch 1958); NAMIBIA: Kaoko

Otavi, Gautscha Pan, Opuwo, Windhoek (Koch 1958).

6. *Psectes porti* Koch, 1958 (Fig. 17D)
Psectes porti Koch 1958: 200.

Type Data. Syntypes. 13 specimens (Ditsong Museum): “Kaokoveld: Swartboois Drift. VII.1951. B. Carp Exped.”.

Distribution. ANGOLA: Swartbooisdrift (Koch 1958).

7. *Psectes transvaalensis* Koch, 1958 (Fig. 17E)
Psectes transvaalensis Koch 1958: 200.

Type Data. Syntypes, eight specimens (Ditsong Museum): “Northern Transvaal: Queensdale, Zoutpansberg District, VIII.1949, C. Koch & C. van Son”.

Distribution. SOUTH AFRICA: Queensdale (Koch 1958).

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REFERENCES CITED

- Agassiz, L. 1846.** *Nomenclatoris zoologici index universalis, continens nomina systematica classium, ordinum, familiarum et generum animalium omnium, tam viventium quam fossilium, secundum ordinem alphabeticum unicum disposita, adjectis homonymiis plantarum, nec non variis adnotacionibus et emendationibus.* Jent & Gassmann, Soloduri [= Solothurn, Switzerland], viii + 393 pp.
- Ancey, C. M. F. 1881.** Description de coléoptères nouveaux d'Aden. *Le Naturaliste* 1: 397.
- Ancey, C. M. F. 1882.** Contributions à la faune de l'Afrique Orientale. *Le Naturaliste* 2: 54–55.
- Ardoine, P. 1979.** Mission Balachowsky-Menier dans l'ancien Territoire français des Afars et des Issas. Coleoptera Tenebrionidae. *Bulletin de la Société Entomologique de France* 84: 58–61.
- Baudi di Selve, F. 1881.** Heteromerum species ex Aegypto, Syria et Arabia. *Deutsche Entomologische Zeitschrift* 25: 273–296.
- Blair, K. G. 1933.** Tenebrionidae from central Arabia collected by Major H. Sr. J.B. Philby. *The Entomologist's Monthly Magazine* 69: 4–7.
- Bouchard, P., Y. Bousquet, R. L. Aalbu, M. A. Alonso-Zarazaga, O. Merkl, and A. E. Davies. 2021.** Review of genus-group names in the family Tenebrionidae (Insecta, Coleoptera). *ZooKeys* 1050: 1–633. doi.org/10.3897/zookeys.1050.64217.
- Bouchard, P., J. F. Lawrence, A. Davies, and A. F. Newton. 2005.** Synoptic classification of the world Tenebrionidae (Insecta: Coleoptera) with a review of family-group names. *Annales Zoologici* 55: 499–530.
- Bousquet, Y., and P. Bouchard. 2013.** The genera in the second catalogue (1833–1836) of Dejean's Coleoptera collection. *ZooKeys* 282: 1–219. doi.org/10.3897/zookeys.282.4401.
- Brancsik, K. 1914.** Coleoptera nova. Bericht des Museumsvereines für das Comitat Trencsén 1914: 58–69.
- Dejean, P. F. M. A. 1834.** Catalogue des Coléoptères de la collection de M. le Comte Dejean. [Livraison 3]. Méquignon-Marvis, Paris, 177–256.
- Doyen, J. T., and W. R. Tschinkel. 1982.** Phenetic and cladistic relationships among tenebrionid beetles (Coleoptera). *Systematic Entomology* 7: 127–183.
- Erichson, W. F. 1843.** Beitrag zur Insecten-Fauna von Angola, in besonderer Beziehung zur geographischen Verbreitung der Insecten in Africa. *Archiv für Naturgeschichte* 9: 199–267.
- Fabricius, J. C. 1801.** *Systema eleutherorum secundum ordines, genera, species, adiectis synonymis, locis, observationibus, descriptionibus, Tomus I.* Bibliopolii Academicii Novii, Kiliae, xiv + 506 pp.
- Fähraeus, O. I. 1870.** Coleoptera Caffrariae, annis 1838–1845 a J. A. Wahlberg collecta. Heteromera descriptis. Öfversigt af Kongliga Vetenskaps-Akademiens Förhandlingar 27: 243–358.
- Fairmaire, L. 1882a.** [new taxa]. In: Mission G. Révoil aux Pays Comalis. Faune et Flore. Coléoptères recueillis par MG Révoil chez les Comalis. Descriptions (L. Fairmaire, V. Lansberge, and J. Bourgeois, editors). Challamel Ainé, Paris, iv + 104 pp, 1 pl. [VI–1882].
- Fairmaire, L. 1882b.** Comptes rendus des séances. Rapports, lectures, communications. Bulletin de la Société Entomologique de Belgique, Bruxelles (3ème série) 16: xlili–lvi.
- Fairmaire, L. 1882c.** Diagnoses de Coléoptères Abyssins. *Le Naturaliste* 4: 48–68.
- Fairmaire, L. 1884.** Diagnoses de Coléoptères de l'Afrique Orientale. Comptes-rendus des Séances de la Société Entomologique de Belgique 3(42): LXX–LXXVIII.
- Fairmaire, L. 1887.** Coléoptères des voyages de M. G. Révoil chez les Somalis et dans l'intérieur du Zanguebar. *Annales de la Société Entomologique de France* 7: 69–186.
- Fairmaire, L. 1888a.** Enumération des Coléoptères recueillis par M. le Dr. Hans Schinz dans le Sud de l'Afrique. *Annales de la Société Entomologique de France* 6: 173–202.
- Fairmaire, L. 1888b.** Description of new species of South-African Tenebrionidae. *Transactions of the South African Philosophical Society* 4: 197–199.
- Fairmaire, L. 1891.** Coléoptères de l'Afrique Orientale. Comptes-rendus des Séances de la Société Entomologique de Belgique 4(20): CCLXXIX–CCCVII.

