

Mosquito Fauna Associated with Aedes aegypti (Diptera: Culicidae) in Yucatán State of Southeastern México, and Checklist with New Records

Authors: Baak-Baak, Carlos M., Cigarroa-Toledo, Nohemi, Arana-Guardia, Roger, Chim, Wilberth A. Chi, Orilla, Julio A. Chan, et al.

Source: Florida Entomologist, 99(4): 703-709

Published By: Florida Entomological Society

URL: https://doi.org/10.1653/024.099.0420

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Mosquito fauna associated with *Aedes aegypti* (Diptera: Culicidae) in Yucatán State of southeastern México, and checklist with new records

Carlos M. Baak-Baak¹, Nohemi Cigarroa-Toledo¹, Roger Arana-Guardia¹, Wilberth A. Chi Chim¹, Julio A. Chan Orilla¹, Carlos Machain-Williams¹, Oswaldo M. Torres-Chable², Aldo I. Ortega-Morales³, David A. Moo-Llanes⁴, Armando Elizondo-Quiroga⁵, and Julián E. García-Rejón^{1,*}

Abstract

As part of our routine surveillance for arboviruses transmitted by *Aedes (Stegomyia) aegypti* (L.) (Diptera: Culicidae) in Yucatán State, México, we regularly monitor the mosquito fauna that occurs in the region. In this study, immature and adult mosquitoes were collected at 60 study sites (including residential premises, churches, tourist areas, cemeteries, forested areas, and garbage dump sites) in Yucatán State from Jul 2014 to Dec 2015. In total, 4,062 larvae, 344 pupae, and 3,812 adults representing 7 genera and 27 species were collected. Three species had never before been identified in Yucatán State, and these are *Aedes (Ochlerotatus) euplocamus* Dyar & Knab, *Aedes (Protomacleaya) podographicus* Dyar & Knab, and *Culex (Culex) declarator* Dyar & Knab. The identification of these 3 species brings the current total of mosquito species recognized in Yucatán State to 52 (with 11 genera). Many of the mosquitoes collected in this study were temporally and spatially associated with *Ae. aegypti*.

Key Words: Aedes euplocamus; Aedes podographicus; Culex declarator

Resumen

Como parte de la vigilancia de arbovirus transmitidos por *Aedes (Stegomyia) aegypti* L., en el estado de Yucatán, México; regularmente monitoreamos la fauna de mosquitos presentes en la región. En este estudio, los estadios de inmaduras y adultos de mosquitos fueron colectados en 60 localidades en el estado de Yucatán durante julio de 2014 y diciembre de 2015. Un rango amplio de lugares urbanos y rurales fueron examinados, incluyendo casas, iglesias, áreas turísticas, cementerios, selva y un basurero municipal. Un total de 4,062 larvas, 344 pupas y 3,812 adultos fueron colectados, representando siete géneros y 27 especies. Tres especies fueron identificadas como nuevas para la fauna de mosquitos de Yucatán: *Aedes (Ochlerotatus) euplocamus* Dyar and Knab, *Aedes (Protomacleaya) podographicus* Dyar and Knab y *Culex (Culex) declarator* Dyar and Knab. Adicionando estas tres especies, actualmente hay 52 especies (con 11 géneros) de mosquitos en Yucatán. Muchos de los mosquitos colectados en el presente estudio fueron temporal y espacialmente asociados con *Ae. aegypti*.

Palabras Clave: Aedes euplocamus; Aedes podographicus; Culex declarator

Aedes (Stegomyia) aegypti (L.) (Diptera: Culicidae) is the main vector of dengue and chikungunya viruses, both of which have been associated with human disease in Yucatán State, México (García-Rejón et al. 2008; Diaz-Gonzalez et al. 2015). Aedes aegypti mosquitoes are common in Yucatán State, and larvae and pupae are often found inside disposable containers, buckets, tires, flower pots, vases, and storm-

water drains/catch basins where present (Winch et al. 1992; García-Rejón et al. 2011; Arana-Guardia et al. 2014). *Aedes aegypti* occurs in a diverse range of habitats including residential premises, vacant lots, parking lots, and cemeteries (Baak-Baak et al. 2014a,b).

Immatures and adults of *Ae. aegypti* usually cohabit with other mosquito species (Baak-Baak et al. 2014b). During surveillance activi-

¹Laboratorio de Arbovirología. Centro de Investigaciones Regionales "Dr. Hideyo Noguchi". Universidad Autónoma de Yucatán, 97069, Mérida, Yucatán, México. Calle 43 No. 613 x Calle 90 Colonia Inalámbrica; E-mail: carlos.baak@correo.uady.mx (C. M. B.-B.), nohemi.cigarroa@gmail.com (N. C.-T.), roger.arana.guardia@gmail.com (R. A.-G.), willynet.a@gmail.com (W. A. C. C.), chan-orilla-julio@hotmail.com (J. A. C. O.), carmachain@gmail.com (C. M.-W.), julian.garcia@correo.uady.mx (J. E. G.-R) ²Laboratorio de Enfermedades Tropicales y Transmitidas por Vector, Universidad Juárez Autónoma de Tabasco. División Académica de Ciencias Agropecuarias. Carretera Villahermosa – Teapa. Km 25, México; E-mail: oswaldo.torres.chable@gmail.com (O. M. T.-C.)

Departamento de Parasitología, Universidad Autónoma Agraria "Antonio Narro" Unidad Laguna, Torreón Coahuila, México. CP 27084; E-mail: agrortega@hotmail.com (A. I. O.-M.)

⁴Centro Regional de Investigación en Salud Pública, Instituto Nacional de Salud Pública. Calle 19 Poniente esquina 4ta Norte. Tapachula, Chiapas, México, CP 30700; E-mail: davidmooll@gmail.com (D. A. M.-L.)

