

# Capillaria hepatica IN SMALL MAMMALS COLLECTED FROM SHOA PROVINCE, ETHIOPIA

Authors: FARHANG-AZAD, A., and SCHLITTER, D. A.

Source: Journal of Wildlife Diseases, 14(3): 358-361

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-14.3.358

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <a href="https://www.bioone.org/terms-of-use">www.bioone.org/terms-of-use</a>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

# Capillaria hepatica IN SMALL MAMMALS COLLECTED FROM SHOA PROVINCE, ETHIOPIA

A. FARHANG-AZAD, Department of Microbiology, School of Medicine, University of Maryland, 660 W. Redwood Street, Baltimore, Maryland 21201, USA.

D. A. SCHLITTER, Museum of Natural History, Carnegie Institute, 4400 Forbes Avenue, Pittsburg, Pennsylvania 15213. USA.

Abstract: A total of 1,110 small mammals has been examined for Capillaria hepatica in Shoa Province, Ethiopia. Nineteen of 308 (6.2%) Rattus rattus and 1 of 212 (0.5%) Praomys albipes were infected. The data indicate that C. hepatica was essentially limited to commensal rats (R. rattus).

#### INTRODUCTION

Capillaria hepatica has been reported from numerous mammalian hosts throughout the world. Although rodents are the principal hosts, 2,3 the parasite has been found in primates, insectivores, lagomorphs, carnivores, and artiodactyls.6,10 Although both genuine and spurious human infections with C. hepatica have been reported, presently it is difficult to estimate the zoonotic implications. Reports of human infections with C. hepatica in different parts of the world in the last few years could indicate that infection with this parasite is more common in humans than presently supposed.5,10 Even though documented human cases are relatively few, the majority of proven infections have ended fatally.5 Furthermore, it has been suggested that infection with a few C. hepatica may cause the "Eosinophilia of Undetermined Origin" which frequently is reported throughout the world.10

Since commensal rodents are the principal hosts, association between human and rodent reservoirs can be considered a potential health hazard, especially to children. Infection in domestic rodents in the United States and in many other countries is well-documented. 2,4,6,10 However, data on the occurrence of *C. hepatica* in commensal or sylvatic rodents is still lacking for much of the world, especially for Africa. In January-

March, 1976 and again in 1977, during the field studies on the ecology of murine typhus in Ethiopia, we examined the livers of small mammals for *C. hepatica*.

This paper reports the prevalence and distribution of *C. hepatica* in commensal and sylvan populations of small mammals in the Ethiopian highlands.

#### **MATERIALS AND METHODS**

A total of 1,110 small mammals was collected from five study areas within the Shoa Province, Ethiopia. The majority was collected by live trapping, the remainder by means of snap traps. Data recorded for each specimen usually included locality, date of collection, habitat, sex, age, body weight, external measurements and reproductive condition.

Screening for infection in the field was based primarily on gross examination of the liver for the *C. hepatica* lesions. However, all specimens judged to be infected by gross examination were preserved in 70% ethyl alcohol. Confirmation of infection, performed in Baltimore, Maryland, was accomplished by microscopic examination for the presence of *C. hepatica* eggs or fragments of adult worms. Severity of infection was judged by the appearance and abundance of lesions as described by Luttermoser.

#### **RESULTS**

Twenty-eight species of mammals were examined, but only 19 of 308 (6.2%) Rattus rattus and one of 212 (0.5%) Praomys albipes were infected. The prevalence of infection among rats collected from the different study areas is shown in Table 1. These varied from 1.9% for Koka in the Rift Valley, to 17.4% for rats collected from houses in Makanissa Village, a suburb of Addis Ababa. Of 75 juvenile rats examined, only two were infected (2.6%), whereas 17 of 233 adults were infected (7.3%).

Species not infected, together with numbers examined (in parentheses) were: three species of Crocidura (16); Procavia capensis (2); Tachyoryctes splendens (14); Graphiurus murinus (4); Tatera robusta (12); Acomys sp. (2); Arvicanthis abyssinicus and A. dembeensis (161); Dendromus lovati (12); Dendromus sp. (4); Desmomys harringtoni (84); Lophuromys flavopunctatus (43); Mastomys natalensis (119); Muriculus imberbis (1); Mus musculus (18); Mus sp. (67); Praomys fumatus (16); Praomys sp. (2); Stenocephalomys albocaudata (6); Otomys typus (2); Lepus capensis (1); Herpestes ichneumon (3); Genetta tigrina (2); and Ictonyx striatus (2).

Lesions vary from punctate to stringlike. The diffuse type lesion, containing eggs and considerable necrotic tissue, was observed in only two cases. Eggs were recovered from the livers of all the infected R. rattus and P. albipes.

