

## **A molecular phylogenetic analysis of the Oedipodinae and their intercontinental relationships**

Authors: Fries, Megan, Chapco, William, and Contreras, Daniel

Source: Journal of Orthoptera Research, 16(2) : 115-125

Published By: Orthopterists' Society

URL: [https://doi.org/10.1665/1082-6467\(2007\)16\[115:AMPAOT\]2.0.CO;2](https://doi.org/10.1665/1082-6467(2007)16[115:AMPAOT]2.0.CO;2)

---

BioOne Complete ([complete.BioOne.org](https://complete.BioOne.org)) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at [www.bioone.org/terms-of-use](https://www.bioone.org/terms-of-use).

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

---

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

# A molecular phylogenetic analysis of the Oedipodinae and their intercontinental relationships

Accepted September 12, 2007

MEGAN FRIES, WILLIAM CHAPCO AND DANIEL CONTRERAS

Department of Biology, University of Regina, Regina, Saskatchewan, S4S 0A2, Canada. Email: William.Chapco@uregina.ca

## Abstract

Oedipodine grasshoppers occur throughout the major continents, making them the most widely distributed of the 30 subfamilies that comprise the Acrididae. Most species have been allocated to one of 15 tribes; some remain unassigned. The subfamily, according to Vickery, had an ancient origin, just after the breakup of Pangaea but before the separation of Laurasia from Africa. Thereafter, Oedipodinae continued to evolve in separate continental centers; some Nearctic species apparently descended more recently from Palearctic ancestors when land bridges still connected the two continents.

Our objectives are to independently assess these biogeographic accounts, to examine the validity of several tribal constructs, and to shed light on problematic taxa such as *Stethophyma* and *Machaerocera* which have had, over the years, an ambivalent affiliation with Oedipodinae. To realize these goals, we sequenced and phylogenetically analyzed portions of four mitochondrial genes (coding for cytochrome oxidase subunits I and II, cytochrome b, and NADH dehydrogenase subunit V), totaling up to 2254 bp, in specimens collected in the Americas, Eurasia, Africa and Australia. Methodology entailed applying weighted and unweighted maximum parsimony, maximum likelihood and Bayesian techniques. A member of the Pyrgomorphidae served as the outgroup. The ages of evolutionary divisions were estimated using the program "r8s"; the date of 100 Mya, previously estimated as the time of divergence between the subfamilies Oedipodinae and Gomphocerinae, was used to calibrate our chronogram.

In general, taxa appear to assort themselves according to continental land mass, rather than by tribe. Aioloipini, Bryodemini, Oedipodini and Spingonotini proved to be nonmonophyletic, whereas there was no evidence to reject monophyly in Acrotilyni, Chortophagini, Locustini and Psinidini. Phylogenetically, both *Machaerocera* and *Stethophyma* were well-positioned within the Oedipodinae, with *Machaerocera* closely aligned with *Chortophaga* and *Encoptolophus*, and *Stethophyma* tightly linked to *Aiolopus*. *Duroniella*, presently regarded as an oedipodinid, emerged strongly connected to the Gomphocerinae.

The current biogeographical distribution of Oedipodinae is the result of widespread intercontinental dispersion. In particular, with the assistance of DIVA analysis, we argue that Asiamerica was the center of initial oedipodinid radiation about 94 Mya. Through a series of early dispersals, the remaining clusters of taxa were established. Somewhat surprisingly, this includes the branch leading to the Australian genera *Austroicetes* and *Chortoicetes*. In contrast, the multiple dispersals to the African continent occurred more recently. It would appear that North American oedipodinids had both an ancient and a more recent ancestry. The single South American species analyzed evolved very recently from North American ancestors.

## Key words

Orthoptera, Oedipodinae, Gomphocerinae, phylogeny, biogeography, mitochondrial DNA, molecular clock

## Introduction

Molecular phylogenetic studies of intercontinental taxa above the species level provide opportunities for investigating the evolutionary impact of geological and climatological processes in the distant past. For the past decade, the number of such studies has steadily increased for several insect orders, for example: Coleoptera (e.g., Pearson & Vogler 2001, Davis *et al.* 2002), Diptera (e.g., Barrio & Ayala 1997, Martin *et al.* 2002), Hemiptera (e.g., Buckley *et al.* 2002, von Dohlen *et al.* 2006), Hymenoptera (e.g., Leys *et al.* 2002, Kawakita *et al.* 2004), Lepidoptera (e.g., Zakharov *et al.* 2004, Hundsdoefer *et al.* 2005), to name just a few. Challenges to traditional views on place and time of origin and directionality of migration (e.g., Pearson & Vogler 2001, Costa *et al.* 2003, Hundsdoefer *et al.* 2005, von Dohlen *et al.* 2006) have often been the result.

Within the Orthoptera, however, there have been few comparable investigations. In one recent example, a mitochondrial DNA (mtDNA) phylogenetic analysis (Lovejoy *et al.* 2005) of Old and New World *Schistocerca* species demonstrated that the genus originated in Africa and not the Western Hemisphere, as had been proposed in an earlier morphological investigation (Song 2004). Rather, a single east-west, trans-Atlantic dispersal event took place, eventually leading to the establishment of the numerous species presently distributed throughout the Americas. Rowell and Flook (2004), also employing mtDNA, speculated on the place of origin of the neotropical subfamily Proctolabinae, identifying proto-Central and South American land areas as alternative locations. Recent findings (Chapco *et al.* 2001, Amédégnato *et al.* 2003) challenged the prevailing view (Vickery 1989) that the subfamily Melanoplinae originated in Laurasia and that during the Pliopleistocene Great Interchange, incursions from the north led to the establishment of taxa in South America. Instead, molecular phylogenetic analyses of mitochondrial genes showed the reverse, that the subfamily originated in South America, probably in the Early Cenozoic, and subsequently, via island-hopping, progressed to establish the Holarctic fauna. A similar analysis (Contreras & Chapco 2006) of Holarctic Gomphocerinae supported Vickery's (1989) contention that there were at least three dispersal events from Eurasia to North America. More recently, with the inclusion of taxa from the southern hemisphere, preliminary analyses (unpub.) have not contradicted that conclusion; however, they further suggest the possibility that the subfamily originated still earlier in Gondwanaland.

Another subfamily, whose distribution surpasses even those of Gomphocerinae and Melanoplinae, is Oedipodinae (= Locustinae), or the band-winged grasshoppers. Its over 900 species and 185 genera occur throughout the major continents, making the subfamily the most cosmopolitan among the 30 subfamilies of the Acrididae

(Vickery & Kevan 1985, Otte 1995). Among their numbers can be counted several infamous pests, such as the migratory locust (*Locusta migratoria*), the Australian plague locust (*Chortoicetes terminifera*), and the clear-winged grasshopper (*Camnula pellucida*).

Taxonomically, oedipodine grasshoppers over the years, have been grouped as a tribe, as a subfamily and at times, as a family [see Guliaeva *et al.* (2005) for a summary]. The subfamily designation is now generally accepted (Otte 1984), but not by all (see Rentz 1996). In the most recent version of the Orthoptera Species File (OSF2) (Eades *et al.* 2007), Oedipodinae is subdivided into 15 tribes, of which a few such as Locustini and Sphingonotini occur on two or more continents. Most however, are restricted to only one land mass. Morphological similarities among continentally separated taxa have led to speculations about the subfamily's historical origins, a topic of interest to orthopterists for about half a century, starting with Rehn's (1958) seminal paper on North American species. Rehn made brief reference to connections with Eurasian taxa, but on the whole focused on identifying probable centers of origin in the New World. Vickery (1987, 1989) proposed that initially the subfamily had evolved over 100 Mya, before the complete sundering of Pangaea; subsequently, diversification continued in separate Nearctic, Palearctic and Ethiopian centers. Vickery (1989) also viewed some Nearctic elements as descendants of more recent invaders from the Old World, entering North America via one of several land bridges that had connected the two land masses. The subfamily is poorly represented in the Neotropics (about seven genera), which would suggest that the incursion from the north was fairly recent (Rehn 1958, Carbonell 1977). The subfamily is well represented on the African continent (Otte 1984), but apart from Vickery's brief statement cited above, very little (see Ritchie 1981, 1982) has been proposed on the origin of that continent's oedipodine fauna.

