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# Preference of *Orius insidiosus* and *Orius tristicolor* (Hemiptera: Anthocoridae) for host plants in olfactometry and free-choice experiments

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## Abstract

The western flower thrips, *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae), is a key pest of sweet pepper cultivation, where it causes feeding damage, excretes phytotoxic substances, and transmits important viruses. Control with chemical insecticides often is ineffective because endophytic oviposition and the cryptic habits of the pest provide protection. In Uruguay, the biological control program of this pest in sweet pepper crops is at risk due to the low settlement rate and high dispersal of releases of predator *Orius insidiosus* (Say) (Hemiptera: Anthocoridae). Previous studies have ruled out an antibiosis effect as the cause of dispersal; therefore, we hypothesized antixenosis (non-preference) for the sweet pepper plants as the cause of poor biological control by *O. insidiosus*. The effect of olfactory stimuli from different structures of strawberry, corn, and sweet pepper plants (lamuyo and blocky type) on the behavior of *O. insidiosus* was evaluated in olfactometry and free-choice cage experiments. Since *Orius tristicolor* (White) (Hemiptera: Anthocoridae) occurs naturally in the area, it was included also in the study with the aim of assessing whether there are differences in behavior between the species. *Orius tristicolor* may act as a complementary biocontrol agent or competitor on sweet pepper. Y-tube experiments showed no preference for plant volatiles in any combination, and response to volatile stimuli generally was poor. However, in the free-choice cage experiment, females of both species of *Orius* preferred the flowering strawberry plants over the flowering sweet pepper plants, which could explain the low establishment of *O. insidiosus* when released on pepper with neighboring strawberry fields. Given that horticultural greenhouses in Uruguay and in many other countries are open, this information can be very useful in designing the spatial and temporal management of different crops on a production field, which enhances the effectiveness of these predatory species.

Key Words: thrips; antixenosis; plant volatile; sweet pepper; strawberry

## Resumen

El trips occidental de las flores, *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae), es una plaga clave para el cultivo de pimiento dulce, donde causa daños al alimentarse, excreta sustancias fitotóxicas y transmite importantes virus. El control con insecticidas químicos no resulta efectivo, debido que la oviposición endofítica y los hábitos crípticos le brindan protección. En Uruguay, el programa de control biológico de esta plaga en el cultivo de pimiento dulce está en riesgo debido a la baja tasa de establecimiento y la alta dispersión de las liberaciones del depredador *Orius insidiosus* (Say) (Hemiptera: Anthocoridae). Estudios previos permiten descartar un efecto de antibiosis como causa de la dispersión, por lo cual se planteó como hipótesis la existencia de antixenosis o no preferencia por la planta de pimiento dulce como la causa de un control biológico deficiente por parte de *O. insidiosus*. Se evaluó el efecto de estímulos olfativos provenientes de diferentes estructuras de plantas de frutilla, maíz y pimiento dulce (tipo lamuyo y california) sobre el comportamiento de *O. insidiosus* en experimentos de olfactometría, y de libre elección en microparcela. Dado que *Orius tristicolor* (White) (Hemiptera: Anthocoridae) también está presente en el área, se incluyó en el estudio con el objetivo de evaluar si existen diferencias en el comportamiento entre ambas especies. *Orius tristicolor* podría actuar como un agente controlador complementario o competidor en pimiento dulce. Los experimentos con tubo en Y no mostraron preferencia por los compuestos volátiles de las plantas en ninguna combinación, y la respuesta a los estímulos volátiles en general fue pobre. Sin embargo, en el experimento de libre elección en microparcela, las hembras de ambas especies de *Orius* prefirieron las plantas con flores de frutilla sobre las plantas con flores de pimiento dulce, lo que podría explicar el bajo establecimiento de *O. insidiosus* cuando se libera en pimiento dulce con los campos de frutilla cercanos. Dado que los invernaderos hortícolas en Uruguay y en muchos otros países están abiertos, esta información puede ser muy útil para diseñar el manejo espacial y temporal de diferentes cultivos en un campo de producción, lo que mejora la efectividad de estas especies depredadoras.

Palabras Claves: trips; antixenosis; volátiles de planta; pimiento dulce; frutilla

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Western flower thrips, *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae), is considered one of the most important pests of greenhouse crops (Gillespie & Vernon 1990; Salguero Navas et al. 1991; Bitterlich & McDonald 1993), limiting the productivity of these

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crops worldwide. In several countries in South America, it is designated as one of the most limiting pest insects of sweet pepper crops (*Capsicum annum* L.; Solanaceae) in greenhouses (Carrizo 1998). Adults and immature stages of this species cause damage to plants by puncturing and then sucking the cellular content of plant tissues (Hunter & Ullman 1989), producing white surface lesions in the epidermis. In addition, they secrete phytotoxic substances when they feed, causing deformation of the foliage in all plant structures and organs (Bosco et al. 2008; Castresana et al. 2008). They also can cause indirect damage by transmitting viruses such as tomato spotted wilt virus (tomato black plague virus) and impatiens necrotic spot virus (Whitfield et al. 2005). Tomato spotted wilt virus is one of the viruses that causes major damage to a wide range of crops (Fanigliulo et al. 2014) and *F. occidentalis* is one of the most efficient species in its transmission (Whitfield et al. 2005). Tomato spotted wilt virus has a wide host range and worldwide distribution (Sherwood et al. 2000). This virus may lead to major economic losses on tomato, lettuce, pepper, eggplant, french beans, broad beans, celery, and different ornamental plants (Roselló et al. 1996). Insecticide applications are not effective in controlling thrips in these systems because females place their eggs under the epidermis of plant leaves, and both immature and adult stages have cryptic habits that give them some protection against toxic substances (Hansen et al. 2003; Helyer & Brobyn 2008). Frequent applications of chemical controls harm the complex of their natural enemies, promote the rapid development of resistance (Castresana et al. 2008), and generate risks to human health from acute and chronic exposure (Damalas & Eleftherohorinos 2011).

As an alternative to chemical control, the use of generalist natural enemies in sweet pepper crops is used widely and has been proven successful (van Lenteren et al. 2018). The release of the predator *Amblyseius swirskii* (Athias-Henriot) (Acari: Phytoseiidae) together with *Orius insidiosus* (Say) (Hemiptera: Anthocoridae) may manage the pepper pests *Bemisia tabaci* Gennadius (Hemiptera: Aleyrodidae) and *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae) successfully (Funderburk et al. 2000; Chow et al. 2010; Calvo et al. 2014).

