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Authors: Grasela, James J., and McIntosh, Arthur H.

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Cross-species investigation of *Helicoverpa armigera* microsatellites as potential markers for other related species in the *Helicoverpa* - *Heliothis* complex

James J. Grasela and Arthur H. McIntosh

USDA, ARS, Biological Control of Insects Research Laboratory, 1503 S. Providence Rd., Research Park Columbia, Missouri USA 65203-3535

Abstract

Primers previously designed to amplify microsatellite DNA markers in the Old World bollworm, *Helicoverpa armigera*, larvae were tested in three closely related species: the corn earworm, *Helicoverpa* zea, tobacco budworm, *Heliothis virescens*, and *Heliothis subflexa*. Of the fourteen loci surveyed, only four loci (HaB60, HaC14, HaC87, HarSSR1) consistently demonstrated scorable single-copy microsatellite bands. Of these four, length polymorphism was identified only in the HaB60 marker (160 bp, 140 bp) of the *H. virescens* and *H. subflexa* sampled laboratory populations. Partial DNA sequences of all the identified single-copy microsatellites are presented as well as alignments to their respective *H. armigera* microsatellite.

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Introduction

The lengthy process and expertise required to isolate and identify potential microsatellite markers often precludes the use of this valuable technique in studies to determine genetic variation in natural populations. If microsatellite markers identified and developed from one biological source could be applied to other similar species, the usefulness of these genetic markers could be broadened. Fortunately, a number of microsatellites markers have been developed to study the population genetic variation in the Old World bollworm, Helicoverpa armigera, a serious insect pest of several agriculturally important grain and fiber crops (Tan et al., 2001; Ji et al., 2003; Scott et al., 2004). We therefore undertook a survey of some of the available lepidopteran species that are used routinely in our laboratory, namely Helicoverpa zea, Heliothis virescens, and Heliothis subflexa, to determine if previously designed microsatellite markers for H. armigera from several published sources could be applied to these closely related lepidopteran species. Selection of these species for study was also contingent on their importance as field crop pests. The host range of H. zea, the corn earworm, includes over 100 plants with the most significant crops being corn, cotton and tomato. Occasional hosts include bean, broccoli, cabbage, chrysanthemum, eggplant, head cabbage, green bean, lettuce, okra, pea, pepper, soybean, strawberry and watermelon. The tobacco budworm, H. virescens, is also principally a field crop pest, attacking such crops as alfalfa, clover, cotton, flax, soybean, corn, and tobacco. However, it sometimes feeds on such vegetables as cabbage, cantaloupe. lettuce, pea, pepper, pigeon pea, squash, and tomato, especially when cotton or other favored crops are not abundant. H. virescens is a common pest of geranium and other flower crops such as ageratum, bird of paradise, chrysanthemum, and gardenia, to name a few. In contrast, H. subflexa is of minor agricultural importance feeding on a few plant species such as Solanum nigrum and Physalis spp, but serves as a unique laboratory subject in studies to determine and compare host range infectivity and genetic resistance to baculoviruses.

Materials and Methods

Based on a previously published protocol (McIntosh et al., 1996), genomic DNA was extracted from 2nd or early 3rd instar *H. zea* and *H. virescens* larvae obtained from the North Carolina