- Fairmaire, L.** 1892. Coléoptères d'Obock. Troisième partie. Revue d'Entomologie (Caen) 11: 77–127.
- Fairmaire, L.** 1893a. Matériaux pour la faune coléoptérique du Sénégal. Annales de la Société Entomologique de France 62: 147–158.
- Fairmaire, L.** 1893b. Note sur quelques Coléoptères des pays Somalis. Annales de la Société Entomologique de Belgique 37: 144–156.
- Fairmaire, L.** 1894a. Coléoptères de l'Afrique intertropicale et Australie (deuxième note). Annales de la Société Entomologique de Belgique 38(6): 314–335.
- Fairmaire, L.** 1894b. Coleopteres de l'Afrique intertropicale et australe. Annales de la Société Entomologique de Belgique 38: 651–679.
- Fairmaire, L.** 1897. Coléoptères nouveaux de l'Afrique intertropicale et australie. Annales de la Société Entomologique de France 66: 109–152.
- Fairmaire, L.** 1899. La faune entomologique du Delagoa. In: Missionnaire avec la collaboration du Prof. E Bugnion (H. A. Junod, editor). Bulletin de la Société Vaudoise des Sciences Naturelles 35: 162–188.
- Ferrer, J.** 1995. Contribution to the knowledge of the Tenebrionidae of Somalia. Frustula Entomologica (N. S.) 18(31): 1–76.
- Ferrer, J.** 1996. Contributions to the knowledge of the Tenebrionidae of Kenya (Insecta: Coleoptera). Frustula Entomologica 19: 51–109.
- Ferrer, J.** 1999. Contribución al conocimiento del género *Drosochrus* Erichson sensu Koch 1958 y descripción de una nueva especie de Tanzania. Coleoptera, Tenebrionidae, Drosochrini. La Rivista Piemontese di Storia Naturale 20: 55–60.
- Ferrer, J.** 2004. Tenebrionidae de Namibia, avec descriptions de 12 espèces nouvelles. Mitteilungen aus dem Museum für Naturkunde in Berlin. Zoológische Reihe 80(2): 181–250.
- Gahan, C. J.** 1896. On Coleoptera from Aden and Somaliland. Annals and Magazine of Natural History 6(18): 448–461.
- Gebien, H.** 1910a. Diagnosen der von Dr. Shefield Neave im südlichen Kongo-Gebiet gesammelten Tenebrioniden nebst Beschreibungen neuer Arten aus Deutsch-Ostafrika. Annales de la Société Entomologique de Belgique 54: 144–182.
- Gebien, H.** 1910b. Pars 15. Tenebrionidae I [pp. 1–166]. In: Coleopterorum Catalogus, Volumen XVIII (S. Schenkling, editor). W. Junk, Berlin.
- Gebien, H.** 1910c. 7. Coleoptera. 19. Tenebrionidae [pp. 363–396]. In: Wissenschaftliche Ergebnisse der Schwedischen Zoologischen Expedition nach dem Kilimandjaro, dem Meru und den umgebenden Massaisteppen Deutsch-Ostafrikas 1905–1906 unter Leitung von Prof. Dr. Yngve Sjöstedt. 1. Band. Abteilung 7. Coleoptera (Y. Sjöstedt, editor). Stockholm, 442 pp. www.biodiversitylibrary.org/page/1371416#page/827.
- Gebien, H.** 1911. Pars 15. Tenebrionidae III [pp. 355–585]. In: Coleopterorum Catalogus, Volumen XXVIII (S. Schenkling, editor). W. Junk, Berlin.
- Gebien, H.** 1920. Käfer aus der Familie Tenebrionidae gesammelt auf der "Hamburger deutsch-südwestafrikanischen Studienreise 1911". Hamburgische Universität Abhandlungen aus der Auslandskunde Band 5. Reihe C Naturwissenschaften Band 2. L. Friederichsen & Co., Hamburg, 168 pp.
