



## **Evaluation of Foliar Damage by *Spodoptera frugiperda* (Lepidoptera: Noctuidae) to Genetically Modified Corn (Poales: Poaceae) in Mexico**

Authors: Aguirre, Luis A., Hernández-Juárez, Agustín, Flores, Mariano, Cerna, Ernesto, Landeros, Jerónimo, et al.

Source: Florida Entomologist, 99(2) : 276-280

Published By: Florida Entomological Society

URL: <https://doi.org/10.1653/024.099.0218>

---

BioOne Complete ([complete.BioOne.org](https://complete.BioOne.org)) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at [www.bioone.org/terms-of-use](https://www.bioone.org/terms-of-use).

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

---

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

# Evaluation of foliar damage by *Spodoptera frugiperda* (Lepidoptera: Noctuidae) to genetically modified corn (Poales: Poaceae) in Mexico

Luis A. Aguirre<sup>1,\*</sup>, Agustín Hernández-Juàrez<sup>1</sup>, Mariano Flores<sup>1</sup>, Ernesto Cerna<sup>1</sup>, Jerónimo Landeros<sup>1</sup>, Gustavo A. Frías<sup>1</sup>, and Marvin K. Harris<sup>2</sup>

---

## Abstract

The fall armyworm, *Spodoptera frugiperda* Smith & Abbot (Lepidoptera: Noctuidae), is a key pest of corn, *Zea mays* L. (Poales: Poaceae), in Mexico. The development of genetically modified (GM) corn hybrids for resistance to this insect, with the inclusion of several genes coding for proteins Cry1Ab, Vip3Aa20, and mCry3A of *Bacillus thuringiensis* Berliner (Bacillales: Bacillaceae) (*Bt*), offer an alternative to conventional insecticides to control this pest. Resistance to fall armyworms of the GM corn hybrids Agrisure 3000 GT, Agrisure Viptera 3110, and Agrisure Viptera 3111 was evaluated in 4 locations at Sinaloa for a 3 yr period. Damage evaluation showed that the maize hybrids with the *Bt* gene insertion were not affected by the fall armyworm as compared with their respective isolines, which were seriously damaged. The results reaffirm the insect control benefits provided by this technology and provide a baseline for resistance management.

Key Words: *Bacillus thuringiensis*;  $\delta$ -endotoxin; fall armyworm; leaf damage

## Resumen

El gusano cogollero *Spodoptera frugiperda* Smith & Abbot (Lepidoptera: Noctuidae), es la plaga de mayor importancia económica del maíz *Zea mays* L. (Poales: Poaceae) en México. El desarrollo de híbridos de maíz genéticamente modificados para resistencia a este insecto, con la inserción de diversos genes que codifican para las proteínas Cry1Ab, Vip3Aa20 y mCry3A de *Bacillus thuringiensis* Berliner (Bacillales: Bacillaceae) (*Bt*), ofrecen una alternativa a los insecticidas convencionales de control de esta plaga. Se evaluó durante tres años, el daño foliar del gusano cogollero en maíz GM con los híbridos Agrisure 3000 GT, Agrisure Viptera 3110 y Agrisure Viptera 3111 en cuatro localidades del estado de Sinaloa. La evaluación del daño demostró que el maíz con la inserción de genes de *Bt* son eficaces para contrarrestar o no ser afectado por el daño provocado por la plaga, en comparación con sus respectivas líneas convencionales que fueron seriamente dañadas. Los resultados reafirman los beneficios del control de insectos que ofrece esta tecnología y proporciona una línea base para el manejo de la resistencia.

Palabras Clave: *Bacillus thuringiensis*;  $\delta$ -endotoxina; gusano cogollero; daño foliar

---

The fall armyworm, *Spodoptera frugiperda* Smith & Abbot (Lepidoptera: Noctuidae), is indigenous to the American continent (Sena et al. 2003) and has been reported to infest 186 host plant species in North and Central America (Casmuz et al. 2010). Corn, *Zea mays* L. (Poales: Poaceae), is the primary host of economic importance wherever it is grown in Mexico (Sena et al. 2003). Tropical and subtropical areas are most seriously affected (Ortega 1987; Rodríguez & Marín 2008) with losses incurred from post-emergence to maturity (Ortega 1987). Yield losses over 30% are common (Herrera 1979; García-Gutiérrez et al. 2012), and in some cases total crop loss occurs (Silva-Aguayo et al. 2010).

