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# First record of *Aegyptobia* Sayed (Acari: Tenuipalpidae) from South America: a new species and redescription of *A. pennatulae* Baker & Tuttle from Peru

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### Abstract

The genus Aegyptobia is recorded from South America for the first time, with the description of a new species collected from the branches of a native forest tree, Neltuma piurensis (Fabaceae), from Department of Piura in north-western Peru. The species A. pennatulae Baker & Tuttle was also collected from the same host, and is redescribed based on the types and new material. Observations during summer and autumn revealed A. pennatulae as the predominant species, and that N. piurensis is likely an accidental host for A. peruensis sp. nov.

Key words: Trombidiformes, flat mites, phytophagous mites, range extension, host extension, algarrobo

## Introduction

The Tenuipalpidae is distributed throughout the world and contains many economically significant pest species (Mesa et al. 2009; Beard et al. 2012; Castro et al. 2020). This family is currently comprised of 41 genera and 1105 described species (Mesa et al. 2009; Castro et al. 2024). Aegyptobia Sayed, 1950, with 111 described species (107 valid), is the third biggest in the family (Mesa et al. 2009; Castro et al. 2024). The genus has been reported from 30 countries in association with 50 families of host plants, mostly from Cupressaceae, Poaceae, Asteraceae, and Amaranthaceae, and many species were described from arid habitats with high temperatures (Castro et al. 2024). The 55 species of Aegyptobia reported in the Americas were described from the USA (35), Mexico (19), and Canada (1), with no species yet recorded from South America (Pritchard & Baker 1952, 1958; Baker & Pritchard 1953; Baker & Tuttle 1964; Baker & Tuttle 1972; Baker et al. 1975; Bury & Krantz 1977; Baker & Tuttle 1987; Mesa et al. 2009; Castro et al. 2024). Here we record the first two species from South America, by describing one new species A. peruensis sp. nov. from Peru, and redescribing the Mexican species A. pennatulae Baker & Tuttle based on types and new material collected in Peru.

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Currently, only six species of Tenuipalpidae are known from Peru (Escobar-Garcia & Andrade 2020; Escobar-Garcia et al. 2021; Huanca et al. 2021; Castro et al. 2024). The two newly recorded species were collected from Neltuma piurensis (L. Vásquez, Escurra & Huamán) C.E. Hughes & G.P. Lewis (Fabaceae; previously placed in Prosopis L.), locally known as algarrobo, a native forest tree in Peru recognized for its important social, economic and environmental benefits (SERFOR 2021; Hughes et al. 2022). The taxonomic placement of the genera Prosopis and Neltuma (and their relatives) varies globally, with names like Fabaceae, Mimosaceae, Leguminosae, and Caesalpiniaceae in use. Recent studies have suggested that Prosopis (approx. 57 species) is polyphyletic, and subsequently lead to the establishment of various segregate genera, including Neltuma. These genera are distributed mostly in the Americas, with a few outlying species in the Indian and north African regions (Hughes et al. 2022; Estrada-Castillon et al. 2024).

Previously, only one flat mite species, *Aegyptobia vannus* Pritchard & Baker, had been recorded from the *Prosopis/Neltuma* group, being described from *Neltuma glandulosa* (Torr.) in Mexico. *Aegyptobia vannus* was later recorded in Arizona, USA on *N. juliflora* (Sw.) Raf. [previously *Prosopis juliflora* (Sw.) DC.], and *Strombocarpa pubescens* (Benth.) A. Gray [previously *P. pubescens* Benth] (Baker & Tuttle 1964; Hughes *et al.* 2022; Castro *et al.* 2024). Algarrobo or *Neltuma piurensis*, was vital as a wild food source for pre-hispanic cultures, and some species underwent long distance translocation by humans and their livestock within the Americas in pre-Colombian times (McRostie *et al.* 2017). The collection of *A. pennatulae* in Peru and the resultant range extension from Mexico may reflect this historical context.

# **Materials and Methods**

Mites were collected from branches (20cm long) of the host plant N. piurensis at the campus of the Universidad Nacional de Piura (UNP), Peru (5°10'46.89"S; 80°37'04.31"W; 36 m.a.s.l.), between December 2020 to June 2021. Sampling involved collecting 10 branches with necrotic spots every 2 weeks. Samples were placed in paper bags and transferred to the laboratory of Entomology (SL01LA68), of the UNP, Peru. Mites were collected with a fine-tipped brush under the stereomicroscope (ZEISS Stemi 508) directly from the branches, mounted dorso-ventrally on glass microscope slides with 7µl Hoyer's medium, and dried in an oven at 50°C for 7 days (Walter and Krantz 2009). The coverslips were subsequently ringed with nail polish, and the morphological details of the mites were examined under a Nikon Eclipse E200 phase-contrast compound microscope. Specimens were measured with an ocular micrometer, and measurements are presented in micrometers (µm). Setal measurements are presented as a range for all specimens followed by the value for the holotype in square brackets. The lengths of setae were measured from the setal base to the tip; distances between setae bases were measured as the distance from center to center of setal bases. Body length was measured between the posterior border of the idiosoma and the tip of the rostrum. Setal terminology follows Mesa et al. (2009). The images were taken using an AxioCam MRc5 camera mounted on a Zeiss Axioscope AX10 Lab.A1 phase-contrast compound microscope or a Nikon Eclipse 80i with Capture 2.3 imaging software. The type specimens are deposited at the following institutions: SL01LA68—Acarology collection of the laboratory of Entomology of the UNP, Piura, Peru; IAM—Acarology Laboratory of the Instituto Agroforestal Mediterráneo of the Universitat Politècnica de València (UPV), Valencia, Spain; ESALQ-Department of Entomology and Acarology, Luiz de Queiroz College of Agriculture, University of São Paulo (USP), Brazil; NMNH—National Insect and Mite Collection, National Museum of Natural History, Smithsonian Institution, located at the Systematic Entomology Laboratory (SEL), USDA, Beltsville, MD, USA; QM—Queensland Museum, PO Box 3300, South Brisbane, Queensland 4101, Australia; and DZB—Collection of Acari, Departamento de Zoologia e Botânica, UNESP, São José do Rio Preto, State of São Paulo, Brazil.