State University- Entomology Insectary, Raleigh, North Carolina, whereas 2nd or 3rd instar *H*. subflexa larvae were obtained in-house at the USDA, ARS, Biological Control of Insects Research Laboratory, Columbia, Missouri. Sample sizes are shown in Tables 1 and 2. Fourteen published primer sets designed to amplify the following microsatellite loci of H. armigera were employed in this study: (1) (HaB60) -- (CTG)₂ (TTG)₃ (CTG)₅ $(TTG)_2$, (2) (HaC14) -- $(ATTT)_5$, (3) (HaD47) --(CA)₅ (TCA)₄, (4) (HaC87) - (TC)₅ (Scott et al., 2004); (5) Ham2 -- (TTTTGA)9, (6) Ham3 --(TAAA)₂ (TAAAT)₄, (7) Ham₄ -- (TCTG)₆ TCTT (TCTG)₆, (8) Ham₅ -- (T)_n (G)_n, (9) Ham₆ --(GAT)₂ TT (GAT)₂ TT.....(AATA)₅ (Tan et al., HarSSR1 (TGC)₂GAT 2001); (10) $(TGY)_4GAT(TGY)_{35}(TGA)_2$ AGC(TGY)8 (11)HarSSR2 - (ATG)7, (12) HarSSR3 - (TCA)6, (13) HarSSR4 - (GYT)₂₅, and (14) HarSSR5 - [T(T)AA]₆ (Ji et al., 2003). DNA microsatellite amplification was conducted under the following two polymerase reaction conditions using a Hybaid OmniGene thermal cycler (Midwest Scientific, www.midsci.com) in 25 l of puReTag Ready-To-GoTM PCR bead reaction mixture (Amersham Biosciences, www.apbiotech.com), including 100-200 ng of genomic DNA template. First, after initial denaturing at 94° C for 5 min, the reaction mixture underwent 35 cycles at 94° C for 1 min, variable annealing temperature (see Tables 1 and 2) for 30 sec, 72° C for 40 sec, and a final extension at 72° C for 5 min (Tan et al., 2001). Second, after initial denaturing at 940 C for 1 min, the reaction mixture underwent 35 cycles at 94° C for 1 min, 50° C for 1 min, 73° C for 1 min, and a final extension at 72° C for 5 min (Scott et al., 2004). These two previously published PCR conditions with their respective primers were used to establish a comparative baseline for the three lepidopteran species examined in this study. However, if the expected fragment size(s) was not detected under the original PCR conditions for a particular microsatellite, empirical studies with various annealing temperatures were conducted in attempt to resolve these problematic microsatellite markers (Table 1). A 10 l aliquot of each amplified sample was run on a 2.5% Metaphor™ agarose gel (10 mM Sodium hydroxide-Boric acid buffer, pH 8.5) for ca. 1 h at a constant 120 v using a Bio-Rad Wide Mini-Sub Cell-gel system.

3

Results

Initially. annealing temperatures previously published for the various microsatellites detected in H. armigera were employed in this study with resulting mixed success. Therefore, as indicated in Table 1, several annealing temperatures were tested for each locus in all three species in an attempt to determine the optimal running conditions for successful microsatellite amplification. Table 2 shows the microsatellite loci that failed to show distinct single-copy bands under the various PCR amplification conditions tested. Of the fourteen loci surveyed, only four loci (HaC14, HaB60, HaC87, and HarSSR1) consistently demonstrated scorable single-copy microsatellite bands that might lead to the potential detection of population polymorphism in subsequent studies (Fig.1). The phrase "potential detection" must be emphasized since the samples tested were limited to only laboratory reared insects. Of the four loci that consistently demonstrated scorable single-copy microsatellite bands, length polymorphism was identified only in the HaB60 marker (160 bp and 140 bp). The remaining microsatellites investigated showed multiple banding patterns, which have typically been observed in a number of lepidopteran species during the process of microsatellite clone development, and further indicate the repetitive nature of the flanking regions of microsatellites throughout the genome of Lepidoptera (Zhang, 2004). The HaC14 270 bp band (Fig. 2) detected among all three species, and first thought to be a microsatellite repeat variation, appears to be actually caused by a duplication of the downstream

primer sequence used to amplify the microsatellite (sequence data not shown).

To obtain a more accurate picture of the nucleotide base composition of some of the detected single-copy microsatellites that showed either the expected allele size or a variant, direct DNA sequencing of PCR products was performed at the University of Missouri DNA Core Facility, Columbia, Missouri using an Applied Biosystems (www.appliedbiosystems.com) 3730 DNA Analyzer. Because of the known potential for amplification errors during the PCR reaction due the inherent nature of the *Taq* polymerase, 2-3 replicate samples of each locus were sequenced from individual insects and a single consensus sequence was generated employing VisCoSe (Spitzer et al., 2004). Partial sequence alignments of five alleles from four microsatellites are indicated in Figure 3. In addition to the generated sequence alignments, the T-coffee program also provides an index of Consistency of the Overall Residue Evaluation (CORE), an objective measure that identifies which regions of the compared sequences are correctly aligned by averaging the scores of each of the aligned pairs involving a base within a column (Notredame et al., 2000). A CORE value > = 3would indicate a properly aligned base position and is considered the best compromise between a level of sensitivity and specificity required for proper base alignment. All of the aligned portions of the expected microsatellites showed reasonably high CORE scores for their individual alleles (70% for HaC14; 70%, 51% for HaB60 (160 bp, 140 bp, respectively); 91% for HaC87 (118bp); and 71% for

Table 1. Microsatellite markers previously published for *Helicoverpa armigera* found to successfully amplify similar microsatellite loci in three other related lepidopteran species.