⁵CONACYT Research Fellow – Centro Regional de Investigación en Salud Pública, Instituto Nacional de Salud Pública. Calle 19 Poniente esquina 4ta Norte.

Tapachula, Chiapas, México, CP 30700; E-mail: aelizondog@gmail.com (A. E.-Q.)

^{*}Corresponding author; E-mail: julian.garcia@correo.uady.mx (J. E. G.-R)

ties for dengue virus vectors, other mosquito species that are collected are not identified or registered. Approximately 18 to 20 genera and 225 to 247 species of mosquitoes are known to occur in México (Bond et al. 2014). Several species reported in México are important vectors of arboviruses that affect humans and domestic animals. For example, West Nile virus was detected in Culex quinquefasciatus Say in Nuevo León, northern México (Elizondo-Quiroga et al. 2005), and in Cx. nigripalpus Theobald and Cx. interrogator Dyar and Knab in Chiapas, southern México (Ulloa et al. 2009). Saint Louis encephalitis virus was detected in Cx. tarsalis Coquillett in Durango, northwestern México (Sudia et al. 1975), and Venezuelan equine encephalitis virus was identified in Cx. coronator Dyar and Knab and Anopheles pseudopunctipennis Theobald in Veracruz State (Scherer et al. 1971). A novel flavivirus, designated T'Ho virus, was detected in Cx. quinquefasciatus in Yucatán State (Farfán-Ale et al. 2009). Several orthobunyaviruses (Cache Valley, Cholul, South River, and Kairi viruses) were isolated from Ae. taeniorhynchus (Wiedemann) in Yucatán State (Farfán-Ale et al. 2009, 2010).

To obtain recent information on the culicid fauna of Yucatán State, all mosquitoes collected during our routine surveillance for *Ae. aegypti* and its associated viruses were identified and documented. Here, we provide an updated checklist of mosquito species that occur in Yucatán State.

Materials and Methods

Yucatán State (20.9678°N, 89.6217°W) is located in the Yucatán Peninsula of México and is bordered by Quintana Roo State and Campeche State (http://www.merida.gob.mx/turismo/contenido/informacion/geografia.htm; last accessed Dec 2015). Yucatán State has a subtropical climate, and rainfall is highest from Jun to Oct (typically >100 mm rainfall per month) with sporadic rainfall occurring during the remainder of the year (García-Rejón et al. 2012).

Immature and adult mosquitoes were collected from Jul 2014 to Dec 2015 in 60 localities of Yucatán State (Fig. 1). A wide range of urban and rural settings were examined, including residential premises (48), cemeteries (5), tourist areas (4, including an archaeological site and a "cenote"), churches (3), forests (3), and a garbage dump site (1) located close to black mangroves (*Avicennia germinans*; Acanthaceae). Some sites were visited more frequently because we had previously shown that *Ae. aegypti* was particularly abundant at these locations. In this study, every study site located at a residential premise or tourist area was visited on a single occasion whereas all other sites were visited on two to more occasions. Locations of study sites were recorded using a global positioning system receiver (Garmin, Olathe, Kansas).

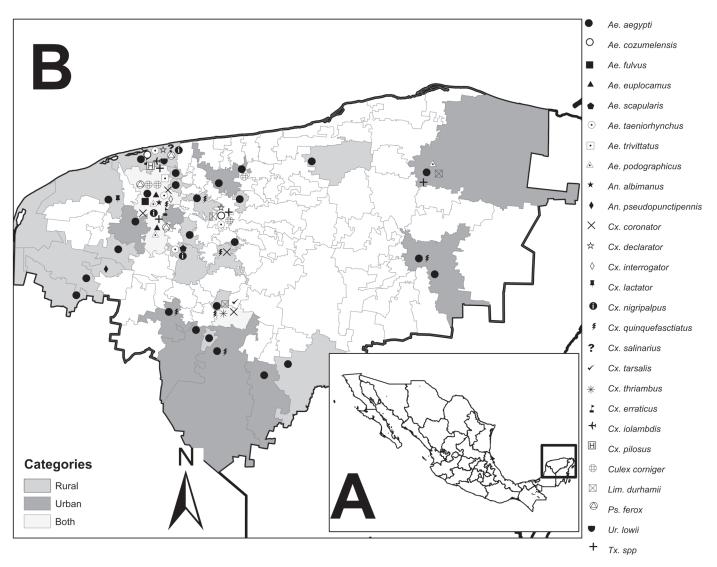


Fig. 1. Geographical distribution of mosquito species from Yucatán State surveyed during Jul 2014 to Dec 2015.

SAMPLING OF IMMATURE AND ADULT MOSQUITOES

Methods used to collect mosquitoes and the classification of container types have been described in previous studies (Nájera-Vázquez et al. 2004; García-Rejón et al. 2011). Nets, turkey basters, or pipettes were used to collect immatures from water-holding containers. Larvae and pupae were placed in plastic containers, labeled according to date, study site, and sample identification number, and transported to the Laboratorio de Arbovirología at Universidad Autónoma de Yucatán. A portion of the larvae were killed in hot water (60 °C); the remaining larvae were individually reared to obtain adults. Pupae were allowed to emerge as adults, and the adults were identified to species. Adult mosquitoes were collected using Centers for Disease Control and Prevention backpack-mounted aspirators. Stereomicroscopes and taxonomic keys were used to identify species (Carpenter & LaCasse 1955; Schick 1970; Arnell 1976; Clark-Gil & Darsie 1983; Ibáñez-Bernal & Martínez-Campos 1994; Darsie & Ward 2005).

SEARCH CHECKLIST OF MOSQUITO SPECIES OF YUCATÁN STATE

We carried out a literature review on mosquito species in Yucatán State in the following databases: Google Scholar, PubMed Health [National Center for Biotechnology Information at the National Library of Medicine], SciELO (Scientific Electronic Library Online), and Web of Science (Thompson Reuters). The search was conducted with the introduction of keywords such as Culicidae, mosquito fauna, mosquito surveillance, new records, mosquito distribution, Yucatán, and México.