#### Results

Family Tenuipalpidae Berlese, 1913
Genus Aegyptobia Sayed, 1950
Diagnosis —Based on Mesa et al. 2009, and Seeman & Beard 2011.
Type species—Aegyptobia tragardhi Sayed, 1950
Species group tragardhi
Type species—Aegyptobia tragardhi Sayed, 1950

Diagnosis—Based on Meyer (1979), and Kamran et al. (2016) with tarsal claws uncinate, claw-like.

# Aegyptobia pennatulae Baker & Tuttle, 1987

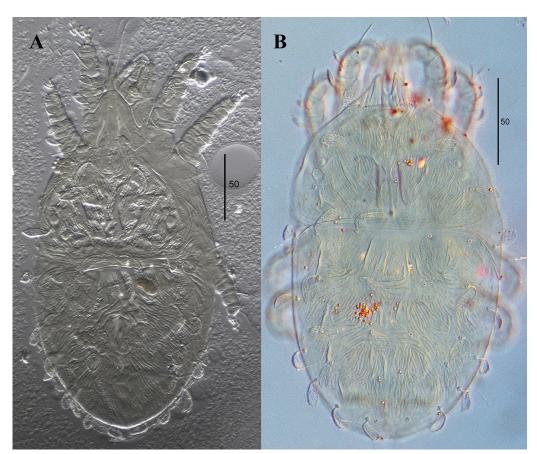
**Type material examined: Holotype:** female (Fig. 1A), Chapala, Jalisco, Mexico, ex. *Acacia pennatula* (Cham. & Schlecht.) Benth. (Mimosaceae), 29.vi.1974, coll. D.M. Tuttle, E.W. Baker and M.J. Abbatiello. **Paratypes:** female, on same slide as holotype; one female, Fresnillo, Zacatecas, Mexico, ex. *Mimosa biuncifera* Benth. (Mimosaceae), 03.vii.1974, coll. D.M. Tuttle, E.W. Baker and M.J. Abbatiello; one protonymph (poor condition), 19.2 km east of Morelia, Michoacan, Mexico, ex. *Mentha sp.* L., (Lamiaceae), 03.vii.1974, coll. D.M. Tuttle, E.W. Baker and M.J. Abbatiello. All in USNM.

Other material examined: 50 females, nine deutonymphs, and one protonymph, Piura, Peru, campus of the UNP, 05°10'46.89"S 80°37'04.31"W, 36 m.a.s.l., ex. *Neltuma piurensis* (Fabaceae), collected 18.i.2020 to 21.xi.2021, coll. H.A. Escobar-Garcia. Deposited in SL01LA68 (26 females, seven deutonymphs, and one protonymph), IAM (14 females), ESALQ (two females), NMNH (three females), QM (two females, one deutonymph), DZB (three females, one deutonymph).

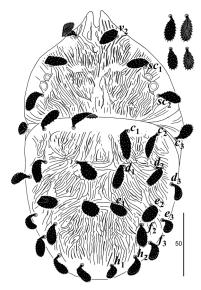
General diagnosis. Female: Tarsal claws uncinate. Anterior margin of prodorsal shield with deep broad notch; propodosomal and opisthosomal setae elongate obovate with weakly developed basal stalk, minute barbs on dorsal surface and fanned ridges on ventral surface; setae strongly curled to form scoop-shape; length of dorsal setae in range 12–25. Numerous minute pores on dorsal idiosoma; large pores apparently absent but with pore-like depressions between  $d_1$  and  $d_2$ . Genital and anal plates developed, smooth. Cuticle between 3a-4a with transverse striae becoming longitudinal laterally. Palp (trochanter to tarsus) 0-0-0-2-3(1 $\omega$ ). Legs I–IV setal counts: coxae 2–2-1-1; trochanters 1-1-2-1; femora 4-4-2-1; genua 2-2-0-0; tibiae 4-4-3-3; tarsi  $8(1\omega)-8(1\omega)-4-4$ . Tarsi I–IV with tc'' absent. Seta l'' absent on tibiae I–II. Seta d on femora I–III and genua I–II similar in form to dorsal setae (as in many species in the genus). Ratio  $v_2/v_2-v_3$ :  $0.6 \pm 0.0$  (0.5–0.7).

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Description. Female (n=13): (Figs. 1–8)
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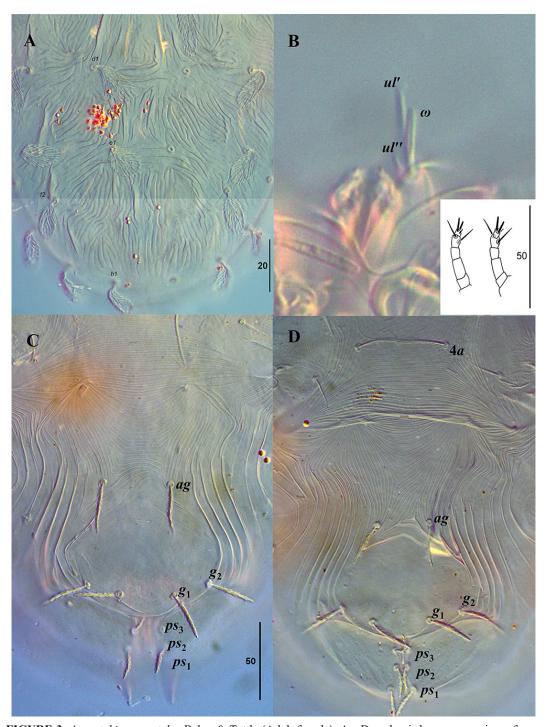
**Body.** Idiosoma oval in shape, with pair of lateral eyes on each side of propodosoma; length 215–240 [220].