- Gebien, H.** 1921 ["1920"]. Die Tenebrioniden Westafrikas. Archiv für Naturgeschichte 86(A): 1–256.
- Gebien, H.** 1937. Ueber neue Tenebrioniden Ostafrikas aus den Sammlungen des Museo Civico di Storia Nautrale di Trieste. Atti del Museo Civico di Storia Naturale di Trieste 14(2): 21–56.
- Gebien, H.** 1943. Katalog der Tenebrioniden (Col. Heteromera). Teil 3. Mitteilungen der Münchener Entomologischen Gesellschaft 33: 839–926 (810–841).
- Gemminger, M., and E. von Harold.** 1870. Catalogus coleopterorum hucusque descriptorum synonymicus et systematicus. Tom. VII. Tenebrionidae, Nilionidae, Pythidae, Melandryidae, Lagriidae, Pedilidae, Anthicidae, Pyrochroidae, Mordellidae, Rhipiphoridae, Cantharidae, Oedemeridae. E. H. Gummi, Monachium [= München], 1801–2179.
- Gerstaeker, C. E. A.** 1854. Bearbeitung der diagnosen der Von Peters in Mossambique gesammelten käfer und Hymenopteren, aus der familie der Melasomen. Monastbericht der Deutschen Akademie der Wissenschaften zu Berlin 1854: 530–534.
- Gerstaeker, C. E. A.** 1871. Beitrag zur Insektenfauna von Zanzibar. III. Coleoptera. Archiv für Naturgeschichte 37(1): 42–86.
- Gerstaeker, C. E. A.** 1884 ["1883"]. Bestimmung der von Herrn Dr. G. A. Fischer während seiner Reise nach dem Massai-Land gesammelten Coleopteren. Jahrbuch der Hamburgischen Wissenschaftlichen Anstalten 1: 43–63.
- Gestro, R.** 1873. Note sopra alcuni coleotteri appartenenti alle collezione del Museo di Genova. Annali del Museo Civico di Storia Naturale di Genova 4: 353–365. www.biodiversitylibrary.org/page/7915473.
- Gestro, R.** 1878. Diagnosi di alcune specie nuove di Coleotteri dell'Abssinia e del paese dei Somali. Annali del Museo Civico de Storia Naturale di Genova 13: 318–322.
- Gridelli, E.** 1939a. Coleotteri dell'Africa orientale italiana 8. contributo. Revisione delle species riferite dagli autori ai generi *Helopinus*, *Drosochrus*, *Emyon* (Coleopt. Tenebrionidae). Atti del Museo Civico di Storia Naturale Trieste 14: 187–286.
- Gridelli, E.** 1939b. Coleotteri dell'Africa Orientale Italiana. 9e contributo. Revisione delle specie dell'Africa Orientale francese ed inglese riferite dagli autori al genere *Micrantereus*. Memorie della Società Entomologica Italiana 18: 5–52.
- Guérin-Méneville, F. E.** 1834. Matériaux pour une classification des mélasomes. (Extraits d'une monographie de cette famille). Magasin de Zoologie 4: 1–39.
- Haag-Rutenberg, G.** 1871. Beiträge zur Familie der Tenebrioniden (II. Stück). Coleopterologische Hefte 7: 21–111.
- Haag-Rutenberg, G.** 1875. Beiträge zur Familie der Tenebrioniden (V. Stück). Coleopterologische Hefte 14: 67–92.
- Haag-Rutenberg, G.** 1879. Fernere Nachträge zu den Heteromeren-Monographien der Moluriden, Eurychoriden und Adesmiden. Deutsche Entomologische Zeitschrift 23: 289–296.