In Uruguay, the incidence of *F. occidentalis* affects more than 80% of the sweet pepper production area, with significant losses in production (Maeso et al. 2013). Additional damage is caused by the tobacco whitefly, *B. tabaci*, which in combination have traditionally forced growers to apply chemical insecticides repeatedly on the sweet pepper crop with little effect. In the current decade, biological control in protected horticultural crops has made great progress in Uruguay, where the release of *A. swirskii* has reduced the incidence of *B. tabaci* on sweet peppers (Buenahora & Basso 2015). At the same time, the release of *O. insidiosus* in numerous greenhouses reached a low settlement rate and did not result in a reduction in the populations of *F. occidentalis* and their crop damage. These results contrast with the statements of Funderburk et al. (2000) indicating that *O. insidiosus* is effective in suppressing populations of thrips in field sweet pepper crops. It is even indicated that *O. insidiosus* remains in that crop for up to 6 mo after having eliminated *F. occidentalis* (van den Meiracker & Ramakers 1991). Uruguay is part of the distribution area of *O. insidiosus* (Bentancourt & Scatoni 2001; Carpintero 2002), but not of *Orius laevigatus* (Fieber) (Hemiptera: Anthocoridae), which is the most widely used species in Europe for biological control of thrips with successful results in sweet pepper (Jacas et al. 2008; van Lenteren 2012). In Europe, the varieties of sweet pepper planted belong preferably to the blocky type, unlike Uruguay, where all varieties are of the lamuyo type (*Orius* works well in both types of sweet pepper).

Studies conducted by Lorenzo et al. (2019) made it possible to rule out that settlement and effectiveness difficulties of *O. insidiosus* in the pepper greenhouses in Uruguay were due to an antibiosis effect. Discounting that the low settlement of *O. insidiosus* is associated with

the use of chemical insecticides for the control of other pests because there was a strict control of the products used for these purposes, it was hypothesized that an effect of antixenosis or non-preference was operating in relation to the sweet pepper plant. It should be noted also that the greenhouses used in Uruguay are open and are surrounded by 2 other common hosts of *O. insidiosus* (strawberry and corn) which are cultivated simultaneously with sweet pepper crops and coincide in their period of flowering in the field.

It is known that the olfactory system allows insects to distinguish the specific volatile compounds emitted by plants (Visser 1986; Bernays & Chapman 1994). These chemical compounds are useful cues for the insect to locate and recognize the plant as a suitable host, which are used to act as inhibitors or stimulants for feeding or oviposition (Campbell et al. 1986; Smith & Boyko 2006). The preferred behavior of females for oviposition derives from particular chemical, physical, and nutritional characteristics of plants (Lundgren & Fergen 2006; Lundgren et al. 2008). In this way, antixenosis or non-preference allows certain genotypes of plants to be incompatible with a specific insect, preventing insect establishment, feeding, or oviposition (Painter 1951). Anthocorids display this behavior, which allows them to select the oviposition substrates by physical and chemical factors (Richards & Schmidt 1996; Griffin & Yeorgan 2002). It should be considered that *Orius*, although being highly appreciated as entomophagous, lay their eggs inside plant tissues (Lundgren & Fergen 2006; Lundgren et al. 2008) and may even complete their development on certain foods of plant origin (Cocuzza et al. 1997). In recent studies, the phytophagous feeding behavior of *O. laevigatus* on sweet pepper was shown to trigger defensive responses in the plant. Plant induced defenses may contribute to the repellence or attraction of pests or natural enemies. Specifically, *O. laevigatus*-punctured sweet pepper plants induced repellency for the whitefly *B. tabaci* and the thrips *F. occidentalis* (Bouagga et al. 2018).

Based on the above, the study of preference or non-preference (antixenosis) for possible hosts may help explain the possible causes of low settlement and reproduction of *O. insidiosus* on sweet pepper. At the same time, it was of interest to compare the behavior of this species with *O. tristicolor*, which is normally found in corn (*Zea mays* L.; Poaceae) and strawberry (*Fragaria* sp.; Rosaceae) crops (Carpintero 2002; Ribeiro & Castiglioni 2008) near the sweet pepper greenhouses. As with *O. insidiosus*, it is common to find *O. tristicolor* in corn panicles due to the abundance of pollen, and in the styles of female flowers near stigmas (Corey et al. 1998), and in strawberry associated with flowers (Saini et al. 2003). Therefore, knowing the behavior of *O. tristicolor* in the cultivation of sweet pepper will let us know if it can be released together with *O. insidiosus* and contribute to the control of thrips in this crop.

In view of these questions, the purpose of this study was to characterize the preference of *O. insidiosus* and *O. tristicolor* for 3 of their host plants. Preference for cultivated varieties of sweet pepper, strawberry, and corn were evaluated under Y-tube olfactometer conditions and in free-choice experiments.

## Materials and Methods

### PLANT MATERIAL AND INSECTS

Studies were carried out in the laboratory and in a greenhouse of Plant Protection-Entomology at the Experimental Station of the School of Agronomy, Universidad de la República in Salto, Uruguay. *Orius insidiosus* and *O. tristicolor* were obtained from the field and maintained in our laboratory at 25 ± 1 °C, 65 ± 10% RH, fed with eggs of *Ephesia*

*kuehniella* Zeller (Lepidoptera: Pyralidae) ad libitum with bean pods (*Phaseolus vulgaris* L.; Fabaceae) as an oviposition substrate (Castañé & Zalom 1994; Richards & Schmidt 1996; Mendes et al. 2005). For the experiments, a photoperiod of 16:8 h (L: D) was used pursuant to Richards & Schmidt (1996).