Results

A summary of the mosquitoes collected in this study is in Table 1. We collected 8,218 specimens representing 7 genera and 27 species. These species represent ~52% of all mosquito species known to occur in Yucatán State (references can be seen in Table 2). Three species had never before been identified in Yucatán State, and these are Aedes (Ochlerotatus) euplocamus Dyar & Knab, Aedes (Protomacleaya) podographicus Dyar & Knab, and Culex (Culex) declarator Dyar & Knab. We collected 4,406 immatures (4,062 larvae and 344 pupae) in 220 waterholding containers or natural breeding sites. We also collected 3,812 adult mosquitoes. The most common species were Ae. aegypti and Cx. quinquefasciatus (Fig. 1).

BIOLOGICAL NOTES

We provide notes for the 3 newly reported species. Information on their localities and ecological information is also provided.

Aedes euplocamus occurs in Colombia, Peru, Venezuela, Costa Rica, El Salvador, Guatemala, Guyana, Honduras, Panama, and México (Arnell 1976; Burkett-Cadena et al. 2013; Rozo-Lopez & Mengual 2015). In México, this species has been identified in the states of Veracruz (Arnell 1976), Quintana Roo (Ortega-Morales et al. 2010), and Oaxaca (Bond et al. 2014). Arnell (1976) performed a complete review of this species group (Aedes: Ochlerotatus: Scapularis): Larvae and pupae have been collected from water covered in partial shade and in rock-

Table 1. Number of mosquito species registered in Yucatán State from Jul 2014 to Dec 2015.

		Adult			Immature		
#	Taxon	Female	Male	Habitat	Larva	Pupa	Breeding
1	Ae. aegypti	550	880	House	1,700	150	Disposable container, tree hole, tire, bucket
2	Ae. cozumelensis	17	5	Forest, black mangrove ^a	25	7	Vase, tire, rock hole
3	Ae. fulvus	6	_	House	_	_	_
4	Ae. euplocamus⁵	2	_	Forest	_	_	_
5	Ae. scapularis	3	_	Forest	7	_	Rock hole
6	Ae. taeniorhynchus	150	15	Forest, house, black mangrove ^a	50	8	Rock hole, temporary pool
7	Ae. trivittatus	30	_	Forest, house, church	8	4	Rock hole
8	Ae. podographicus⁵	3	_	Forest	21	5	Tree hole
9	An. albimanus	5	_	House	8	_	Temporary pool, artificial lake (aguada)
10	An. pseudopuctipennis	_	_	_	7	3	Cenote
11	Cx. coronator	_	_	_	720	25	Bucket, rock hole
12	Cx. declarator ^b	_	_	_	120	8	Tire, rock hole
13	Cx. interrogator	35	5	House, church	300	5	Tire, vase, bucket
14	Cx. lactator	_	_	_	211	9	Vase, bucket
15	Cx. nigripalpus	150	25	Church	250	30	Vase, bucket
16	Cx. quinquefasciatus	800	1,100	House, church	500	70	Disposable container, bucket, tire
17	Cx. salinarius	_	_	_	4		Tire
18	Cx. tarsalis	2		House	_	_	_
19	Cx. thriambus	2		House	_	_	_
20	Cx. erraticus	_	_	_	15		Temporary pool, artificial lake (aguada)
21	Cx. iolambdis	12	3	Forest, black mangrove ^a	11	3	Temporary pool
22	Cx. pilosus	_	_	_	5		Temporary pool
23	Cx. corniger	_	_	_	20	5	Bucket, rock hole
24	Li. durhamii	2	_	Church	60	8	Vase, disposable container, bucket
25	Ps. ferox	3	_	Forest, church	_	_	_
26	Ur. Iowii	7	_	Forest	5	1	Temporary pool
27	Toxorhynchites spp.	_	_	_	15	3	Rock hole, bucket, tire

^{*}Garbage collection site near of black mangrove (Avicennia germinans).

^bNew records for Yucatán State.

^{(-):} not found.

Table 2. Checklist of the mosquito species known from Yucatán State and new records. Classification according to Knight & Stone (1977). Previous occurrence records are abbreviated: MI: Martini (1935); MP: Martínez-Palacios (1952); VA: Vargas (1956); VA-MP: Vargas & Martínez-Palacios (1956), WI: Winch et al. (1992); IB: Ibáñes-Bernal & Martínez-Campos (1994); NV: Nájera-Vázquez et al. (2004); ZP: Zapata-Peniche et al. (2007); GR-1: García-Rejón et al. (2008); FA-9: Farfán-Ale et al. (2009); FA-0: Farfán-Ale et al. (2010); GR-2: García-Rejón et al. (2011), OM-1: Ortega-Morales et al. (2011); GR-3: García-Rejón et al. (2012); MS: Manrique-Saide et al. (2012); AG: Arana-Guardia et al. (2014); BB: Baak-Baak et al. (2014a); NR: never reported in Yucatán.