Species	Locus	Annealing temp (Co) - DNA band fragment profile	Number of replicates	Sample size (n)	Reference for PCR running conditions
H. zea	HaB60	50° - multiple bands > 200 bp marker	1	7	
		55° - single 160 bp band	3	18	Tan et al., 2001(modified)*
		60° - multiple bands > 200 bp marker and a single 160 bp band	2	12	
H. zea	HaC87	50° - single-copy band 118 bp band	2	14	Scott et al., 2004;
		55°- multiple bands	1	3	Tan et al., 2001(modified)
		60° - single-copy 118 bp band	2	12	
H. zea	HaC14	50° - single-copy 160 bp band	1	12	Scott et al., 2004
		55° - single-copy 160 bp band	1	3	Tan et al., 2001(modified)
H. zea	HarSSR1	50° - single-copy 240 bp band	1	2	Scott et al., 2004
		58° - single-copy 240 bp band	2	5	
		50° - multiple bands	2	12	
H. virescens	HaC14	55° - single-copy 160 bp	2	15	Scott et al., 2004
		50° - multiple bands	2	4	Scott et al., 2004
H. virescens	HaB60	55° - 160 bp and 140 bp single-copy bands	3	11	Tan et al., 2001(modified)
		50° - 240 bp single-copy band	1	5	
H. virescens	HarSSR1	58° - multiple bands	2	4	Scott et al., 2004
		60° - multiple bands	1	2	
H. subflexa	HaB60	50° - multiple bands	2	4	Scott et al., 2004
		55° - 160 bp and 140 bpsingle-copy bands	3	14	
H. subflexa	HaC14	50° - 160 bp single-copy band	3	14	Scott et al., 2004
		55° - multiple bands	1	2	
H. subflexa	HaSSR1	58° - 240 bp single-copy band	2	4	Scott et al., 2004

Table 2. Microsatellite markers previously published for Helicoverpa armigera found to unsuccessfully amplify similar microsatellite loci in three other related lepidopteran species.

Species	Locus	Annealing temperature (Co) - DNA band fragment profile	Number of replicates	Sample size	PCR running conditions
H. zea	HaD47	500 - single-copy 240 bp band	1	2	Scott et al., 2004; Tan et al. 2001 (modified)
ii. zeu	11404/	550 - multiple bands	3	18	Scott et al., 2004, Tall et al. 2001 (modified)
		530 - multiple bands	1	6	
I. zea	Ham3	600 - multiple bands > 200 bp marker	1	6	Tan et al., 2001; Tan et al. 2001 (modified)
		660 - multiple bands > 200 bp marker	2	12	
I. zea	HarSSR2	520 - fb	1	2	Scott et al., 2004; Tan et al. 2001 (modified)
1. zcu	114100142	600 - nb	1	4	Scott et al., 2004, Tall et al. 2001 (modified)
		590 - nb	1	2	
I. zea	HarSSR3	600 - multiple bands	1	4	Scott et al., 2004; Tan et al. 2001 (modified)
		550 - multiple bands > 200 bp marker	1	3	
H. zea	HarSSR4	60 - single-copy 240	1	2	Scott et al., 2004; Tan et al. 2001 (modified)
		660 - multiple bands > 240 bp band	1	3	
I. zea	HarSSR5	660 - multiple bands	1	4	Tan et al., 2001
I. zea	Ham5	520 -multiple bands	1	2	Tan et al., 2001
		500 - multiple bands	1	2	
1. virescens	HaC87	550 - multiple bands	1	4	Scott et al., 2004; Tan et al. 2001 (modified)
		600 - nb	1	4	
I. virescens	HarSSR2	520 - fb	1	2	Tan et al. 2001(modified); Scott et al., 2004
11 011 0000110	11410014	660 - multiple bands	2	7	7 un et un 2001 (mounteu), beote et un, 2004
		590 - multiple bands	1	2	
H. virescens	HarSSR3	600 - multiple bands	1	3	Scott et al., 2004
		500 - multiple bands	2	7	
H. virescens	HarSSR4	600 - multiple bands	1	2	Scott et al., 2004
	•	530 - multiple bands	1	2	· •
1. virescens	Ham3	550 - multiple bands	1	4	Tan et al., 2001
H. virescens	Ham5	520 - multiple bands	1	2	Tan et al., 2001
	· ·	550 - nb	1	4	· ·
1. virescens	Ham6	660 - nb	1	3	Tan et al., 2001
	** 00	500 - multiple bands	1	2	a 1
H. subflexa	HaC87	550 - multiple bands > 200 bp marker and strongly stained bands at 55, 70, 75 bp markers	2	18	Scott et al., 2004
I. subflexa	HaD47	500 - multiple band	1	2	Scott et al., 2004
,	• * *	550 - suspected single-copy band at 140 bp	2	10	, , ,
H. subflexa	HaD47	500 - multiple bands	1	2	Scott et al., 2004
I. subflexa	Ham3	530 - multiple bands >180 bp marker band	2	16	Tan et al., 2001; Tan et al. 2001 (modified)
п. ѕиојіеха	Haing	590 - multiple bands	1	4	ran et al., 2001, Tan et al. 2001 (modified)
I. subflexa	Ham6	530 - multiple bands	1	4	Tan et al., 2001
I. subflexa	HaSSR2	520 - nb	1	2	Tan et al., 2001
	HaSSr3	590 - fb	1	2	Tan et al., 2001
H. subflexa	HaSSR4	600 - fb	1	2	Tan et al., 2001
H. subflexa	HaSSR5	540 - fb	1	2	Tan et al., 2001