The plants used for tests were propagated in the Station at the School of Agronomy, Universidad de la República in Salto, Uruguay. The cultivation conditions were optimized in order to obtain good production of flowers (Corey et al. 1998; Saini et al. 2003). Five liter pots (Media Grow, Montevideo, Uruguay) were used with a sterile substrate (Plagron allmix, Bertels, Netherlands) that is a mix of the types of peat which contain various types of fiber and perlite, along with localized and automated irrigation. Plants were propagated from seed and fertilized based on optimal rates for the region (MGAP 2016). The plants evaluated were sweet pepper (*Capsicum annuum*) lamuyo type (cultivar 'Bilano' by Syngenta) and blocky type (cultivar 'Phycol' by Agritec), strawberry (*Fragaria × ananassa* Duch [Rosaceae], cultivar 'Guapa' by INIA), and corn (*Zea mays*, cultivar 'IPB 2650' by IPB seeds).

#### Y-TUBE EXPERIMENTS

A glass Y-tube olfactometer was used, with an internal diameter of 0.5 cm, which is a modified version of the one developed by Sabelis & van de Baan (1983), applying the methodology proposed by Koschier et al. (2000). The olfactometer consisted of a central chamber (long arm 12 cm) from which 2 arms of choice are derived (6 cm). At the far ends of the Y-shaped glass tube, PVC tubes (10 cm) were connected that contain either the presumed odor-emitting plants or clean air. The apparatus also contained an activated carbon filter, a pump for air circulation, and a rotameter for the measurement of air flow. The airflow in each arm of the olfactometer was calibrated to 0.5 m per s (VelociCalc® *Air Velocity Meter 9545-A*, TSI, Shoreview, Minnesota, USA). Through this system it is possible to observe and subsequently record the movement of each insect within the central chamber and in the arms of the Y olfactometer. The tests were conducted in a climate-controlled room at  $25 \pm 2$  °C and  $65 \pm 5$  % RH.

The response of adult females of *O. insidiosus* and *O. tristicolor* was evaluated to stimuli from different structures of foliage and flowers (Visser & Piron 1998; Drukker et al. 2000; López-Ávila & Rincón 2006; Yoneya et al. 2009, 2010; Bosa et al. 2011), with an approximate weight of 1 g each (Reid & Lampman 1989). Whole leaves and flowers were used, washed with water and weighed. The materials were used in individual experiments for 30 min and were not used for more than 2 h altogether. The stimuli compared were strawberry, sweet pepper, or corn leaves versus control (clean air); strawberry, sweet pepper, or corn flowers versus control (clean air); or sweet pepper flower versus strawberry flower. For sweet pepper, both blocky and lamuyo type sweet peppers, foliage, and flowers were tested versus control (clean air).

In each test, a single female of *O. insidiosus* or *O. tristicolor* was placed in the central part of the olfactometer main chamber with the help of a fine tip brush to avoid damaging the insect. The chamber was then sealed with cotton. The experiment was carried out with 40 females of the 2 species of *Orius* less than 72 h old. The females were placed individually in plastic containers without food for a period of 8 to 10 h before the test. Forty females were evaluated ( $n = 40$ ) in each experiment, 20 predators per d.

The initial choice of the insect was recorded, and a response was considered positive when, within 10 min, the insect remained inside 1 of the olfactometer arms after moving past the center of the glass tube (Pivnick et al. 1990; Peraza 2011). Upon reaching the Y-junction

in the glass tube, the *Orius* could choose between the air flow coming from either arm. When it reached the far end of an arm, the choice was noted, and the insect was removed from the tube. If the insect did not enter the decision zone within the established period, the test was recorded as null.

All treatments were evaluated in a darkened space to focus the experiment on the volatile substances generated by the plants introduced into the Y-tube 5 min before the start of each test. Between tests, the olfactometer was cleaned with 70% acetone to remove possible traces of contaminants or pheromones. The acetone was left to evaporate for 1 min at ambient conditions. In addition, to avoid any bias, the sources of odors were alternately switched to the other arm of the olfactometer after every 5 females tested, and the plant material was replaced after every 10 females.

#### CHOICE-TEST EXPERIMENTS

Antixenosis (non-preference) was evaluated by the free host selection test following the methodology of Castro et al. (2001). The study was carried out in a research greenhouse made of polyethylene (Naka SA, Montevideo, Uruguay) measuring 10 × 5 m, under controlled conditions of temperature and humidity (22–26 °C, 60–70% RH). The sweet pepper and strawberry plants were kept in plastic pots 20 cm in diam and 10 cm deep (Biriden, Montevideo, Uruguay) until sprouting of first flowers.

A micro-plot of 4 m<sup>2</sup>, covered with a non-woven polypropylene mesh (MC Agril P 17) (Naka SA, Montevideo, Uruguay) was placed inside the greenhouse to prevent dispersal of the insects. Twelve-wk old sweet pepper plants (30 cm high) and 11-wk old strawberry plants (25 cm high) were arranged alternatively with 20 cm distance between them, with 6 rows and 6 columns (36 plants). In the center of each plot, 72 females (two 1–5 d old *Orius* per plant) of each species of *Orius* to be evaluated (*O. insidiosus* or *O. tristicolor*) were released. Separate experiments were performed for each species of *Orius*, and the insects were allowed free choice of the host plants. In a second pair of experiments, both species of insects was tested regarding their preference for lamuyo or blocky type sweet pepper plants using the same procedure described above. At 24 h after release, the number of females present per plant was counted. Each experiment attempted to simulate the cultivation conditions in the area, matching time of transplantation, flowering, and release of natural enemies as would occur in the field.

#### STATISTICAL ANALYSIS

In both experimental designs, olfactometer and micro-plot, the preference for a plant or plant organ, was analyzed using a Chi-square goodness of fit test corrected for continuity (Yates-correction). The null hypothesis was that *Orius* spp. distribution was 50:50 across the 2 arms of the olfactometer and on the 2-plant species (sweet pepper and strawberry) or pepper cultivar (lamuyo and blocky). Analyses were performed using the R version 3.3.2 (R Core Team 2016).

## Results

#### Y-TUBE EXPERIMENTS

Females of *O. insidiosus* preferred stimuli from strawberry flowers compared to those from sweet pepper flowers ( $\chi^2 = 5.565$ ;  $P = 0.018$ ), while females of *O. tristicolor* preferred the stimuli originated in the sweet pepper leaves over the control (pure air) ( $\chi^2 = 9.520$ ;  $P = 0.002$ ). In the other combinations tested, no differences were found in the response to odor stimuli (Tables 1, 2).