#	Taxon	Immature	Adult
1	Aedes (Howardina) cozumelensis Díaz Nájera	IB, GR-3, BB, present study	Present study
2	Aedes (Stegomyia) aegypti (Linnaeus)	WI, IB, NV, ZP, GR-2, MS, AG, BB, present study	GR-1, FA-1, FA-2, GR-2, present study
3	Aedes (Ochlerotatus) angustivittatus Dyar and Knab	NR	MI
ļ	Aedes (Georgecraigius) epactius Dyar and Knab	IB	NR
5	Aedes (Ochlerotatus) euplocamus Dyar and Knab ^a	NR	Present study
5	Aedes (Ochlerotatus) fulvus Wiedemann	NR	FA-2, present study
7	Aedes (Ochlerotatus) infirmatus Dyar and Knab	NR	FA-1
3	Aedes (Ochlerotatus) scapularis (Rondani)	IB, NV, ZP, present study	
)	Aedes (Ochlerotatus) serratus Theobald	NR	MI
.0	Aedes (Ochlerotatus) sollicitans (Walker)	VA, IB, NV, ZP	NR
1	Aedes (Ochlerotatus) taeniorhynchus (Wiedemann)	VA, IB, NV, AG, BB, present study	MI, GR-1, FA-1, FA-2, GR-2, present stud
2	Aedes (Ochlerotatus) tortilis Theobald	NR	MI
.3	Aedes (Ochlerotatus) trivittatus (Coquillett)	BB, present study	GR-1, FA-1, FA-2, GR-2, present study
4	Aedes (Protomacleaya) podographicus Dyar and Knab ^a	Present study	Present study
.5	Anopheles (Nyssorhynchus) albimanus Wiedemann	IB, NV, present study	VA-MP, FA-1, FA-2, present study
.6	Anopheles (Anopheles) apicimacula Dyar and Knab	IB	VA-MP
7	Anopheles (Anopheles) bradleyi King	IB	VA-MP
.8	Anopheles (Anopheles) crucians Wiedemann	IB	VA-MP, FA-1, FA-2
9	Anopheles (Anopheles) pseudopuctipennis Theobald	IB, present study	VA-MP, FA-1
20	Anopheles (Anopheles) vestitipennis Dyar and Knab	IB	VA-MP, FA-1, FA-2
1	Culex (Culex) bidens (Dyar)	IB, NV, ZP	, =, =
2	Culex (Culex) coronator s.l. Dyar and Knab	VA, WI, IB, NV, ZP, MS, AG, BB, present study	FA-1, FA-2
3	Culex (Culex) declarator Dyar and Knab ^a	Present study	NR
4	Culex (Culex) interrogator Dyar and Knab	IB, NV, ZP, MS, AG, BB, present study	FA-1, FA-2, GR-2, present study
5	Culex (Phenacomyia) lactator Dyar and Knab	ZP, AG, BB, present study	NR
:6	Culex (Culex) nigripalpus Theobald	IB, NV, ZP, BB, present study	FA-2, present study
27	Culex (Culex) migriparpus medidad Culex (Culex) quinquefasciatus Say	VA, WI, IB, NV, ZP, MS, AG, BB, present study	MI, GR-1, FA-1, FA-2, present study
.7	Culex (Culex) quinque jusciatus say Culex (Culex) restuans Theobald	IB	NR
.0			NR
	Culex (Culex) salinarius Coquillett	IB, ZP, MS, AG, BB, present study	
0	Culex (Culex) stigmatosoma Dyar	IB, OM-1	Present study
1	Culex (Culex) tarsalis Coquillett	IB, AG, BB	FA-1, present study
2	Culex (Culex) thriambus Dyar	IB, NV, ZP, MS, AG, BB, present study	NR NB
3	Culex (Melanoconion) erraticus (Dyar and Knab)	MP, VA, IB, present study	NR
4	Culex (Melanoconion) iolambdis Dyar	IB, present study	Present study
5	Culex (Melanoconion) pilosus (Dyar and Knab)	IB, present study	NR
6	Culex (Phenacomyia) corniger Theobald	IB, present study	Present study
7	Haemagogus (Haemagogus) anastasionis Dyar	IB, NV	NR
8	Haemagogus (Haemagogus) equinus Theobald	IB, NV, BB	NR
9	Haemagogus (Haemagogus) mesodentatus Komp & Kumm	IB, NV	FA-1, FA-2
0	Limatus durhamii Theobald	BB, present study	Present study
1	Mansonia (Mansonia) titillans (Walker)	IB	FA-1, FA-2
2	Psorophora (Grabhamia) confinnis s.s. (Lynch-Arribalzaga)	IB, NV	FA-1
3	Psorophora (Janthinosoma) albipes Theobald	NR	FA-1
4	Psorophora (Janthinosoma) cyanescens (Coquillett)	VA, IB	MI, FA-1, FA-2
5	Psorophora (Janthinosoma) ferox (von Humboldt)	IB	MI, FA-1, FA-2, present study
6	Psorophora (Psorophora) ciliata (Fabricius)	IB	FA-1
7	Psorophora (Psorophora) howardii Coquillett	NV	FA-1, FA-2
8	Deinocerites cancer Theobald		MI
9	Uranotaenia (Uranotaenia) lowii Theobald	VA, IB, present study	MI, FA-2, present study
0	Toxorhynchites (Lynchiella) theobaldi (Dyar and Knab) b	VA, ZP	NR
51	Toxorhynchites spp.	Present study	
52	Wyeomyia (Wyeomyia) celaenocephala	NR	MI

^aNew records.

holes without vegetation. Aedes euplocamus often cohabits with Ae. aegypti and various Culex and Uranotaenia species (Arnell 1976). It is not considered to be a vector of pathogens of medical or veterinary

importance. In the present study, we collected 2 females of *Ae. euplocamus* in a forest in Xmatkuil (20.8501747°N, 89.6002139°W), which is a small town close to Mérida City.

^bCited by Vargas (1956) as *Megarhinus theobaldi* Dyar and Knab.