nb= no bands detected; fb = faint bands

Repetitive sequences are defined as repeated genomic regions containing microsatellite motifs and their flanking regions.

HarSSR1 (240bp), indicating at least for the most part a good portion of the base positions were properly aligned. Based on the aligned regions generated bv the T-coffee program (www.ch.embnet.org/software/TCoffee.html), identity of the nucleotide sites of the partially sequenced microsatellites relative to H. armigera was found to be 78% for all three species at the HaC14 160 allele, 83% for all three species at the HaB60 160 allele, 41% for H. virescens and H. zea at the HaB60 140 allele, and 84% for H. zea and H. virescens at the HaC87 118 allele, and 76% for H. zea and H. virescens at the HarSSR1 240 allele. As indicated in Fig. 3 (A-D) only the downstream primer used in PCR amplification for each locus appeared in the sequence along with the microsatellite marker. However, the upstream primer that would typically be included as part of the 5'-end of the microsatellite marker was not sequenced during the automatic analysis.

Several reports have shown that comparing allele sizes can result in inaccurate allele size differences for microsatellites (Estoup et al., 1995; Haberl and Tautz, 1999). One can approach this potential problem of size homoplasy by either employing single-strand conformation polymorphism analysis (SSCP) or sequence analysis of the DNA fragments. However, Liepelt et al. (2001) has shown that even sequenced, aligned microsatellites can show differences in repeat numbers occurring among clones and samples from the same individual. Their solution was to split the analyzed complex locus into two new loci. Nevertheless, we chose sequence analysis to determine if our unknown fragments contained not only the microsatellite but also to obtain an overall view of the alignment patterns of the fragments relative to the H. armigera markers.

Overall, the alignments of the four microsatellite loci detected in the three species, but with the

Figure 1. An assortment of PCR amplifications depicting several potential microsatellite primer pairs. (A) PCR amplification of three single-copy microsatellites from 10 individual Heliothis subflexa larvae; (B) PCR amplification of two single-copy microsatellites from eight Heliothis virescens larvae; (C) three single-copy microsatellites detected in H. zea, the more closely related of the three species to Helicoverpa armiger a. Base pair markers are indicated on the left of each gel. The size of specific bands that were sequenced is indicated for each of the microsatellite loci. nc = negative control.

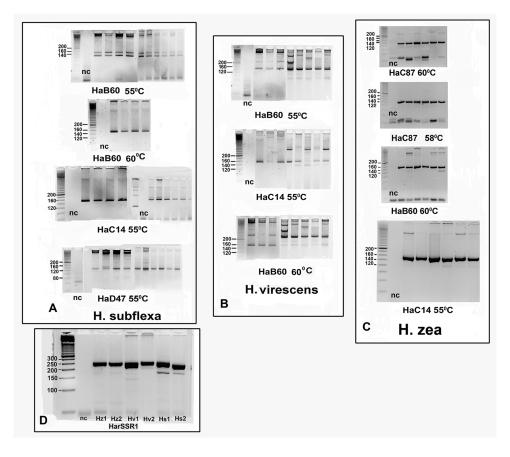


Figure 2. Successful identification of PCR amplified single-copy microsatellites from sampled individuals of the three species. (A) HaC14; (B) HaB60; (C) HaC87; and (D) HarSSR1. Hz = *Helicoverpa zea*; Hv = *Heliothis virescens*; Hs = *Heliothis subflexa*. Base pair markers are indicated on the left of each gel. nc = negative control. The size of specific bands that were sequenced is also indicated for each of the microsatellite loci.