Unfortunately, the few published phylogenies — both morphologically based (*e.g.*, Otte 1984) and molecularly based (*e.g.*, Chapco *et al.* 1997, Rowell & Flook 2004, Guliaeva *et al.* 2005, Lu & Huang 2006) — are inadequate for testing these biogeographic hypotheses. Trees appear either somewhat arbitrary in their construct (*e.g.*, Otte 1984), or they include too few oedipodine genera (Rowell & Flook 2004, Guliaeva *et al.* 2005), or they focus on only one continent (Chapco *et al.* 1997, Lu & Huang 2006). The present study is a phylogenetic analysis of mtDNA sequences from a selected group of oedipodine grasshoppers sampled from both Old and New-World continents. Our objectives are 1) to shed light on the subfamily's origins and thereby test the aforementioned biogeographic hypotheses, and 2) to add to our ongoing understanding of taxonomic relationships and organization within the subfamily Oedipodinae. Of possible interest to orthopterists, the Australian oedipodine genera, *Austroicetes* and *Chortoicetes* — neither of which has been assigned to tribe — are included. We also provide further insight into the phylogenetic affinities of *Machaerocera* and *Stethophyma*, two genera that have had a somewhat uncertain relationship with Oedipodinae (Otte 1984).

## Materials and Methods

Species, along with sources, are listed in Table 1. Included are 22 species from Eurasia/Africa, 12 from North America, one from South America and two from Australia. Collectively, these represent 12 of the tribes listed in the OSF2. Two tribes, Bryodemini and Sphingonotini, contain both New and Old-World genera. In order to assist in estimating times of divergence (see below), five members of the closely related subfamily, Gomphocerinae, were included. *Pyrgomorpha conica* was employed as the outgroup. An earlier in-

vestigation (Flook & Rowell 1997) had established that within the Acridoidea, the Pyrgomorphae are basal to the Acrididae.

DNA was extracted from specimens using either the DTAB/CTAB method outlined in Philips and Simon (1995) or using a QIAGEN DNeasy tissue kit (Mississauga, Canada). Portions of the mitochondrial genes encoding NADH dehydrogenase subunit V (ND5), cytochrome oxidase subunit I (CO1) and II (CO2), and cytochrome b (cytb) were amplified and sequenced. Primer sequences, PCR gene amplification conditions, as well as DNA sequencing methods, are described elsewhere (Litzenberger & Chapco 2001a, 2001b; Contreras & Chapco 2006). [Two additional primers used for amplifying cytb sequences are:

mtd26 5'-TATGTACTACCATGAGGACAAATATC-3' and  
mtd28 5'-ATTACACCTCTAATTATTAGGAAT-3'].

Sequences were easily aligned by visual inspection, imported into MacClade (Maddison & Maddison 2004) and analyzed using the software packages PAUP\* (version 4.0b8 — Swofford 2003) and MrBayes (MB) (Version 3.0b4 — Huelsenbeck & Ronquist 2001). Both standard maximum parsimony (MP) and weighted maximum parsimony (wMP), following Farris' (1969) iterative reweighting scheme, were used. In addition to using parsimony and Bayesian methods, maximum likelihood (ML) was applied, also available in PAUP\*. In order to reduce the run-time for ML, parameter estimates provided by the program Modeltest (Version 3.6 — Posada & Crandall 1998) were used as input values. Levels of support for parsimony-derived relationships were estimated through 1000 bootstrap replicates. Bayesian analyses provided measures of nodal support in the form of posterior probabilities (PP). For all analyses, the four sequences were treated as a combined unit, a procedure that, as in our previous studies (Chapco *et al.* 2001; Litzenberger & Chapco 2001a, 2001b), always yielded trees with greater resolution and support when compared to those based on single genes.

In order to place biogeographic events within a geological context, it was important to estimate the times of divergence for various nodes. We initially applied the maximum likelihood ratio test (Page & Holmes 1998) to determine whether sequences evolved in a clock-like manner. Because sequences did not in fact conform to a model of rate constancy, we estimated divergence times by employing a semiparametric penalized-likelihood (PL) method, which can accommodate rates that vary over lineages (Sanderson 2002). To this end, the program r8s, version 1.70 (Sanderson 2004) was used. As recommended by Sanderson, the TN (Truncated Newton) algorithm was applied in conjunction with PL. A cross-validation analysis was first performed to determine the most likely smoothing parameter (a measure of the relative contributions of parametric and nonparametric models that underlie PL), necessary for estimating optimal divergence times. Zero-length branches were collapsed. A more extensive description of the method and theory is given by Sanderson (2002). The program yields estimates of absolute times of divergence if at least one known divergence date is provided as input. Usually these times are based on the fossil record, which in the case of Acrididae is rather poor (Vickery 1989) [To date, the earliest fossil on record that is unequivocally an oedipodine dates to the Miocene (Stidham & Stidham 2000), too recent for the specimen to be ancestral]. Instead, we relied on the work of Gaunt and Miles (2002) who, calibrating their molecular clock using dated ancient cockroach fossils, estimated the time of split between subfamilies Oedipodinae and Gomphocerinae at about 100 Mya. Accordingly we have used this value to calibrate our chronogram.

**Table 1.** Species analyzed, locations and GenBank Accession numbers of mtDNA sequences.

| Subfamily/Tribe* | Species                            | Source                                | Accession Nos<br>CO1, CO2, cytb, nd5   |
|------------------|------------------------------------|---------------------------------------|--|
| Oedipodinae      |                                    |                                       |  |
| /Acrotylini      | <i>Acrotylus blondeli</i>          | Torodi, Niger <sup>ab</sup>           | EF151860, --, EF151895, EF151922       |
|                  | <i>Acrotylus insubricus</i>        | Malatya, Turkey                       | EF151836, EF151816, EF151870, EF151904 |
|                  | <i>Acrotylus patruelis</i>         | Salobrena, Spain                      | EF151859, --, EF151894, EF151921       |
| /Aiolopini       | <i>Aiolopus simulatrix</i>         | Niamey, Niger <sup>abc</sup>          | EF151862, --, EF151897, --             |
|                  | <i>Aiolopus strepens</i>           | Sierra Nevada, Spain                  | EF151841, EF151821, EF151875, EF151907 |
|                  | <i>Aiolopus thalassinus</i>        | Nerja, Spain                          | EF151842, EF151822, EF151876, --       |
|                  | <i>Duroniella fracta</i>           | Malatya, Turkey <sup>ab</sup>         | DQ230738, DQ230807, DQ230827, DQ230774 |
|                  | <i>Heteropternis coulöniana</i>    | Mt Nimba, Guinea <sup>abc</sup>       | EF151858, EF151830, EF151892, EF151920 |
| /Arphiini        | <i>Arphia conspersa</i>            | Jameson, SK <sup>d</sup>              | EF151839, EF151819, EF151873, --       |
| /Bryodemini      | <i>Angaracris barabensis</i>       | Gansu Prov., China <sup>a</sup>       | EF151856, --, EF151890, --             |
|                  | <i>Bryodema luctuosum</i>          | Gansu Prov., China <sup>a</sup>       | EF151854, EF151829, EF151888, EF151917 |
|                  | <i>Circotettix carlinianus</i>     | Condie, SK <sup>d</sup>               | EF151845, --, EF151879, EF151909       |
| /Chorthophagini  | <i>Chorthophaga viridifasciata</i> | Jameson, SK <sup>d</sup>              | EF151864, --, EF151899, EF151924       |
|                  | <i>Encoptolophus costalis</i>      | Regina, SK <sup>d</sup>               | EF151850, EF151827, EF151884, EF151913 |
| /Locustini       | <i>Gastrimargus africanus</i>      | Niamey, Niger <sup>abc</sup>          | --, EF151831, EF151893, --             |
|                  | <i>Locusta migratoria</i>          | Genbank <sup>abc</sup>                | X80245                                 |
|                  | <i>Oedaleus decorus</i>            | Massif Central, France <sup>abc</sup> | EF151834, EF151814, EF151868, EF151903 |
|                  | <i>Oedaleus asiaticus</i>          | Gansu Prov., China                    | EF151865, --, EF151900, --             |
| /Macherocerini   | <i>Machaerocera mexicana</i>       | Oaxaca, Mexico <sup>d</sup>           | EF151861, --, EF151896, EF151923       |
| /Oedipodini      | <i>Celes variabilis</i>            | Massif Central, France <sup>a</sup>   | EF151855, --, EF151889, EF151918       |
|                  | <i>Oedipoda caerulea</i>           | Massif Central, France <sup>ab</sup>  | EF151835, EF151815, EF151869, --       |
|                  | <i>Oedipoda miniata</i>            | Erzingan, Turkey                      | EF151840, EF151820, EF151874, EF151906 |
| /Psinidiini      | <i>Metator pardalinus</i>          | Last Mountain, SK <sup>d</sup>        | EF151849, EF151826, EF151883, EF151912 |
|                  | <i>Trachyrhachys kiowa</i>         | Condie, SK <sup>d</sup>               | EF151846, --, EF151880, EF151910       |
| /Sphingonotini   | <i>Dissosteira carolina</i>        | Bimidji, WISC <sup>d</sup>            | EF151851, EF151828, EF151885, EF151914 |
|                  | <i>Spharagemon collare</i>         | Jameson, SK <sup>d</sup>              | EF151852, --, EF151886, EF151915       |
|                  | <i>Spharagemon campestris</i>      | Condie, SK                            | EF151838, EF151818, EF151872, --       |
|                  | <i>Sphingonotus caeruleus</i>      | Sierra Nevada, Spain <sup>abc</sup>   | EF151844, EF151824, EF151878, EF151908 |
|                  | <i>Sphingonotus yenchihensis</i>   | Gansu Prov., China                    | EF151853, --, EF151887, EF151916       |
|                  | <i>Sphingonotus nebulosus</i>      | Malatya, Turkey                       | EF151866, EF151832, EF151901, --       |
|                  | <i>Trimerotropis pistrinaria</i>   | Findlater, SK <sup>de</sup>           | EF151848, EF151825, EF151882, EF151911 |
|                  | <i>Trimerotropis pallidipennis</i> | Uspallata, Argentina                  | EF151863, --, EF151898, --             |
| /Unassigned      | <i>Austroicetes sp.</i>            | Canberra, Australia <sup>c</sup>      | EF185880, --, EF185881, --             |
|                  | <i>Chortoicetes terminifera</i>    | Merriwah, Australia <sup>c</sup>      | EF185877, EF185876, EF185878, EF185879 |
|                  | <i>Wernerella pachecoi</i>         | El Llano, Canary Is. <sup>b</sup>     | EF151857, --, EF151891, EF151919       |
| Acridinae        | <i>Stethophyma gracile</i>         | Last Mountain, SK <sup>ad</sup>       | DQ230737, DQ230806, DQ230826, DQ230773 |
|                  | <i>Stethophyma grossum</i>         | Massif Central, France                | DQ230735, DQ230804, --, DQ230771       |
| Gomphocerinae    | <i>Aeropedellus clavatus</i>       | Dilke, SK                             | DQ230708, DQ230777, EF565468, DQ230741 |
|                  | <i>Chorthippus curtipennis</i>     | Pinawa, MB                            | DQ230709, DQ230779, DQ230809, DQ230743 |
|                  | <i>Dociostaurus maroccanus</i>     | Montpellier, France                   | DQ230714, DQ230784, DQ230814, DQ230748 |
|                  | <i>Paracyptera brevipennis</i>     | Montpellier, France                   | DQ230713, DQ230783, DQ230813, DQ230747 |
|                  | <i>Stenobothrus zubowskyi</i>      | Malatya, Turkey                       | DQ230716, DQ230786, DQ230816, DQ230750 |
| Outgroup         | <i>Pyrgomorpha conica</i>          | Malatya, Turkey                       | EU031777, EU031776, EU031778, EU031779 |

