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Source: Journal of Insect Science, 14(6) : 1-8

Published By: Entomological Society of America

URL: <https://doi.org/10.1673/031.014.06>

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***Wolbachia* infection and Lepidoptera of conservation concern**

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Abstract

Conservation of at-risk species requires multi-faceted and carefully-considered management approaches to be successful. For arthropods, the presence of endosymbiotic bacteria, such as *Wolbachia* (Rickettsiales: Rickettsiaceae), may complicate management plans and exacerbate the challenges faced by conservation managers. *Wolbachia* poses a substantial and underappreciated threat to the conservation of arthropods because infection may induce a number of phenotypic effects, most of which are considered deleterious to the host population. In this study, the prevalence of *Wolbachia* infection in lepidopteran species of conservation concern was examined. Using standard molecular techniques, 22 species of Lepidoptera were screened, of which 19 were infected with *Wolbachia*. This rate is comparable to that observed in insects as a whole. However, this is likely an underestimate because geographic sampling was not extensive and may not have included infected segments of the species' ranges. *Wolbachia* infections may be particularly problematic for conservation management plans that incorporate captive propagation or translocation. Inadvertent introduction of *Wolbachia* into uninfected populations or introduction of a new strain may put these populations at greater risk for extinction. Further sampling to investigate the geographic extent of *Wolbachia* infections within species of conservation concern and experiments designed to determine the nature of the infection phenotype(s) are necessary to manage the potential threat of infection.

Keywords: captive rearing, endangered species, endosymbiont, translocation

Abbreviations: CI, cytoplasmic incompatibility; fbpA, fructose-bisphosphate aldolase

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Editor: Patricia S. Stock was editor of this paper.

Received: 29 October 2012 **Accepted:** 31 July 2013 **Published:** 14 January 2014

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ISSN: 1536-2442 | Vol. 14, Number 6

Cite this paper as:

Hamm CA, Handley CA, Pike A, Forister ML, Fordyce JA, Nice CC. 2014. *Wolbachia* infection and Lepidoptera of conservation concern. *Journal of Insect Science* 14:6. Available online: <http://www.insectscience.org/14.6>

Introduction

Conservation managers often adopt active management strategies when confronted by complex and interconnected problems. Captive propagation and translocation programs are increasingly being incorporated into management plans to augment endangered populations or repopulate formerly occupied habitats (Crone et al. 2007). These programs present their own challenges and must be carefully designed to minimize the possibility of disease transmission and maintain genetic diversity (Snyder et al. 1996; Van Oosterhout et al. 2007). When working with arthropods of conservation concern, an additional and under-appreciated challenge arises in the form of endosymbiotic bacteria that may manipulate the reproductive biology of their hosts (Werren et al. 2008; Nice et al. 2009).

The α -proteobacteria *Wolbachia* (Rickettsiales: Rickettsiaceae) is a maternally inherited endosymbiont of many arthropods and is estimated to occur in up to 66% of all insects (Hilgenboecker et al. 2008). *Wolbachia* has been the focus of intense research efforts due to its potentially significant effects on the reproduction of its host (Werren et al. 2008; Nice et al. 2009). In short, *Wolbachia* manipulates its host's reproduction to facilitate its own by inducing one of four phenotypes: (1) feminization occurs when a *Wolbachia* infection transforms genetically male embryos into fully functional females, leading to production of progeny that are all functionally female; (2) male-killing strains eliminate all male embryos so only female progeny are produced; (3) parthenogenesis-induction occurs in species with haplo-diploid sex determination in which infected females do not need to mate and therefore produce only female progeny from unfertilized eggs; (4) cytoplasmic incompatibility (CI) prevents in-

fecting males from reproducing with uninfected females or females infected with a different strain of *Wolbachia*, and this is perhaps the most common phenotype (Werren et al. 2008). Should CI-infected individuals be introduced to an uninfected population, the consequences may be dire, especially for small populations (Nice et al. 2009). These CI infections may induce a population bottleneck as its frequency spreads; if the population is not large enough to pass through this bottleneck, it may be extirpated. *Wolbachia* infection can be difficult to detect by demographic observations, and laboratory experiments to determine infection phenotype can be costly and time consuming (Weeks et al. 2007). The most common method for detecting *Wolbachia* infection utilizes the cost effective PCR, and strains may be identified using standardized protocols (Baldo et al. 2006).

Within the insects, Lepidoptera are disproportionately represented among the threatened and endangered species in the United States (28 of 62 species; U.S. Fish & Wildlife Service). Lepidoptera have also been the subject of captive rearing programs and have been discussed as candidates for translocation (Tolson 2008; Richardson et al. 2009; Landis et al. 2011). While the threat of *Wolbachia* to Lepidoptera of conservation concern has been demonstrated theoretically (Nice et al. 2009), it remains to be seen if *Wolbachia* infections are present in Lepidoptera of conservation concern. Therefore, 22 lepidopteran species from the United States, including federal and state listed threatened and endangered taxa, were surveyed for *Wolbachia* to document the presence of the bacterium and draw attention to the role that infection may play in recovery efforts.

Materials and Methods

A total of 296 individuals from 22 at-risk lepidopteran species were sampled (Table 1). The very nature of threatened and endangered species prohibits the use of large and random samples, thus these taxa were utilized because the material was either at immediate disposal or was kindly donated by colleagues. Genomic DNA (gDNA) was extracted from all individuals following standard methods (Brookes et al. 1997). Screening for *Wolbachia* in gDNA samples required two PCRs and followed the methods used by Nice et al. (2009). Primers for the ribosomal rDNA gene 16S (WSpecF and WSpecR) were used to detect *Wolbachia* and were run in concert with arthropod-specific 28S rDNA primers (28sF3633 and 28sR4076) to act as a positive control for each reaction. Standard positive and negative controls were run simultaneously during the *Wolbachia* screens. PCR products were visualized on a 1% agarose gel and scored for the presence of *Wolbachia*.

