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Phenotypic Variation in Brook Trout *Salvelinus fontinalis* (Mitchill) at Broad Spatial Scales Makes Morphology an Insufficient Basis for Taxonomic Reclassification of the Species

Shannon L. White¹, David C. Kazyak¹, Richard C. Harrington², Matt A. Kulp³, Jacob M. Rash⁴, T. Casey Weathers⁵, and Thomas J. Near²

It was recently proposed that there are three new species of *Salvelinus* with microendemic distributions in the Great Smoky Mountains National Park, Tennessee, USA. The three species of *Salvelinus* were hypothesized to be distinct from their congener Brook Trout *S. fontinalis* based on three meristic traits—pored lateral-line scales, vertebral counts, and number of basihyal teeth. After analyses that included specimens sampled from a larger portion of the geographic range of *S. fontinalis*, we conclude that the three populations of *Salvelinus* recently described as new species are not morphometrically distinct from Brook Trout and consider all three to be synonyms of *S. fontinalis*. Moreover, the low number of specimens originally examined conflates morphological differences among populations with sexual dimorphism and/or phenotypic plasticity, both of which are documented extensively in Brook Trout but were not controlled for in the species descriptions. While there is currently insufficient phenotypic or genotypic evidence to support the hypothesis of three new species that are distinct from *S. fontinalis*, we acknowledge the need to understand the unique selection pressures that shape evolutionary trajectories in small, isolated populations of Brook Trout and to conserve evolutionarily significant sources of genotypic and phenotypic diversity. To that end, we provide comments on research opportunities to support Brook Trout conservation, including the importance of collaborative, range-wide phylogenetic studies to identify the most appropriate scales of management efforts.

BROOK Trout, *Salvelinus fontinalis*, is a widely distributed species of coldwater fish that has a native distribution spanning from eastern Canada to northern Georgia, USA (Fig. 1). Recently, Stauffer (2020) proposed that three tributaries to the Pigeon River in the Great Smoky Mountains National Park (GSMNP), Tennessee, USA, each contain a unique and previously undescribed species of *Salvelinus* that is phenotypically distinct from *S. fontinalis*. These species designations were based primarily on three meristic traits, including pored lateral-line scales, vertebral counts, and number of basihyal teeth. Stauffer (2020) described these traits as being diagnosable, derived characters indicative (*sensu* Cracraft, 1983) of distinct species.

We have considerable concerns about the analysis and interpretation of the data used by Stauffer (2020) and question the validity of the three proposed species. Most notably, the proposed species in Tennessee were described from a sample size of just ten individuals, which were compared only to Brook Trout populations in New York described in Stauffer and King (2014). In addition to lacking sufficient statistical power, limited comparisons to published species accounts disregard the natural variation in phenotypes that occurs within populations and at regional and range-wide scales. When this variation is ignored, it is not possible to characterize the distribution of a trait, and small samples are more likely to erroneously suggest significant differences in trait values between groups.

Given the small sample sizes, we hypothesized that a more inclusive analysis that compared data presented in Stauffer (2020) to additional regional and range-wide species accounts would not support the distinctiveness of the three proposed species from *S. fontinalis*. Moreover, given that the clade of *Salvelinus* has undergone multiple adaptive radiations across its Holarctic distribution (Muir et al., 2016), we consider a comprehensive analysis of morphology a minimum requisite for evaluating the distinctiveness of new species. In particular, careful examination is needed to account for the effects of phenotypic plasticity and sexual dimorphism, both of which are well documented in species of *Salvelinus* (Klemetsen, 2010; Muir et al., 2016) but were not considered by Stauffer (2020). In Brook Trout, phenotypic plasticity is frequently noted among different life history morphs, with highly migratory anadromous and potamodromous populations displaying significant differences in size, body shape, coloration, and fin morphology among individuals (Power, 1980; Morinville and Rasmussen, 2008; Varian and Nichols, 2010). However, even within homogenous habitats, individual morphology can be correlated to physical and chemical habitat (Zimmerman et al., 2006; Zastavniouk et al., 2017), temperature, feeding mode (Woods et al., 2013), and interspecific competition (Nakano et al., 2020), and these factors can vary across short temporal scales as a result of environmental and/or demographic stochasticity. Significant plasticity has been noted in two of the diagnostic

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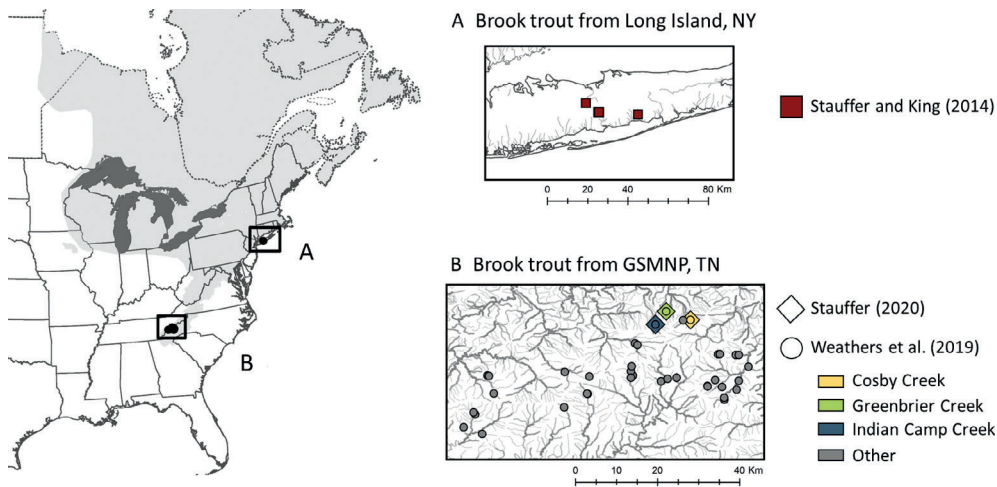


Fig. 1. Native distribution of Brook Trout (shaded gray area) in the United States and Canada, with Brook Trout used in our comparative analyses originating from survey locations located in panels A and B. The three streams in Long Island, NY, surveyed by Stauffer and King (2014) are shown in panel A. Panel B shows streams from the Great Smoky Mountains National Park (GSMNP) surveyed by Weathers et al. (2019; circles) and Stauffer (2020; diamonds), with the three streams included in both studies symbolized with matching colors (Cosby Creek: yellow; Greenbrier Creek: green; Indian Camp Creek: blue). Streams included in Weathers et al. (2019) but not included in Stauffer (2020) are shown in gray circles.

traits proposed by Stauffer (2020)—pored lateral-line scales and vertebral counts—which are known to exhibit gradual latitudinal variation in many taxa with variation even noted within single populations due to differences in incubation temperature (Garside, 1966; Fowler, 1970; Wallace, 1973).