\* - according to the OSF2; MB = Manitoba, SK = Saskatchewan, WISC = Wisconsin; Distribution of genera: a – Eurasia, b – Africa, c – Australia, d – North America, e – South America



Ancestral geographic areas were reconstructed with the assistance of the program "DIVA" (Version 1.1) (Ronquist 1996, 1997). Analysis was simplified by collapsing the dataset to genera. Each genus was coded as 0 or 1 according to its presence in Africa, Australia, Eurasia, North America or South America (even though the particular species analyzed was restricted to one continent). For "maxareas", a parameter that limits the range of ancestral distributions, two sets of values were employed: the default (which favors vicariance) and 2 (which favors dispersal). While the results from DIVA proved helpful, some artifacts appeared. For instance land masses assigned to certain nodes proved unlikely as ancestral areas, given the tectonic events during the times suggested by our temporal analysis; consequently output was interpreted and modified, taking those features into account.

## Results

The overall A+T content in Oedipodinae is about 69.6%, virtually the same as that obtained for subfamilies Melanoplineae (Litzenberger & Chapco 2001a) and Gomphocerinae (Contreras & Chapco 2006). Base compositions did not differ significantly among the 37 taxa, averaging 31.6% (A), 16.2% (C), 14.2% (G) and 38.0% (T). Across the four genes, spanning up to 2254 bp, 1254 sites were variable, of which 873 were phylogenetically informative.

Based on parsimony analysis, maximum resolution was achieved when sites were weighed according to their rescaled consistency indices (Fig. 1A). Bayesian methods (Fig. 1B) recovered the same associations and branching orders (Fig. 1B), with a couple of minor differences (see below). The ML tree (presented as a chronogram in Fig. 2) is essentially the same as the Bayesian topology, with the exception that the *Aiolopus-Stethophyma* clade is positioned as in Fig. 1A. Bootstrapping using ML was abandoned owing to excessive run-times. Modeltest identified the general time reversal model (GTR) with variable rates (G) and invariable sites (I) as the one best fitting the data; Modeltest parameter estimates were very similar to Bayesian values.

In broad terms, all approaches agree in identifying groups of Old and New World genera, labeled in the figures A to F (note that, between methods, configurations of taxa within some groups vary slightly – see below). On the whole, the Bayesian tree is somewhat less resolved: *Oedipoda* occupies an unresolved position within groups A to C and cluster D occupies an unresolved position within A to E. In the parsimony tree, *Oedipoda* is basal to groups B and C, and in the ML tree it is basal to A. Another difference between the outcomes of the methods concerns group C. It is monophyletic in the Bayesian and ML trees, but paraphyletic (to B) in the parsimony tree. Within group C, the association of *Celes* and *Sphingonotus nebulosus* with other taxa differs among methods. All approaches show that clades D and E (the Australian group) are external to groups A to C. Clade F, consisting of *Encoptolophus* to *Machaerocera*, is outside all remaining oedipodids, excepting *Duroniella*. The latter is external to the five gomphocerine genera clustered as a monophyletic group.

The following tribes emerged as monophyletic assemblages (the numbers of genera sampled from the complete list in OSF2 are indicated in brackets): Locustini (4/12), Acrotylini (1/2) and Chortophagini (2/4). Remaining tribes: Aiolopini, Bryodemini, Oedipodini, Psinidini and Sphingonotini proved to be nonmonophyletic. Trees in which each tribe was constrained to be monophyletic were constructed and all, except Psinidini, had significantly lower likelihood values compared to that of the ML tree [comparisons

were based on Kishino-Hasegawa and Shimodair-Hasegawa tests, available in PAUP\*]. Monophyly on the part of Psinidini, here consisting of *Metator* and *Trachyrhachys*, could not be statistically rejected. Members of some tribes are widely separated phylogenetically. For example, *Heteropternis*, listed among the Aiolopini, belongs to group A, whereas another member, *Aiolopus*, connects with *Stethophyma* within group D.

## Discussion

### Monophyly or lack thereof

Previously considered part of Acridinae (Bei-Bienko & Mischenko 1964), *Duroniella* is now listed in the OSF2 among the Oedipodinae. However, according to our analysis, *Duroniella* is very closely associated with the Gomphocerinae (99% bootstrap and PP). Whether this position remains invariant will depend on the outcome of studies underway which include members of Acridinae. In any case, without this genus, Oedipodinae can be regarded as monophyletic. Tribal groupings are another matter.

Given a lack of support for monophyly of most tribes, it would appear that the very traits used to define this category are the result of convergence, probably brought about by natural selection factors imposed by similar habitats and conditions. Morphological similarities between continentally-separated, but phylogenetically unrelated, taxa have been noted in other acridids as well and have been similarly attributed to ecological convergence (Amédégno 1993, Rowell 2005).

It is entirely possible that, with further sampling, Locustini, Acrotylini, Chortophagini, and perhaps Psinidini, may also prove to be nonmonophyletic. Indeed, this is probably true of Locustini, Guliaeva *et al.* (2005) having discovered that one member, *Psophus*, was far removed from *Locusta* and *Oedaleus*. In an earlier work Otte (1984) included the Old World genus *Acrotylus* in Psinidia, but our data provide no support for that association.

### Sphingonotini and Bryodemini

Sphingonotini is the largest oedipodine tribe with 22 genera listed in the OSF2, and, according to Rehn (1958) and Otte (1984), has an affinity with Bryodemini. Although the present study does (strongly) support a connection between the two tribes, neither is monophyletic, a conclusion also reached by Guliaeva *et al.* (2005). However, there is a clean separation between North American and Eurasian members (some genera also occur in Africa and Australia; see below) of Sphingonotini and Bryodemini. On the Eurasian side, these elements together with *Wernerella* and *Celes*, form a monophyletic or a paraphyletic group according to the method used. Internal within the North American group is *Circotettix*, which, according to Otte (1984) and the OSF2, is the single North American representative of the otherwise Eurasian Bryodemini. Our results question that particular allocation, and appear to confirm earlier cytological findings (White 1973, Weissman & Rentz 1980, Weissman 1984) which showed that *Circotettix*'s chromosomes greatly resemble those of *Trimerotropis*, and in particular, of those species belonging to "Section B" (in White's scheme). This resemblance is reflected in our phylogeny in which *Circotettix* is directly linked to *T. pallidipennis*, a Section B member. The morphological similarity between *Circotettix* and some members of Bryodemini is probably the result of convergent evolution.