The geographic distribution of Ae. podographicus encompasses Belize, Nicaragua, Panama, Costa Rica, Ecuador, El Salvador, Guatemala, Honduras, México, and Venezuela (Schick 1970). In México, Ae. podographicus has been reported in the states of Campeche (Schick 1970), Chiapas (Bond et al. 2014), Jalisco (Martini 1935), Nayarit (Schick 1970), Oaxaca (Martini 1935; Schick 1970), Quintana Roo (Ortega-Morales et al. 2010), Guerrero (Cortés-Guzmán et al. 2013), and Tamaulipas (Ortega-Morales et al. 2015). Larvae and pupae have commonly been collected from tree holes (Schick 1970). Aedes podographicus is not known to vector any pathogens of medical or veterinary importance. Although Martini (1935) and Ibáñez-Bernal & Martínez-Campos (1994) reported the presence of Ae. terrens in Yucatan, there have not been further reports of this particular mosquito's species in México (Zavortink 1972; Darsie 1996). Therefore, it is arguable that the above mentioned authors could have mistakenly identified the more common mosquito Ae. podographicus, which has extensively been reported in Mexican territories. In the present study, we collected 3 adult females of Ae. podographicus in a forest in Xmatkuil (20.8501747°N, 89.6002139°W). We also collected larvae and pupae of Ae. podographicus in a tree hole in Tizimín City (21.1169106°N, 88.1334789°W), in the east of Yucatán. Immatures of Ae. aegypti were also present in the same collections.

Culex declarator has a very wide distribution ranging from southern Texas through to México, Central America, and South America. Older literature, such as Carpenter & LaCasse (1955), has named this species Cx. virgultus Theobald. In México, Cx. declarator has been identified in the states of Quintana Roo (Ortega-Morales et al. 2010), Guerrero (Bond et al. 2014), and Tamaulipas (Ortega-Morales et al. 2015). The larvae are found in a variety of habitats, including rock pools, swamps, cement drains, rot cavities in trees, and coconut husks (Carpenter & LaCasse 1955). Culex declarator is of public health importance. St. Louis encephalitis virus was detected in Cx. declarator in Trinidad (Aitken et al. 1964) and Brazil (Monath et al. 1980). Culex declarator is also considered to be a potential vector of Dirofilaria immitis, dog heartworm (Labarthe et al. 1998). In the present study, immatures of Cx. declarator were collected in rock holes in a rural area in Ekmul (20.9501358°N, 89.3336006°W), Tixkokob, and from a tire at a trash collection site (21.2667072°N, 89.7577778°W) located near black mangroves in Progreso City.

CHECKLIST OF MOSQUITO SPECIES KNOWN FROM YUCATÁN

Between 1935 (Martini 1935) and 2014 (Arana-Guardia et al. 2014; Baak-Baak et al. 2014b), we found 17 works related to the record and distribution of mosquito species in Yucatán State, representing 11 genera and 49 species. The addition of 3 more species brings the species count to 52. The highest richness of culicid taxa belongs to *Aedes* (17 species), *Culex* (16 species), *Anopheles* (7 species), and *Psorophora* (6 species) (Table 2).

Discussion

Dengue virus is endemic in Yucatán State, and recently chikungunya virus has been associated with human morbidity in this region (García-Rejón et al. 2008; Diaz-Gonzalez et al. 2015, Cigarroa-Toledo et al. 2016). Both of these viruses are primarily vectored by *Ae. aegypti* (García-Rejón et al. 2008; Diaz-Gonzalez et al. 2015). For this reason, we routinely monitor the activity of *Ae. aegypti* and its associated viruses in Yucatán State. In the present study, we identified 27 mosquito species that occupy the same niche as *Ae. aegypti*, including 3 species never before reported in Yucatán State: *Ae. euplocamus*, *Ae. podographicus*, and *Cx. declarator*. Globally, the family Culicidae comprises

3,549 species distributed among 112 genera (Harbach 2013). It was recently reported that 217 to 239 species and 15 to 16 genera of mosquitoes have been documented in México (Bond et al. 2014).

In an entomological investigation performed by Ibáñez-Bernal & Martínez-Campos (1994), 36 species of mosquitoes were identified in Yucatán State. Additionally, Vargas (1956) reported 9, Nájera-Vázquez et al. (2004) reported 16, and Farfán-Ale et al. (2009, 2010) reported up to 20 species of mosquitoes in Yucatán State.

It should be noted that most studies on mosquitoes have been carried out in Mérida, because it is the largest city in Yucatán State and the incidence of dengue is high in this location (García-Rejón et al. 2008). Previous studies have shown that immature mosquitoes of various species coexist with immatures of *Ae. aegypti* in disposable containers, buckets, tires, and storm-water drains in Mérida (Winch et al. 1992; Zapata-Peniche et al. 2007; Ortega-Morales et al. 2011; Manrique-Saide et al. 2012; Arana-Guardia et al. 2014; Baak-Baak et al. 2014b). Adult mosquitoes of various species often coexist with adults of *Ae. aegypti* inside houses in Mérida (García-Rejón et al. 2008). Currently, there are 33 species of mosquitoes registered in Mérida City (Baak-Baak et al. 2014b).

At the state level, Tamaulipas and Quintana Roo have the highest richness of culicid taxa with 82 and 76 species, respectively (Ortega-Morales et al. 2010, 2015), followed by Chiapas with 54 species (Bond et al. 2014). As noted earlier, the revised total for Yucatán State is now 52 species. However, additional species could be present within the state but have remained undetected because we usually use trapping methods and focus on sites that allow for the collection of high numbers of *Ae. aegypti*. Other states where entomological investigations have recently been performed are Guerrero with 38 species, Oaxaca with 34 species (Bond et al. 2014), and Colima with 28 species (Espinoza-Gomez et al. 2013).

In the present study, many of mosquitoes identified are capable of serving as vectors of pathogens affecting humans or domestic animals in México. A notable absence is Ae. (Stegomyia) albopictus (Skuse), previously recorded in the Yucatán Peninsula (Salomón-Grajales et al. 2012) but not found in the present study. Martini (1935) reported the identification of Anopheles intermedius (Peryassu), but Vargas & Martínez-Palacios (1956) considered it to be a misclassification and suggested that An. apicimacula Dyar and Knab had been collected. Notably, we collected An. albimanus Wiedemann and An. pseudopunctipennis; these are the main vectors of the parasite that causes malaria in southeastern México (Martini 1935; Vargas & Martínez-Palacios 1956). Additionally, we recorded Ae. taeniorhynchus, which is a major vector of D. immitis (dog heartworm) in Yucatán State (Manrique-Saide et al. 2010).