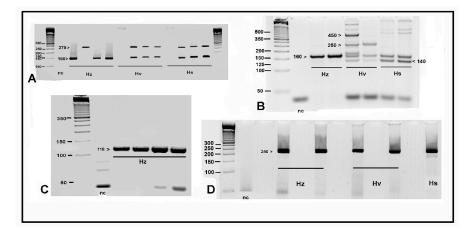


Figure 3. Partial sequences of the four simple sequence loci. All sequences were aligned employing the T-Coffee multiple sequence alignment package. Microsatellite alleles are shown for (A) HaC14, (B,C) HaB60, (D) HaC87 and (E) HarSSR1. Bold letters indicate the location of the simple sequence repeat and the box-shaded regions indicate identities. A CORE index for each base position is indicated in the outlined box below each alignment. The primer sequences flanking the loci are shown in lowercase letters.

A.	HaC14		10			40	50
HA HV		TAGTcca	acacagtt! 		AAATGGGTT	.!!! CGTTACTGTTTT	
HZ HS							A
		04					
			60	70	80	90	100
НА		TTTATT	TATTTATT	r-atttcagi	rggccgcggt	.!!! CCCATTCGAGGI	TGGGA
HV HZ HS		TTTGTT	TATTTATT	rgatttcag1	rggacgcggt	CCCATTCGAGGT CCCATTCGAGGT CCCATTCGAGGT	TGGGA
	555466			77777778			.100011
НА				! !	.!!	140 .!!!	
1 '		CC-TCAC CCATCAC 67-56666	CCT-TTTT:	rgtt-ttgaj	rttgtttt rgtatgtttt	AACCCTTCTC AACCCTTCTC TAAACCCTTCTC	CTCGTT
_7	776777	7777					
		!			180 .!!	190	200
HA HV HZ		TGAATCA	AA-TAGGA:	TTATGGCAG-		GATTACCTATAT	
HS	778878	TGAATCA	AACTAGGA:		3		
			70000121				
HA HV HZ HS			!	! !		240 .!!! AATACTGTGGTA	
		,	260				
HA HV			!! ATCTATAGO				
HZ HS		 		 			

Figure 3 (B).

В. НаВ60		10	20	30	40	50
НА		ccacctga	cataacgcT(CACAGGTTG	!! CTGCAACTGTI	GTTGTTG
HV HZ					GCT T	
HS					CT 	
233334						
		60	70	80	90	100
НА					!! GTTGTTGCTGC	
HV	CTGCTG	TTGCAGTT	GCGCCACTT	GCTGCTGTT	GTTGTTGCTGC	CTGCTGCT
HZ HS					GTTGCTGCTGC GTTGTTGCTGC	
			677777777		01101100100	71001001
	1	110	120	_00	140	150
НА					CGCCTGTTGCT	
HV					CGCTTGTTGCT	
HZ HS					IGCCTGTTGCI CGCTTGTTGCI	
	66666676		77777-677			
777777	<i></i>	1.00	170	100	100	000
	! .		170		190 !!	200
НА	cttgcaa	attgctgc			TTTTCCTCGAG	
HV						
HZ HS			TCCTTAAA			
		75554222				
		210	220	230		
	!.		! !	.!!	•	
HA	TGTATT	TTGTATAT	CTTTTCGGC	AAGTAGATG	A	
HV HZ					_	
HS					_	

Figure 3 (C).