A discord between morphology and molecules, of course, is

not universal, as illustrated by *Wernerella*'s close relationship to *Sphingonotus*. In this case, both genera are very similar morphologically (Bland & Gangwere 1998). *Wernerella* has yet to be assigned to tribe.

A case could therefore be made for redefining the two tribes, along continental lines: a New World group consisting of *Spharagemon*, *Trimerotropis*, *Dissosteira* and *Circotettix* and (if the Bayesian tree proves to be correct) a largely Old World group, consisting of *Sphingonotus*, *Wernerella*, *Bryodema*, *Angaracris* and perhaps *Celes* (Guliaeva *et al.* also connected this genus to *Sphingonotus*, but with moderate bootstrap support). [It should be noted that three species of *Sphingonotus* also occur on some Caribbean islands (Otte 1984), but at least one of them seems to have been recently introduced by humans. The one species of *Sphingonotus* that occurs in Australia (Rentz *et al.* 2004), like most congeners with Eurasian relatives (see below), is regarded as a recent invader (Key 1959).] Notwithstanding these exceptions, the above suggestion of dividing these taxa into Old and New World groups is worth further consideration.

### *Aiolopus* and *Stethophyma*

Rowell and Flook (2004), employing mitochondrial rDNA sequences, positioned the genus *Mecostethus*, a member of the same tribe as *Stethophyma* (Storozhenko & Otte 1994), externally to eight species of Oedipodinae; within the latter, the most basal was *Aiolopus*. However, given that no members of Gomphocerinae or Acridinae (at various times *Stethophyma* had been assigned to one or other subfamily) were included in that study, the subfamily affiliation of *Stethophyma*/*Mecostethus* remained unclear. Curiously, in a recent ordination analysis (Petit *et al.* 2006) of several tegminal characters in Acrididae, *Aiolopus* proved to be far removed from *Stethophyma* but much closer to *Mecostethus*. In contrast to both analyses, our results link *Stethophyma* directly with *Aiolopus*, well within the Oedipodinae. For now, it would appear that the question of *Stethophyma*'s subfamily affiliation has been resolved.

Phylogenetic relationships uncovered among the three species of *Aiolopus* aid in interpreting some earlier findings. In crossing experiments (Fuzeau-Braesch & Chapco 1977), the three species mate quite readily, deposit egg pods, but produce no viable offspring. Among the six possible two-way crosses, those involving *A. strepens* yielded the fewest number of egg pods (none in one reciprocal cross). The extent of reproductive isolation is thus reflected phylogenetically, with *A. strepens* occupying a position external and therefore, more genetically distant to the other two species. Our topology also makes sense biogeographically. *A. strepens* has the most restricted geographical distribution, occurring along the Mediterranean coast; in contrast, *A. simulatrix* and *A. thalassinus* occur throughout Eurasia, Africa and, in the case of *thalassinus*, Australia (Bei-Bienko & Mishchenko 1964, Rentz *et al.* 2004). The basal position of *A. strepens* in our phylogeny would indicate that the common ancestor (of at least these three species) occupied the Mediterranean region and subsequently spread out and diversified.

### *Austroicetes* and *Chortoicetes*

According to Rentz *et al.* (2004), these genera are often confused with one another. They also state that *Austroicetes* superficially resembles *Aiolopus* and *Heteropternis*. Based on their morphological similarities and biogeographic distributions, Key (1954) constructed a provisional phylogeny that first linked eight species of *Austroicetes* to *Chortoicetes* and then that group to *Aiolopus*. Despite these ap-

parent affinities, neither Australian genus has yet been assigned to a tribe. The two genera are most certainly closely related (Fig. 1A, B), but no direct relationship with any of the current tribes emerged in our study. As a pair, they are basal to groups A, B, C and perhaps D (parsimony tree).

### *Machaerocera* and *Chortophagini*

This clade constitutes the most ancient offshoot at the base of the oedipodini tree. We are however, unaware of other studies that would suggest this outcome, at least for the Chortophagini. *Machaerocera* does appear to share ecological and morphological features with both Oedipodinae and Acridinae and as a result, Otte (1984) regards the genus as a link between the two subfamilies. Nevertheless, the OSF2 lists *Machaerocera* among the Oedipodinae, as the sole member of the tribe Machaerocerini. [*Machaerocera* is known to possess an unusual set of multiple chromosome associations during meiosis (Helwig 1942) compared to most Acrididae; this feature, however, seems to have very little phylogenetic value.]

### Early history and biogeography of Oedipodinae

The chronogram presented in Fig. 2 is topologically the maximum likelihood tree with lengths between internal and external nodes replaced by estimated times of divergence. The earliest time of oedipodine radiation is the late Cretaceous, about 94 Mya, a date considerably older than Flook and Rowell's (2004) estimate, but in remarkable agreement with Vickery's (1989) date. This initial split gave rise to the basal North American clade and the remaining taxa. Thereafter, major lineages branched off in relatively rapid succession, giving rise to the Australian pair *Austroicetes* and *Chortoicetes* (88 Mya), the cluster consisting of *Aiolopus* and *Stethophyma* (86 Mya), followed by the lineage leading to cluster A, composed of *Locusta* to *Oedipoda* (76 Mya). About 65 Mya, a split between the (largely) Eurasian group C and the (largely) North American cluster B occurred. Thus, the North American oedipodinids, as suspected by Vickery (1989), had both ancient and recent origins.

What then can we say about the place of origin of the subfamily? As a first approximation, DIVA proved useful in identifying ancestral areas and probable dispersal events, but as we shall see, some adjustments were required after geological events were taken into account. DIVA analysis identified three widespread distributional areas as possible roots of Oedipodinae: 1) Eurasia-North America - Australia, 2) North America - Australia, and 3) Eurasia - North America. The first area was recovered when the default "maxareas" option was used, but clearly it can be dismissed, given that by 100 Mya Pangaea had already split into two supercontinents. The last two area outcomes resulted when the "maxareas" was set equal to 2. As with 1), the second possibility is unlikely for the same reason, thus leaving Eurasia - North America (Laurasia) as a logical choice for the place of initial radiation of the subfamily.

DIVA then places Eurasia at each backbone node, except for the ancestor leading to the Australian genera. For that node, DIVA identifies both Australia and Eurasia as sites occupied by that ancestor. Since these two land masses were not conjoined at the time, the implication is that Australia was reached by dispersal. While this is a possibility, it is more likely that movement to the southern continent took place more recently (see below). We therefore decided to assign Eurasia to all the backbone nodes, with North America-Eurasia (Laurasia) at the root. Also, as another adjustment, we decided to regard dispersal as playing the primary role in the establishment of