**Table 1.** Olfactory response of females of *Orius insidiosus* and *Orius tristicolor* to stimuli from foliage and flower of strawberry, corn, and pepper plants (in the arm [+]), when offered against clean air (in the arm [-]) of a Y-tube olfactometer.

Source smell on the arm (+)	<i>n</i> (+)	<i>n</i> (-)	<i>n</i> (0)	% (+)	$\chi^2$	<i>P</i>
<i>Orius insidiosus</i>						
Strawberry foliage	6	12	22	15	2.778	0.096
Corn foliage	9	15	16	22.5	2.083	0.148
Pepper foliage	7	13	20	17.5	2.500	0.114
Strawberry flower	18	12	10	45	1.667	0.197
Corn flower	10	17	13	25	2.667	0.102
Pepper flower	12	16	12	30	0.276	0.599
<i>Orius tristicolor</i>						
Strawberry foliage	7	13	20	17.5	2.500	0.114
Corn foliage	10	14	16	25	0.750	0.386
Pepper foliage	16	5	8	40	9.520	0.002
Strawberry flower	14	14	12	35	0.071	0.789
Corn flower	19	12	9	47.5	2.323	0.128
Pepper flower	14	15	11	35	0.069	0.790

*n* (+, -) = number of predators that reached the end of the (+) or (-) arm of the Y-tube; *n* (0) = number of predators that did not respond within a 10 min test; % (+) = percentage of individuals that reached the end of the arm (+); *P* = critical level.

Females of *O. insidiosus* showed no preference between blocky or lamuyo type sweet pepper foliage ( $\chi^2 = 0.320$ ; *P* = 0.572), and the same result was obtained when evaluating blocky sweet pepper flower compared to lamuyo sweet pepper flower ( $\chi^2 = 3.769$ ; *P* = 0.052;  $\alpha = 0.05$ ), although the value obtained is at the limit, being almost significant (Table 3). The females of *O. tristicolor* also did not show a preference for blocky or lamuyo type sweet peppers, either in foliage ( $\chi^2 = 0.077$ ; *P* = 0.782) or flower ( $\chi^2 = 1.796$ ; *P* = 0.181) (Table 3).

#### CHOICE TESTS EXPERIMENTS

After 24 h of being released, females of both species preferred strawberry plants over sweet pepper plants (*O. insidiosus*,  $\chi^2 = 4.721$ , *p* = 0.030; *O. tristicolor*,  $\chi^2 = 4.321$ ; *p* = 0.038) (Table 4).

The females of the two species of *Orius* did not show a different preference between lamuyo or blocky type sweet pepper plants (*O. insidiosus* –  $\chi^2 = 0.450$ ; *P* = 0.371; *O. tristicolor* –  $\chi^2 = 1.389$ , *P* = 0.239) (Table 4).

## Discussion

In the process of colonization of a host, the behavior of females, including those belonging to Anthocoridae, usually includes a sequence of patterns consisting of orientation, landing, and recognition under different selection pressures. Currently, the mechanisms *Orius* spp. employ to locate their host in complex mixed-vegetation habitats re-

main poorly understood. All steps involve different senses and occur at varying distances from the plant surface; because olfactory and visual cues are perceptible at distance, they play a determinant role in plant location by foraging insects (Schoonhoven et al. 1998).

It is worth emphasizing the importance of visual search cues in Hemiptera, such as aphid and whitefly (Pickett et al. 1992; Isaacs et al. 1999). Ohno & Takemoto (1997) and Furihata et al. (2019) established that *Orius* spp. responded to different colors of traps, when they demonstrated that blue sticky traps were more effective for monitoring studies than white or yellow ones. Studies by Patt et al. (2011) showed that visual stimuli improve olfactory responses to the volatile compounds of the host plant. In fact, to initiate the process of colonization of a host, an olfactory stimulus may not be sufficient to generate a search for resources and oviposition (Visser 1986; Bernays & Chapman 1994; Finch & Collier 2000; Awmack & Leather 2002). The low response rate obtained in the Y-tube bioassays was due possibly to insufficient olfactory stimulation. In general, when a high percentage of test insects do not show a response in the Y tubes, the species does not respond to odor stimuli (this is not the case with *Orius* spp.) or the olfactory stimulus is not sufficient to induce search behaviors (Drukker et al. 2000; Bernardo 2015; Ardanuy et al. 2016).

It also should be noted that the size of the leaf or flower offered (amount of plant material = 1 g) as a source of odor may not have been sufficient. This could explain why the females of *O. insidiosus* showed no preference for the foliage and flowers of sweet pepper, strawberry, or corn in relation to fresh air (control). Another aspect to consider is that cutting leaves or using structures and not whole plants could

**Table 2.** Olfactory response of females of *Orius insidiosus* and *Orius tristicolor* to stimuli from strawberry flowers (on the arm [+]), when offered against pepper flowers (on the arm [-]) of an olfactometer of Y-tube.

Source smell on the arm (+)	<i>n</i> (+)	<i>n</i> (-)	<i>n</i> (0)	% (+)	$\chi^2$	<i>P</i>
<i>Orius insidiosus</i>						
Strawberry flower	16	7	17	40	5.565	0.018
<i>Orius tristicolor</i>						
Strawberry flower	14	9	17	35	1.391	0.238

*n* (+, -) = number of predators that reached the end of the (+) or (-) arm of the Y tube; *n* (0) = number of predators that did not respond within a 10 min test; % (+) = percentage of individuals that reach the end of the arm (+); *P* = critical level.



**Table 3.** Olfactory response of females of *Orius insidiosus* and *Orius tristicolor* to stimuli from foliage pepper type blocky (on the arm [+]) when offered against foliage pepper type lamuyo (on the arm [-]) or flower pepper type blocky (on the arm [+]) when offered against flower pepper type lamuyo (on the arm [-]) of an olfactometer of Y-tube.