- Hesse, A. J.** 1935. Scientific results of the Vernay-Lang Kalahari Expedition, March to September, 1930. Tenebrionidae (Coleoptera). Annals of the Transvaal Museum 16: 525–579.
- ICZN (The International Commission on Zoological Nomenclature).** 1999. International Code of Zoological Nomenclature. Fourth Edition. The International Trust of Zoological Nomenclature, London, xxix + 306 pp.
- Kamiński, M. J.** 2020. Distribution of the subtribe Helopinina (Tenebrionidae: Blaptinae). Harvard Dataverse. doi.org/10.7910/DVN/OBNL8G.
- Kamiński, M. J., and Iwan, D.** 2017. Revision of the subtribe Pedinina (Tenebrionidae: Pedinini). Annales Zoologici 67(3): 585–607. doi.org/10.3161/00034541ANZ2017.67.3.006.
- Kamiński, M. J., K. Kanda, R. Lumen, A. D. Smith, and D. Iwan.** 2019a. Molecular phylogeny of Pedinini (Coleoptera, Tenebrionidae) and its implications for higher-level classification. Zoological Journal of the Linnean Society 185: 77–97. doi.org/10.1093/zoolinnean/zly033.
- Kamiński, M. J., R. Lumen, K. Kanda, D. Iwan, M. A. Johnston, G. Kergoat, P. Bouchard, X.-L. Bai, X.-M. Li, G.-D. Ren, and A. D. Smith.** 2021. Reevaluation of Blapimorpha and Opatrinae: Addressing a major phylogeny-classification gap in darkling beetles (Coleoptera: Tenebrionidae: Blaptinae). Systematic Entomology 46(1): 140–156. doi.org/10.1111/syen.12453.
- Kamiński, M. J., R. Lumen, M. Kubicz, W. Steiner, Jr., K. Kanda, and D. Iwan.** 2019b. Immature stages of beetles representing the ‘Opatrinoïd’ clade (Coleoptera: Tenebrionidae): An overview of current knowledge of the larval morphology and some resulting taxonomic notes on Blapstinina. Zoomorphology 138: 349–370. doi.org/10.1007/s00435-019-00443-7.
- Kamiński, M. J., and C. S. Schoeman.** 2018. Taxonomic revision of a darkling beetles genus *Anaxius* (Tenebrionidae: Pedinini: Helopinina). Zootaxa 4455: 471–485.
- Kaszab, Z.** 1971. The scientific results of Hungarian Zoological Expedition to Tanganyika, 15. Coleoptera: Tenebrionidae. Annales Historico-Naturales Musei Nationalis Hungarici Pars Zoologica 63: 225–238.
- Kaszab, Z.** 1972. Missione 1965 del Prof. Giuseppe Scortecci nello Yemen (Arabia meridionale) Coleoptera Tenebrionidae includendo materiale di viaggi nello Yemen del Sig. Ing.-Agr. A. Szalay-Marzso (1969–1971). Atti della Società Italiana di Scienze Naturali e del Museo Civico di Storia Naturale di Milano 113: 366–384.
- Koch, C.** 1958. Tenebrionidae of Angola. Publicações Culturais da Companhia de Diamantes de Angola 39: 11–231.
- Koch, C.** 1962. Vierter taxonomischer Beitrag zur Kenntnis der Tenebrioniden Somalias: Über die von Prof. G. Scortecci 1953 und 1957 in der Miguurtinia-Provinz gesammelten Arten. Atti della Società Italiana di Scienze Naturali e del Museo Civico di Storia Naturale 101: 237–270.
- Koch, C.** 1963. Fünfter taxonomischer Beitrag zur Kenntnis der Tenebrioniden Somalias (1): Über neue *Micrantereus* aus der Sammlung des Museums Triest. Atti del Museo Civico di Storia Naturale di Trieste 23: 165–173.