A reliance on chemical control to manage pest populations has become increasingly ineffective as *S. frugiperda* now expresses resistance to several toxicological groups of insecticides (Georghiou & Mellon 1983; Yu 1991; Pacheco-Covarrubias 1993; Morillo & Notz 2001; Yu et al. 2003).

The development of new control techniques led to the elaboration of genetically modified corn hybrids expressing a *Bacillus thuringiensis* Berliner (Bacillales: Bacillaceae) (*Bt*) crystal protein that, when consumed by lepidopterous larvae, proved fatal to the European corn borer, *Ostrinia nubilalis* Hübner (Lepidoptera: Crambidae), the southwestern corn borer, *Diatraea grandiosella* Dyar (Lepidoptera: Crambidae), the sugarcane borer, *Diatraea saccharalis* (F.) (Lepidoptera: Crambidae), the corn earworm, *Helicoverpa zea* Boddie (Lepidoptera: Noctuidae), and *S. frugiperda* (Abel et al. 2000; Castro et al. 2004). A pyramided strategy that combines 2 or more *Bt* genes deployed in the same corn plant is now used to conserve insecticidal efficacy (Burkness et al. 2010; Niu et al. 2013; Yang et al. 2013). Genetically modified corn (GM) hybrids with *Bt* genes have also been developed to resist a wider range of pests within Lepidoptera and Coleoptera (Buntin et al. 2004a,b; Buntin 2008; Duan et al. 2008; Hardke et al. 2011). These hybrids support a

---

<sup>1</sup>Universidad Autónoma Agraria Antonio Narro, Departamento de Parasitología, Calzada Antonio Narro 1923, Buenavista, Saltillo, Coahuila, 25315, México

<sup>2</sup>Texas A&M University, Department of Entomology, College Station, Texas 77843, USA

\*Corresponding author; E-mail: luisaguirreu@yahoo.com.mx

pest management strategy in modern agriculture (Fernandes et al. 2007), although concerns from an economic, scientific, and social standpoint remain in Mexico.

Mexico is the center of origin of over 61 native races of corn (Reyes 1990; Matsuoka 2005; CONABIO 2006; Kato et al. 2009), and there is concern that GM corn could jeopardize those races (Kato-Yamakake 2004; Serratos-Hernández et al. 2004; Turrent et al. 2010); however, Baltazar et al. (2015) suggest that measures such as spatial isolation could minimize contamination risks. More information is needed in Mexico to validate if the Bt Cry proteins of GM corn are effective in controlling the crop pests under various environmental conditions beyond those reported by Aguirre et al. (2015a) to control corn earworm in the state of Sinaloa and by Aguirre et al. (2015b) to control *S. frugiperda* in the state of Tamaulipas. Thus, the objective of this study was to evaluate foliar damage in corn hybrids with Cry1Ab, Vip3Aa20, and mCry3A toxins from *B. thuringiensis* to control *S. frugiperda* larvae in Sinaloa, Mexico, during 3 growing seasons.

## Materials and Methods

Research was conducted at Oso Viejo, El Dorado, and Camalote in the city of Culiacan and the city of Navolato, both in the state of Sinaloa, during the 2011–2013 fall–winter growing seasons. Plots were planted under biosafety conditions, isolated at least 500 m from commercial corn plots and planted at least 21 d later than recommended to

avoid cross-pollination with non-GM hybrids in accordance with government regulations for field tests with GM corn (Halsey et al. 2005; LBOGM 2005).

Three Bt corn hybrids (Agrisure™ 3000 GT with Cry1Ab and mCry3A proteins; Agrisure® Viptera™ 3111 with Cry1Ab, Vip3Aa20, and mCry3A; and Agrisure® Viptera™ 3110 with Cry1Ab and Vip3Aa20) were used in this research and compared with their respective non-GM isolines provided by Syngenta Agro S.A de C.V. de México (San Lorenzo 1009, Primer Piso, Colonia Del Valle, 03100, México, D.F.). The first two hybrids are resistant to Lepidoptera and Coleoptera and the last one is resistant to Lepidoptera.