Trait variation can be further exacerbated by sex and ontogeny (Zastavniouk et al., 2017), with mature males developing deeper body forms and more exaggerated jaw and head morphologies, particularly during spawning season (Kazyak et al., 2013; Stauffer et al., 2016). However, it can be difficult to reliably sex Brook Trout using nonlethal measures (but see Kazyak et al., 2013), and dissection is often not viable given the conservation status of many populations. While plasticity and sexual dimorphism do not preclude the use of morphology for species delineation, it does emphasize the need for robust sample sizes across relevant temporal and spatial scales, and ideally the use of morphology in conjunction with genetic information when attempting to infer phylogenetic relationships (Kuparinen and Hutchings, 2019).

Given concerns about small sample sizes and limited consideration for extragenetic sources of variability, we sought to independently investigate the strength of the evidence for three new species of *Salvelinus*. We conclude that the author's species designations are unfounded due to: 1) the inability to discriminate the three proposed species of *Salvelinus* from *S. fontinalis*, 2) broad overlap of the meristic traits that were proposed as being diagnostic of the three proposed species with values observed in *S. fontinalis*, and 3) methodological insufficiencies that do not account for confounding sources of phenotypic variation.

Here, we provide results from our analyses that support our assessment that there is presently insufficient evidence for the distinctiveness of the three species proposed by Stauffer (2020). However, it is not our intent to diminish the role that processes such as isolation, local adaptation, and genetic drift can have on the evolutionary trajectories of Brook Trout populations, including the potential for speciation. Rather, we feel these topics are best discussed within the context of ongoing, range-wide efforts and the large body of existing literature that seeks to understand the unique selection pressures shaping the evolutionary trajectories of popula-

tions of *Salvelinus*. Accordingly, we provide comments about future research opportunities that would assist in the identification of evolutionary significant sources of variation and the conservation of Brook Trout at appropriate spatial scales.

MATERIALS AND METHODS

Data sources used for comparisons.—We compare data presented in Stauffer (2020) to: 1) data presented in Stauffer and King (2014) that were used to describe the neotype of *S. fontinalis* from Long Island, New York (Fig. 1A), 2) independent meristic counts from 38 Brook Trout populations located in the GSMNP described by Weathers et al. (2019; Fig. 1B), and 3) trait data reported in peer-reviewed literature and governmental agency reports. We provide a more detailed summary of these three data sources in the Supplemental Appendix (see Data Accessibility).

We direct readers to Stauffer (2020) for a description of the author's sample collection and data analysis. Briefly, Stauffer (2020) measured 23 morphometric and 10 meristic traits for ten fish from each of Cosby, Indian Camp, and Greenbrier Creeks within the GSMNP. These three collection locations are all located within the same 12-digit Hydrologic Unit Code (HUC 12) watershed and are separated by a waterway distance of no more than 19 km (Fig. 1B).

We contacted the author, who provided the original, individual-level data used in Stauffer (2020) and the individual-level data from Stauffer and King (2014) from populations in New York. However, analysis of the individual-level data failed to reproduce many of the summary statistics reported in the data tables presented in Stauffer (2020), and the number of vertebrae and basihyal teeth were absent. There were also inconsistencies between the individual-level data from populations in Long Island, New York and data presented in Stauffer and King (2014). Accordingly, we report two sets of values when summarizing data from Stauffer (2020) and Stauffer and King (2014)—one that provides the data as reported in the respective manuscript and another set of values calculated from the individual-level data provided by the author.

We also include data from Weathers et al. (2019), which described the morphometric and meristic variation of Brook Trout from 35 streams in the GSMNP (Table S1; see Data Accessibility). Although not included in the original publication, Weathers et al. (2019) also independently collected data from the three Stauffer (2020) sites (Cosby, Indian Camp, and Greenbrier Creeks; Fig. 1B) that we included in our analyses for a total of 38 populations from the GSMNP. Because Weathers et al. (2019) included an independent analysis from more sites, we can compare Weathers et al. (2019) and Stauffer (2020) to infer the amount of within- and between-stream variation in trait values relative to other streams that occur within the same region (i.e., HUC 4 watershed). Notably, though both studies included morphometric and meristic data, we limit our comparisons to meristic data because morphometric data were not indicated by Stauffer (2020) to be diagnostic, and because there is less subjectivity in collection methods for meristic data.

Stauffer and King (2014) and Weathers et al. (2019) did not include vertebral or basihyal tooth counts in their studies. To understand how vertebral counts compare to other Brook Trout species accounts, we compared the summary data presented in Stauffer (2020) to published species descriptions and experimental studies (sources provided in the Table S2; see Data Accessibility). Vertebral counts are infrequently reported in species accounts, and so the spatial extent of our comparisons is limited with most reported values originating from populations that occur at more northern latitudes. We were unable to locate independent records for basihyal tooth counts and discuss the limitation of this variable in more detail below.

Data analysis.—To evaluate claims of unique meristic identity in the proposed new species of *Salvelinus*, we quantified patterns of phenotypic variation among putative species using the ten meristic traits measured in Stauffer and King (2014), Weathers et al. (2019), and Stauffer (2020; see original publications for variable descriptions). Although specimens from all localities outside of Cosby, Greenbrier, and Indian Camp Creeks would currently be classified as *S. fontinalis* under the delimitation proposed by Stauffer (2020), we expected broad differences in phenotype between New York and GSMNP populations due to the separation of northern Atlantic and southern Atlantic populations into distinct phylogenetic assemblages (Danzmann et al., 1998; Stauffer and King, 2014). Accordingly, we treated samples from New York and the GSMNP as two separate populations of *S. fontinalis*.

We used a principal components analysis (PCA) to make multivariate comparisons of meristic traits among populations and calculated pairwise Mahalanobis distances, which are unitless measurements that summarize the differences between group means along all axes of the PCA. To evaluate the ability of meristic traits to correctly classify species of *Salvelinus* into distinct categories, we conducted a cross-validation linear discriminant analysis (LDA) using the MASS package (Venables and Ripley, 2002) in R v. 4.0.2 (R Core Team, 2020). For these analyses, we used a classification model based on five categories, which included each of three populations surveyed by Stauffer (2020) from Cosby, Greenbrier, and Indian Camp Creeks, *S. fontinalis* from other watersheds in the GSMNP, and *S. fontinalis* from New York.

For this analysis, we combined data collected by Weathers et al. (2019) with data from Stauffer (2020) for Cosby, Indian Camp, and Greenbrier Creeks. Our assumption was that if fish from the three study streams did represent new species as proposed by Stauffer (2020), then Weathers et al. (2019) would have inadvertently collected data from the three new species of *Salvelinus*, rather than Brook Trout. If so, adding the additional data from Weathers et al. (2019) would have either no effect, or potentially increase the power to detect significant differences among populations through increased sample size.

To account for the potential confounding effects of regional population structure, which could have occurred when comparing the individual streams from Stauffer (2020) to the collective GSMNP group that included the 35 other sites surveyed by Weathers et al. (2019), we reran this analysis and treated each stream as an independent factor (i.e., 41 total streams including 38 from the GSMNP and 3 from Long Island, New York). Inferences from the two analyses were congruent, and, for concision, we report the results of the individual stream analysis in the supplement (Table S3; see Data Accessibility).