 $\textbf{FIGURE 1.} \ \textit{Aegyptobia pennatulae} \ \text{Baker \& Tuttle adult female dorsal habitus: } \textbf{A---} holotype; \ \textbf{B---} specimen from Peru.$ 



**FIGURE 2.** Aegyptobia pennatulae Baker & Tuttle adult female from Peru. Dorsal habitus, with detail of dorsal and ventral aspects of setae.



**FIGURE 3.** Aegyptobia pennatulae Baker & Tuttle (Adult female). **A**—Dorsal opisthosoma, specimen from Peru; **B**—Palp tarsi, indicating three phaneres present [two eupathidia (ul' and ul'') and a solenidion ( $\omega$ )] with insertion of detail of the palp; **C** and **D**—Posterior venter, indicating variation in striae pattern anterior to setae ag. Note that the setae are barbed.

**Idiosomal dorsum.** Idiosoma oval in shape, with corrugated-rugose sculpturing, and a total of 16 pairs of elongate obovate, barbed setae (Figs. 1, 2). Dorsal cuticle with fine parallel sulci aligned in varying directions to form fine corrugate-rugose sculpturing. Prodorsum with regions of mostly oblique narrow parallel corrugations. Opisthosoma with a patchwork pattern of regions of narrow oblique and longitudinal corrugations (Fig. 3A); corrugations mostly longitudinal centrally becoming oblique laterally; pygidial region between  $e_1$ - $h_1$  with mostly longitudinal corrugations. Anterior margin of prodorsal shield with deep broad notch (Figs. 1B, 2); two pairs of eyes anterior to  $sc_2$  setae, and three pairs of setae  $(v_2, sc_1)$  and  $sc_2$ ; ratio  $v_2/v_2-v_2$ : 0.5–0.7 [0.7]. Numerous minute pores present, and pair pore-like depressions between setae  $d_1$  and  $d_2$ . Thirteen pairs of setae ( $c_{1-3}$ ,  $d_{1-1}$ )  $a_{1}, e_{1-3}, f_{2-3}, h_{1-2}$ ). Length of dorsal setae:  $v_{2}$  15–23 [23],  $sc_{1}$  18–22 [22],  $sc_{2}$  15–20 [20],  $c_{1}$  21–25 [24],  $c_2\ 18-22\ [21],\ c_3\ 12-17\ [17],\ d_1\ 16-21\ [21],\ d_2\ 17-21\ [21],\ d_3\ 12-15\ [15],\ e_1\ 13-16\ [16],\ e_2\ 15-18$ [17],  $e_3$  12–16 [16],  $f_2$  12–16 [16],  $f_3$  13–16 [16],  $h_1$  13–15 [14],  $h_2$  13–16 [16]. Distance between setae:  $v_2 - v_2$  28-34 [33],  $sc_1 - sc_1$  67-75 [75],  $sc_2 - sc_2$  84-94 [90],  $c_1 - c_1$  40-45 [43],  $c_2 - c_2$  82-92 [90],  $c_3 - c_3 \ 100 - 115 \ [110], d_1 - d_1 \ 32 - 38 \ [38], d_2 - d_2 \ 80 - 90 \ [88], d_3 - d_3 \ 104 - 112 \ [111], e_1 - e_1 \ 17 - 24 \ [23], e_2 - d_3 \ 104 - 112 \ [111], e_3 - e_4 \ 17 - 24 \ [23], e_4 - d_3 \ 104 - 112 \ [111], e_5 - e_6 \ 17 - 24 \ [23], e_5 - d_5 \ 104 - 112 \ [111], e_6 - e_6 \ 17 - 24 \ [23], e_7 - d_7 \ 104 - 112 \ [23], e_8 - d_8 \ 104 - 112 \ [24], e_8 - d_8 \$  $e_2$  72–82 [80],  $e_3$ – $e_3$  90–100 [95],  $f_2$ – $f_2$  62–68 [65],  $f_3$ – $f_3$  75–85 [80],  $h_1$ – $h_1$  23–26 [24],  $h_2$ – $h_2$  52–60 [60].

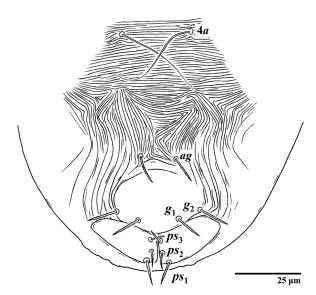


FIGURE 4. Aegyptobia pennatulae Baker & Tuttle adult female from Peru, posterior venter.

**Gnathosoma.** Ventral subcapitulum with a pair of short, slender and simple setae: m 5–8; distance between m–m 10–11. Palp with a pair of supracoxal setae ep; palp trochanter, femur, genu without setae; palp tibia with two tactile setae (d, l''); palp tarsus with three phaneres - two eupathidia: ul' (6–7) [5], and ul'' (3) and one solenidion  $\omega$  (6) (Fig. 3B).