A randomized complete block design was used in each locality and date. In 2011, Agrisure 3000 GT and Agrisure Viptera 3110, plus their isolines, were planted at Oso Viejo. In addition, each variety had a corresponding treatment that included a foliar insecticide control (see Table 1). There were 4 replicate blocks per treatment, and they were planted on 28 Jan. In 2012, Agrisure Viptera 3111 and Agrisure 3000 GT hybrids, with and without insecticide treatments, were planted on 15 Feb at Navolato. Agrisure Viptera 3111 was planted at El Dorado on 19 Feb, also with and without insecticide applications (see Table 1). Only 3 replicates were planted in these areas. In 2013, Agrisure Viptera 3111 was planted at Camalote and Oso Viejo on 14 and 15 March, respectively, with 3 treatments (GM hybrid, isolate, isolate plus insecticide) and 4 replicates (see Table 1). In addition, experimental plots during the 3 yr period received an insecticide treatment for *S. frugiperda* if plants less than 20 cm tall reached a 10% infestation level, or plants 20

**Table 1.** Insecticide treatments used to evaluate leaf damage by fall armyworms in genetically modified (GM) corn during 2011–2013 fall–winter growing seasons in localities of Culiacan and Navolato, Sinaloa, Mexico.

Year	Hybrid <sup>a</sup>	Locality	Insecticide <sup>b,c</sup>
2011	Agrisure 3000 GT	Oso Viejo	without insecticide application
	Agrisure 3000 GT + ic	Oso Viejo	permethrin—lambda cyhalothrin—emamectin benzoate
	Isoline + ic	Oso Viejo	permethrin—lambda cyhalothrin—emamectin benzoate
	Isoline	Oso Viejo	check
	Agrisure Viptera 3110	Oso Viejo	without insecticide application
	Agrisure Viptera 3110 + ic	Oso Viejo	permethrin—lambda cyhalothrin—emamectin benzoate
	Isoline + ic	Oso Viejo	permethrin—lambda cyhalothrin—emamectin benzoate
	Isoline	Oso Viejo	check
2012	Agrisure Viptera 3111	El Dorado	without insecticide application
	Agrisure Viptera 3111+ ic	El Dorado	permethrin—lambda cyhalothrin—emamectin benzoate
	Isoline + ic	El Dorado	permethrin—lambda cyhalothrin—emamectin benzoate
	Isoline	El Dorado	check
	Agrisure Viptera 3111	Navolato	without insecticide application
	Agrisure Viptera 3111 + ic	Navolato	permethrin—lambda cyhalothrin—emamectin benzoate
	Isoline + ic	Navolato	permethrin—lambda cyhalothrin—emamectin benzoate
	Isoline	Navolato	check
	Agrisure 3000 GT	Navolato	without insecticide application
	Agrisure 3000 GT + ic	Navolato	permethrin—lambda cyhalothrin—emamectin benzoate
Isoline + ic	Navolato	permethrin—lambda cyhalothrin—emamectin benzoate	
Isoline	Navolato	check	
2013	Agrisure Viptera 3111	Camalote	without insecticide application
	Isoline + ic	Camalote	emamectin benzoate
	Isoline	Camalote	check
	Agrisure Viptera 3111	Oso Viejo	without insecticide application
	Isoline + ic	Oso Viejo	emamectin benzoate
	Isoline	Oso Viejo	check

<sup>a</sup>ic = insecticide control

<sup>b</sup>check = isolate without insecticide application

<sup>c</sup>Insecticides were applied at the following rates: permethrin, 400 mL/ha; lambda cyhalothrin, 500 mL/ha; emamectin benzoate, 200 mL/ha.

cm or taller reached a 20% infestation level, at the rate of 2 applications per year (Table 1).

Each experimental plot consisted of 10 rows, each 5 m long, with 0.8 m between rows with a 40 to 50 seed planting density. The seedlings were later thinned to 34 plants per row. The experimental plot was surrounded with a buffer area of 6 rows of conventional corn, and other buffer areas were planted between replicates, which were planted the same time as the experimental material as required by official regulations. Agricultural management of the plot followed the technical guide for corn growers developed by the Instituto Nacional de Investigaciones Forestales, Agrícolas y Pecuarias (INIFAP 2010).

Foliar damage under natural infestation by fall armyworms was evaluated by sampling 10 plants randomly in the 4 central rows at V6–V8 (2011), and V2–V4, V6–V8, and V10–V12 (2012 and 2013) phenological stages of the plant. A numerical scale (1–9), also known as the Davis scale, was used to evaluate foliar feeding damage (Davis et al. 1989, 1992; Mihm 1983) ranging from 1 = no foliar damage (highly resistant) to 9 = severe foliar damage (totally susceptible).