RESULTS

Discriminatory ability of meristic traits.—Principal components analysis of meristic traits showed a clear distinction between *S. fontinalis* in New York and all southern Appalachian populations from the GSMNP on the first axis of the PCA (Fig. 2). This axis, which explained 22.3% of the total variation, was most closely associated with parr marks (variable loading = -0.31), number of teeth on lower left jaw (0.53), and pored scales posterior to the lateral-line scales (0.44). There was considerable overlap on PC1 and PC2 among all GSMNP populations, including the three sites surveyed by Stauffer (2020). In accordance with this finding, pairwise Mahalanobis distances were smaller among GSMNP populations than were Mahalanobis distances between GSMNP populations and populations from New York (Table 1).

Results from the LDA also suggested that the three populations described in Stauffer (2020) were not phenotypically distinct. While the LDA was able to correctly identify Brook Trout from New York, it correctly classified only 50% of individuals from Cosby Creek, 60% from Greenbrier Creek, and 55% from Indian Camp Creek (Table 2).

Comparison of diagnostic traits to Brook Trout.—Stauffer (2020) proposed that the specimens from Cosby, Greenbrier, and Indian Camp Creeks were diagnosable using three traits: pored lateral-line scales, vertebral counts, and number of basihyal teeth. However, when data presented in Stauffer (2020) are compared to additional populations, the three meristic traits do not appear to be diagnostic at regional and range-wide scales and rather support a pattern of gradual variation in phenotype across the distribution of Brook Trout. For example, individual pored lateral-line scale counts documented in Weathers et al. (2019) for 38 streams in the GSMNP differed by up to 39 scales, and this range generally encompassed the variation reported by Stauffer (2020; Fig. 3A, B). Moreover, direct comparison of pored lateral-line scale counts from the three populations surveyed by both studies suggests that there may be significant individual and/or

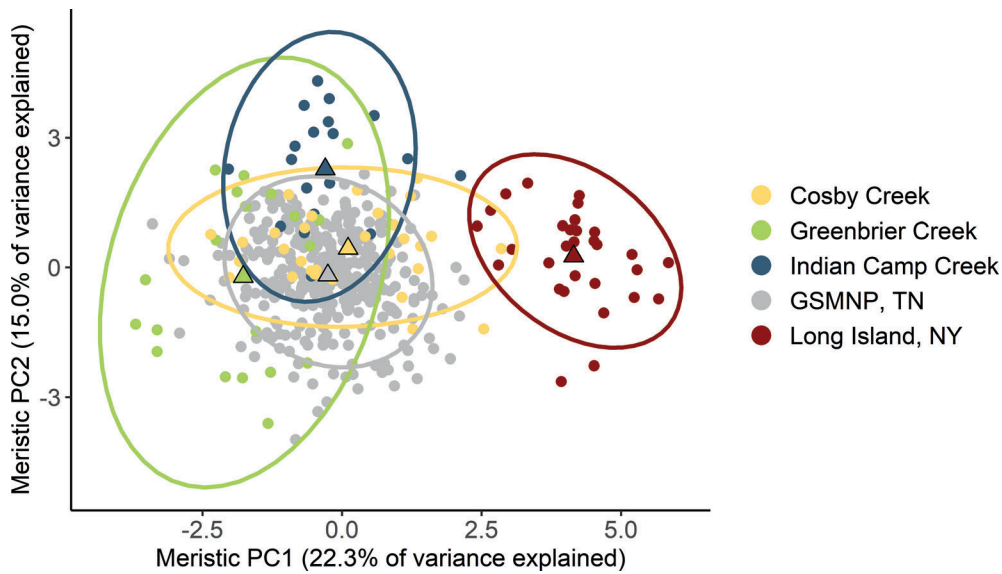


Fig. 2. First two dimensions of principal components analysis (PCA) of ten meristic traits for five populations of *Salvelinus*. The populations analyzed included the three surveyed by Stauffer (2020) and Weathers et al. (2019) from Cosby (yellow), Greenbrier (green), and Indian Camp (blue) Creeks, collections from Weathers et al. (2019) for 35 additional streams in the Great Smoky Mountains National Park (GSMNP; gray), and three populations from Long Island, NY described by Stauffer and King (2014; red). Ellipses envelop 95% of variation for each population, and population centroids are indicated by a triangle.

interannual variation within streams, in which case the range of values reported by Stauffer (2020) is not a full representation of the trait's distribution within the populations studied. For instance, data for Cosby Creek are similar between Stauffer (2020) and Weathers et al. (2019); however, counts from Indian Camp and Greenbrier Creeks, which were noted as having diagnostically low and high counts, respectively, in Stauffer (2020; but see Fig. 3B for discrepancies between the provided and published datasets), are close to the median value in the dataset collected by Weathers et al. (2019). Findings from Weathers et al. (2019) are consistent with those of Harris et al. (1978), who indicated that counts of total lateral-line scales—which are presumably correlated to counts of pored lateral-line scales—for Indian Camp Creek were approximately average relative to other populations within the GSMNP and across the range of Brook Trout (Cosby and Greenbrier Creeks were not evaluated by Harris et al. [1978]). Discrepancies between the datasets demonstrate the spatial and temporal variability in morphological and meristic traits and highlight the need for larger sample sizes across space and time in order to characterize intra- and inter-specific variation.

Weathers et al. (2019) did not collect vertebral or basihyal tooth counts, and we do not have individual-level data for these traits from Stauffer (2020) or Stauffer and King (2014). However, the three populations of *Salvelinus* surveyed by Stauffer (2020) did not differ significantly from one another in either trait. (Fig. 4). Additionally, vertebral counts reported in Stauffer (2020) were comparable to the range of values

reported for Brook Trout in other studies (Fig. 4A; Supplemental Table S2; see Data Accessibility). While we were unable to locate independent records of basihyal tooth counts in Brook Trout, the range of values reported by Stauffer (2020) was small with significant overlap among each of the populations surveyed (Fig. 4B). Overall, the limited data available for vertebrae and basihyal tooth counts in Brook Trout reflect the inherent challenges in quantifying these traits in a species of conservation concern, where researchers seek to minimize individual mortality when conducting field studies.

Given data insufficiencies, the use of vertebral and basihyal tooth counts for diagnosing species of *Salvelinus* is problematic as it is not possible to compare the distribution of these traits across many populations. Moreover, vertebral counts have also been shown to be a plastic trait in Brook Trout, with up to a three vertebrae difference in populations incubated at different temperatures and dissolved oxygen concentrations (Garside, 1966; Fowler, 1970). Less is known about variation in the extent and number of basihyal teeth, but, given that it is a trait that is related to foraging success, and others have documented rapid changes in traits associated with foraging kinematics (Adams et al., 2003), we would expect the variability in habitat across streams and the fine-scale partitioning of within-stream habitat that occurs from the realization of social dominance hierarchies to create similar levels of variability in Brook Trout. Accordingly, basihyal tooth counts are unlikely to be diagnostic across thousands of populations of Brook Trout

Table 1. Pairwise Mahalanobis distances between the three populations of *Salvelinus* from Cosby, Greenbrier, and Indian Camp Creeks (combined data from Stauffer [2020] and Weathers et al. [2019]; see text for details), *S. fontinalis* from 35 streams in the Great Smoky Mountains National Park (GSMNP) described in Weathers et al. (2019), and *S. fontinalis* from three populations in New York used by Stauffer and King (2014) in the description of the Brook Trout neotype. Larger distances are indicative of more substantial differentiation in meristic traits between populations.