- Koch, C.** 1965. Missione 1962 del Prof. Giuseppe Scortecci nell’ Arabia meridionale Coleoptera Tenebrionidae Includendo materiale di viaggi in Arabia del Sig. G. Popov (1962) e del Dr. G. Benardelli (1962–63). Atti della Società Italiana di Scienze Naturali e del Museo Civico di Storia Naturale 104: 99–154.
- Kulzer, H.** 1951. Über die Gattung *Nicandra* Fairm. (Helopinini) und einige Neue Arten derselben. Neue Arten der Gattung *Nicandra*. Entomologische Arbeiten aus dem Museum G. Frey 2: 563–573. www.biodiversitylibrary.org/item/172545#page/913/mode/1up.
- Lacordaire, J. T.** 1859. Histoire naturelle des insectes. Genera des Coléoptères ou exposé méthodique et critique de tous les genres proposés jusqu’ici dans cet ordre d’insectes. Tome cinquième contenant les familles des ténebrionides, cistélides, nilionides, pythides, méländryides, lagriides, pédilides, anthicides, pyrochroïdes, mordellides, rhipiphorides, stylopides, meloides et oedémérides. Première partie. Librairie Encyclopédique de Roret, Paris, pp. 1–400.
- Lacordaire, J. T.** 1876. Histoire naturelle des insectes. Genera des coléoptères ou exposé méthodique et critique de tous les genres proposés jusqu’ici dans ce ordre d’Insectes. Atlas. Librairie Encyclopédique de Roret, Paris, 47 pp., 134 pls.
- Lucas, R.** 1920. Catalogus alphabeticus generum et subgenerum Coleopterorum orbis terrarum totius (famili., trib., subtr., sect. incl.). Pars I. R. Stricker, Berlin, xxxi, 696 pp.
- Merkl, O., G. Aranka, and S. Zoltán.** 2015. Type catalogue of darkling beetles (Tenebrionidae) preserved in the Hungarian Natural History Museum. Hungarian Natural History Museum, 735 pp., 96 pls.
- Müller, C. L.** 1887. Vierzehn neue Heteromeren von Bradshaw im Zambezi-Gebiete aufgefunden, mit Abbildungen von van de Poll. Tijdschrift voor Entomologie 30: 297–308.
- Pascoe, F. P.** 1882. IV.—Notes on Coleoptera, with descriptions of new genera and species—Part IV. Journal of Natural History 9: 25–37. doi.org/10.1080/00222938209458985.
- Péringuey, L. A.** 1892a. Third contribution to the South-African coleopterous fauna. Transactions of the South African Philosophical Society 6: 1–94.
- Péringuey, L. A.** 1892b. Fourth contribution to the South African coleopterous fauna. Transactions of the South African Philosophical Society 6: 95–135.
- Péringuey, L. A.** 1896. Descriptions of new genera and species of Coleoptera from South Africa, chiefly from Zambezia. Transactions of the Entomological Society of London 1896: 149–189.
- Péringuey, L. A.** 1899. Fifth contribution to the South-African coleopterous fauna. Annals of the South African Museum 1: 240–330.
- Péringuey, L. A.** 1904a. Sixth contribution to the South African coleopterous fauna. Annals of the South African Museum 3: 167–300.
- Péringuey, L. A.** 1904b. Sixth contribution of the South African Coleopterous fauna. Description of new