Source smell on the arm (+)	<i>n</i> (+)	<i>n</i> (-)	<i>n</i> (0)	% (+)	$\chi^2$	<i>P</i>
<i>Orius insidiosus</i>						
Foliage pepper type blocky	13	11	13	32.5	0.320	0.572
Flower pepper type blocky	17	9	11	42.5	3.769	0.052
<i>Orius tristicolor</i>						
Foliage pepper type lamuyo	14	12	14	35	0.077	0.782
Flower pepper type lamuyo	15	11	15	37.5	1.796	0.181

*n* (+, -) = number of predators that reached the end of the (+) or (-) arm of the Y tube; *n* (0) = number of predators that did not respond within a 10 min test; % (+) = percentage of individuals that reach the end of the arm (+); *P* = critical level.

affect the chemical composition and influence the behavior of the species observed (Madadi et al. 2008). Cut plants may have different odor profiles; the volatile flowers, undamaged leaves, and leaves attacked by herbivores often exhibit different emission patterns from each other (Dudareva & Negre 2005), i.e., each plant structure has a specific odor profile (Karlsson et al. 2009).

The release of volatile defense compounds by plant substrates should not be ruled out either, because it has been reported that in other species of Solanaceae, such as tomatoes, substances derived from sesquiterpenes can repel insects (Frelichowski & Juvik 2005; Álvarez Gil 2015). Recent results by Bouagga et al. (2018) indicate that *O. laevigatus* punctures in the sweet pepper plants induced the release of an altered blend of volatiles and activation of the jasmonate acid and salicylic acid signaling pathways. These results highlight a potential tritrophic interaction in which the predator is able to induce defensive responses in the plant that reduces feeding by the pest.

In free-choice experiments, the difference found in the preference of females of *O. insidiosus* and *O. tristicolor* for the volatiles emitted by flowering strawberry plants compared to those of flowering sweet pepper plants would confirm that the females use chemical and olfactory cues to recognize their plant host, and that these cues favored strawberry. In the case of strawberry, volatile compounds have been studied widely in fruits; however, there are no studies that evaluate the aromatic differences between the organs of the plant or the whole plant at different phenological stages (Pérez-Rubio 1994). It should be considered that in polyphagous insects, the developmental stage of the host plant can determine the preference, where the main hosts emit a set of very similar volatiles, but both host and non-host plants are more attractive to many species during the flowering stage (Rajapakse 2006). The positive response behavior to the volatile emitted by plants in the flowering stage may be dependent on the response of the anthocorids to this stage of the plant where they feed on pollen and oviposit. It is known that females, during the search for hosts for oviposition, seek to ensure that the plant guarantees the best development of immature stages (Schoonhoven et al. 2005; Carroll et al. 2006), a principle known as “mother knows best” (Singer et al. 1988;

Schoonhoven et al. 2005). As various authors indicated, sweet pepper produces numerous volatile compounds and at least 14 of them are released during flowering and fructification (Addesso et al. 2011), phenological stages when sweet pepper plants are highly attractive to *Orius* spp. adults. The principal compound emitted during these stages is (E)-beta-ocimene (Addesso et al. 2011). This monoterpene has been reported to affect the behavior of adults of some species Coleoptera (Magalhães et al. 2012). Other volatile compounds from *Capsicum* spp. also induce behavioral or electrophysiological activity in different species (Muñiz et al. 2014).

Although the results of this study arise from experiments under controlled conditions in a laboratory or greenhouse, they can be considered useful for informing the management of these predators in the field. These results suggest that females of *Orius* would be able to discriminate between different host plants, and that in particular both *O. insidiosus* and *O. tristicolor* show greater preference for strawberry crops than sweet pepper crops, which could explain the low rate of establishment of the *Orius* females in the sweet pepper crop when they are released on it. It should be noted that in Uruguay, unlike in other horticultural areas, greenhouses are not closed and sweet pepper often is surrounded by other crops, including strawberry. The evaluated host plants coincide with each other in their period of cultivation and flowering in the field. Sweet pepper under greenhouse (113 ha in the country) and strawberry under micro or macro tunnel (55 ha in the country) are produced from Mar to Dec and coincide in their flowering periods (MGAP 2016). The first pepper and strawberry blossoms coincide with the end of corn cultivation, and it is very common to find corn in the surroundings of the greenhouses with the presence of *Orius*. Although these results contrast with those stated by van den Meiracker & Ramakers (1991) and Funderburk et al. (2000), where these authors described the establishment and effectiveness of *O. insidiosus* to control *F. occidentalis* on sweet pepper, they do not indicate whether the greenhouses in these systems were closed or open, or whether there were crops in the vicinity of the sweet peppers that could have interfered with predator establishment.

It should be reiterated that, as evidenced by Lorenzo et al. (2019), there is no effect of sweet pepper antibiosis on *O. insidiosus*; therefore,

**Table 4.** Response of females *Orius insidiosus* and *Orius tristicolor* to sweet pepper plant type blocky and type lamuyo, and strawberry plant in the free-choice test.

	Sweet pepper plant type lamuyo	Strawberry plant	$\chi^2$	<i>P</i>
Female <i>O. insidiosus</i> choice rate ( <i>n</i> = 72)	33	51	4.721	0.030
Female <i>O. tristicolor</i> choice rate ( <i>n</i> = 72)	30	47	4.321	0.038
	Sweet pepper plant type lamuyo	Sweet pepper plant type blocky		
Female <i>O. insidiosus</i> choice rate ( <i>n</i> = 72)	25	30	0.450	0.371
Female <i>O. tristicolor</i> choice rate ( <i>n</i> = 72)	20	29	1,389	0.239

if there were no other repellent stimuli present it would remain and reproduce on the sweet pepper crop. The type of sweet pepper grown in Uruguay (lamuyo) did not have an influence on this non-preference either, because no differences were observed in preference for lamuyo or blocky sweet peppers. Although females of the species *O. tricolor* were attracted to stimuli from sweet pepper foliage as opposed to pure air, they also were attracted to strawberry plants in the micro-plot test, which could explain their absence on sweet pepper crops while still being found in other hosts surrounding pepper greenhouses.

As a complement, a tool to improve the establishment and colonization by *Orius* could be banker plants located inside the greenhouse. Native species with good flowering and good acceptance by *Orius* should be investigated. Determining a suitable flowering banker plant could improve opportunities for using the predator to control thrips by providing a source of supplementary food in the form of pollen (Huang et al. 2011).

Given that the horticultural greenhouses in Uruguay and in many other countries are open, these results regarding *Orius* preference for different crops may be very useful to aid in the design of spatial and temporal management of different horticultural crops on a production field in order to enhance the effectiveness of these predatory species.