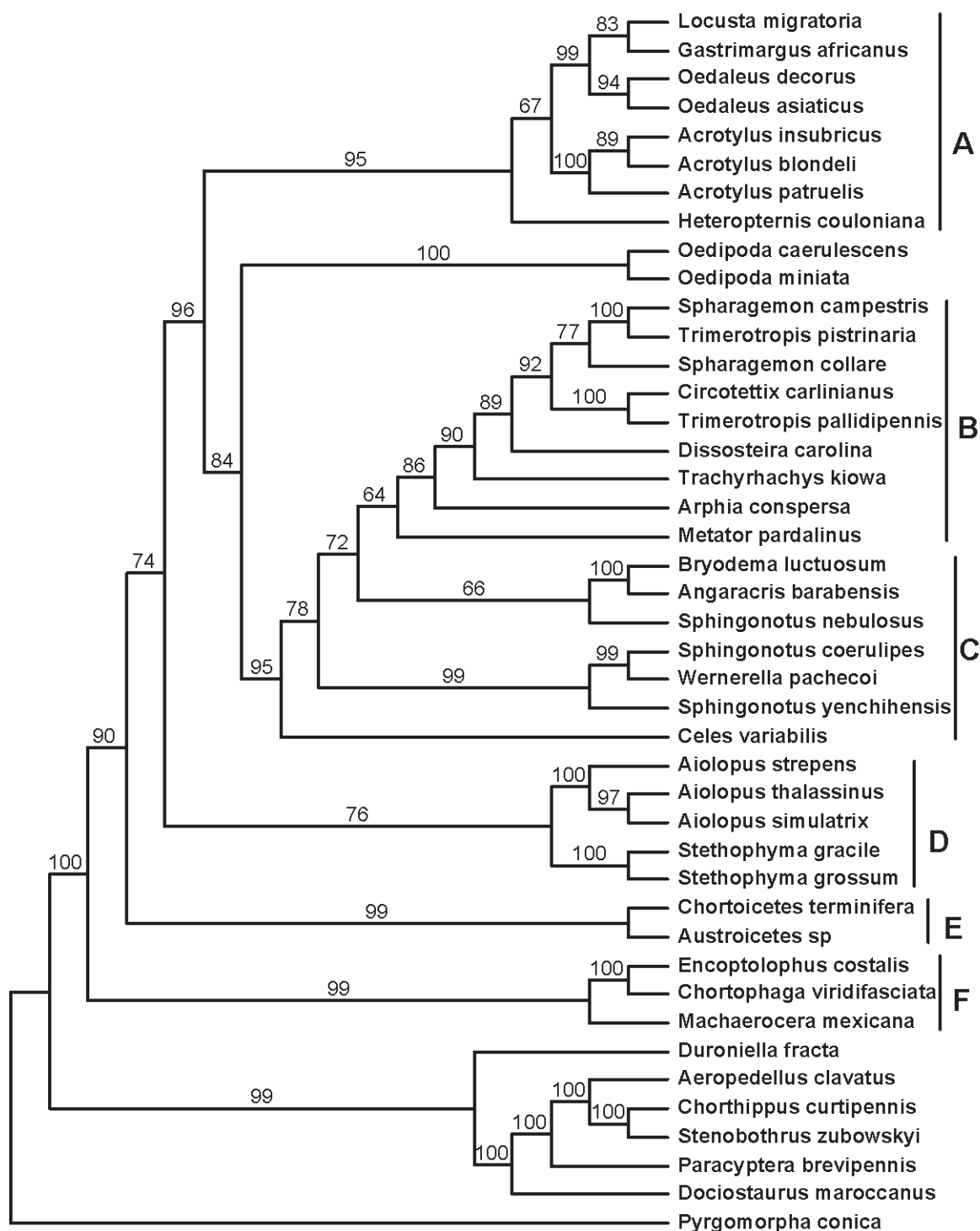
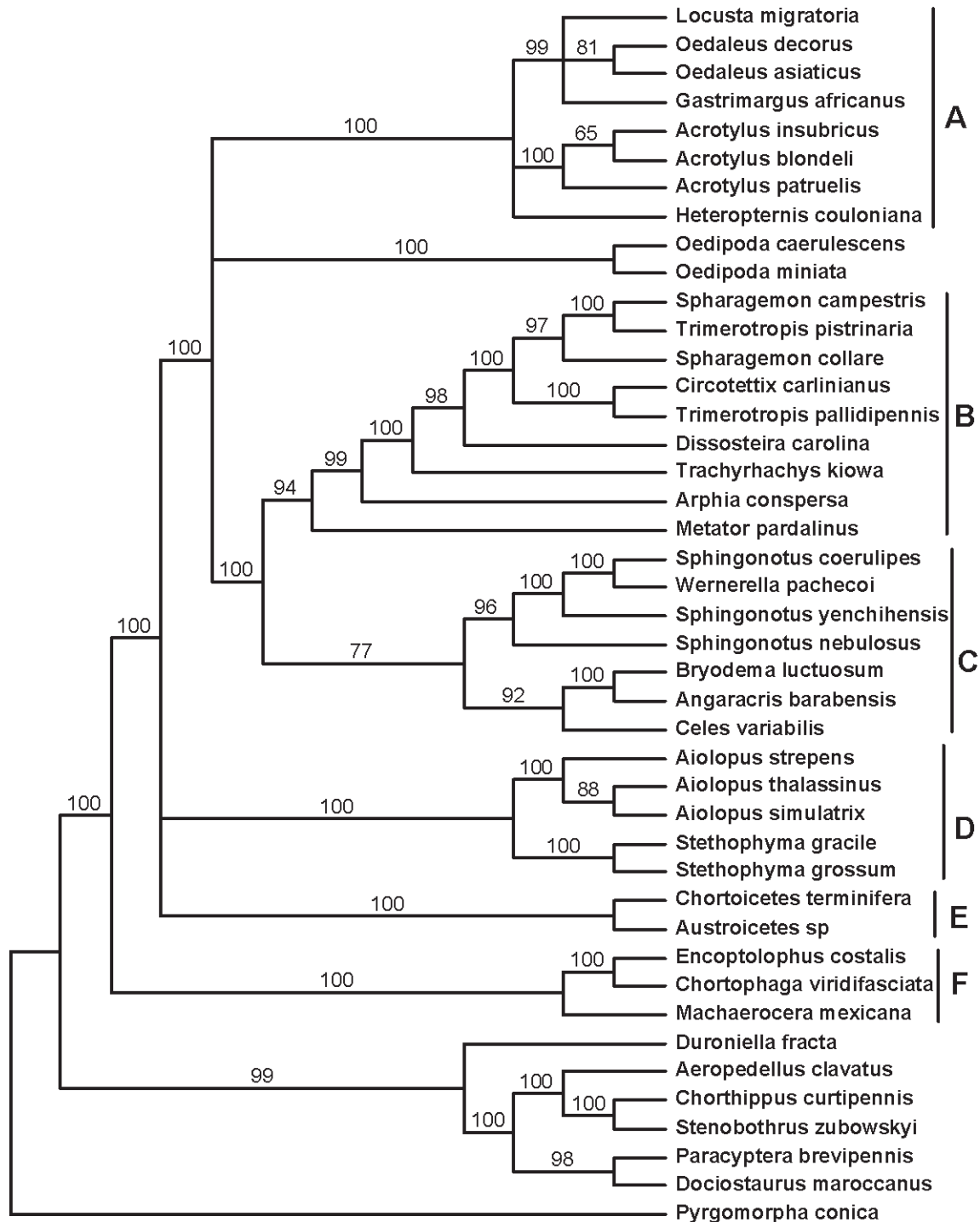


Fig. 1a. Relationships recovered using weighted parsimony. Letters A to F refer to groups of taxa identified in text. Numbers indicate bootstrap levels of support using 1000 replicates.



**Fig. 1b.** Bayesian tree based on model: GTR + G + I. Eight Monte Carlo Markov chains, one cold and seven heated, were run simultaneously for  $1 \times 10^6$  generations. Trees were saved every 200 generations, yielding 5000 trees; the last 2500 were used to estimate the topology, parameter values and posterior probabilities (indicated in figure). Parameter estimates are:  $R_{AC} = 0.0555$ ,  $R_{AG} = 0.2543$ ,  $R_{AT} = 0.0914$ ,  $R_{CG} = 0.0477$ ,  $R_{CT} = 0.5169$ ,  $R_{GT} = 0.0341$ ,  $\pi_A = 0.3170$ ,  $\pi_C = 0.1409$ ,  $\pi_G = 0.1129$ ,  $\pi_T = 0.4290$ ,  $\alpha = 0.7767$ ,  $p_{inv} = 0.3110$ . Letters A to F refer to groups of taxa identified in text.



species distributions because the (two) vicariant events identified by DIVA analysis occurred at a time when the participating continents (e.g., Eurasia and North America) were clearly still together or at least, passage between them was possible. A thoughtful discussion of DIVA's shortcomings when dispersal events predominate can be found in Cook and Crisp (2005). Focusing on dispersal as a major factor is not an unreasonable inference to make considering the flying proclivities of many insect species (e.g., Prüser & Mossakowski 1996, Fuller *et al.* 2005). Indeed, long-distance dispersal is not unheard of in grasshoppers, as illustrated for example by *Chortoicetes* (700 km, Uvarov 1977) and more recently, by the remarkable transatlantic invasion by *Schistocerca* in 1988 (5000 km, Rosenberg & Burt 1999).

As noted, DIVA analysis suggests a vicariant event took place at the base, leading to a split between North America and Eurasia, but contact between these land masses, together constituting Laurasia, was not broken until much later (Askevold 1991). Moreover, approximately 90 Mya, Laurasia consisted of two land masses, Euramerica (eastern North America and Europe) and Asiamerica (western North America and Asia) separated by two epicontinental seas (San Martin *et al.* 2001). Given that the present distribution of the Mexican genus *Machaerocera* overlaps the southern region of the western North American component of Asiamerica, one might provisionally suggest that of the two Laurasian land masses, Asiamerica was the site of oedipodine radiation. [By about 41 Mya, the common ancestor of *Chortophaga* and *Encoptolophus* had appeared, by which time dispersal into eastern North America was made possible by the disappearance of the Mid-Continental Seaway (San Martin *et al.* 2001)]. Additional support for Asiamerica as the center of initial oedipodine radiation is that this possibility presents a reasonable explanation for how an early passage to Australia may have occurred (see below).