In the historical literature, it appears that some species of mosquitoes in Yucatán State were erroneously reported. For example, Vargas (1956) apparently collected Aedes (Ochlerotatus) punctor (Kirby), but this species is not considered to occur in México (Darsie 1996). Additionally, Ibáñez-Bernal & Martínez-Campos (1994) reported immatures of Aedes (Protomacleaya) triseriatus (Say). Recently, Sánchez-Trinidad et al. (2014) revised the geographical distribution of this species and concluded that it is only distributed in northern and central México. It is currently unclear as to which Toxorhynchites species occur in Yucatán State. Toxorhynchites (Lynchiella) theobaldi (Dyar and Knab) was reported by Vargas (1956) and Zapata-Peniche et al. (2007). However, Zavortink & Chaverri (2009) examined the status of Toxorhynchites species by their distinctive tarsal markings from several collections and suggested that Tx. moctezuma (Dyar & Knab) and Tx. hypoptes (Knab) had been collected instead. Additionally, the Mosquitoes of Middle America Project makes no mention of these species occurring in México (Heinemann & Belkin 1977). For convenience, Toxorhynchites species collected in our study were identified only to the genus level and will later be conclusively identified using molecular tools. We did not find mosquitoes of the genera *Haemagogus*, *Mansonia*, *Deinocerites*, and *Wyeomyia*, which have all previously been reported in Yucatán State. However, we provide valuable information on mosquito fauna temporally and spatially associated with *Ae. aegypti* in this region.

Acknowledgments

We thank the laboratory staff at Arbovirología of Universidad Autónoma de Yucatán for assistance with mosquito collections. The study was supported in part by the Consejo Nacional de Ciencia y Tecnología de México grant INFR-2014-01-225046. We are very grateful to Bradley J. Blitvich for his review of the English grammar in this manuscript.

References Cited

- Aitken TH, Downs WG, Spence P, Jonkers AH. 1964. St. Louis encephalitis virus isolations in Trinidad, West Indies, 1953–1962. The American Journal of Tropical Medicine and Hygiene 13: 450–451.
- Arana-Guardia R, Baak-Baak CM, Loroño-Pino MA, Machain-Williams C, Beaty BJ, Eisen L, García-Rejón JE. 2014. Stormwater drains and catch basins as sources for production of *Aedes aegypti* and *Culex quinquefasciatus*. Acta Tropica 134: 33–42.
- Arnell JH. 1976. Mosquito studies (Diptera, Culicidae) XXXIIL. A revision of the Scapularis group of *Aedes* (*Ochlerotatus*). Contributions of the American Entomological Institute 13: 1–144.
- Baak-Baak CM, Arana-Guardia R, Cigarroa-Toledo N, Loroño-Pino MA, Reyes-Solis G, Machain-Williams C, Beaty BJ, Eisen L, García-Rejón JE. 2014a. Vacant lots: productive sites for *Aedes (Stegomyia) aegypti* (Diptera: Culicidae) in Merida City, Mexico. Journal of Medical Entomology 51: 475–483.
- Baak-Baak CM, Arana-Guardia R, Cigarroa-Toledo N, Puc-Tinal M, Coba-Tun C, Rivero-Osorno V, Lavalle-Kantun D, Loroño-Pino MA, Machain-Williams C, Reyes-Solis GC, Beaty BJ, Eisen L, García-Rejón JE. 2014b. Urban mosquito fauna in Merida City, Mexico: immatures collected from containers and storm-water drains/catch basins. Southwestern Entomologist 39: 291–306.
- Bond JG, Casas-Martinez M, Quiroz-Martinez H, Novelo-Gutierrez R, Marina CF, Ulloa A, Orozco-Bonilla A, Munoz M, Williams T. 2014. Diversity of mosquitoes and the aquatic insects associated with their oviposition sites along the Pacific coast of Mexico. Parasites and Vectors 7: 41.
- Burkett-Cadena N, Graham SP, Giovanetto LA. 2013. Resting environments of some Costa Rican mosquitoes. Journal of Vector Ecology 38: 12–19.
- Carpenter SJ, LaCasse WJ. 1955. Mosquitoes of North America (North of Mexico). University of California Press, Berkeley, California.
- Cigarroa-Toledo N, Blitvich BJ, Cetina-Trejo RC, Talavera-Aguilar LG, Baak-Baak CM, Torres-Chable OM, Hamid MN, Friedberg I, Gonzalez-Martinez P, Alonzo-Salomon G, Rosado-Paredes EP, Rivero-Cardenas N, Reyes-Solis GC, Farfán-Ale JA, García-Rejón JE, Machain-Williams C. 2016. Chikungunya virus in febrile humans and *Aedes aegypti* mosquitoes, Yucatan, Mexico. Emerging Infectious Diseases 22. doi: 10.3201/eid2210.152087.
- Clark-Gil S, Darsie Jr RF. 1983. The mosquitoes of Guatemala. Their identification, distribution and bionomics, with keys to adult females and larvae. Mosquito Systematics 15: 151–284.
- Cortés-Guzmán AJ, Sánchez-Casas RM, Ibarra-Juárez LA, Ortega-Morales AI, García-Rejón JE, Contreras-Cordero JF, Fernández-Salas I. 2013. West Nile virus survey of birds, horses, and mosquitoes of the Pacific Coast, southern Mexico. Southwestern Entomologist 38: 231–240.
- Darsie Jr RF. 1996. A survey and bibliography of the mosquito fauna of México (Diptera: Culicidae). Journal of the American Mosquito Control Association 12: 298–306.
- Darsie Jr RF, Ward RA. 2005. Identification and Geographical Distribution of the Mosquitoes of North America, North of Mexico. University of Florida Press, Gainesville, Florida.
- Diaz-Gonzalez EE, Kautz TF, Dorantes-Delgado A, Malo-Garcia IR, Laguna-Aguilar M, Langsjoen RM, Chen R, Auguste DI, Sanchez-Casas RM, Danis-Lozano R, Weaver SC, Fernández-Salas I. 2015. First report of *Aedes aegypti* transmission of chikungunya virus in the Americas. The American Journal of Tropical Medicine and Hygiene 93: 1325–1329.