#### DISCUSSION

The data indicate that in the areas surveyed, C. hepatica infection was essentially limited to commensal R. rattus; only a single specimen of the other 27 species of mammals examined was positive. The infected P. albipes was collected in a house; moreover all the R. rattus were collected in buildings, and these were only in the vicinity of Addis Ababa and at Koka, in the Rift Valley. Sylvatic murines, in contrast, were abundant in the fields and other outdoor areas; however, in certain places they were common in domiciles and other buildings, as was the case in the area where the infected Praomys was taken. C. hepatica infection was present in all the sites where R. rattus occurred, but the prevalence of infection was relatively low, ranging only from about 1.9% in Koka, to 17% in a suburb of Addis Ababa. In contrast, infection rates of 25-30% have been reported for R. rattus from the island of Ponape in the South Pacific

TABLE 1. Prevalence of Capillaria hepatica infection among rodents collected from human settlements.

	Addis Ababa City and Suburbs					
		Makanissa		Intoto	Intoto	
	Town Houses	Village Houses	Barn	Kedani Mehrat	Kedani Mariam	Koka (Houses)
Rattus rattus:						
No. examined	84	23	47	48	0	106
No. positive	7	4	3	3		2
% infected	8.3	17.4	6.3	6.3		1.9
Praomys albipes:						
No. examined	0	0	0	38	80	0
No. positive				1	0	
% infected				2.6	0	

(Jackson in Storer<sup>11</sup>), 90% or more for *R. norvegicus* in Baltimore, Maryland, USA,<sup>2</sup> and 48% for *Mastomys natalensis* in Johannesburg, South Africa. In South Africa, *C. hepatica* has been reported in other mammals such as mice, rabbits, hares, meerkats, and the gerbils; moreover, three cases of human infection have been reported in South Africa.<sup>1,8</sup>

Since C. hepatica infection in Shoa was virtually restricted to commensal rats living indoors, and a gamut of mammals can naturally acquire this nematode, the dearth of infection in native mammals in Ethiopa was due to

either the absence of Rattus in the areas examined or else the other small mammals had little contact with rat-infested foci. C. hepatica infection apparently is perpetuated primarily by ingestion of infective ova released from the liver after 1) death and decomposition of the host, 2) cannibalism, or 3) predation. Accordingly, the low population density of Rattus in the Ethiopian areas studied, and the low rate of infection in those hosts, may be factors contributing to the absence of C. hepatica in the indigenous mammals there. However, the present data are too limited to resolve that important point.

### Acknowledgements

We gratefully acknowledge the cooperation, suggestions and contribution of Dr. C.L. Wisseman, Jr. and Dr. R. Traub, Department of Microbiology, University of Maryland, who participated in the field work in Ethiopia and helped in various other ways.

Research supported in part by Contract N00014-76-C00393 of the Office of Naval Research and the Navy Medical Research and Development Command with the Department of Microbiology, University of Maryland, School of Medicine. The opinions and assertions herein are not to be construed as necessarily reflecting the views of the Department of the Navy.

## LITERATURE CITED

- COCHRANE, J.C., L. SAGORIN and M.G. WILCOCKS. 1957. Capillaria hepatica infection in man. S. Afr. Med. J. 31: 751-755.
- FARHANG-AZAD, A. 1977. Ecology of Capillaria hepatica (Bancroft, 1893) (Nematoda). I. Dynamics of infection among Norway rat populations of the Baltimore Zoo, Maryland. J. Parasit. 63: 117-122.
- ——. 1977. Ecology of Capillaria hepatica (Bancroft, 1893) (Nematoda). II. Eggreleasing mechanisms and transmission. J. Parasit. 63: 701-706.
- FREEMAN, R.S. and K.S. WRIGHT. 1960. Factors concerned with the epizootology of Capillaria hepatica (Bancroft, 1893) (Nematoda) in a population of Peromyscus maniculatus in Algonquin Park, Canada. J. Parasit. 44: 373-382.
- LAMMLER, G., H. ZAHNER, VOLLERTHUN, H. and R. RUDOLPH. 1974. Egg production and host reaction in *Capillaria hepatica* infection of *Mastomys* natalensis. In: Parasitic Zoonoses. E.J.L. Soulsby, ed. pp. 327-341 Academic Press, New York.
- LAYNE, J.N. 1968. Host and ecological relationship of the parasitic helminth Capillaria hepatica in Florida mammals. Zoologica 53: 107-123.
- LUTTERMOSER, G.W. 1938. An experimental study of Capillaria hepatica in the rat and mouse. Am. J. Hyg. 37: 321-340.

- 8. SILVERMAN, N.H., J.S. KATZ, and S.E. LEVIN. 1973. Capillaria hepatica infection in a child. S. Afr. Med. J. 47: 219-221.
- 9. SOLOMON, G.B., and C.O. HANDLEY, Jr. 1971. Capillaria hepatica (Bancroft, 1893) in Appalachian mammals. J. Parasit. 57: 1142-1144.
- and E.J.L. SOULSBY. 1973. Granuloma formation to Capillaria hepatica eggs. I. Descriptive definition. Exp. Parasit. 33: 458-467.
- STORER, T.J. (ed.) 1962. Pacific island rat ecology. Bernice P. Bishop Museum, Honolulu Bull. 225, 274 pp.

Received for publication 28 October 1977