C. HaB60 (140)

HA HV HS 11112 73333333	ACGCcaccacctg-acata ACATC-TCGGTACTTCA-TCGGTACTTC	aacgcTCACAGGT GGG, GG	TGCTGCAA-CT(AGCATCTAGCC(GTTGTTGT CACGGGGA
HA HV HS	60 70!!! TGCTGCTGTTGCAATTGCGC GAC-GCTATTACCATGGC-C GAC-GCTATTACCATGGC-C	!!! Caacttg ctgctg Cttcat-ct-ctg.	!! TTGTTGTTGCTC	GCTGCTG
444-4555	444-45555544445444-3655444444-44	1-4443		
HA HV HS	CTGCTGTTGTTGCTGTTGCTCAACAGGAACTACAA-AGGAACTA222234444443	IGCTGCTGTTGCG	!! CCGCCTGTTGCT GATT	.!! IGTTGAg -GTTGC- -GTTGCG
HA HV HS -6666666	160 17!!! cttgcaattgctgctCCTTG -TTGCAATTGCTGCTCCTTA -TGCAATTGCTGCTCCTTA -66666666777777765432 57777777654322 210 22!!! TGTATTTTGTATATCTTTTC	!!! CCGTTTTTGTCTC'AA	!! TTTTCCTCGAG	TTCTTTC
HV HS			- -	

Figure 3 (D).

D. HaC87 (118)

НА			! !	30 !! ACCAGGAACT		
HZ						
				80		
HA HZ		ATATTACT(CTCATTTTAT	!! GCCGCTCTTC -CAGCTCTTC	CGAACTTTCT	TCACTTT
999999999	999		66999999	999999		
	!			130		
HA HZ	TTCTTTT	TTTTCTCT	CTCAACTCCT	TG-TTATTTI TGGTCATTTI		
9999999 6666666-6		99999999	999-996696	666966		
				180		
HA HZ	ttggtct		AAGGTCTTCA	!! ATCATTATTA		
66666666	666666 					
	!			240		
HA HZ	TAAATTO	CACTTCAAA	AGACTCTCAT	TACACACCAC	STATTCTATT	

Figure 3 (E).

E. HarSSR1 (240)

HZ	!.	10 !	!	20 !	!.	30 !	!	40	!	50
HV HA	AAACAA	GGACAT	AGGTI	 FAACAA	AGTT	 ATTTAC	 ATCA(GTAGTT	TGTTO	TGG
HZ	!.	60 !	!	70 !	!.	0 0	!.	90 !	!	100
HV HA	GACTCC'	IGAGTT	CCCAT	TACTG	GTTag	gtgatt	gtgga	ctcagT	TTTTC	GAA
	!.	110	!.	120		130	!.	140	!	150 !
HZ HV HA	TTTGAT	 -CTG ICTGCT	TGI	GATGG	GTTGC'		TTGCT	GTTGT	TGTGA	
66777	 222 766488	3444	43356	66665	55545	566666	6			
	!.	160	!	170	!.	180	!	190	!	200
HZ HV	TTGTTG(CTGCTG	CTGTI	GTTGT	TGTT	GCTGCT	GCTG	CTGCTG	CTGTI	GTT
HA 57777	TTGTTG 6				66666		77	CTGCTG	TTGTT	'GCT
	!.	210		220	!.	230	!	240		250 !
HZ HV HA	GCTGCT(GTTGTT)	GTTGC-	TGI	TGTTG	CTGC'		TGCT			
777777 555555	777777 55666	7777	77666	66666	66666	666555	55			

Figure 3 (E, con't).

		260	270	280	290	300
	! .	!	! !	.!!	.!!!	!
ΗZ	-GCTGA	TGAAGTTG'	TTGTTGTTG	CTGATGCTGTT	GTTGGTATTGC	CTGAAC
HV	TGCTGA	TGAAGCTG'	TTGTTGCTG	CTGTTGT7	GTTGGTATTGC	CTGAAC
HA	TGCTGA	TGAAGCTG'	TTGTTGTTG	CTGATGCTGTT	GT TGGTATTGC	CTGAAC
67777	777777	77788888	77777576	7678888888]	
			8	8999999888		
					_	
		310	320	330	340	350
	! .	!	! !	.!!	. ! ! !	!
ΗZ	TTGATG	TGCCTGTT	GCATTTGCT	GATGGGTTTG-		
HV	TTGATG	TGCCTGTT	GCATTTGCT	GATGGGTTTG-		
НА	TTGATG	TGCCTgtt	gcatttgct	gatgggtttg(CTGCATGTGCTA	AAATA
99999	9999999	999888999	988887766	6666		
		360	370	380	390	400
	! .	!	! !	.!!	. ! ! !	!
ΗZ						
HV						
НА	TAGTTT	AATATAAT'	TAACTGGCA	GCCATATTGCT	TACGTTTTACGT	TTTTA:
		410	420	430	440	
	! .	!	! !	.!!	. ! ! !	
ΗZ						-
HV						-
HA	TTAAAA	AACAGATA:	AAGCTATAT.	AGAATCATGTO	GAAATAGTATTI	1