- Elizondo-Quiroga D, Davis CT, Fernández-Salas I, Escobar-Lopez R, Velasco Olmos D, Soto Gastalum LC, Aviles Acosta M, Elizondo-Quiroga A, Gonzalez-Rojas JI, Contreras Cordero JF, Guzman H, Travassos da Rosa A, Blitvich BJ, Barrett AD, Beaty BJ, Tesh RB. 2005. West Nile virus isolation in human and mosquitoes, México. Emerging Infectious Diseases 11: 1449–1452.
- Espinoza-Gomez F, Arredondo-Jimenez JI, Maldonado-Rodriguez A, Perez-Renteria C, Newton-Sanchez OA, Chavez-Flores E, Gomez-Ibarra E. 2013. Geographic distribution of adult mosquitoes (Diptera: Culicidae) in rainforest areas of Colima, Mexico. Revista Mexicana de Biodiversidad 84: 685–689.
- Farfán-Ale JA, Loroño-Pino MA, García-Rejón JE, Hovav E, Powers AM, Lin M, Dorman KS, Platt KB, Bartholomay LC, Soto V, Beaty BJ, Lanciotti RS, Blitvich BJ. 2009. Detection of RNA from a novel West Nile-like virus and high prevalence of an insect-specific flavivirus in mosquitoes in the Yucatán Península of México. The American Journal of Tropical Medicine and Hygiene 80: 85–95
- Farfán-Ale JA, Loroño-Pino MA, García-Rejón JE, Soto V, Lin M, Staley M, Dorman KS, Bartholomay LC, Hovav E, Blitvich BJ. 2010. Detection of flaviviruses and orthobunyaviruses in mosquitoes in the Yucatán Península of México in 2008. Vector Borne and Zoonotic Diseases 10: 777–783.
- García-Rejón JE, Loroño-Pino MA, Farfán-Ale JA, Flores-Flores L, del Pilar Rosado-Paredes E, Rivero-Cardenas N, Najera-Vazquez MR, Gomez-Carro S, Lira-Zumbardo V, Gonzalez-Martinez P, Lozano-Fuentes S, Elizondo-Quiroga D, Beaty BJ, Eisen L. 2008. Dengue virus–infected *Aedes aegypti* in the home environment. The American Journal of Tropical Medicine and Hygiene 79: 940–950.
- García-Rejón JE, López-Uribe MP, Loroño-Pino MA, Farfán-Ale JA, Najera-Vazquez MR, Lozano-Fuentes S, Beaty BJ, Eisen L. 2011. Productive container types for *Aedes aegypti* immatures in Mérida, México. Journal of Medical Entomology 48: 644–650.
- García-Rejón JE, López-Uribe MP, Loroño-Pino MA, Arana-Guardia R, Puc-Tinal M, Lopez-Uribe GM, Coba-Tun C, Baak-Baak CM, Machain-Williams C, Reyes-Solis GC, Lozano-Fuentes S, Saavedra-Rodriguez K, Black WC, Beaty BJ, Eisen L. 2012. Aedes (Stegomyia) aegypti and Aedes (Howardina) cozumelensis in Yucatán State, México, with a summary of published collection records for Ae. cozumelensis. Journal of Vector Ecology 37: 365–372.
- Harbach RE. 2013. Mosquito Taxonomic Inventory, http://mosquito-taxonomic-inventory.info/ (last accessed 23 Jan 2016).
- Heinemann SJ, Belkin JN. 1977. Collection records of the project "Mosquitoes of Middle America" 9. Mexico (MEX, MF, MT, MX). Mosquito Systematics 9: 483–535.
- Ibáñez-Bernal S, Martínez-Campos C. 1994. Clave para la identificación de larvas de mosquitos comunes en las áreas urbanas y suburbanas de la República Mexicana (Díptera: Culicidae). Folia Entomológica Mexicana 92: 43–73.
- Knight KL, Stone A. 1977. Catalog of the Mosquitoes of the World (Diptera: Culicidae), 2nd Edition. Volume VI. Thomas Say Foundation. College Park, Maryland.
- Labarthe N, Serrao ML, Melo YF, de Oliveira SJ, Lourenco de Oliveira R. 1998. Mosquito frequency and feeding habits in an enzootic canine dirofilariasis area in Niteroi, State of Rio de Janeiro, Brazil. Memorias do Instituto Oswaldo Cruz 93: 145–154.
- Manrique-Saide P, Escobedo-Ortegon J, Bolio-Gonzalez M, Sauri-Arceo C, Dzib-Florez S, Guillermo-May G, Ceh-Pavia E, Lenhart A. 2010. Incrimination of the mosquito, Aedes taeniorhynchus, as the primary vector of heartworm, Dirofilaria immitis, in coastal Yucatán, México. Medical and Veterinary Entomology 24: 456–460.
- Manrique-Saide P, Uc V, Prado C, Carmona C, Vadillo J, Chan R, Dzib-Florez S, Che-Mendoza A, Barrera-Perez M, Sanchez EC, Arredondo-Jimenez JI. 2012. Storm sewers as larval habitats for *Aedes aegypti* and *Culex* spp. in a neighborhood of Mérida, México. Journal of the American Mosquito Control Association 28: 255–257.
- Martínez-Palacios A. 1952. Nota sobre la distribución de los mosquitos *Culex* en México (Diptera: Culicidae). Revista de la Sociedad Mexicana de Historia Natural 13: 75–87.
- Martini E. 1935. Los mosquitos de México. Boletin Técnico. Imprenta del Departamento de Salubridad Pública 1: 66.
- Monath TP, Cropp CB, Bowen GS, Kemp GE, Mitchell CJ, Gardner JJ. 1980. Variation in virulence for mice and rhesus monkeys among St. Louis encephalitis virus strains of different origin. The American Journal of Tropical Medicine and Hygiene 29: 948–962.
- Nájera-Vázquez R, Dzul F, Sabido M, Tun-Ku E, Manrique-Saide P. 2004. New distribution records of mosquitoes (Diptera: Culicidae) for Yucatán, México. Entomological News 115: 181–190.
- Ortega-Morales Al, Mis-Avila P, Elizondo-Quiroga A, Harbach RE, Siller-Rodríguez QK, Fernández-Salas I. 2010. The mosquitoes of Quintana Roo State, México (Diptera: Culicidae). Acta Zoológica Mexicana 26: 33–46.