	Cosby Creek	Greenbrier Creek	Indian Camp Creek	<i>S. fontinalis</i> (GSMNP)	<i>S. fontinalis</i> (NY)
Cosby Creek	*				
Greenbrier Creek	3.30	*			
Indian Camp Creek	3.06	3.56	*		
<i>S. fontinalis</i> (GSMNP)	1.60	2.93	2.92	*	
<i>S. fontinalis</i> (NY)	4.19	6.24	5.15	4.55	*

Table 2. Cross-validation linear discriminant analysis applied to meristic traits from five populations of *Salvelinus*, including the combined data from Stauffer (2020) and Weathers et al. (2019) from Cosby, Greenbrier, and Indian Camp Creeks, collections from Weathers et al. (2019) for 35 streams in the Great Smoky Mountains National Park (GSMNP), and three populations from NY. Values indicate the number of individuals classified to each source population, and the overall proportion of correctly classified individuals for each population. The low assignment success for individuals from Cosby, Greenbrier, and Indian Camp Creeks reflects the lack of unique meristic identity in those populations relative to other populations from the GSMNP.

Correct class	Predicted class					Percent correct classification
	Cosby Creek	Greenbrier Creek	Indian Camp Creek	<i>S. fontinalis</i> (GSMNP)	<i>S. fontinalis</i> (NY)	
Cosby Creek	10	2	0	7	1	50%
Greenbrier Creek	0	12	2	6	0	60%
Indian Camp Creek	1	1	11	7	0	55%
<i>S. fontinalis</i> (GSMNP)	7	2	0	342	0	97%
<i>S. fontinalis</i> (NY)	0	0	0	0	30	100%

across eastern North America, and at a very minimum would require data collected from populations at broader spatial scales before the trait could be considered diagnostic of a putative species.

Although Stauffer (2020) relied primarily on meristic traits for species delimitation, the author noted qualitative differences in mouth shape when compared to the Brook Trout neotype—specifically the presence of a more retrognathous versus isognathous mouth position. However, no morphological differences are evident in the sheared PCA in Stauffer (2020), and it is difficult to evaluate the potential for sex-based differences in the trait as the sex of the neotype is not noted by Stauffer and King (2014). Mouth morphology is difficult to compare across populations as jaw shapes are

rarely reported in Brook Trout species accounts, are highly subjective, and often vary considerably within a single population (Zastavniouk et al., 2017; Fig. 5). In addition to the significant effects of sexual dimorphism and ontogeny, jaw morphology can change within a few generations in response to competition and the relative abundance of autochthonous and allochthonous prey sources in other species of *Salvelinus* (Nakano et al., 2020). Overall, given the potential for substantial phenotypic plasticity from multiple, interactive factors, the use of morphological traits associated with foraging has been cautioned against when conducting phylogenetic studies in *Salvelinus* (Reist et al., 2013; Muir et al., 2016).

Significant methodological insufficiencies given confounding

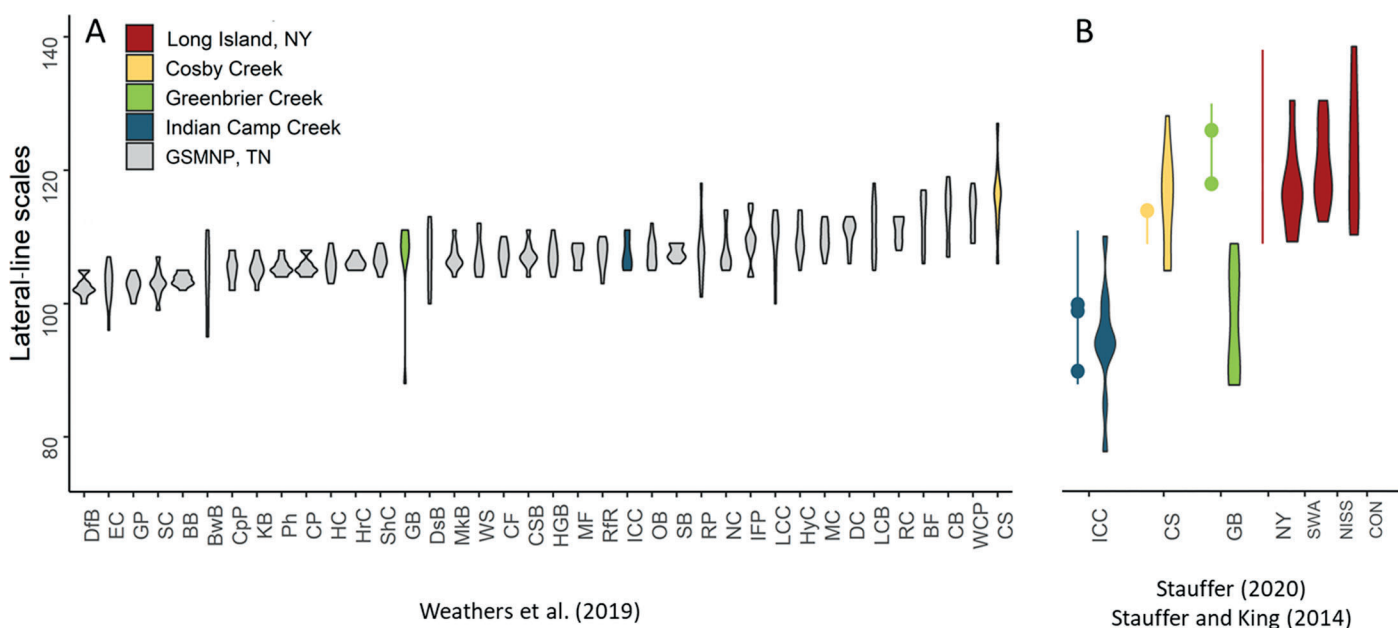


Fig. 3. Comparison of pored lateral-line scale counts for specimens collected from (A) 38 streams in the Great Smoky Mountains National Park (GSMNP) by Weathers et al. (2019) and (B) three streams surveyed by Stauffer (2020) and three populations described by Stauffer and King (2014) in Long Island, NY. Individual-level data collected by Weathers et al. (2019) are displayed with violin plots, with the width of the violin plot for each stream demonstrating the density of the distribution for a given value and the minimum and maximum values indicated by the tails of the distribution. Due to discrepancies between published and raw data, values from Stauffer (2020) and Stauffer and King (2014) are shown using two methods. Data from the publication appear as the mode(s) (circle) and range (lines), and the raw, individual-level data appear as violin plots. Streams appear on the x-axis by ascending average trait value, and streams included in both Weathers et al. (2019) and Stauffer (2020) are plotted with the same color (Cosby Creek [CS]: yellow; Greenbrier Creek [GB]: green; Indian Camp Creek [ICC]: blue). Data from populations in NY are shown in red and all other sites from GSMNP, TN in gray.