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## References Cited

- Adesso KM, McAuslane HJ, Alborn HT. 2011. Attraction of pepper weevil to volatiles from damaged pepper plants. *Entomologia Experimentalis et Applicata* 138: 1–11.
- Álvarez Gil M. 2015. Resistencia de insectos en tomate (*Solanum* spp.). *Cultivos Tropicales* 36: 100–110.
- Ardanuy A, Albajes R, Turlings TC. 2016. Innate and learned prey-searching behavior in a generalist predator. *Journal of Chemical Ecology* 42: 497–507.
- Awmack CS, Leather SR. 2002. Host plant quality and fecundity in herbivorous insects. *Annual Review of Entomology* 47: 817–844.
- Bentancourt CM, Scatoni IB. 2001. Enemigos naturales. Manual ilustrado para la agricultura y la forestación. Universidad de la República (Uruguay). Facultad de Agronomía, PREDEG-GTZ, Montevideo, Uruguay.
- Bernardo A. 2015. Alternative food and learning as a promising strategy for biological control. M.Sc. thesis, Federal University of Viçosa, Viçosa, Minas Gerais, Brazil.
- Bernays E, Chapman R. 1994. Behavior: the process of host-plant selection, pp. 95–165 *In* Bernays E, Chapman R [eds.], *Host Plant Selection by Phytophagous Insects*. Chapman & Hall, New York, USA.
- Bitterlich I, McDonald LS. 1993. The prevalence of tomato spotted wilt virus in weeds and crops in southwestern British Columbia. *Canadian Plant Disease Survey* 73: 137–142.
- Bosa CF, Clavijo A, Karlsson FM, Cotes AM, Witzgall P. 2011. Respuesta de *Tecia solanivora* (Lepidoptera: Gelechiidae) a compuestos volátiles de papa, *Solanum tuberosum* L. *Revista Colombiana de Entomología* 37: 1–7.
- Bosco L, Giacometto E, Tavella L. 2008. Colonization and predation of thrips (Thysanoptera: Thripidae) by *Orius* spp. (Heteroptera: Anthocoridae) in sweet pepper greenhouses in Northwest Italy. *Biological Control* 44: 331–340.
- Bouagga S, Urbaneja A, Rambla JL, Granell A, Pérez-Hedo M. 2018. *Orius laevigatus* strengthens its role as a biological control agent by inducing plant defenses. *Journal of Pest Science* 91: 55–64.
- Buenahora J, Basso C. 2015. Utilización de *Amblyseius swirskii*, un enemigo natural clave para el manejo integrado de plagas en el cultivo de pimiento en invernadero, *In* 4tas. Jornadas de Enfermedades y Plagas en Cultivos Bajo Cubierta. Facultad de Ciencias Agrarias y Forestales de la Universidad Nacional de La Plata, La Plata, Argentina.
- Calvo FJ, Knapp M, van Houten YM, Hoogerbrugge H, Belda JE. 2014. *Amblyseius swirskii*: what made this predatory mite such a successful biocontrol agent? *Experimental and Applied Acarology* 65: 419–433.
- Campbell BC, Jones CK, Dreyer DL. 1986. Discriminative behavioral responses by aphids to various plant matrix polysaccharides. *Entomologia Experimentalis et Applicata* 41: 17–24.
- Carpintero DL. 2002. Catalogue of the Neotropical Anthocoridae (Heteroptera). *Revista Sociedad Entomológica Argentina* 61: 25–44.
- Carrizo PL. 1998. Hospederas naturales para trips vectores de peste negra: propuesta de calificación de riesgo. *Boletín de Sanidad Vegetal Plagas* 24: 155–166.
- Carroll M, Schmelz E, Meagher R, Teal P. 2006. Attraction of *Spodoptera frugiperda* larvae to volatiles from herbivore-damaged maize seedlings. *Journal of Chemical Ecology* 32: 1911–1924.
- Castañé C, Zalom FG. 1994. Artificial oviposition substrate for rearing *Orius insidiosus* (Hemiptera, Anthocoridae). *Biological Control* 4: 88–91.
- Castresana J, Gagliano E, Puhl L, Bado S, Vianna L, Castresana M. 2008. Atracción del trips *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae) con trampas de luz en el cultivo de *Gerbera jamesonii* (G.). *Idesia* 26: 51–56.
- Castro AM, Ramos S, Vasicek A, Worland A, Giménez D, Clúa A, Suárez E. 2001. Identification of wheat chromosomes involved with different types of resistance against greenbug and Russian wheat aphid (*Diuraphis noxia* Mordvilko). *Euphytica* 118: 321–330.
- Chow A, Chau A, Heinz KM. 2010. Compatibility of *Amblyseius* (Typhlodromips) *swirskii* (Athias-Henriot) (Acari: Phytoseiidae) and *Orius insidiosus* (Hemiptera: Anthocoridae) for biological control of *Frankliniella occidentalis* (Thysanoptera: Thripidae) on roses. *Biological Control* 53: 188–196.
- Cocuzza GE, De Clercq P, van de Veire M, De Cock A, Degheele D, Vacante V. 1997. Reproduction of *Orius laevigatus* and *Orius albidipennis* on pollen and *Ephesia kuehniella* eggs. *Entomologia Experimentalis et Applicata* 82: 101–104.
- Corey D, Kamhampati S, Wilde GE. 1998. Electrophoretic analysis of *Orius insidiosus* (Hemiptera: Anthocoridae) feeding habits in field corn. *Journal of the Kansas Entomological Society* 71: 11–17.
- Damala CA, Eleftherohorinos IG. 2011. Pesticide exposure, safety issues, and risk assessment indicators. *International Journal of Environmental Research and Public Health* 8: 1402–1419.
- Dudareva N, Negre F. 2005. Practical applications of research into the regulation of plant volatile emission. *Current Opinion in Plant Biology* 8: 113–118.
- Fanigliulo A, Viggiano A, Gualco A, Crescenzi A. 2014. Control of viral diseases transmitted in a persistent manner by thrips in pepper (tomato spotted wild virus). *Communications in Agricultural Applied Biological Science* 79: 433–437.
- Finch S, Collier R. 2000. Host-plant selection by insects – a theory based on ‘appropriate/ inappropriate’ landings by pest insects of cruciferous plants. *Entomologia Experimentalis et Applicata* 96: 91–102.
- Frelichowski JE, Juvik AJ. 2005. Inheritance of sesquiterpene carboxylic acid synthesis in crosses of *Lycopersicon hirsutum* with insect-susceptible tomatoes. *Plant Breeding* 124: 277–281.
- Funderburk J, Stavisky J, Olsen S. 2000. Predation of *Frankliniella occidentalis* (Thysanoptera: Thripidae) in field peppers by *Orius insidiosus* (Hemiptera: Anthocoridae). *Environmental Entomology* 29: 376–382.
- Furihata S, Tabuchi K, Okudera S, Takahashi A, Hinomoto N, Shimoda M, Yamaguchi T. 2019. An efficient method for monitoring predatory minute pirate bugs *Orius* spp. (Hemiptera: Anthocoridae) populations using blue-colored sticky traps. *Environmental Entomology* 48: 1–8.
- Gillespie DR, Vernon RS. 1990. Trap catch of western flower thrips (Thysanoptera: Thripidae) as affected by color and height of sticky card traps in mature greenhouse cucumber crops. *Journal of Economic Entomology* 83: 971–975.
- Griffin ML, Yeagan KV. 2002. Oviposition site selection by the spotted lady beetle *Coleomegilla maculata* (Coleoptera: Coccinellidae): choices among plant species. *Environmental Entomology* 31: 107–111.
- Hansen EA, Funderburk JE, Reitz SR, Ramachandran S, Eger JE, McAuslane H. 2003. Within-plant distribution of *Frankliniella* species (Thysanoptera: Thripidae) and *Orius insidiosus* (Heteroptera: Anthocoridae) in field pepper. *Environmental Entomology* 32: 1035–1044.
- Helyer NL, Brobyn PJ. 2008. Chemical control of western flower thrips (*Frankliniella occidentalis* Pergande). *Annals of Applied Biology* 121: 219–231.
- Huang N, Enkegaard A, Osborne LS, Ramakers PMJ, Messelink G, Pijnakker J, Murphy G. 2011. The banker plant method in biological control. *Critical Reviews in Plant Sciences* 30: 259–278.
- Hunter WB, Ullman DE. 1989. Analysis of mouthpart movements during feeding of *Frankliniella occidentalis* (Pergande) and *F. schultzei* trybom (Thysanoptera: Thripidae). *International Journal of Insect Morphology and Embryology* 18: 161–171.