The lineage giving rise to the Australian clade split off about 88 Mya. Presumably between that time and 44 Mya, when the common ancestor of *Austroicetes* and *Chortoicetes* appeared, their progenitors had somehow dispersed from Laurasia. [While there are several other oedipodines in Australia, a large proportion of them, such as *Gastrimargus* and *Oedaleus*, have congeners in the Old World (Rentz *et al.* 2004) and are probably the result of recent invasions via New Guinea (Key 1959, Ritchie 1981).] One route may have been via Asia, Southeast Asia and then on to Australia. If passage took place during the earlier part of the time span, long distance dispersal would have been necessary in the latter step, because it was not until 25 to 30 Mya that Australia and Southeast Asia were connected by a series of islands, making island-hopping possible (Jönsson & Fjeldså 2006). It could be argued that, given the vagaries associated with dating molecular clocks and geological events, the upper limit of 30 Mya is not that far removed from 44 Mya. The Australian continent could therefore have been reached in that manner. During 88 to 44 Mya, a range that encompasses a time when climate was favorable to insects in Australia (Raven & Axelrod 1972), alternative pathways might also have been feasible. Two routes, apparently used by other insects, are: Laurasia – South America – Australia via Antarctica (Hundsdoefer *et al.* 2005) and Laurasia – Africa – India – Australia (Fuller *et al.* 2005). The problem with the first scenario is that, as already stated, there are not very many South American oedipodines, and in any case the latter were most likely derived from North American ancestors only recently (Carbonell 1977, Confalonieri *et al.* 1998, this study: see *Trimerotropis pallidipennis* below). It is possible, but unlikely, that at the time there were ancestral oedipodines in South America, these

having since become extinct. Conditions in the Neotropics were certainly favorable during the Early Cenozoic to other subfamilies of Acrididae (Carbonell 1977), and it is therefore difficult to envisage how or why early Oedipodinae in particular might have perished. For the second scenario, that involving Africa and India, at least two transoceanic dispersal events would be required, depending on the timing and positioning of India relative to Africa and Australia as it moved northward toward the Asian continent. For now, our working hypothesis, pending analysis of additional Old-World genera (in particular those endemic to the African continent), is that Oedipodinae originated in Laurasia (probably Asiamerica), and sometime between 88 and 44 Mya, via southeast Asia, reached Australia.

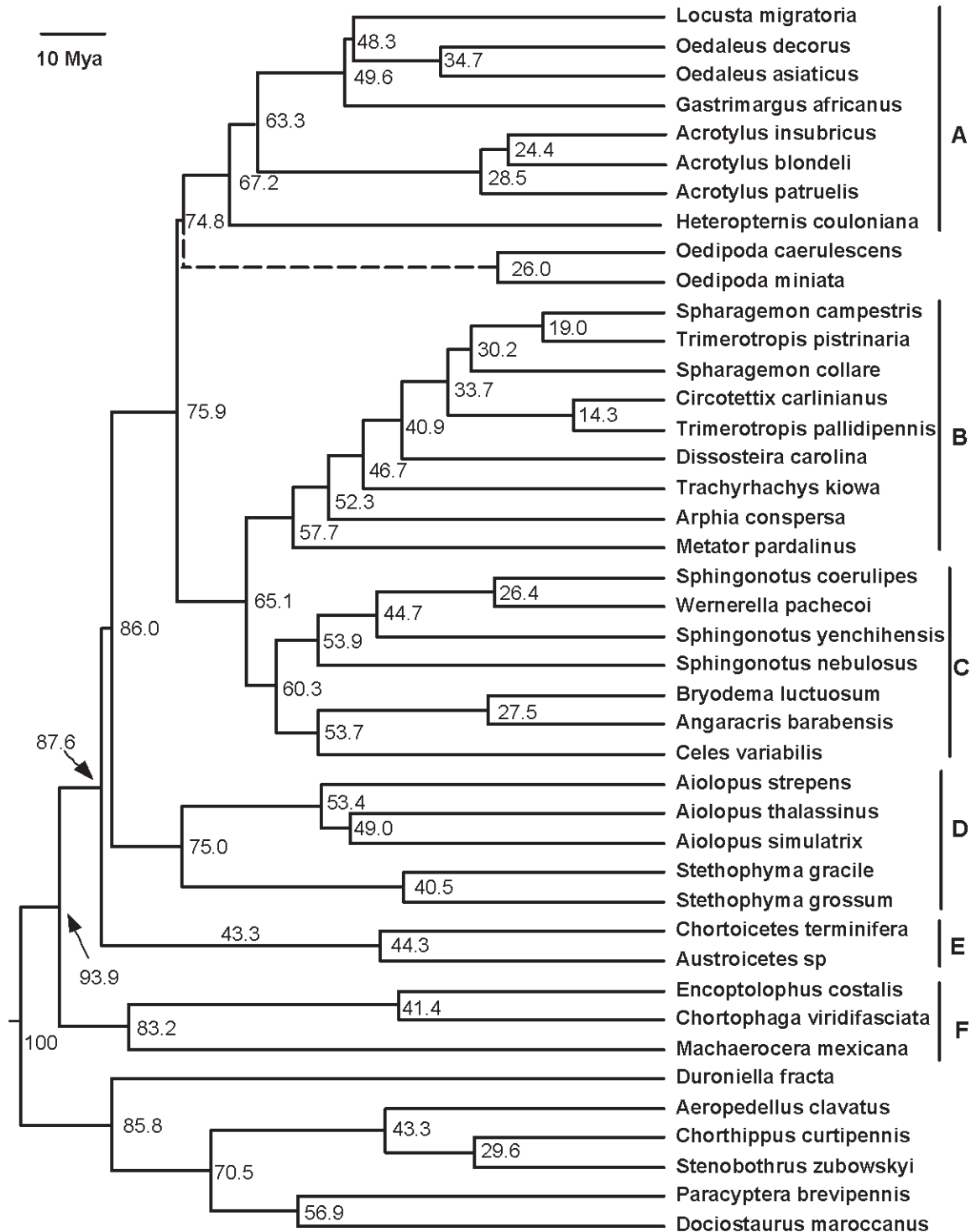
From about 75 to 58 Mya, a series of radiations led to groups D, A, C and B. That Eurasian/African taxa are basal and paraphyletic to the North American clade B, supports the conclusion that this second New World ensemble evolved from Old World ancestors. According to the chronogram, this occurred in the early Tertiary, about 65 Mya. During this period, climatic conditions for insect activity were favorable and both Atlantic and Bering Land Bridges offered possible pathways of incursion (Tiffney 1985, Askevold 1991).

Two sets of species with African connections were analyzed here. The first consists of five species (Table 1) collected in Africa: *Acrotylus blondeli*, *Aiolopus simulatrix*, *Heteropternis coulouanensis*, *Gastrimargus africanus*, and *Wernerella pachecoi*. Except for the latter, all these species, or their genera, occur elsewhere in the Old World, and, in some cases, Australia (Dirsh 1965). The second set consists of *Locusta*, *Oedaleus*, *Oedipoda*, and *Sphingonotus*, collected in Europe or Asia. All these genera also occur in Africa and except for *Oedipoda*, in Australia.

DIVA analysis indicates that several dispersals from Eurasia to Africa and Australia probably took place recently. We cannot provide dates of entry into Africa for the second set, but we can for the first group of species. Our chronogram (Fig. 2) indicates that *Acrotylus blondeli* and *Wernerella pachecoi* diverged from Eurasian ancestors some time in the late Oligocene, a period when the Arabian Bridge linked Asia and Africa (Jolivet & Faccenna 2000) and when a substantial number of intermigrations took place (Hallam 1994). It is somewhat more difficult to account for the earlier divergence of *Gastrimargus* and *Heteropternis* from Eurasian ancestors, respectively about 50 and 67 Mya, in light of the vast (open-water) distance from Africa during those times (Smith *et al.* 2004). Moreover, if ancestors had come from Asia, as suggested above, then two gaps would have had to be traversed: the Turgai Sea (which did not disappear until 30 Mya (Sanmartin *et al.* 2001) and the still present Tethys Sea, although the latter separating southern Europe and northern Africa would have been rather narrow (Smith *et al.* 2004). Still another avenue from Asia to Africa may have been via India during the latter's northern migration (Briggs 1987). Clearly a wider geographical sampling of Oedipodinae needs to be assayed to help decide which passageway was more likely.

*Trimerotropis pallidipennis* is the only South American species examined (other museum specimens from that continent were refractory to DNA extraction methods). It branched off from a common ancestor shared with *Circotettix* about 14 Mya, well within the period in which passage between the two Americas would have been possible (Vickery 1989), but somewhat before the estimate of 3 Mya produced by Confalonieri *et al.* (1998) in their analysis of mitochondrial RFLP patterns.