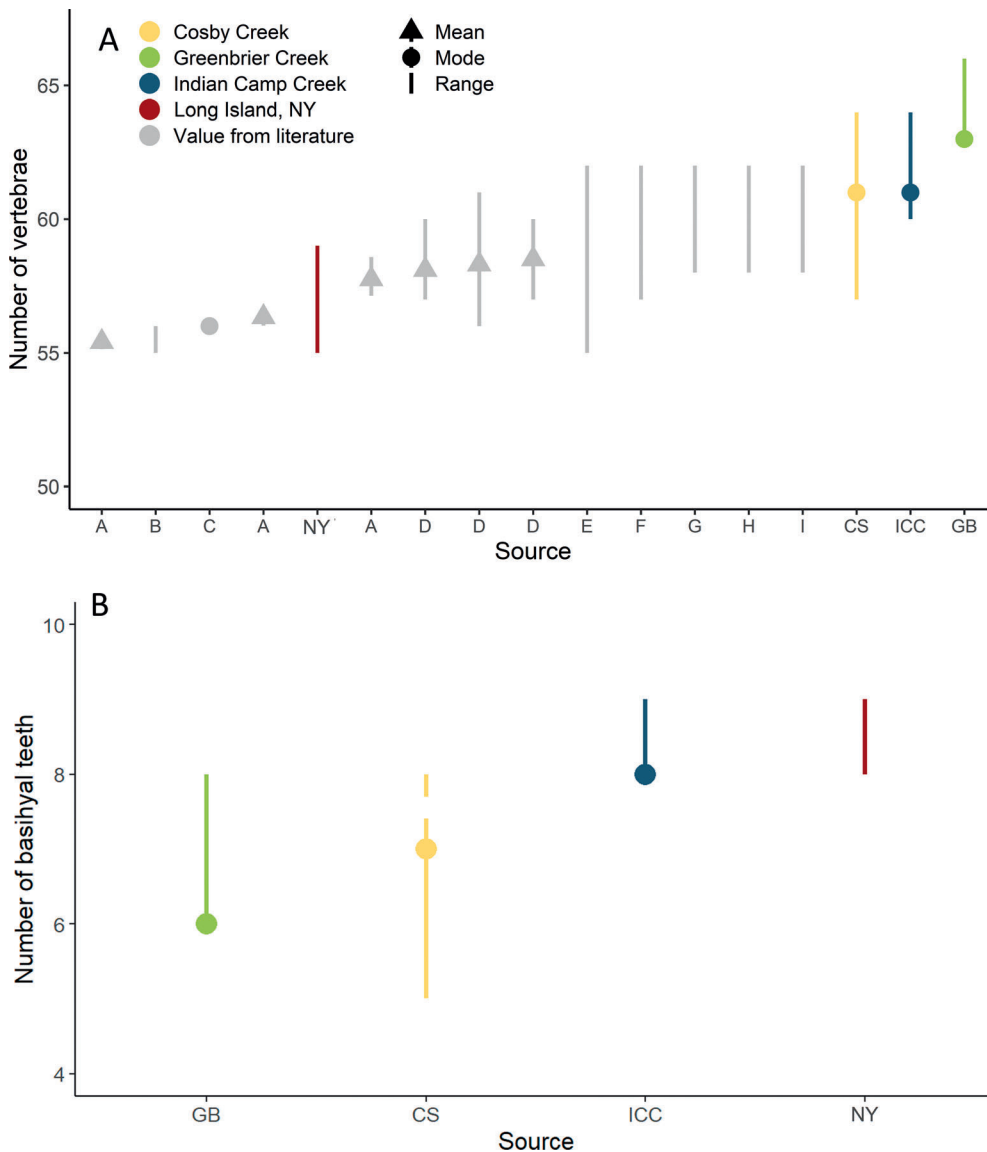


Fig. 4. The number of vertebrae (A) and basihyal teeth (B) reported by Stauffer (2020) for Cosby (CS; yellow), Indian Camp (ICC; blue), and Greenbrier (GB; green) Creeks. Vertebrae counts from peer-reviewed literature are also shown (see the supplement for sources), and, where appropriate, mean (triangle), mode (circle), and/or range (line) are indicated. The dashed line in panel B reflects values that were reported for basihyal tooth count from Cosby Creek in Stauffer (2020) but which were inconsistent with the requested data.

sources of variation.—As previously discussed, small sample sizes are inadequate for controlling for the effects of phenotypic plasticity and sexual dimorphism, and the minimal comparisons to regional and/or range-wide data risk a spurious conclusion of diagnosability when traits display widespread continua across the Brook Trout range. Moreover, the available data suggest broad overlap in the morphometrics and meristics of the three populations surveyed by Stauffer (2020) from the GSMNP. This shortcoming is apparent in the statistical analyses of Stauffer (2020) where, despite the author's conclusions of significant differentiation in meristic characters, the three populations broadly overlapped one another in multivariate ordination space. This result is unsurprising given that trait variation can be large in small, isolated populations (Zastavniouk et al., 2017), which highlights the need for more robust sample sizes.

The absence of sex data in Stauffer (2020) is problematic given the commonality of sexual dimorphism, including the potential for stronger phenotype-by-environment interactions in females and stronger selection for primary sex characters in males (Zastavniouk et al., 2017). Given that

specimens used by Stauffer (2020) were sacrificed, it seems reasonable that fish could have been sexed directly (via dissection or using molecular sex markers [Schill et al. 2016]) or indirectly using secondary sexual characteristics (Kazyak et al., 2013). In the absence of reliable sex data, the robust literature on sexual dimorphism in *Salvelinus* could have been used to evaluate the potential influence of ontogeny and sexual dimorphism on inferred patterns of variation (Proulx and Magnan, 2004; Nitychoruk et al., 2013; Zastavniouk et al., 2017).

While Stauffer (2020) asserts that all individuals were sexually mature, the range of individual sizes from each stream makes this assertion speculative, and sexually dimorphic characters tend to become exaggerated with size (Kazyak et al., 2013; Nitychoruk et al., 2013). Acknowledging that food and habitat limitations often limit the size structure of Brook Trout populations within the species' southern range (Ensign et al., 1990), and that Stauffer (2020) measured standard length (SL) rather than a total length (TL), a threshold of 100 to 120 mm TL is commonly applied for distinguishing adults and subadults/juveniles (Etnier and Starnes, 1993; Whiteley et al., 2012; Davis et al., 2015). The