PROC ANOVA and Tukey's multiple rank test ( $P < 0.05$ ) were used to compare among treatments. SAS/STAT (SAS version 9.0; SAS Institute, Cary, North Carolina) software was used to analyze the percentage of injury to plants injured and the damage ratings (Davis scale).

## Results

Genetically modified hybrids at Oso Viejo in 2011 did not show significant ( $P > 0.05$ ) damage by fall armyworms. The isolines with insecticide treatment also had low damage scores, 1.85 and 1.22 (Davis scale) for Agrisure 3000 GT and Agrisure Viptera 3110, respectively, with only some pinhole feeding marks on leaves and no significant differences between treatments ( $P > 0.05$ ). In contrast, isolines without chemical treatment had an average of damage score of 4.20 in both hybrids, which possessed large holes and long lesions on leaves. *Spodoptera frugiperda* did not damage Agrisure Viptera 3110 and Agrisure 3000 GT with insecticide treatment, and 5.0% of plants were damaged in the latter without treatment. The non-GM hybrids displayed damaged plants from 15.0 to 90.0%, including those in which chemical control was used (Table 2).

At El Dorado and Navolato in 2012, results were similar to those in 2011. Agrisure Viptera 3111 and Agrisure 3000 GT, including those with

insecticide treatment, showed little foliar damage (<1.4 on the Davis scale) and few injured plants, with some plants with pinholes and only 13.3% of plants injured in the Agrisure 3000 GT treatment. The isolines of the GM hybrids were significantly different, with 52.2 to 81.1% of plants injured and plant damage that varied from 2.89 to 4.97 on the Davis scale, including large holes and long leaf lesions (Table 3).

In 2013, Agrisure Viptera 3111 did not show feeding signs by *S. frugiperda* at any of the test sites, whereas the isolines were heavily infested including those in which chemical control was applied. Foliar injury was 1.72 to 3.22 on the Davis scale and 35.8 to 74.2% of plants injured, including large lesions (Table 4).

## Discussion

Agrisure 3000 GT, Agrisure Viptera 3110, and Agrisure Viptera 3111 hybrids were resistant to *S. frugiperda* as compared with their severely injured respective isolines during the 3 yr research study in the Sinaloa corn-growing areas. These results are similar to those found by Aguirre et al. (2015b), testing the same *Bt* hybrids in the state of Tamaulipas. Also, Piña & Solleiro (2013) indicated that experimental tests of GM corn in various areas of Mexico are consistently efficacious in controlling key pests.

This research showed that use of GM *Bt* corn hybrids provides season-long protection from *S. frugiperda*. In contrast, chemical control only protects the plant when the insecticide residue is present, and a failure in timing of application(s) represents a risk in control efficacy. Piña & Solleiro (2013) reported that protecting non-*Bt* corn in several areas of Mexico from infestations of corn earworms and fall armyworms required from 3 to 5 insecticide applications per season, and from 720 g to 3.6 kg of active ingredient per ha. In addition, researchers in other countries reported similar results of *Bt* corn with the Cry1Ab toxin for controlling the fall armyworm with respect to conventional hybrids with and without insecticide control (Buntin et al. 2001, 2004a,b; Buntin 2008; Hardke et al. 2011; Huang et al. 2011; Ríos-Díez et al. 2012).

Hybrids with the *Bt* toxin used in this research consistently demonstrated reduction in foliar damage. However, in all areas tested, Agrisure 3000 GT displayed more injured plants and larger lesion size than Agrisure Viptera 3110 and Agrisure Viptera 3111. The higher level of damage is thought to be due to Agrisure 3000 GT having only 1 *Bt* toxin (Cry1Ab) for Lepidoptera control, whereas the other tested hybrids have 2 pyramided *Bt* genes. Multiple genes for resistance are thought to provide better resistance to the pest. Agrisure Viptera 3110 and Agrisure Viptera 3111 have 2 toxins (pyramid events) for Lepidoptera control, the  $\delta$ -endotoxin Cry1Ab and the vegetative insecticide protein Vip3Aa20, which provide excellent protection to the crop, not only from *S. frugiperda* but also from other Lepidoptera (Burkness et al. 2010; Niu et al. 2013; Yang et al. 2013).