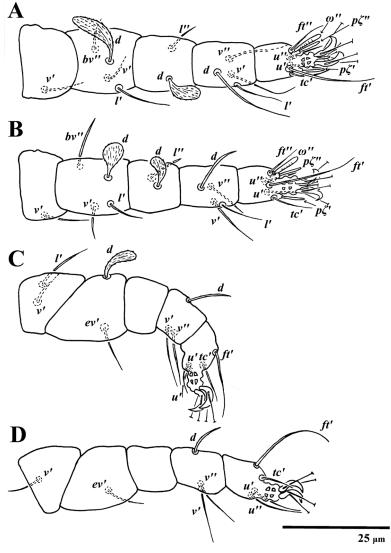
**Idiosomal venter.** Intercoxal area anterior to setae 1b-1a with fine longitudinal striae (can be indistinct); cuticle with strong transverse striae between setae 1a-3a; area between 3a to 4a with fine transverse striae becoming longitudinal laterally; area posterior to setae 4a with coarse transverse striations; area anterior to aggenital setae ag with variable pattern of fine striae: weak concave to transverse anteriorly becoming arching between ag-ag (Figs. 3C, 3D); strongly concave or broadly V-shaped anteriorly becoming oblique and longitudinal towards ag-ag (Figs. 3D, 4). Genital plate indicated by strong longitudinal folds in lateral cuticle, smooth, with setae  $g_{1-2}$  inserted along posterior margin. Pseudanal setae  $ps_{1-3}$  inserted longitudinally along medial margins of well-defined smooth anal plates (Figs. 3C, 3D, 4). Most ventral setae fine, smooth to weakly barbed, except 2c,

 $ag, g_{1-2}, ps_{1-3}$  thickened and densely barbed. Ratio 4a/ag: 2.3–3.1. Length of ventral setae: 1a 30–45 [45], 1b 13–27 [13], 1c 9–10 [10], 2b 15–18 [17], 2c 15–18 [16], 3a 10–13 [12], 3b 12–13 [13], 4a 28–46 [38], 4b 11–14 [13], ag 11–15 [14],  $g_1$  11–15 [15],  $g_2$  11–16 [16],  $ps_1$  8–10 [9],  $ps_2$  7–9 [8],  $ps_3$  6–7 [7]. Distance between setae: 1a–1a 18–22 [18], 3a–3a 34–36 [35], 4a–4a 22–27 [25], ag–ag 12–16 [15],  $g_1$ – $g_1$  14–18 [16],  $g_2$ – $g_2$  29–35 [30].

# Spermatheca. not visible

**Legs**. Legs I–IV short (Figs. 5A–D). Dorsal setae on femora I–III and genua I–II similar in form to dorsal body setae. Supracoxal setae eI. Number of setae on leg segments with solenidia (on tarsi) given in parentheses and included in setal counts: coxae 2-2-1-1; trochanters 1-1-2-1; femora 4-4-2-1; genua 2-2-0-0; tibiae 4-4-3-3; tarsi  $8(1\omega)$ - $8(1\omega)$ -4-4. Tarsi I–IV with tc'' absent. All apoteles terminate in two uncinate claws and pad-like empodium, each with tenent hairs. Solenidion  $\omega''$  ta I 7–8 [8], ta II 6–7 [7].

Male. Unknown.



**FIGURE 5.** *Aegyptobia pennatulae* Baker & Tuttle adult female from Peru, detail of left side legs: **A**—leg I; **B**—leg II; **C**—leg III; **D**—leg IV.

Deutonymph (n=9): (Fig. 6)

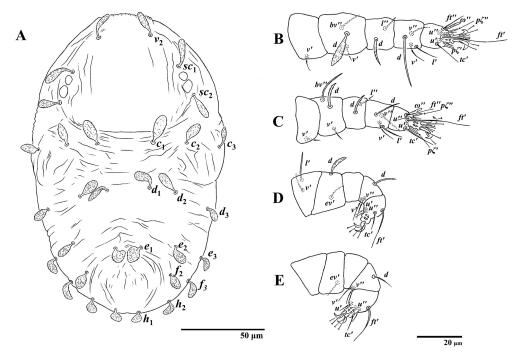
Body. Ovoid body, smaller than female; length (207–225).

**Idiosomal dorsum.** Weak prodorsal shield indicated, smooth; opisthosoma without sculpturing, with widely spaced transverse cuticular folds between  $c_1$  and  $e_1$ ; weak pygydial shield indicated by arching folds between setal pairs  $e_1$ ,  $f_2$ ,  $h_1$  (Fig. 6A). Anterior margin of prodorsal shield smoothly rounded, without projection or notch; ratio  $v_2/v_2-v_2$ : 0.4–0.5. Length of dorsal setae:  $v_2$  13–16,  $sc_1$  14–16,  $sc_2$  15–16,  $c_1$  15–20,  $c_2$  12–15,  $c_3$  10–13,  $d_1$  14–16,  $d_2$  14–16,  $d_3$  10–12,  $e_1$  10–12,  $e_2$  12–14,  $e_3$  9–12,  $f_2$  10–12,  $f_3$  10–14,  $h_1$  8–9,  $h_2$  9–10. Distance between setae:  $v_2-v_2$  30–33,  $sc_1-sc_1$  65–68,  $sc_2-sc_2$  78–85,  $c_1-c_1$  34–35,  $c_2-c_2$  70–72,  $c_3-c_3$  105–113,  $d_1-d_1$  26–29,  $d_2-d_2$  58–62,  $d_3-d_3$  92–103,  $e_1-e_1$  16–18,  $e_2-e_2$  58–63,  $e_3-e_3$  80–93,  $f_2-f_2$  48–53,  $f_3-f_3$  68–78,  $h_1-h_1$  15–17,  $h_2-h_2$  44–50.

**Gnathosoma.** Similar to that of female. Ventral subcapitulum with a pair of slender and simple setae: m 5–6 short. Distance between m–m 9–10. Palps as in adult female.