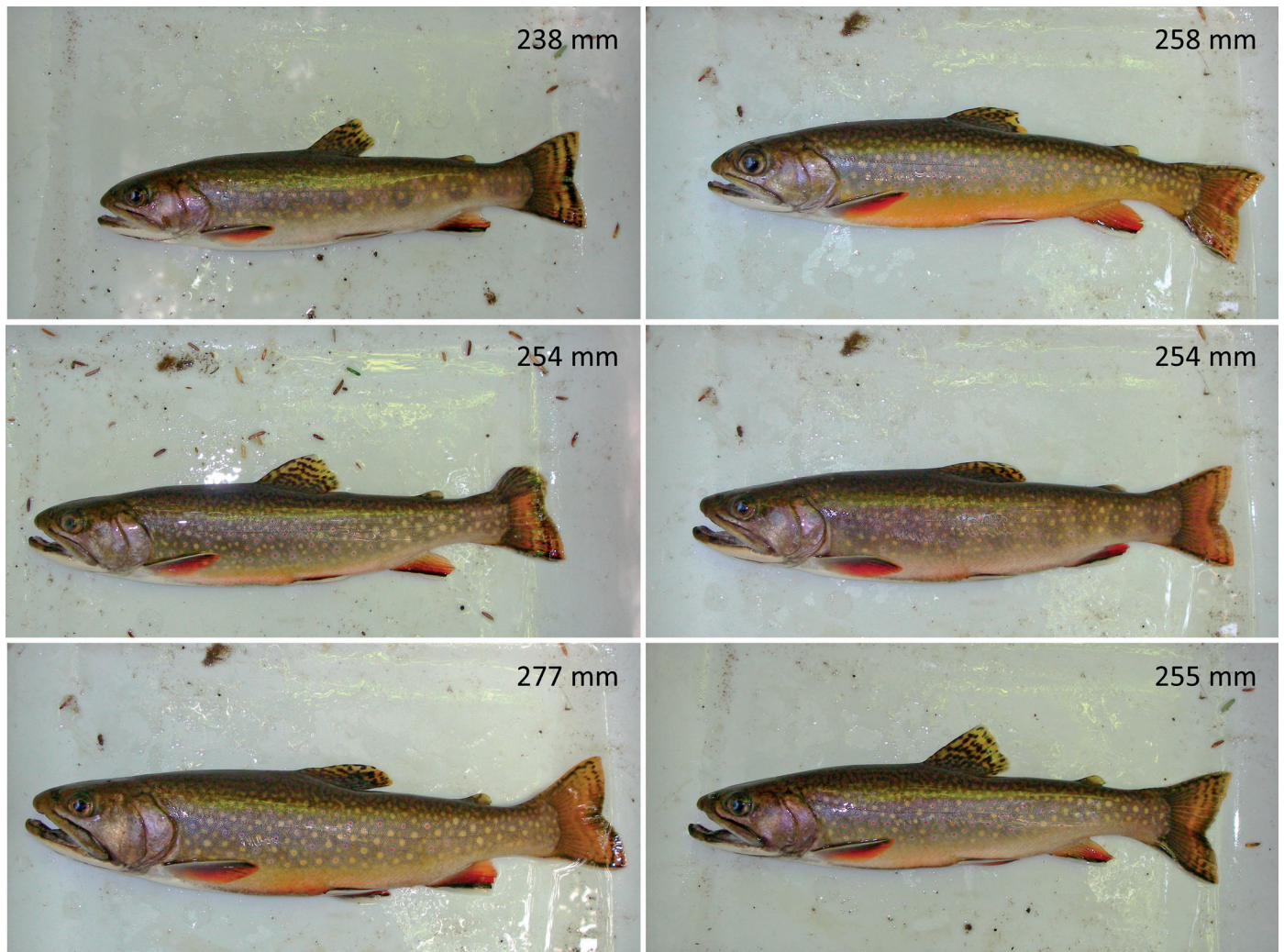


Fig. 5. Representative examples of diverse morphology, particularly in mouth shape and position, observed within a single stream-dwelling Brook Trout population. Fish on the first row display more inferior mouth positions, whereas fish on the last row show more isognathous and prognathic jaws with a terminal/superior mouth position. All fish were captured from Crabtree Creek in the Savage River Watershed of western Maryland (39°27'47.25"N, 79°12'36.08"W). Fish total length is noted in the upper right corner of each photograph. A full description of collection and photography protocols is provided in Kazyak et al. (2015).

size range of individuals included in Stauffer (2020), particularly individuals from one stream (which we believe to be Greenbrier Creek, but inconsistent data reporting makes it unclear which stream used individuals that ranged in size from 77.0 to 107.7 mm), raises concerns that a significant proportion of individuals in the analysis were not sexually mature. Moreover, the sex for the holotype from Greenbrier Creek was not indicated, but the specimen was only 97.2 mm suggesting the holotype may have been described from a sexually immature individual. The inclusion of subadults would likely have limited effects on meristics, but it could impart significant biases on morphometric relationships, particularly those related to mouth and jaw shape/size and body depth.

The potential for familial relationships within the samples considered by Stauffer (2020) was also not considered. In small headwater streams, full-sibling families are often restricted to short reaches of habitat (Hudy et al., 2010), which suggests the small sample sizes may represent a limited number of families. If morphology and meristics are heritable, which is an implicit assumption of using these

traits to delimit species, then undocumented family structure could bias assessments of taxonomy based on phenotypic characters.

The potential for intraspecific variation and plasticity highlights the importance of molecular data for a thorough taxonomic investigation in this clade (Muir et al., 2016; Kuparinen and Hutchings, 2019). Stauffer (2020) did reference neutral microsatellite genetic analyses by Stauffer and King (2014) and suggested significant population structure and isolation. While microsatellites provide valuable insights on recent population demographic parameters (Wan et al., 2004; Selkoe and Toonen, 2006), they are not informative for inferring relationships at deeper evolutionary time scales. When microsatellites are appropriately applied, it is not uncommon to find significant spatial structuring at scales commensurate to, or sometimes finer than, those detected by Stauffer (2020). Accordingly, the finding of population genetic structure is unsurprising given the ecology of Brook Trout and the limited number of populations analyzed (Kelson et al., 2015; Kazyak et al., 2016; Weathers et al., 2019).

DISCUSSION

While there are currently not sufficient data to support the distinctiveness of additional species currently classified as *Salvelinus fontinalis*, we acknowledge that populations of Brook Trout, particularly those in the southern extent of the species' range, are not interchangeable. Genetic and phenotypic differences do exist (Kazyak et al., 2015; Weathers et al., 2019), sometimes at fine scales, and identifying the causes and consequences of this variation is critical to the ecology and conservation of the species. Taxonomic resolution may prove challenging as Brook Trout are frequently characterized by small, isolated populations. In these scenarios, phenotypic variation can be a weak indicator of natural selection due to the stronger influences of drift, founder effects, or the decoupling of phenotypic correlations due to interspecific competition (Lowe et al., 2012; Zastavniouk et al., 2017). Conversely, small, isolated populations may be under stronger selective pressures than larger populations and can harbor unique phenotypic and genotypic variation that is important for species conservation at larger, range-wide scales (Stelkens et al., 2012; Fraser et al., 2014). Ultimately, it is important to consider how neutral and adaptive variation can confound inferences about speciation. Stauffer (2020) suggested that reduced offspring fitness observed by Kulp et al. (2017) after artificially crossing individuals from the three GSMNP populations was suggestive of speciation; however, the potential for outbreeding depression due to the breakup of coadapted gene complexes that evolved in differentially locally adapted populations was not considered (Edmunds and Timmerman, 2003).