- Ortega-Morales Al, Huerta H, Strickman D, Sánchez Ramos FJ, Landeros Flores J, Chávez EC. 2011. Registros de mosquitos en México: *Culex stigmatosoma* Dyar y *Cx. thriambus* Dyar (Diptera: Culicidae) con notas taxonómicas para ambas especies. Southwestern Entomologist 36: 177–196.
- Ortega-Morales Al, Zavortink TJ, Huerta-Jiménez H, Sánchez-Rámos FJ, Valdés-Perezgasga MT, Reyes-Villanueva F, Siller-Rodríguez QK, Fernández-Salas I. 2015. Mosquito records from Mexico: the mosquitoes (Diptera: Culicidae) of Tamaulipas State. Journal of Medical Entomology 52: 171–184.
- Rozo-Lopez P, Mengual X. 2015. Updated list of the mosquitoes of Colombia (Diptera: Culicidae). Biodiversity Data Journal 9: e4567.
- Salomón-Grajales J, Lugo-Moguel GV, Tinal-Gordillo VR, de la Cruz-Velazquez J, Beaty BJ, Eisen L, Lozano-Fuentes S, Moore CG, García-Rejón JE. 2012. *Aedes albopictus* mosquitoes, Yucatán Península, México. Emerging Infectious Diseases 18: 525–527.
- Sanchez-Trinidad A, Ordonez-Sanchez F, Valdes-Perezgasga MT, Sanchez-Ramos FJ, Zavortink TJ, Cortes-Guzman AJ, Ortega-Morales Al. 2014. Geographical distribution of the Aedes Triseriatus Group (Diptera: Culicidae) in México. Journal of Vector Ecology 39: 134–137.
- Scherer WF, Dickerman RW, Diaz-Najera A, Ward BA, Miller MH, Schaffer PA. 1971. Ecologic studies of Venezuelan encephalitis virus in southeastern México. 3. Infection of mosquitoes. The American Journal of Tropical Medicine and Hygiene 20: 969–979.
- Schick RX. 1970. Mosquito studies (Diptera, Culicidae) XX. The Terrens group of *Aedes (Finlaya*). Contributions of the American Entomological Institute 5: 1–96.
- Sudia WD, Fernández L, Newhouse VF, Sanz R, Calisher CH. 1975. Arbovirus vector ecology studies in México during the 1972 Venezuelan equine encephalitis outbreak. American Journal of Epidemiology 101: 51–58.

- Ulloa A, Ferguson HH, Méndez-Sanchez JD, Danis-Lozano R, Casas-Martínez M, Bond JG, Garcia-Zebadua JC, Orozco-Bonilla A, Juarez-Ordaz JA, Farfán-Ale JA, García-Rejón JE, Rosado-Paredes EP, Edwards E, Komar N, Hassan HK, Unnasch TR, Rodriguez-Perez MA. 2009. West Nile virus activity in mosquitoes and domestic animals in Chiapas, México. Vector Borne and Zoonotic Diseases 9: 555–560.
- Vargas L. 1956. Especies y distribución de mosquitos mexicanos no Anofelinos (Insecta
- Diptera). Revista del Instituto de Salubridad y Enfermedades Tropicales 15: 19–36.
- Vargas L, Martínez-Palacios A. 1956. Anofelinos mexicanos: taxonomía y distribución, Secretaría de Salubridad y Asistencia, Comisión Nacional para la Erradicacion del Paludismo 16: 19–36.
- Winch PJ, Barrientos-Sanchez G, Puigserver-Castro E, Manzano-Cabrera L, Lloyd LS, Méndez-Galván JF. 1992. Variation in *Aedes aegypti* larval indices over a one year period in a neighborhood of Mérida, Yucatán, México. Journal of the American Mosquito Control Association 8: 193–195.
- Zapata-Peniche A, Manrique-Saide P, Rebollar-Téllez EA, Che-Mendoza A, Dzul-Manzanilla F. 2007. Identificación de larvas de mosquitos (Diptera: Culicidae) de Mérida, Yucatán, México y sus principales criaderos. Revista Biomedica 18: 3–17.
- Zavortink TJ. 1972. Mosquito studies (Diptera, Culicidae) XXVIII: the New World species formerly placed in *Aedes* (*Finlaya*). Contributions of the American Entomological Institute 8: 1–206.
- Zavortink TJ, Chaverri LG. 2009. Resurrection of the names *Toxorhynchites moctezuma* (Dyar & Knab) and *Toxorhynchites hypoptes* (Knab) from synonymy with *Toxorhynchites theobaldi* (Dyar & Knab) (Diptera: Culicidae). Proceedings of the Entomological Society of Washington 111: 890–897.