**Idiosomal venter.** Similar to female except with longitudinal striae between ag to  $g_1$  becoming oblique around genitoanal region; anal plates weakly developed. Pseudoanal setae  $ps_{1-3}$  inserted longitudinally along medial margins of anal plates. All ventral setae simple and finely barbed except 2c, ag,  $g_{1-2}$ , and  $ps_{1-3}$  densely barbed. Ratio 4a/ag: 0.9–1.5. Length of ventral setae: 1a 28–37, 1b 17, 1c 7, 2b 7, 2c 17–18, 3a 7–8, 4a 10–15, ag 9–11,  $g_1$  8–11,  $ps_1$  5–6,  $ps_2$  5–6,  $ps_3$  3–4. Distance between setae: 1a–1a 18–20, 3a–3a 32–35, 4a–4a 22–27, ag–ag 14–15,  $g_1$ – $g_1$  12–15.

**Legs.** Setal formula for legs I–IV and form of setae as in adult female except: trochanters 1-1-2-0; femora 3-3-2-1 (Figs. 6B–E). Solenidion  $\omega''$  on ta I 6–7 and ta II 5–6. Ontogenetic additions: setae  $\nu'$  added to tr I–III; d, l'' added to ge I–II; tc' added to ta I, II, IV (n.b. tc' added to ta III in protonymph).



**FIGURE 6.** *Aegyptobia pennatulae* Baker & Tuttle (Deutonymph); **A**—dorsal idiosoma; **B**—left leg I; **C**—left leg II; **E**—left leg IV.

Protonymph (n=1): (Fig. 7)

Body. Ovoid body, smaller than deutonymph; length 207.

**Idiosomal dorsum.** Similar to deutonymph but with less developed shields (Fig. 7A). Length of dorsal setae:  $v_2$  14,  $sc_1$  14,  $sc_2$  12,  $c_1$  15,  $c_2$  11,  $c_3$  11,  $d_1$  15,  $d_2$  12,  $d_3$  10,  $e_1$  9,  $e_2$  9,  $e_3$  9,  $f_2$  9,  $f_3$  8,  $h_1$  8,  $h_2$ 10. Distance between setae:  $v_2$ – $v_2$  30,  $sc_1$ – $sc_1$  55,  $sc_2$ – $sc_2$  70,  $c_1$ – $c_1$  30,  $c_2$ – $c_2$  65,  $c_3$ – $c_3$  98,  $d_1$ – $d_1$  20,  $d_2$ – $d_2$  52,  $d_3$ – $d_3$  84,  $e_1$ – $e_1$  14,  $e_2$ – $e_2$  55,  $e_3$ – $e_3$  68,  $f_2$ – $f_2$  40,  $f_3$ – $f_3$  48,  $h_1$ – $h_1$  12,  $h_2$ – $h_2$  35.

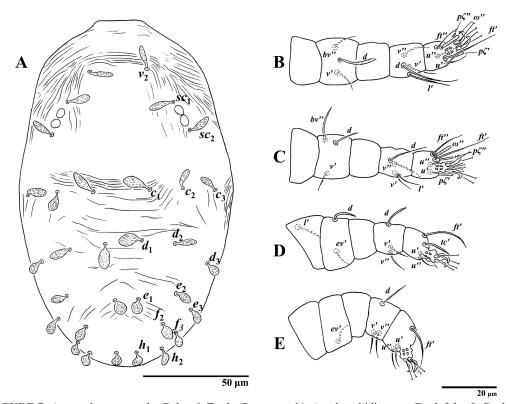
**Gnathosoma.** Similar to that of deutonymph. Ventral subcapitulum with a pair of slender and simple setae: m 5. Distance between m–m 10.

**Idiosomal venter.** Similar to that of deutonymph in all respects but with setae 2c, 4a, 4b,  $g_1$ , and  $g_2$  absent. Length of ventral setae: 1a 30, 3a 10, ag 8,  $ps_1$  5,  $ps_2$  5,  $ps_3$  3. Distance between setae: 1a-1a 18, 3a-3a 30, ag-ag 15.

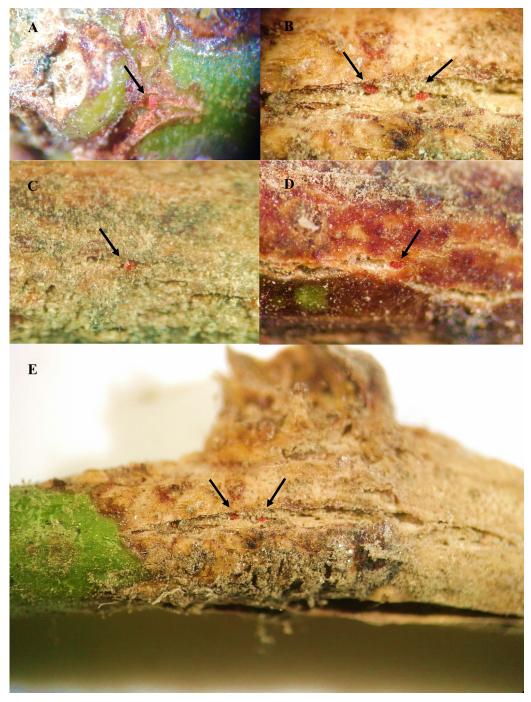
**Legs.** Setal formula for legs I–IV (Figs. 7B–E): coxae 2-1-1-0; trochanters 0-0-1-0; femora 3-3-2-1; genua 0-0-0-0; tibiae 4-4-3-3; tarsi  $7(1\omega)$ - $7(1\omega)$ -4-3. Solenidion  $\omega''$  ta I 6 and ta II 5.

Distribution. Mexico, Peru.