- species of Coleoptera in the collection of the South African Museum. Annals of the South African Museum 3: 167–299.
- Quedenfeldt, G. 1885.** Verzeichniss der von Herrn Major D. von Mechow in Angola und am Quango-Strom 1878–1881 gesammelten Tenebrioniden und Cisteliden. Berliner Entomologische Zeitschrift 29(1): 1–38.
- Reitter, E. 1917.** Bestimmungs-Schlüssel für die Unterfamilien und Tribus der paläarktischen Tenebrionidae [81. Heft]. Wiener Entomologische Zeitung 36(3/5): 51–66, (9/10): 296 [Berichtigung].
- Robiche, G. 2001.** Contribution à l'étude de la famille Tenebrionidae de la région afrotropicale. Description de deux nouvelles espèces appartenant au genre *Oncosoma* Westwood, 1842 (Coleoptera, Tenebrionidae). Lambillionea 101: 77–80.
- Robiche, G. 2003.** Description d'une nouvelle espèce du Kenya, appartenant au genre *Oncosoma* Westwood, 1842. Entomologia Africana 8(1): 15–18.
- Robiche, G. 2004a.** Contribution à la connaissance de la tribu Drosochrini, genre *Oncosoma* Westwood, 1842: Description d'un nouveau sous-genre, *Strophia*, et de quatre nouvelles espèces afrotropicales (Coleoptera, Tenebrionidae). Bulletin de la Société Entomologique de France 109: 129–140.
- Robiche, G. 2004b.** Contribution à la connaissance de la tribu Drosochrini: Description d'un nouveau genre et de son espèce. Lambillionea 104: 736–741.
- Robiche, G. 2005.** Rectification homonymique du sous-genre *Strophia* (Col., Tenebrionidae). Bulletin de la Société Entomologique de France 110(3): 358.
- Robiche, G. 2008a.** Contribution à l'étude du genre *Drosochrus* Erichson, 1843, au Mozambique (Coleoptera, Tenebrionidae). Bulletin de la Société Entomologique de France 113(3): 389–395.
- Robiche, G. 2008b.** Contribution à l'étude du genre *Oncosoma* Westwood, 1842 de la région afrotropicale (Coleoptera, Tenebrionidae). Bulletin de la Société Entomologique de France 113(4): 525–532.
- Robiche, G. 2008c.** Description d'une nouvelle espèce appartenant au genre *Oncosoma* Westwood, 1842. (Col. Tenebrionidae). Lambillionea 108(4): 525–528.
- Robiche, G. 2009.** Deux nouvelles espèces appartenant au genre *Oncosoma* Westwood, 1842 (Coleoptera, Tenebrionidae). Entomologia Africana 14: 2–8.
- Robiche, G. 2010.** Nouvelles espèces appartenant au genre *Drosochrus* Erichson, 1843 (Coleoptera, Tenebrionidae). Bulletin de la Société Entomologique de France 115: 367–374.
- Robiche, G. 2012a.** Nouvelles espèces appartenant au genre *Oncosoma* Westwood, 1842 (Coleoptera, Tenebrionidae). Entomologia Africana 17(2): 11–22.
- Robiche, G. 2012b.** Une nouvelle espèce du Kenya appartenant au genre *Micrantereus* Solier, 1848 (Coleoptera, Tenebrionidae). Bulletin de la Société Entomologique de France 117(2): 219–223.
- Robiche, G. 2013.** Un nouveau statut pour le genre *Conophthalmus* Quedenfeldt (Coleoptera, Tenebrionidae, Tenebrioninae). Bulletin de la Société Entomologique de France 118(4): 521–525.
- Robiche, G. 2019.** Une nouvelle espèce de *Micrantereus* Solier, 1848, de la République du Somaliland. Bulletin de la Société Entomologique de France 124(4): 357–360.
- Robiche, G., P. Le Gall, and G. Goergen. 2002.** Contribution à l'étude de la biodiversité des coléoptères Tenebrionidae de la République du Bénin: Premier inventaire. Lambillionea (Supplément 4) 102(4): 381–431.
- Schulze, L. 1968.** The Tenebrionidae of South Africa, XL: Three new Drosochrini (Coleoptera) from Transvaal. Annals of the Transvaal Museum 25: 177–188.
- Smith, A. D., J. S. Wilson, and A. I. Cognato. 2014.** The evolution of Batesian mimicry within the North American Asidini (Coleoptera: Tenebrionidae). Cladistics 31: 441–454. doi.org/10.1111/cla.12101.
- Solier, A. J. J. 1843.** Essai sur les collaptérides de la tribu des Molurites. Imprimerie Royale, Turin, 127 pp. [4 pls.] [extract of Memorie della Reale Accademia delle Scienze di Torino (2)6 [1844]: 213–339].
- Solier, A. J. J. 1848.** Essai sur les collaptérides, 14e tribu Blapsites [pp. 149–370]. In: Studi Entomologici, Volume 2 (F. Baudi and E. Truqui, editors). Stamperia degli Artisti Tipografi, Torino, Italy.
- Westwood, J. O. 1843.** Description of some coleopterous insect from tropical Africa, belonging to the section Heteromera. Proceedings of the Zoological Society of London 10: 117–123. www.biodiversitylibrary.org/item/96442#page/7 mode/1up.

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