A future challenge for Brook Trout conservation will be to decouple the relative influence of genotype and phenotypic plasticity in shaping local morphology and behavior. Plasticity to local conditions can have long-term and/or transgenerational effects, as demonstrated by consistent differences in spawning timing in the three populations examined by Stauffer (2020) even after being held in captivity under identical conditions for 1.5 years. While plasticity can be particularly beneficial for surviving in highly stochastic habitats (Miner et al., 2005), it can be misconstrued as evidence of divergent evolution (Price et al., 2003). For example, the phenology of spawning in stream salmonids is often adapted to location conditions, to the extent that spawning occurs across multiple months, and sometimes all year (Rinne, 1980; USFWS, 2009). However, phenological plasticity may not be as strongly correlated with ecological speciation as previously believed (Hendry, 2009), and populations often return to less variable behavior when environmental conditions stabilize.

Phylogenetic uncertainty stemming from vast intraspecific variation is not unique to the discussion of Brook Trout taxonomy and has long been part of conversations related to the conservation of *Salvelinus* and other stream salmonids (Guinand et al., 2021). Repeated radiation through increasingly diverse environments has led to significant uncertainty in how to appropriately classify the extensive taxonomic and functional diversity in many species of *Salvelinus* (Reist et al., 2013). Given these challenges, insular efforts to describe microendemic species that ignore the larger body of research and literature are likely to yield greater uncertainty. Ultimately, this highlights the need for holistic phylogenies that incorporate equally the influence of biology, ecology,

geography, and genetics for inferring past and present relationships among Brook Trout and its congeners.

Addressing questions about genotypic-by-phenotypic interactions and the appropriate scales of conservation, particularly as it relates to conservation under future climatic and anthropogenic disturbance regimes, will be aided by large-scale studies of Brook Trout phylogenetics and genomics. Range-wide assessments would be particularly beneficial for investigating the adaptive significance of broad patterns of genetic and morphometric differences that have been documented between regions and, if warranted, modifying conservation objectives to meet the unique phylogenetic histories and future conservation concerns at these larger spatial scales. Widespread interest in Brook Trout conservation has already motivated the establishment of a unique, range-wide collaboration among non-profits, universities, and state and federal agencies. Using these partnerships to investigate Brook Trout phylogenetic relationships offers great potential for developing management plans that effectively balance the need to conserve ecological processes across multiple temporal and spatial scales with the challenges of executing conservation efforts given finite resources. Ultimately, while conservation of important sources of genetic and phenotypic diversity is of utmost priority, poorly supported taxonomic revisions can negatively impact conservation initiatives. As such, caution is warranted when applying species concepts that allow for microendemic distributions when characterizing a broadly distributed taxon that is noted for high levels of phenotypic plasticity.

DATA ACCESSIBILITY

Supplemental material is available at <https://www.ichthyologyandherpetology.org/i2020154>. Unless an alternative copyright or statement noting that a figure is reprinted from a previous source is noted in a figure caption, the published images and illustrations in this article are licensed by the American Society of Ichthyologists and Herpetologists for use if the use includes a citation to the original source (American Society of Ichthyologists and Herpetologists, the DOI of the *Ichthyology & Herpetology* article, and any individual image credits listed in the figure caption) in accordance with the Creative Commons Attribution CC BY License.

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LITERATURE CITED

- Adams, C. E., C. Woltering, and G. Alexander. 2003. Epigenetic regulation of trophic morphology through feeding behaviour in Arctic charr, *Salvelinus alpinus*. *Biological Journal of the Linnean Society* 78:43–49.
- Cracraft, J. 1983. Species concepts and speciation analysis, p. 159–187. *In*: Current Ornithology. R. Johnston (ed.). Springer, Boston.

- Danzmann, R. G., R. P. Morgan, M. W. Jones, L. Bernatchez, and P. E. Ihssen. 1998. A major sextet of mitochondrial DNA phylogenetic assemblages extant in eastern North American brook trout (*Salvelinus fontinalis*): distribution and postglacial dispersal patterns. *Canadian Journal of Zoology* 76:1300–1318.
- Davis, L. A., T. Wagner, and M. L. Bartron. 2015. Spatial and temporal movement dynamics of brook *Salvelinus fontinalis* and brown trout *Salmo trutta*. *Environmental Biology of Fishes* 98:2049–2065.
- Edmands, S., and C. C. Timmerman. 2003. Modeling factors affecting the severity of outbreeding depression. *Conservation Biology* 17:883–892.
- Ensign, W. E., R. J. Strange, and S. E. Moore. 1990. Summer food limitation reduces brook and rainbow trout biomass in a southern Appalachian stream. *Transactions of the American Fisheries Society* 119:894–901.
- Etnier, D. A., and W. C. Starnes. 1993. *The Fishes of Tennessee*. University of Tennessee Press, Knoxville, Tennessee.
- Fowler, J. A. 1970. Control of vertebral number in teleosts—an embryological problem. *The Quarterly Review of Biology* 45:148–167.
- Fraser, D. J., P. V. Debes, L. Bernatchez, and J. A. Hutchings. 2014. Population size, habitat fragmentation, and the nature of adaptive variation in a stream fish. *Proceedings of the Royal Society B: Biological Sciences* 281: 20140370.
- Garside, E. T. 1966. Developmental rate and vertebral number in salmonids. *Journal of the Fisheries Board of Canada* 23:1537–1551.
- Guinand, B., M. Oral, and C. Tougard. 2021. Brown trout phylogenetics: a persistent mirage towards (too) many species. *Journal of Fish Biology*. DOI: 10.1111/jfb.14686.
- Harris, J., R. Estes, M. Van Den Avyle, W. Brandes, E. Shipp, and D. White. 1978. Final Report. Systematics of the Brook Trout *Salvelinus fontinalis* (Mitchill) in the Great Smoky Mountains National Park. Part II—Meristics. Tennessee Cooperative Fishery Research Unit, Cookeville, Tennessee.
- Hendry, A. P. 2009. Ecological speciation! Or the lack thereof? *Canadian Journal of Fisheries and Aquatic Sciences* 66:1383–1398.
- Hudy, M., J. A. Coombs, K. H. Nislow, and B. H. Letcher. 2010. Dispersal and within-stream spatial population structure of brook trout revealed by pedigree reconstruction analysis. *Transactions of the American Fisheries Society* 139:1276–1287.
- Kazyak, D. C., R. H. Hilderbrand, and A. E. Holloway. 2013. Rapid visual assessment to determine sex in brook trout. *North American Journal of Fisheries Management* 33:665–668.
- Kazyak, D. C., R. H. Hilderbrand, S. R. Killer, M. C. Colaw, A. E. Holloway, R. P. Morgan, and T. L. King. 2015. Spatial structure of morphological and neutral genetic variation in brook trout. *Transactions of the American Fisheries Society* 144:480–490.
- Kazyak, D. C., R. H. Hilderbrand, T. L. King, S. R. Keller, and V. E. Chhatre. 2016. Hiding in plain sight: a case for cryptic metapopulations in Brook Trout (*Salvelinus fontinalis*). *PLoS ONE* 11:e0146295.
- Kelson, S. J., A. R. Kapuscinski, D. Timmins, and W. R. Ardren. 2015. Fine-scale genetic structure of brook trout in a dendritic stream network. *Conservation Genetics* 16:31–42.
- Klemetsen, A. 2010. The charr problem revisited: exceptional phenotypic plasticity promotes ecological speciation in postglacial lakes. *Freshwater Reviews* 3:49–74.
- Kulp, M. A., S. Mitchell, D. Kazyak, B. Kuhajda, J. Henegar, C. Weathers, A. George, J. Ennen, and T. King. 2017. The southern Appalachian Brook Trout management conundrum: what should restoration look like in the 21st century, p. 65–75. *In: Wild Trout XII: Who's Driving and Where are We Going? Wild Trout Symposium*. R. F. Carline (ed.). Bozeman, Montana.
- Kuparinen, A., and J. A. Hutchings. 2019. When phenotypes fail to illuminate underlying genetic processes in fish and fisheries science. *ICES Journal of Marine Science* 76: 999–1006.
- Lowe, W. H., M. A. McPeck, G. E. Likens, and B. J. Cosentino. 2012. Decoupling of genetic and phenotypic divergence in a headwater landscape. *Molecular Ecology* 21:2399–2409.
- Miner, B. G., S. E. Sulton, S. G. Morgan, D. K. Padilla, and R. A. Relyea. 2005. Ecological consequences of phenotypic plasticity. *Trends in Ecology & Evolution* 20:685–692.
- Morinville, G. R., and J. B. Rasmussen. 2008. Distinguishing between juvenile anadromous and resident brook trout (*Salvelinus fontinalis*) using morphology. *Environmental Biology of Fishes* 81:171–184.
- Muir, A. M., M. J. Hansen, C. R. Bronte, and C. C. Krueger. 2016. If Arctic charr *Salvelinus alpinus* is 'the most diverse vertebrate', what is the lake charr *Salvelinus namaycush*? *Fish and Fisheries* 17:1194–1207.
- Nakano, S., K. D. Fausch, I. Koizumi, Y. Kanno, Y. Taniguchi, S. Kitano, and Y. Miyake. 2020. Evaluating a pattern of ecological character displacement: charr jaw morphology and diet diverge in sympatry versus allopatry across catchments in Hokkaido, Japan. *Biological Journal of the Linnean Society* 129:356–378.
- Nitychoruk, J. M., L. F. G. Gutowsky, P. M. Harrison, T. J. Hossie, M. Power, and S. J. Cooke. 2013. Sexual and seasonal dimorphism in adult adfluvial bull trout (*Salvelinus confluentus*). *Canadian Journal of Zoology* 91:480–488.
- Power, G. 1980. A systematic review of the genus *Salvelinus*, p. 141–204. *In: Charrs: Fishes of the Genus Salvelinus*. E. K. Balon (ed.). Dr. W. Junk Publishers, The Hague, Netherlands.
- Price, T. D., A. Qvarnström, and D. E. Irwin. 2003. The role of phenotypic plasticity in driving genetic evolution. *Proceedings of the Royal Society of London. Series B: Biological Sciences* 270:1433–1440.
- Proulx, R., and P. Magnan. 2004. Contribution of phenotypic plasticity and heredity to the trophic polymorphism of lacustrine brook charr (*Salvelinus fontinalis* M.). *Evolutionary Ecology Research* 6:503–522.
- R Core Team. 2020. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org/>
- Reist, J. D., M. Power, and J. B. Dempson. 2013. Arctic charr (*Salvelinus alpinus*): a case study of the importance of understanding biodiversity and taxonomic issues in northern fishes. *Biodiversity* 14:45–56.
- Rinne, J. N. 1980. Spawning habitat and behavior of Gila trout, a rare salmonid of the southwestern United States. *Transactions of the American Fisheries Society* 109:83–91.