**Remarks.** All known life stages of these mites are carmine red. The male and larva were not collected. The setae on ge I–II were named d (deutonymphal seta) and l'' (larval seta) instead of l'-l'' (as delayed larval setae) due to the form of seta d present in other species of Aegyptobia (and many other tenuipalpid taxa) with three setae present on ge I–II (d, l', l'') which matches the dorsal setae.



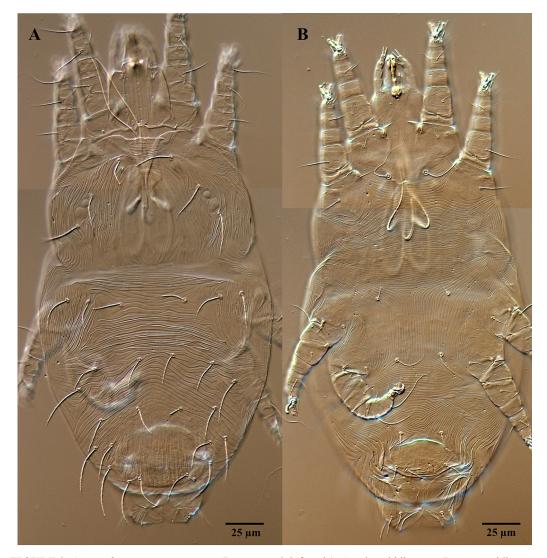
**FIGURE 7.** *Aegyptobia pennatulae* Baker & Tuttle (Protonymph); **A**—dorsal idiosoma; **B**—left leg I; C—left leg II; **D**—left leg IV.



**FIGURE 8.** Aegyptobia pennatulae Baker & Tuttle on the host plant: **A**–**E** symptoms of damage to branches of host tree algarrobo *Neltuma piurensis* (Fabaceae); arrows indicate individual females indicated except in **C** where an egg is indicated.

*Aegyptobia peruensis* Escobar-Garcia & Beard **sp. nov.** urna:lsid:zoobank.org:act:F721FDCA-8E46-41D2-9EB3-3EA85E214F38

**Type material: Holotype** female, Peru, campus of the UNP, Piura, (05°10'46.89"S; 80°37'04.31"W; 36 m.a.s.l.), ex. *Neltuma piurensis*, 1.ii.2021; coll. H.A. Escobar-Garcia. **Paratypes**: same data as holotype except collection dates – one female collected 18.i.2021 (SL01LA68); three females 18.i.2021 and one female 07.vi.2021 (IAM); one female 01.ii.2021 (NMNH); one female 24.v.2021 (QM).



**FIGURE 9.** *Aegyptobia peruensis* **sp. nov.,** (Paratype, adult female); **A**—dorsal idiosoma; **B**—venter idiosoma (image by F. Ferragut).

General diagnosis. Female: Tarsal claws uncinate. Anterior margin of prodorsal shield rounded, without projection or notch; dorsal setae slender lineate, barbed; dorsal shields not developed. Propodosomal cuticle smooth centrally with weak longitudinal striae sublaterally becoming stronger laterally. Dorsal opisthosoma with weak transverse striae between  $c_1$ – $e_1$ ,

increasingly arched towards  $e_1$ ; striae  $e_1$ – $h_1$  longitudinal becoming oblique laterally. Propodosomal and opisthosomal slender, linear, barbed setae; length of dorsal setae in range 17–34. Dorsal opisthosomal pores apparently absent. Genital and anal plates weakly developed, smooth. Cuticle between 3a–4a with longitudinal striae. Palp (trochanter to tarsus) 0-0-0-2-3(1 $\omega$ ). Legs I–IV setal counts: coxae 2-2-1-1; trochanters 1-1-2-1; femora 3-3-2-1; genua 0-0-0-0; tibiae 4-4-3-3; tarsi  $7(1\omega)$ - $7(1\omega)$ -3-3. Tarsi I–IV with tectal pair, tc' and tc'', absent. Ratio  $v_2/v_2$ – $v_2$ :  $0.7 \pm 0.0$  (0.6–0.8).

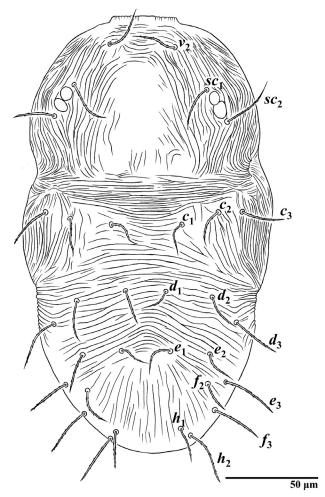


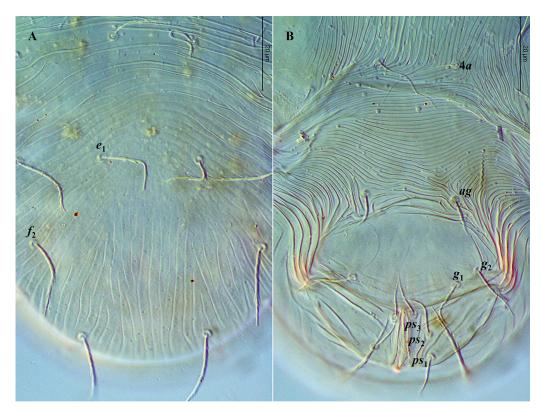
FIGURE 10. Aegyptobia peruensis sp. nov., (Holotype, adult female). Dorsal habitus.

Description. Female (n=8): (Figs. 9–14)

**Body.** Color in life carmine red; a pair of lateral eyes on each side of propodosoma; idiosoma oval in shape; length 250–305 [260].