Planting dates of experimental plots were at least 21 d later than the recommended date in Sinaloa. This requirement was imposed by regulatory authorities in order to avoid cross pollination with conventional corn in the area. This condition put the experimental plots in this research under high pest pressure, which came from surrounding corn fields and sorghum fields. Despite this high level of pest pressure, the *Bt* toxin in the crop reduced the infestation level and damage. If these GM hybrids were planted on the recommended planting date under optimal conditions to the crop, these *Bt* hybrids could be expected to perform well. Such a corn pest management program would reduce the use of chemical insecticides, allow the crop to better express its genetic potential, and conserve yield and grain quality by decreasing foliar damage.

**Table 2.** Foliar damage and percentage of plants injured by fall armyworms on the genetically modified hybrids Agrisure 3000 GT and Agrisure Viptera 3110 and their respective isolines at Oso Viejo, Culiacan, Sinaloa, in 2011.

Hybrid <sup>a</sup>	Leaf damage <sup>b,c</sup>	Plants injured (%) <sup>c</sup>
Agrisure 3000 GT	1.10 a	5.0 a
Agrisure 3000 GT + ic	1.00 a	0.0 a
Isoline + ic	1.85 a	30.0 b
Isoline	4.20 b	82.5 c
	$F = 17.33^{***}$	$F = 38.58^{***}$
Agrisure Viptera 3110	1.00 a	0.0 a
Agrisure Viptera 3110 + ic	1.00 a	0.0 a
Isoline + ic	1.22 a	15.0 b
Isoline	4.20 b	90.0 c
	$F = 13.22^{***}$	$F = 297.00^{***}$

<sup>a</sup>ic = insecticide control

<sup>b</sup>Mean numerical scale

<sup>c</sup>Genetically modified hybrids and their respective isolines followed by the same letter do not differ significantly (ANOVA and Tukey's test;  $P > 0.05$ ). \*\*\* Indicates significant *F* value at  $P < 0.001$ , *df* = 3,15.

**Table 3.** Foliar damage and percentage of plants injured by fall armyworms on the genetically modified corn hybrids Agrisure Viptera 3111 and Agrisure 3000 GT and their respective isolines at El Dorado and Navolato, Sinaloa, in 2012.

Hybrid <sup>a</sup>	Locality	Leaf damage <sup>b,c</sup>	Plants injured (%) <sup>c</sup>
Agrisure Viptera 3111	El Dorado	1.00 a	0.0 a
Agrisure Viptera 3111 + ic	El Dorado	1.00 a	0.0 a
Isoline + ic	El Dorado	3.53 b	64.4 b
Isoline	El Dorado	2.89 b	56.7 b
		$F = 10.35^{***}$	$F = 16.54^{***}$
Agrisure Viptera 3111	Navolato	1.07 a	1.1 a
Agrisure Viptera 3111 + ic	Navolato	1.07 a	1.1 a
Isoline + ic	Navolato	3.20 b	52.2 b
Isoline	Navolato	4.90 c	81.1 c
		$F = 11.52^{***}$	$F = 14.99^{***}$
Agrisure 3000 GT	Navolato	1.34 a	13.3 a
Agrisure 3000 GT + ic	Navolato	1.13 a	7.8 a
Isoline + ic	Navolato	3.87 b	66.7 b
Isoline	Navolato	4.97 b	77.8 b
		$F = 9.65^{***}$	$F = 10.39^{***}$

<sup>a</sup>ic = insecticide control<sup>b</sup>Mean numerical scale<sup>c</sup>Genetically modified hybrids and their respective isolines followed by the same letter do not differ significantly (ANOVA and Tukey's test;  $P > 0.05$ ). \*\*\* Indicates significant  $F$  value at  $P < 0.001$ ,  $df = 11,35$ .**Table 4.** Foliar damage and percentage of plants injured by fall armyworms on the genetically modified corn hybrid Agrisure Viptera 3111 and its respective isoline at Camalote and Oso Viejo, Culiacan, Sinaloa, in 2013.