occurrence of some inversions at HaB60, a substitution in HaC87, and deletions in HaC14 and HarSSR1, showed a high number of identical nucleotide sites with the *H. armigera* repetitive motifs (Table 3). The length polymorphism detected in *H. virescens* and *H. subflexa* at the HaB60 locus revealed a large deletion of the repetitive array in the 140 bp allele of both species. However, with the complete sequence of one primer and a partial of the other 5'-end primer contained in the sequence read, it was still deemed to be a factual allele (Fig.3C).

The occurrence of null alleles in microsatellites is known to be an impediment to their successful application as markers in population genetic studies (Pemberton et al., 1995; Schlötterer and Pemberton, 1998; Liewlaksaneeyanawin et al., 2002), and have been implicated as a possible

cause for the low levels of heterozygosity found in Lepidoptera (Meglecz et al., 2004). Since only samples collected from laboratory populations were employed in this study, we probably restricted ourselves from determining some level of polymorphism, if any, in the loci studied from the three species, though the number of polymorphic microsatellites to date has been found to be typically low in Lepidoptera (Ji and Zhang, 2004). Given the inherent variability of the microsatellite flanking regions in Lepidoptera, further work, in particular controlled mating studies, will be needed to elucidate the frequency of null alleles in these species.

The specific repetitive nature of the microsatellite flanking regions found in Lepidoptera demonstrates the difficulty of isolating similar microsatellites from closely related species.

Table 3. Comparison between four *H. armigera* microsatellites and the repetitive sequences identified in three related lepidopteran species.

Locus	Species	Microsatellite sequence
	H. armigera	ATTT ATTT ATTT ATTT ATTT
HaC14 (160 bp)	H. zea	ATTT GTTT TATT ATTT
	H. virescens	GTT ATTT ATTT
	H. subflexa	ATTT ATTT ATTT ATTT
	H. armigera	CTG CTG TTG TTG (CTG) ₅ (TTG) ₂
HaB60 (160 bp)	H. zea	CTG CTG TTG TTG TTG (CTG) ₄ TTG CTG TTG
	H. virescens	CTG CTG TTG TTG TTG (CTG) ₄ TTG CTG TTG
	H. subflexa	CTG CTG CTG TTG TTG (CTG) ₄ TTG CTG TTG
HaB60 (140 bp)	H. armigera	CTG CTG TTG TTG (CTG) ₅ (TTG) ₂
	H. zea	-
	H. virescens	(CT-) (CTG) (A)() ₆ (CAA)(CAG)(GAA)
	H. subflexa	(CT-) (CTG) (A)() ₆ (CAA) (-AG)(GAA)
	H. armigera	(TC) ₅
HAC87 (118 bp)	H. zea	TT (TC) ₄
	H. virescens	-
	H. subflexa	-
	H. armigera	(TGC) ₂ GAT (TGY) ₄ GAT (TGY) ₃₅ (TGA) ₂ AGC (TGY) ₈
HarSSR1	H. zea	(TGC) ₂ GAAT (TGY) ₄ GAT (TGY) ₃₀ AGT (TGY) ₈
(240 bp)	H. virescens	TGC TGT GAAT (TGY) ₄ GAAT (TGY) ₃₁ AGC (TGY) ₇
	H. subflexa	-

	TGC TGT TGC TGT
(TCV)	TGT TGT TGC TGC TGC TGC TGT TGT TGT TGT
(101)30 -	TGC TGT TGT TGC TGC TGT TGC TGC TGA TGA
$(TGY)_8 =$	TGT TGT TGC TGA TGC TGT TGT
$(TGY)_4 =$	TGC TGT TGT TGT
(TCV) -	TGT TGT TGT TGC TGC TGC TGC TGC TGC TGT TGT
$(TGY)_{31} =$	TGC TGT TGT TGC TGC TGC TGA TGA
$(TGY)_7 =$	TGT TGT TGC TGC TGT TGT
(-) =	not detected

However, some of the data presented here extends the utility of previously developed microsatellites of one species to closely related members, and has the potential to be used as population genetic markers in other related lepidopteran species.

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13

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