- Schill, D. J., J. A. Heindel, M. R. Campbell, K. A. Meyer, and E. R. Mamer. 2016. Production of a YY male brook trout broodstock for potential eradication of undesired brook trout populations. *North American Journal of Aquaculture* 78:72–83.
- Selkoe, K. A., and R. J. Toonen. 2006. Microsatellites for ecologists: a practical guide to using and evaluating microsatellite markers. *Ecology Letters* 9:615–629.
- Stauffer, J. R. 2020. Description of three species of *Salvelinus* (Teleostei: Salmonidae) from the Great Smoky Mountains National Park, Tennessee, USA. *Ichthyological Exploration of Freshwaters* 30:97–110.
- Stauffer, J. R., R. W. Criswell, and D. P. Fischer. 2016. *The Fishes of Pennsylvania*. Cichlid Press, El Paso, Texas.
- Stauffer, J. R., and T. L. King. 2014. Designation of a neotype for brook trout, *Salvelinus fontinalis*. *Proceedings of the Biological Society of Washington* 127:557–567.
- Stelkens, R. B., G. Jaffuel, M. M. Escher, and C. Wedekind. 2012. Genetic and phenotypic population divergence on a microgeographic scale in brown trout. *Molecular Ecology* 21:2896–2915.
- USFWS (U.S. Fish and Wildlife Service). 2009. Apache Trout (*Oncorhynchus apache*) Recovery Plan. U.S. Fish and Wildlife Service Region 2, Albuquerque, New Mexico.
- Varian, A., and K. M. Nichols. 2010. Heritability of morphology in brook trout with variable life histories. *PLoS ONE* 5:e12950.
- Venables, W. N., and B. D. Ripley. 2002. *Modern Applied Statistics with S*. Fourth edition. Springer, New York.
- Wallace, C. R. 1973. Effects of temperature on developing meristic structures of smallmouth bass, *Micropterus dolomieu* Lacépède. *Transactions of the American Fisheries Society* 102:142–144.
- Wan, Q. H., H. Wu, T. Fujihara, and S. G. Fang. 2004. Which genetic marker for which conservation genetics issue? *Electrophoresis* 25:2165–2176.
- Weathers, T. C., D. C. Kazzyak, J. R. Stauffer, M. A. Kulp, S. E. Moore, T. L. King, and J. E. Carlson. 2019. Neutral genetic and phenotypic variation within and among isolated headwater populations of Brook Trout. *Transactions of the American Fisheries Society* 148:58–72.
- Whiteley, A. R., J. A. Coombs, M. Hudy, Z. Robinson, K. H. Nislow, and B. H. Letcher. 2012. Sampling strategies for estimating brook trout effective population size. *Conservation Genetics* 13:625–637.
- Woods, P. J., D. Yound, S. Skúlason, S. S. Snorrason, and T. P. Quinn. 2013. Resource polymorphism and diversity of Arctic Charr *Salvelinus alpinus* in a series of isolated lakes. *Journal of Fish Biology* 82:569–587.
- Zastavniouk, C., L. C. Weir, and D. J. Fraser. 2017. The evolutionary consequences of habitat fragmentation: body morphology and coloration differentiation among brook trout populations of varying size. *Ecology and Evolution* 7: 6850–6862.
- Zimmerman, M. S., C. C. Krueger, and R. L. Eshenroder. 2006. Phenotypic diversity of lake trout in Great Slave Lake: differences in morphology, buoyancy, and habitat depth. *Transactions of the American Fisheries Society* 135: 1056–1067.