Given the basal position of Old World taxa within clade D, it is



**Fig. 2.** A molecular chronogram for Oedipodinae evolution. Relationships are those obtained by maximum likelihood (see text); branch lengths are proportional to times of divergence estimated by r8s. Numbers on the nodes refer to times of divergence. Dashed branch indicates (ML) phylogenetic position is different using MP and MB. Calibration is based on time of divergence between Oedipodinae and Gomphocerinae, set at 100 Mya. Letters A to F refer to groups of taxa identified in text.

logical to conclude that the North American species, *Stethophyma gracile*, evolved from Eurasian ancestors, as hypothesized by Vickery (1989). According to our r8s calculations, this species split from its Eurasian counterpart, *S. grossum*, about 40 Mya. This value is approximately in the lower range of values determined previously (Contreras & Chapco 2006). The time of dispersal into North America still remains considerably earlier than the time-frame envisaged by Vickery (1989), namely before the last glaciation.

## Acknowledgments

We thank C. Amédégnato (CNRS, France), M. Cigliano (Museo de La Plata, Argentina), B. Çiplak (Akdeniz University, Turkey), T. McNary, D. Rentz (CSIRO, Australia), and C. Yong-Lin (Academia Sinica, China) for providing grasshopper specimens. We are especially grateful to J.-P. Camacho (University of Granada, Spain), and A. Foucart and M. LeCoq (both CIRAD, France) for their extensive help in collecting and identifying specimens. We also thank E. Chapco and D. Weissman for their helpful comments on the manuscript. This research was funded by a grant (W.C.) and a postgraduate scholarship (D.C.) from the Natural Sciences and Engineering Research Council of Canada.

## References

- Amédégnato C. 1993. African-American relationships in the Acridians (Insecta, Orthoptera), pp. 59-75. In: George W., Lavocat R. (Eds) The Africa-South America Connection. Clarendon Press, Oxford.
- Amédégnato C., Chapco W., Litzenger G. 2003. Out of South America? Additional evidence for a southern origin of melanopline grasshoppers. *Molecular Phylogenetics and Evolution* 29: 115-119.
- Askevold I. S. 1991. Classification, reconstructed phylogeny, and geographic history of the New World members of *Plateumaris* Thomson, 1959 (Coleoptera: Chrysomelidae: Donaciinae). *Memoirs of the Entomological Society of Canada* 157: 1-175.
- Barrio E., Ayala F.F. J. 1997. Evolution of the *Drosophila obscura* species group inferred from the Gpdh and Sod genes. *Molecular Phylogenetics and Evolution* 7: 79-93.
- Bei-Bienko G. Ya., Mishchenko L.L. 1964. Locusts and Grasshoppers of the USSR and Adjacent Countries. Part II. (Translated from Russian). Israel Program for Scientific Translations. Jerusalem, Israel.
- Bland R.G., Gangwere S.K. 1998. A new species of *Wernerella* Karny (Orthoptera: Acrididae: Oedipodinae) from the Canary Islands, Spain. *Journal of Orthoptera Research* 7: 23-28.
- Briggs J. C., 1987. Biogeography and Plate Tectonics. Elsevier. Oxford, UK.
- Buckley T.R., Arensburger P., Simon C., Chambers G.K. 2002. Combined data, Bayesian phylogenetics, and the origin of the New Zealand cicada genera. *Systematic Biology* 51: 4-18.
- Carbonell C. S. 1977. Origin, evolution, and distribution of the Neotropical acridomorph fauna (Orthoptera): a preliminary hypothesis. *Revista de la Sociedad Entomológica Argentina* 36: 153-175.
- Chapco W., Martel R.K.B., Kuperus W.R. 1997. Molecular phylogeny of North American band-winged grasshoppers (Orthoptera: Acrididae). *Annals of the Entomological Society of America* 90: 555-562.
- Chapco W., Litzenger G., Kuperus W.R. 2001. A molecular biogeographic analysis of the relationship between North American melanopline grasshoppers and their Eurasian and South American relatives. *Molecular Phylogenetics and Evolution* 18: 460-466.
- Confalonieri V.A., Sequeira A.S., Todaro L., Vilardi J.C. 1998. Mitochondrial DNA and phylogeography of the grasshopper *Trimerotropis pallidipennis* in relation to clinal distribution of chromosome polymorphisms. *Heredity* 81: 444-452.
- Contreras D., Chapco W. 2006. Molecular phylogenetic evidence for multiple dispersal events in gomphocerines grasshoppers. *Journal of Orthoptera Research* 15: 91-98.
- Cook L.G., Crisp M.D. 2005. Directional asymmetry of long-distance dispersal and colonization could mislead reconstructions of biogeography. *Journal of Biogeography* 32: 741-754.
- Costa M.A., Del Lama M.A., Melo G.A.R., Sheppard W.S. 2003. Molecular phylogeny of the stingless bees (Apidae, Apinae, Meliponini) inferred from mitochondrial 16S rDNA sequences. *Apidologie* 34: 73-84.
- Davis A.L.V., Scholtz C.H., Phillips T.K. 2002. Historical biogeography of scarabaeine dung beetles. *Journal of Biogeography* 29: 1217-1256.
- Dirsh V. M., 1965. The African Genera of Acridoidea. Cambridge University Press. Cambridge, UK.
- Eades D.C., Otte D., Naskrecki P. Orthoptera Species File Online. Version 2.8/2.8. [2007]. <http://osf2.orthoptera.org>.
- Farris J. S. 1969. A successive approximations approach to character weighting. *Systematic Zoology* 18: 374-385.
- Flook P.K., Rowell C.H.F., 1997. The phylogeny of the Caelifera (Insecta, Orthoptera) as deduced from mtrRNA gene sequences. *Molecular Phylogenetics and Evolution* 8: 89-103.
- Fuller S., Schwarz M., Tierney S. 2005. Phylogenetics of the allodapine bee genus *Braunsapis*: historical biogeography and long-range dispersal over water. *Journal of Biogeography* 32: 2135-2144.
- Fuzeau-Braesch S., Chapco W. 1977. Comparaison sur la biologie de trois espèces de Criquets du genre *Aiolopus*: *simulator* (Walkre, 1870), *thalassinus* (Fabricius, 1781), *strepens* (Latreille, 1804).
- Gaunt M.W., Miles M.A. 2002. An insect molecular clock dates the origin of the insects and accords with palaeontological and biogeographic landmarks. *Molecular Biology and Evolution* 19: 748-761.
- Guliaeva O.N., Vysotskaya L.V., Sergeev M.G. 2005. Taxonomic and phylogenetic relationships of the Holarctic grasshoppers (Orthoptera: Acrididae): a new view on old problems. *Eurasian Entomological Journal* 4: 87-94.
- Hallam A., 1994. An Outline of Phanerozoic Biogeography. Oxford University Press. Oxford, UK.
- Helwig E.R. 1942. Unusual integrations of the chromatin in *Machaerocera* and other genera of the Acrididae (Orthoptera). *Journal of Morphology* 71: 1-33.
- Huelsenbeck J.P., Ronquist F. 2001. MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics* 17: 754-755.
- Hundsdoerfer A. K., Kitching I. J., Wink M. 2005. A molecular phylogeny of the hawkmoth genus *Hyles* (Lepidoptera: Sphingidae, Macroglossinae). *Molecular Phylogenetics and Evolution* 35: 442-458.
- Jolivet L., Faccenna C., 2000. Mediterranean extension and the Africa-Eurasia collision. *Tectonics* 19: 1095-1106.
- Jönsson K.A., Fjeldså J. 2006. Determining biogeographical patterns of dispersal and diversification in oscine passerine birds in Australia, Southeast Asia and Africa. *Journal of Biogeography* 33: 1155-1165.
- Kawakita A., Sota T., Ito M., Ascher J. S., Tanaka H., Kato M., Roubik D.W. 2004. Phylogeny, historical biogeography, and character evolution in bumble bees (*Bombus*: Apidae) based on simultaneous analysis of three nuclear gene sequences. *Molecular Phylogenetics and Evolution* 31: 799-804.
- Key K.H.L. 1954. The Taxonomy, Phases, and Distribution of The Genera *Chortoicetes* Brunn. and *Austroicetes* Uv. (Orthoptera: Acrididae). Commonwealth Scientific and Industrial Research Organization. Canberra, Australia.
- Key K.H.L. 1959. The ecology and biogeography of Australian grasshoppers and locusts. *Monographiae Biologicae* 8: 92-210.
- Leys R., Cooper S.J.B., Schwarz M.P. 2002. Molecular phylogeny and historical biogeography of the large carpenter bees, genus *Xylocopa* (Hymenoptera: Apidae). *Biological Journal of the Linnean Society* 77: 240-266.
- Litzenger G., Chapco W. 2001a. Molecular phylogeny of selected Eurasian podismine grasshoppers (Orthoptera: Acrididae). *Annals of the Entomological Society of America* 94: 505-551.
- Litzenger G., Chapco W. 2001b. A molecular phylogeographic perspective on a fifty-year-old taxonomic issue in grasshopper systematics. *Heredity* 86: 54-59.