Hybrid <sup>a</sup>	Locality	Leaf damage <sup>b,c</sup>	Plants injured (%) <sup>c</sup>
Agrisure Viptera 3111	Camalote	1.00 a	0.0 a
Isoline + ic	Camalote	1.72 b	35.8 b
Isoline	Camalote	3.22 c	74.2 c
		$F = 48.92^{***}$	$F = 58.44^{***}$
Agrisure Viptera 3111	Oso Viejo	1.00 a	0.0 a
Isoline + ic	Oso Viejo	1.80 b	39.2 b
Isoline	Oso Viejo	2.63 c	62.5 c
		$F = 35.46^{***}$	$F = 27.38^{***}$

<sup>a</sup>ic = insecticide control<sup>b</sup>Mean numerical scale<sup>c</sup>Genetically modified hybrid and the respective isoline treatments followed by the same letter do not differ significantly (ANOVA and Tukey's test;  $P > 0.05$ ). \*\*\* Indicates significant  $F$  value at  $P < 0.001$ ,  $df = 8,35$ .

## Acknowledgments

The authors would like to thank Syngenta Agro S.A de C.V. de México for providing the genetic material that made this research possible.

## References Cited

- Abel CA, Wilson RL, Wiseman BR, White WH, Davis FM. 2000. Conventional resistance of experimental maize lines to corn earworm (Lepidoptera: Noctuidae), fall armyworm (Lepidoptera: Noctuidae), southwestern corn borer (Lepidoptera: Crambidae), and sugarcane borer (Lepidoptera: Crambidae). *Journal of Economic Entomology* 93: 982–988.
- Aguirre LA, Hernández A, Flores M, Frías GA, Cerna E, Landeros J, Harris MK. 2015a. Genetically modified maize resistant to corn earworm (Lepidoptera: Noctuidae) in Sinaloa, Mexico. *Florida Entomologist* 98: 821–826.
- Aguirre LA, Hernández A, Flores M, Pérez-Zubiri R, Cerna E, Landeros J, Frías GA. 2015b. Comparación del nivel de daño de *Spodoptera frugiperda* (Lepidoptera: Noctuidae) en plantas de maíz genéticamente modificado y convencional en el Norte de México. *Southwestern Entomologist* 40: 171–178.
- Baltazar BM, Castro EL, Espinoza BA, de la Fuente MJM, Garzón TJA, González GJ, Gutiérrez MA, Guzmán RJL, Heredia DO, Horak MJ, Madueño MJ, Schapagha WA, Stojšin D, Uribe MHR, Zavala GF. 2015. Pollen-mediated gene flow in maize: implications for isolation requirements and coexistence in Mexico, the center of origin of maize. *PLoS One* 10: e0131549.
- Buntin GD. 2008. Corn expressing Cry1Ab or Cry1F endotoxin for fall armyworm and corn earworm (Lepidoptera: Noctuidae) management in field corn for grain production. *Florida Entomologist* 91: 523–530.
- Buntin GD, Dewey LR, Wilson DM, McPherson RM. 2001. Evaluation of Yieldgard transgenic resistance for control of fall armyworm and corn earworm (Lepidoptera: Noctuidae) on corn. *Florida Entomologist* 84: 37–42.
- Buntin GD, All JN, Lee RD, Wilson DM. 2004a. Plant-incorporated *Bacillus thuringiensis* resistance for control of fall armyworm and corn earworm (Lepidoptera: Noctuidae) in corn. *Journal of Economic Entomology* 97: 1603–1611.
- Buntin GD, Flanders KL, Lynch RE. 2004b. Assessment of experimental Bt events against fall armyworm and corn earworm in field corn. *Journal of Economic Entomology* 97: 259–264.
- Burkness EC, Dively G, Patton T, Morey AC, Hutchison WD. 2010. Novel Vip3A *Bacillus thuringiensis* (Bt) maize approaches high-dose efficacy against *Helicoverpa zea* (Lepidoptera: Noctuidae) under field conditions. *GM Crops* 1: 337–343.
- Casmuz A, Juárez ML, Socías MG, Murúa MG, Prieto S, Medina S, Willink E, Gastaminza G. 2010. Revisión de los hospederos del gusano cogollero del maíz, *Spodoptera frugiperda* (Lepidoptera: Noctuidae). *Revista de la Sociedad Entomológica Argentina* 69: 209–231.
- Castro BA, Leonard BR, Riley TJ. 2004. Management of feeding damage and survival of southwestern corn borer and sugarcane borer (Lepidoptera: Cram-