- Isaacs R, Willis MA, Byrne DN. 1999. Modulation of whitefly takeoff and flight orientation by wind speed and visual cues. *Physiological Entomology* 24: 311–318.
- Jacas J, Urbaneja A, García Marí F. 2008. Artrópodos depredadores, pp. 39–56 *In* Jacas J, Urbaneja A [eds.], *Control Biológico de Plagas Agrícolas*. Phytoma, Valencia, Spain.
- Karlsson MF, Birgersson G, Cotes Prado AM. 2009. Plant odor analysis of potato: response of guatemalan moth to above- and belowground potato volatiles. *Journal of Agricultural and Food Chemistry* 57: 5903–5909.
- Koschier EH, de Kogel WJ, Visser JH. 2000. Assessing the attractiveness of volatile plant compounds to western flower thrips (*Frankliniella occidentalis* Pergande). *Journal of Chemical Ecology* 26: 2643–2655.
- López Ávila A, Rincón DF. 2006. Diseño de un olfatómetro de flujo de aire para medir respuestas olfativas de insectos de tamaño mediano y pequeño. *Revista Corpoica — Ciencia y Tecnología Agropecuaria* 7: 61–65.
- Lorenzo ME, Bao L, Mendez L, Grille G, Bonato O, Basso C. 2019. Effect of two oviposition feeding substrates on *Orius insidiosus* and *Orius tristicolor* (Hemiptera: Anthocoridae). *Florida Entomologist* 102: 395–402.
- Lundgren JG, Fergen JK. 2006. The oviposition behavior of the predator *Orius insidiosus*: acceptability and preference for different plants. *BioControl* 51: 217–227.
- Lundgren JG, Fergen JK, Riedell WE. 2008. The influence of plant anatomy on oviposition and reproductive success of the omnivorous bug, *Orius insidiosus*. *Animal Behaviour* 75: 1495–1502.
- Madadi H, Enkegaard A, Brødsgaard HF, Kharrazi-Pakdel A, Ashouri A, Mohaghegh-Neishabouri J. 2008. *Orius albidipennis*: intraguild predation of and prey preference for *Neoseiulus cucumeris* on different host plants. *Entomologica Fennica* 19: 32–40.
- Maeso D, Paullier J, González P, Arboleya J, Alfredo Fernández A, Walasek W. 2013. Seguimiento de “peste negra” en morrón: experimentos en la zona sur de Uruguay. *INIA Serie Actividades de Difusión* 723: 1–44.
- Magalhães DM, Borges M, Laumann RA, Sujii ER, Mayon P, Caulfield JC, Midega CAO, Khan ZR, Pickett JA, Birkett MA, Blassioli Moraes MC. 2012. Semiochemicals from herbivory induced cotton plants enhance the foraging behavior of the cotton boll weevil, *Anthonomus grandis*. *Journal of Chemical Ecology* 38: 1528–1538.
- Mendes SM, Bueno VHP, Carvalho LM. 2005. Adequabilidade de diferentes substratos à oviposição do predador *Orius insidiosus* (Say) (Hemiptera: Anthocoridae). *Neotropical Entomology* 34: 415–421.
- MGAP. 2016. Encuestas Hortícolas 2015-2016 – Zonas Sur y Litoral Norte. <https://www.gub.uy/ministerio-ganaderia-agricultura-pesca/datos-y-estadisticas/estadisticas/encuestas-hortícolas-2015-2016-zonas-sur-litoral-norte-344> (last accessed 28 Jul 2020).
- Muñiz M, Cibrián J, Hidalgo C, Bautista N, Vaquera H, Aldama C. 2014. Volatile compounds attract the pepper (*Capsicum* spp.) weevil (*Anthonomus eugenii* Cano) and synergize its aggregation pheromone. *Agrociencia* 48: 819–832.
- Ohno K, Takemoto H. 1997. Species composition and seasonal occurrence of *Orius* spp. (Heteroptera: Anthocoridae), predacious natural enemies of *Thrips palmi* (Thysanoptera: Thripidae), in eggplant fields and surrounding habitats. *Journal of Applied Entomology and Zoology* 32: 27–35.
- Painter RH. 1951. *Insect Resistance to Crop Plants*. The McMillan Co., New York, USA.
- Patt JM, Meikle WG, Mafra-Neto A, Setamou M, Mangan R, Yang C. 2011. Multimodal cues drive host-plant assessment in Asian citrus psyllid (*Diaphorina citri*). *Environmental Entomology* 40: 1494–1502.
- Peraza AR. 2011. Preferencia de hospedero y parámetros de desarrollo de *Copitarsia decolora* sobre plantas seleccionadas para la diversificación del cultivo de uchuva (*Physalis peruviana*). [bdigital.unal.edu.co/8426/](http://bdigital.unal.edu.co/8426/) (last accessed 28 Jul 2020).
- Pérez Rubio AG. 1994. Composición aromática y biogénesis de compuestos volátiles en fresa (*Fragaria ananassa* Duch.) var. Chandler. Doctoral thesis. Universidad de Sevilla, Seville, Spain.
- Pickett J, Wadhams L, Woodcock C, Hardie J. 1992. The chemical ecology of aphids. *Annual Review of Entomology* 37: 67–90.