**Idiosomal dorsum.** Idiosoma oval in shape, with striate sculpturing, and a total of 16 pairs of slender, linear, barbed setae (Figs. 9A, 10). Prodorsal shield with central region smooth, with weak longitudinal striae sublaterally becoming stronger laterally; anterior margin of prodorsal shield rounded without projection or notch; two pairs of eyes anterior to  $sc_2$  setae, and three pairs of setae  $(v_2, sc_1 \text{ and } sc_2)$ ; ratio  $v_2/v_2-v_2$ : 0.6–0.8 [0.8]. Opisthosomal shield with transverse striate sculpturing between  $c_1-e_1$ , striae increasingly arching towards posterior to  $e_1$ ; with longitudinal sculpturing

between  $e_1-h_1$  becoming oblique laterally (Figs. 10, 11A); cuticle between  $c_1$  to  $c_2$  often smoother or with fewer striae; cuticle lateral to  $c_2$  with longitudinal striae becoming stronger laterally. Large pores apparently absent. Thirteen pairs of setae ( $c_{1-3}$ ,  $d_{1-3}$ ,  $e_{1-3}$ ,  $f_{2-3}$ ,  $h_{1-2}$ ). Length of dorsal setae:  $v_2$  22–25 [25],  $sc_1$  25–26 [25],  $sc_2$  26–30 [26],  $c_1$  18–22 [18],  $c_2$  20–21 [20],  $c_3$  24–30 [24],  $d_1$  18–20 [19],  $d_2$  20–24 [22],  $d_3$  28–30 [30],  $e_1$  17–20 [17],  $e_2$  19–22 [20],  $e_3$  30–34 [30],  $f_2$  20–22 [22],  $f_3$  28–32 [28],  $h_1$  18–21 [18],  $h_2$  25–30 [27]. Distance between setae:  $v_2-v_2$  32–35 [32],  $sc_1-sc_1$  67–72 [67],  $sc_2-sc_2$  85–100 [85],  $c_1-c_1$  38–45 [38],  $c_2-c_2$  78–95 [78],  $c_3-c_3$  100–120 [100],  $d_1-d_1$  21–24 [21],  $d_2-d_2$  70–78 [70],  $d_3-d_3$  93–100 [93],  $e_1-e_1$  25–28 [26],  $e_2-e_2$  66–74 [66],  $e_3-e_3$  80–88 [80],  $f_2-f_2$  60–65 [62],  $f_3-f_3$  66–70 [66],  $h_1-h_1$  30–35 [33],  $h_2-h_2$  40–44 [40].



**FIGURE 11.** Aegyptobia peruensis **sp. nov.** adult female: **A**—posterior dorsal opisthosoma showing detail of striate pattern; **B**—posterior venter.

**Gnathosoma.** Ventral subcapitulum with a pair of short, slender and simple setae: m 5–6 [6]; distance between m–m 9–10[9]; with a pair of supracoxal setae ep on dorsal infracapitulum. Palp trochanter, femur, genu without setae; palp tibia with two tactile setae (d, l''); palp tarsus with three phaneres - two eupathidia (ul') and ul'') and one solenidion  $(\omega)$ .

**Idiosomal venter.** Cuticle between setae 1b to 1a with very fine longitudinal striae (can be indistinct); cuticle between setae 1a-3a with transverse striae; area between 3a-4a with strong longitudinal striations; 4a to ag with transverse striations becoming oblique laterally (Figs. 9B, 11B, 12). Genital plate smooth. Genital setae  $g_{1-2}$  inserted along posterior margin of plate. Pseudoanal setae  $ps_3$  inserted in anteriomedial corners of anal plates, with  $ps_{1-2}$  inserted off medial margins in longitudinal row; plates with oblique striae (Figs. 11B, 12). All ventral setae simple and apparently smooth except 2c, ag,  $g_{1-2}$ , and  $ps_{1-3}$  barbed. Ratio 4a/ag: 2.1–3.0 [2.4]. Length of ventral setae: 1a

30–32 [30], 3a 8–10 [10], 4a 22–45 [22], ag 15–18 [16],  $g_1$  15–20 [15],  $g_2$  18–22 [18],  $ps_1$  12–15 [12],  $ps_2$  12–15 [12],  $ps_3$  10–13 [10]. Distance between setae: 1a–1a 22–25 [22], 3a–3a 40–49 [40], 4a–4a 26–30 [26], ag–ag 25–31 [25],  $g_1$ – $g_1$  20–25 [23],  $g_2$ – $g_2$  35–40 [36].

**Spermatheca** (Fig. 12). Spermathecal tube narrow, sclerotized basally, membranous distally, termination not discernible.

Leg phanerotaxy. Legs I–IV short (Figs. 13A–D). Seta l' on tr III long, barbed; unguinal setae u'-u'' fine with few short barbs. Supracoxal setae eI. Number of setae on leg segments, with solenidia (on tarsi) given in parentheses and included in setal counts: coxae 2-2-1-1; trochanters 1-1-2-1; femora 3-3-2-1; genua 0-0-0-0; tibiae 4-4-3-3; tarsi  $7(1\omega)-7(1\omega)-3-3$ . Tectal pair, tc' and tc'', absent on tarsi I–IV. All apoteles terminate in two uncinate claws and a pad-like empodium, each with tenant hairs. Solenidion  $\omega''$  on ta I 7–8 [8] and ta II 6–7 [6].

**Etymology.** The specific epithet "*peruensis*" is derived from the country of origin, Peru. **Distribution.** Peru, Department of Piura, north-western region, campus of the UNP.