- bidiae) with *Bacillus thuringiensis* transgenic field corn. *Journal of Economic Entomology* 97: 2106–2116.
- CONABIO (Comisión Nacional para el conocimiento y uso de la Biodiversidad). 2006. Elementos para la determinación de centros de origen y centros de diversidad en general y el caso específico de la liberación experimental de maíz transgénico al ambiente en México. Documento base preparado por la Coordinación Nacional de la CONABIO para la Secretaría de Medio Ambiente y Recursos Naturales (SEMARNAT) y la Secretaría de Agricultura, Ganadería, Pesca y Alimentación (SAGARPA). México, D.F. [http://www.biodiversidad.gob.mx/genes/pdf/Doc\\_CdeOCdeDG.pdf](http://www.biodiversidad.gob.mx/genes/pdf/Doc_CdeOCdeDG.pdf) (last accessed 10 Nov 2014).
- Davis FM, Williams WP, Wiseman BR. 1989. Methods used to screen maize for and to determine mechanisms of resistance to the southwestern corn borer and fall armyworm. *In* Proceedings, Toward Insect Resistant Maize for the Third World. International Symposium on Methodologies for Developing Host Plant Resistance to Maize Insects, 9–14 Mar 1987, International Maize and Wheat Improvement Center (CIMMYT), México, D.F.
- Davis FM, Ng SS, Williams WP. 1992. Visual rating scales for screening whorl-stage corn for resistance to fall armyworm. Mississippi Agricultural & Forestry Experiment Station. Technical Bulletin 186.
- Duan JJ, Teixeira D, Huesing JE, Jiang C. 2008. Assessing the risk to nontarget organisms from *Bt* corn resistant to corn rootworms (Coleoptera: Chrysomelidae): tier-I testing with *Orius insidiosus* (Heteroptera: Anthracoridae). *Environmental Entomology* 37: 838–844.
- Fernandes OA, Faria M, Martinelli S, Schmidt F, Ferreira CV, Moro G. 2007. Short-term assessment of *Bt* maize on non-target arthropods in Brazil. *Scientia Agricola (Piracicaba, Brazil)* 64: 249–255.
- García-Gutiérrez C, González-Maldonado MB, Cortez-Mondaca E. 2012. Uso de enemigos naturales y biorracionales para el control de plagas de maíz. *Ra Ximhai* 8: 57–70.
- Georghiou GP, Mellon RB. 1983. Pesticide resistance in time and space, pp. 1–46 *In* Georghiou GP, Saito T [eds.], *Pest Resistance to Pesticides*. Plenum Press, New York, New York.
- Halsey ME, Remund KM, Davis CA, Qualls M, Eppard PJ, Berberich SA. 2005. Isolation of maize from pollen-mediated gene flow by time and distance. *Crop Science* 45: 2172–2185.
- Hardke JT, Leonard BR, Huang F, Jackson RE. 2011. Damage and survivorship of fall armyworm (Lepidoptera: Noctuidae) on transgenic field corn expressing *Bacillus thuringiensis* Cry proteins. *Crop Protection* 30: 168–172.
- Herrera AJ. 1979. Principales Plagas del Maíz. Boletín Especial de la Dirección de Agricultura y Ganadería del Perú.
- Huang F, Andow DA, Buschman LL. 2011. Success of the high dose/refuge resistance management strategy after 15 years of *Bt* crops in North America. *Entomologia Experimentalis et Applicata* 140: 1–16.
- INIFAP (Instituto Nacional de Investigaciones Forestales, Agrícolas y Pecuarias). 2010. Centro de Investigación Regional del Noreste (CIRNO). Campo Experimental Valle de Culiacán (CEVACU). Maíz, pp. 41–47 *In* Guía técnica para el área de influencia del Campo Experimental Valle de Culiacán. Culiacán, Sinaloa, México.
- Kato TA, Mapes C, Mera LM, Serratos JA, Bye RA. 2009. Origen y diversificación del maíz: una revisión analítica. Universidad Nacional Autónoma de México (UNAM), Comisión Nacional para el Conocimiento y Uso de la Biodiversidad (CONABIO), México.
- Kato-Yamakake TA. 2004. Variedades transgénicas y el maíz nativo en México. *Agricultura, Sociedad y Desarrollo* 1: 101–109.