- Pivnick K, Jarvis B, Slater G, Gillot C, Underhill E. 2019. Attraction of the diamondback moth (Lepidoptera: Plutellidae) to volatiles of oriental mustard: the influence of age, sex, and prior exposure to mates and host plants. *Environmental Entomology* 19: 704–709.
- Rajapakse CNK, Walter GH, Moore CJ, Hull CD, Cribb BW. 2006. Host recognition by a polyphagous lepidopteran (*Helicoverpa armigera*): primary host plants, host produced volatiles and neurosensory stimulation. *Physiological Entomology* 31: 270–277.
- R Core Team. 2016. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- Reid CD, Lampman RL. 1989. Olfactory responses of *Orius insidiosus* (Hemiptera: Anthocoridae) to volatiles of corn silks. *Journal of Chemical Ecology* 15: 1109–1111.
- Ribeiro A, Castiglioni E. 2008. Caracterización de las poblaciones de enemigos naturales de *Piezodorus guildinii* (Westwood) (Hemiptera: Pentatomidae). *Agrociencia* 12: 48–56.
- Richards PC, Schmidt JM. 1996. The suitability of some natural and artificial substrates as oviposition sites for the insidious flower bug, *Orius insidiosus*. *Entomologia Experimentalis et Applicata* 80: 325–333.
- Roselló S, Diaz MJ, Nuez F. 1996. Viral diseases causing economic losses to the tomato crop. I. the tomato spotted wilt virus - a review. *Scientia Horticulturae* 67: 117–150.
- Sabelis MW, van de Baan HE. 1983. Location of distant spider mite colonies by phytoseiid predators: demonstration of specific kairomones emitted by *Tetranychus urticae* and *Panonychus ulmi*. *Entomologia Experimentalis et Applicata* 33: 303–314.
- Saini E, Cervantes V, Alvarado L. 2003. Efecto de la temperatura, la dieta y el hacinamiento sobre la fecundidad, fertilidad y longevidad de *Orius insidiosus* (Say) (Hemiptera: Anthocoridae) predador del trips en cultivos protegidos. *Revista de Investigaciones Agropecuarias* 2: 21–32.
- Salguero Navas VE, Funderburk JE, Olson SM, Beshear RJ. 1991. Damage to tomato fruit by the western flower thrips (Thysanoptera: Thripidae). *Journal of Entomological Science* 26: 436–442.
- Schoonhoven LM, Jermy T, van Loon J. 1998. Host-plant selection: how to find a host plant, pp. 121–153 *In* Schoonhoven LM, Jermy T, van Loon J [eds.], *Insect-Plant Biology*. Chapman & Hall, London, United Kingdom.
- Schoonhoven LM, van Loon JJA, Dicke M. 2005. *Insect-Plant Biology*. Oxford University Press, Oxford, United Kingdom.
- Sherwood J, German TL, Moyer JW, Ullman DE, Whitfield AE. 2000. Tomato spotted wilt, pp. 1030–1031 *In* Maloy OC, Murray TD [eds.], *Encyclopedia of Plant Pathology*. John Wiley & Sons, New York, USA.
- Singer MC, Ng D, Thomas CD. 1988. Heritability of oviposition preference and its relationship to offspring performance within a single insect population. *Evolution* 42: 977–985.
- Smith CM, Boyko EV. 2006. The molecular bases of plant resistance and defense responses to aphid feeding: current status. *Entomologia Experimentalis et Applicata* 122: 1–16.
- van den Meiracker RAF, Ramakers PMJ. 1991. Biological control of the western flower thrips, *Frankliniella occidentalis*, on sweet pepper with the anthocorid predator *Orius insidiosus*. *Mededelingen van de Faculteit Landbouwwetenschappen, Rijksuniversiteit Gent* 56: 241–249.
- van Lenteren JC. 2012. The state of commercial augmentative biological control: plenty of natural enemies, but a frustrating lack of uptake. *Biocontrol* 57: 1–20.
- van Lenteren JC, Bolckmans K, Köhl J, Ravensberg WJ, Urbaneja A. 2018. Biological control using invertebrates and microorganisms: plenty of new opportunities. *BioControl* 63: 39–59.
- Visser JH. 1986. Host odor perception in phytophagous insect. *Annual Review of Entomology* 31: 121–144.
- Visser JH, Piron PGM. 1998. An open Y-track olfactometer for recording of aphid behavioural responses to plant odours. *Entomologia Experimentalis et Applicata* 9: 41–46.
- Whitfield AE, Ullman DE, German TL. 2005. Tospovirus-thrips interactions. *Annual Review of Phytopathology* 43: 459–489.
- Yoneya K, Kugimiya S, Takabayashi J. 2009. Do adult leaf beetles (*Plagioderma versicolora*) discriminate between odors from intact and leaf-beetle-infested willow shoots? *Journal of Plant Interactions* 4: 125–129.
- Yoneya K, Ozawa R, Takabayashi J. 2010. Specialist leaf beetle larvae use volatiles from willow leaves infested by conspecifics for reaggregation in a tree. *Journal of Chemical Ecology* 36: 671–679.