- Lovejoy N.R., Mullen S.P., Sword G.A., Chapman R.F., Harrison R.G. 2005. Ancient trans-Atlantic flight explains locust biogeography: molecular phylogenetics of *Schistocerca*. *Proceedings of the Royal Society B* 273: 767-774.
- Lu H.-M., Huang Y. 2006. Phylogenetic relationship of 16 Oedipodinae species (Insects: Orthoptera) based on the 16S rRNA gene sequences. *Insect Science* 13: 103-108.
- Maddison W.P., Maddison D.R. 2004. *MacClade: Analysis of Phylogeny and Character Evolution*. Version 4.05. Sinauer, Sunderland, MA.
- Martin J., Guryev V., Blinov A., Edward D.H.D. 2002. A molecular assessment of the extent of variation and dispersal between Australian populations of the genus *Archaeochlus* Brundin (Diptera: Chironomidae). *Invertebrate Systematics* 16: 599-603.
- Otte D. 1984. *The North American Grasshoppers. Volume II. Acrididae. Oedipodinae*. Harvard University Press. Cambridge, Massachusetts.
- Otte D. 1995. *Orthoptera Species File 4*. Orthopterists' Society and Academy of Natural Sciences of Philadelphia. Philadelphia, PA.
- Page R.D.M., Holmes E.C. 1998. *Molecular Evolution. A Phylogenetic Approach*. Blackwell, Oxford.
- Pearson D.L., Vogler A.P. 2001. *Tiger Beetles*. Cornell University Press. Ithaca, NY.
- Petit D., Picaud F., Elghadraoui L. 2006. Géométrie morphologique des ailes des Acrididae (Orthoptera: Caelifera): sexe, stridulation, caractère. *Annales de la Société Entomologique de France (Nouvelle Série)* 42: 63-73.
- Phillips A. J., Simon C. 1995. Simple, efficient, and non-destructive DNA extraction protocol for arthropods. *Annals of the Entomological Society of America* 88: 281-283.
- Posada D., Crandall K.A. 1998. Modeltest: testing the model of DNA substitution. *Bioinformatics* 14: 817-818.
- Prüser F., Mossakowski D. 1998. Conflicts in phylogenetic relationships and dispersal history of the supertribe Carabitaie (Coleoptera: Carabidae), pp. 297-328. In: Ball G. E., Casale A., Taglianti V. (Eds) *Phylogeny and Classification of Caraboidea* (Coleoptera: Adephaga). Museo Regionale de Scienze Naturali, Torino, Italy.
- Raven P.H., Axelrod D.I. 1972. Plate tectonics and Australasian palaeobiogeography. *Science* 176: 1379-1386.
- Rehn J.A.G. 1958. The origin and affinities of the Dermaptera and Orthoptera of western North America, pp. 253-298. In: Hubbs C.L. (Ed.) *Zoogeography* 12. Horn-Shafer, Baltimore.
- Rentz D.C.F. 1996. *Grasshopper Country: The Abundant Orthopteroid Insects of Australia*. University of New South Wales Press. Sydney, Australia.
- Rentz D.C.F., Lewis R.C., Su Y.N., Upton M.S. 2004. *A Guide to Australian Grasshoppers and Locusts*. Natural History Publications, Borneo.
- Ritchie J.M. 1981. A taxonomic revision of the genus *Oedaleus* Fieber (Orthoptera: Acrididae). *Bulletin of the British Museum of Natural History (Entomology)* 42: 83-183.
- Ritchie J.M. 1982. A taxonomic revision of the genus *Gastrimargus* Saussure (Orthoptera: Acrididae). *Bulletin of the British Museum of Natural History (Entomology)* 44: 239-329.
- Ronquist F. 1996. *DIVA*, Version 1.1 Computer program and manual available by anonymous FTP from Uppsala University at ftp.uu.se or ftp.sysbot.uu.se
- Ronquist F. 1997. Dispersal-vicariance analysis: a new approach to the quantification of historical biogeography. *Systematic Biology* 46: 195-203.
- Rosenberg J., Burt P.J.A. 1999. Windborne displacements of desert locusts from Africa to the Caribbean and South America. *Aerobiologia* 15: 167-175.
- Rowell C.H.F. 2005. A new Ugandan species of *Pterotiltus* (Orthoptera, Acrididae, Oxyinae) with epiphyllid oviposition. *Journal of Orthoptera Research* 14: 33-44.
- Rowell C.H.F., Flook P.K. 2004. A dated molecular phylogeny of the Proctolabinae (Orthoptera: Acrididae), especially the Lithoscirtae, and the evolution of their adaptive traits and present biogeography. *Journal of Orthoptera Research* 13: 35-56.
- Sanderson M.J. 2002. Estimating absolute rates of molecular evolution and divergence times: a penalized likelihood approach. *Molecular Biology and Evolution* 19: 101-109.
- Sanderson M.J. 2004. *r8s*. Published privately, Davis. Available from: <http://ginger.ucdavis.edu/r8s/>
- Sanmartin I., Enghoff H., Ronquist F. 2001. Patterns of animal dispersal, vicariance and diversification in the Holarctic. *Biological Journal of the Linnean Society* 73: 345-390.
- Smith A.G., Smith D.G., Funnell B.M. 2004. *Atlas of Mesozoic and Cenozoic Coastlines*. Cambridge University Press. Cambridge, UK.
- Song H. 2004. Revision of the *Alutacea* group of genus *Schistocerca* (Orthoptera: Acrididae: Cyrtacathacridinae). *Annals of the Entomological Society of America* 97: 420-436.
- Stidham T.A., Stidham J.A. 2000. A new Miocene band-winged grasshopper (Orthoptera: Acrididae) from Nevada. *Annals of the Entomological Society of America* 93: 405-407.
- Storozhenko S., Otte D. 1994. Review of the genus *Stethophyma* Fischer (Orthoptera: Acrididae: Acridinae: Parapleurini). *Journal of Orthoptera Research* 2: 61-64.
- Swofford D.L. 2003. *PAUP\*: Phylogenetic Analysis Using Parsimony (\* and Other Methods)*. Version 4.0b10. Sinauer, Sunderland, MA.
- Tiffney B.H. 1985. The Eocene north Atlantic bridge: its importance in Tertiary and modern phytogeography of the northern hemisphere. *Journal of the Arnold Arboretum* 66: 243-273.
- Uvarov B. 1977. *Grasshoppers and Locusts. Volume II. Centre Overseas Pest Research*. London.
- Vickery V.R. 1987. The northern Nearctic Orthoptera: their origins and survival. In: Baccetti B.C. (Ed.) *Evolutionary Biology of Orthopteroid Insects*. Ellis Horwood Limited, West Sussex, UK.
- Vickery V.R. 1989. The biogeography of Canadian Grylloptera and Orthoptera. *Canadian Entomologist* 121: 389-424.
- Vickery V.R., Kevan D.K. McE. 1985. The grasshoppers, crickets and related insects of Canada and adjacent regions. *Ulonata: Dermaptera, Cheleutoptera, Notoptera, Dictuoptera, Grylloptera and Orthoptera, The Insects and Arachnids of Canada. Part 14*. Research Branch, Agriculture Canada Publications, 1777. Ottawa, ON.
- von Dohlen C.D., Rowe C.A., Heie O.E. 2006. A test of morphological hypotheses for tribal and subtribal relationships of Aphidinae (Insecta: Hemiptera: Aphididae) using DNA sequences. *Molecular Phylogenetics and Evolution* 38: 316-329.
- Weissman D.B. 1984. Notes on the autecology, cytology, morphology, and crepitation of *Trimerotropis* grasshoppers (Orthoptera: Oedipodinae). *Pan-Pacific Entomologist* 60: 269-278.
- Weissman D.B., Rentz D.C.F. 1980. Cytological, morphological, and crepitation characteristics of the trimerotropine (*Aerchoreutes*, *Circotettix*, and *Trimerotropis*) grasshoppers (Orthoptera: Oedipodinae). *Transactions of the American Entomological Society* 106: 253-272.
- White M.J.D. 1973. *Animal Cytology and Evolution*. Third Edition. Cambridge University Press. New York, NY.
- Zakharov E.V., Caterino M.S., Sperling F.A.H. 2004. Molecular phylogeny, historical biogeography, and divergence time estimates for swallowtail butterflies of the genus *Papilio* (Lepidoptera: Papilionidae). *Systematic Biology* 53: 193-215.