- LBOGM (Ley de Bioseguridad de Organismos Genéticamente Modificados). 2005. Diario Oficial de la Federación, 18 marzo 2005, México.
- Matsuoka Y. 2005. Origin matters: lessons from the search for the wild ancestor of maize. *Breeding Science* 55: 383–390.
- Mihm JA. 1983. Efficient Mass-Rearing and Infestation Techniques to Screen for Host Plant Resistance to Fall Armyworm, *Spodoptera frugiperda*. International Maize and Wheat Improvement Center (CIMMYT), México, D.F.
- Morillo F, Notz A. 2001. Resistencia de *Spodoptera frugiperda* (Smith) (Lepidoptera: Noctuidae) a lambdacihalotrina y metomil. *Entomotropica* 16: 79–87.
- Niu Y, Meagher Jr RL, Yang F, Huang F. 2013. Susceptibility of field populations of the fall armyworm (Lepidoptera: Noctuidae) from Florida and Puerto Rico to purified Cry1F and corn leaf tissue containing single and pyramided *Bt* genes. *Florida Entomologist* 96: 701–713.
- Ortega AC. 1987. Insectos nocivos del maíz: una guía para su identificación en el campo. Centro Internacional de Mejoramiento de Maíz y Trigo (CIMMYT), México.
- Pacheco-Covarrubias JJ. 1993. Monitoring insecticide resistance in *Spodoptera frugiperda* populations from the Yaqui Valley, Son., Mexico. *Resistant Pest Management, Newsletter* 5: 3–4.
- Piña S, Solleiro JL. 2013. México, pp. 341–408 *In* Solleiro RJJ, Castañón IR [eds.], *Introducción al ambiente del maíz transgénico: Análisis de ocho casos en Iberoamérica*, México. AgroBio México y CambioTec, México.
- Reyes CP. 1990. El maíz y su cultivo. AGT-EDITOR S.A., México.
- Ríos-Díez JD, Siegfried B, Saldamando-Benjumea CI. 2012. Susceptibility of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) strains from central Colombia to Cry1Ab and Cry1Ac entotoxins of *Bacillus thuringiensis*. *Southwestern Entomologist* 37: 281–293.
- Rodríguez DLA, Marín AJ. 2008. Insectos plaga y su control, pp. 29–46 *In* Rodríguez MR, De León C [eds.], *El cultivo del maíz*. Temas selectos 1. Colegio de postgraduados, Mundi Prensa, México.
- Sena Jr DG, Pinto FAC, Queiroz DM, Viana PA. 2003. Fall armyworm damaged maize plant identification using digital images. *Biosystems Engineering* 85: 449–454.
- Serratos-Hernández JA, Islas-Gutiérrez F, Buendía-Rodríguez E, Berthaud J. 2004. Gene flow scenarios with transgenic maize in Mexico. *Environmental Biosafety Research* 3: 149–157.
- Silva-Aguayo G, Rodríguez-Maciel JC, Lagunes-Tejeda A, Landerl-Cázares C, Alatorre-Rosas R, Shelton AM, Blanco CA. 2010. Bioactivity of boldo (*Peumus boldus* Molina) (Laurales: Monimiaceae) on *Spodoptera frugiperda* (J. E. Smith) and *Helicoverpa zea* (Boddie) (Lepidoptera: Noctuidae). *Southwestern Entomologist* 35: 215–231.
- Turrent FA, Cortés FJ, Espinosa CA, Mejía AH, Serratos HJA. 2010. ¿Es ventajosa para México la tecnología actual de maíz transgénico? *Revista Mexicana de Ciencias Agrícolas* 1: 631–646.
- Yang F, Qureshi JA, Leonard BR, Head GP, Niu Y, Huang F. 2013. Susceptibility of Louisiana and Florida populations of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) to pyramided *Bt* corn containing Genuity® VT Double Pro™ and SmartStax™ traits. *Florida Entomologist* 96: 714–723.
- Yu SJ. 1991. Insecticide resistance in the fall armyworm, *Spodoptera frugiperda* (J. E. Smith). *Pesticide Biochemistry and Physiology* 39: 84–91.
- Yu SJ, Nguyen SN, Abo-Elghar GE. 2003. Biochemical characteristics of insecticide resistance in the fall armyworm, *Spodoptera frugiperda* (J. E. Smith). *Pesticide Biochemistry and Physiology* 77: 1–11.