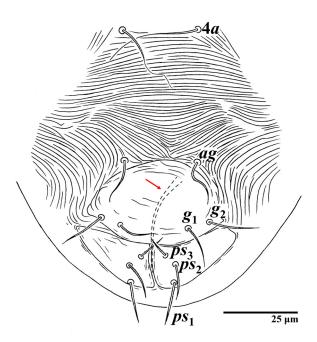
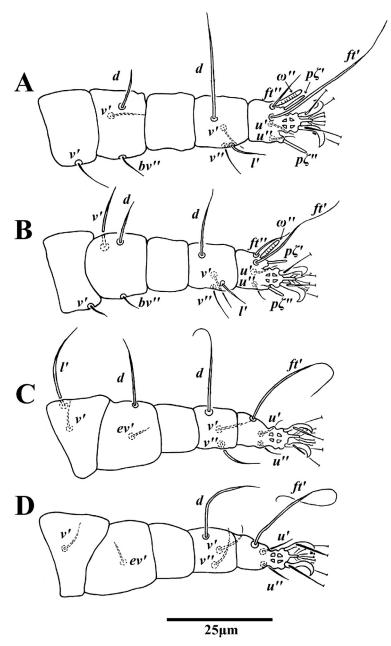


FIGURE 12. Aegyptobia peruensis sp. nov. adult female posterior venter; red arrow indicating duct of spermatheca.

## Differential diagnosis

The closest region to Peru where *Aegyptobia* has been recorded is Mexico, with 19 species. Three Mexican species have been recorded from Fabaceae and/or related host plants (as discussed in Introduction)—*A. cassiae* Baker & Tuttle, *A. pennatulae*, and *A. vannus* Pritchard & Baker.

Based on descriptions only, A. peruensis sp. nov. (Aper) can be easily distinguished from A. cassiae (Acas) using the dorsal sculpture pattern—Aper has striate sculpture (prodorsum mostly longitudinal, opisthosma mostly transverse) compared with Acas which has regular, areolate-rugose sculpture; from A. pennatulae (Apen) using the dorsal sculpturing and shape of dorsal setae—Aper with lineate striate cuticle and slender, linear, barbed setae, compared to Apen with a patchwork of narrow parallel corrugations and elongate obovate setae; and from A. vannus (Avan) using the shape of the dorsal setae—Aper with slender, linear, barbed setae compared to Avan with distinctly orbicular dorsal setae.

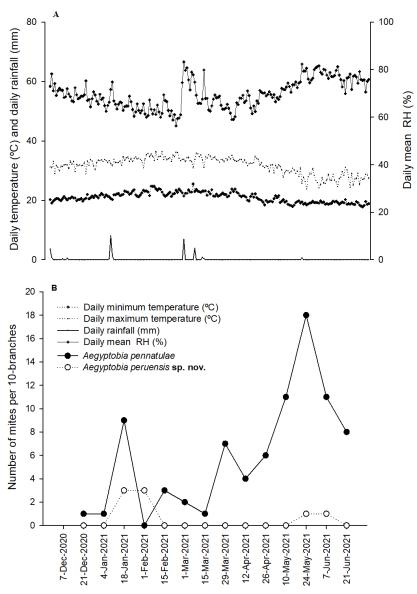


**FIGURE 13.** *Aegyptobia peruensis* **sp. nov.** adult female, detail of left legs: **A**—leg I; **B**—leg II; **C**—leg III; **D**—leg IV.

Further afield, there are three species collected in Arizona, USA, that all share a similar striate dorsal sculpturing with A. peruensis **sp. nov.**—A. acacia Baker & Tuttle (Aaca) from Acacia (Mimosaceae), A. eriogonum Baker & Tuttle (Aeri) from Polygonaceae, and A. fallugia Baker & Tuttle (Afal) from Rosaceae. Based on descriptions available in the literature, Aper is most similar to Aeri, but can be separated based on the number of setae on genua I—II—Aper ge I—II without setae compared with Aeri ge I—II with 1 seta. Aper can be separated from Aaca using dorsal sculpturing and setae on genua I—II: Aper with transverse striae between setae  $c_1$  and  $d_1$  and ge I—II are without

setae, while Aaca is distinctly smooth between setae  $c_1$  and  $d_1$  with a narrow band of transverse striae just anterior to the D row (which Baker & Tuttle 1972) point out is reminiscent of the dorsal morphology generally seen on male tenuipalpids), and ge I–II have setae. Aper can be separated from Afal using dorsal sculpturing—Aper with transverse striae from  $c_1$  to  $e_1$ , while Afal has longitudinal striae centrally between  $c_1$  and  $d_1$ .

Outside of the Americas, Aegyptobia peruensis sp. nov. shares striate dorsal sculpturing with A. nummulus Chaudhri 1972 (Anum) from Cupressaceae in Pakistan, but Aper can be separated by having uncinate claws (tragardhi species group) and the palp femur without setae, while Anum has pad-like claws (macswaini species group) and one seta on the palp femur.



**FIGURE 14.** Climatic and population data: A—climatic variation data; B—population fluctuation of *Aegyptobia* spp. on branches of host tree algorrobo *Neltuma piurensis* (Fabaceae).

# Ecological information

Aegyptobia pennatulae and A. peruensis sp. nov. were sampled and observed during the summer (January–February) and autumn seasons (May–June). The former species was encountered more frequently during the periods of flowering and subsequent fruit development in summer, and populations were negatively affected during periods of precipitation (Fig. 14). Females and immatures were often observed resting or potentially feeding in the cracks and divots in the bark of the host plant (Fig. 8). Irregularly ovoid red eggs were laid in protected areas such as depressions and split bark (Fig. 8C). In contrast, only female A. peruensis sp. nov. were observed and collected during this study, and in much lower numbers than A. pennatulae (Fig. 14). The lack of immatures may indicate that this host-plant association is accidental.

The environmental conditions during the autumn season were as follows: minimum temperatures ranged from 18.9 to 22.3°C, maximum temperatures 27.0 to 33.8°C, and the relative humidity ranged from 65.1 to 79.6%.

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