While the use of *Wolbachia*-specific PCR remains the standard method for the detection of *Wolbachia* (Baldo et al. 2006), certain primers may be prone to cross amplification of other bacteria (Simões et al. 2011). To minimize the likelihood of false positives, additional primers were used to amplify fructose-bisphosphate aldolase (*fbpA*) for a subset of 16 individuals that tested positive for 16S (Baldo et al. 2006) (Table 1); *fbpA* amplification followed the PCR protocols outlined in Simões et al. (2011). This combination of molecular markers, 16S, and *fbpA* should only amplify *Wolbachia* and no other bacterial endosymbiont (Simões et al. 2011). A subset of individuals that tested positive for *Wolbachia* infections were sequenced in both directions for 16S and *fbpA* at the Research Technology Support Facility at Michigan State University

using an ABI Prism 3730 Genetic Analyzer (Applied Biosystems, Life Technologies, www.lifetechnologies.com). The resulting sequences were combined into contigs using Geneious version 5.4 (Biomatters, www.biomatters.com) and submitted for BLAST search to confirm the amplification of *Wolbachia* sequences (Altschul et al 1990).

To further minimize the chances of false positives for individuals that were not sequenced, a restriction digest was conducted. It was possible to predict the size of DNA fragments generated by certain restriction enzymes using the genetic sequences previously produced. The 4-cutter *MseI* (New England Biolabs, www.neb.com) should have two restriction sites in 16S, both at the ends of the sequence, and generate one band of approximately 300 bp and two bands of 100 bp. The gene *fbpA* should contain two restriction sites for *MseI*, both located at the ends of the sequence, and would generate one band of approximately 350 bp and two bands of 75bp. Restriction digests were carried out under manufacturer's specifications and the resulting products were visualized on a 2% agarose gel to confirm that the actual band sizes matched predictions.

Results

19 of the 22 species screened had at least one individual score positive for *Wolbachia* infection, and a subset of individuals positive for 16S were also positive for *fbpA*. Individuals from six species (*Neonympha mitchellii* French (Nymphalidae), *Speyeria idalia* (Drury), *S. diana* (Cramer), *Eupheys dukesii* (Lindsey) (Hesperiidae), *Hesperia ottoe* Edwards, and *Papaipema sciata* Bird (Noctuidae)) were sequenced for 16S and *fbpA*, resulting in reads of ~400 bp in length. All sequences queried with a BLAST search returned hits to the appropriate *Wolbachia* iso-

Table 1. Results of a survey of *Wolbachia* infections in 244 lepidopteran individuals, representing 22 species of conservation concern. The county of collection was not available for some individuals. The total number screened is presented with the number testing positive for *Wolbachia* presence, which is given parentheses. All samples that were positive for 16S also tested positive for *fbpA*.

Family	Species	Vernacular Name	Locality	# Screened for 16S (# Present)	Conservation Status ^a (States)
LYCAENIDAE	<i>Callophrys (Mitoura) muiri</i>	Muir's Hairstreak	California	8(0)	SC (CA), C (Federal)
	<i>Callophrys (Incisalia) irus</i>	Frosted Elfin	Allegan Co., MI	1(1)	E (DE, MD, NH, OH); T (CT, MI, NJ, NY, WI); SC (MA, RI)
	<i>Callophrys (Mitoura) thornei</i>	Thorne's Juniper Hairstreak	San Diego Co., CA	5(0)	SC (Federal), E (CA)
	<i>Euphilotes pallescens arenamontana</i>	Sand Mountain blue	Sand Mountain, Churchill Co., NV	12(3)	C (Federal)
	<i>Plebejus (Lycaeides) idas nabokovi</i>	Northern Blue	Schoolcraft Co., MI	2(1)	T (MI), SC (MN)
	<i>Plebejus saepiolus</i>	White Mountains Saepiolus Blue Butterfly	White Mountains, Mono Co., CA	10(2)	SC (Federal), E (CA)
NYMPHALIDAE	<i>Speyeria diana</i>	Diana Fritillary	North Carolina	1(1)	SC (NC)
	<i>Speyeria idalia</i>	Regal Fritillary	Buena Vista prairie, WI	1(1)*	E (MI, NY, OH, WI); T (IL); SC (IA, MN, MO, OK, PA)
	<i>Speyeria idalia</i>	Regal Fritillary	Hog Back prairie, WI	1(1)	E (MI, NY, OH, WI); T (IL); SC (IA, MN, MO, OK, PA)
	<i>Speyeria idalia</i>	Regal Fritillary	Thompson Prairie, WI	1(1)	E (MI, NY, OH, WI); T (IL); SC (IA, MN, MO, OK, PA)
	<i>Phyciodes batesii</i>	Tawny Crescent	Alger Co., MI	1(0)	SC (MI)
	<i>Neonympha mitchellii francisi</i>	Saint Francis satyr	Fort Bragg, NC	4(4)	E (Federal)
	<i>Neonympha mitchellii mitchellii</i>	Mitchell's satyr	Jackson County Central, MI	180(20)*	E (Federal, MI); T (VA)
HESPERIIDAE	<i>Pyrgus ruralis lagunae</i>	Laguna Mountain Skipper	San Diego Co., CA	7(1)	E (Federal)
	<i>Hesperia ottoe</i>	Ottoe Skipper	Allegan Co., MI	1(1)*	T (IL, MI, MN); SC (IA, MT)
	<i>Polites sabuleti albomontana</i>	White Mountains Sandhill Skipper	White Mountains, Mono Co., CA	1(1)	C (Federal), E (CA)
	<i>Polites mardon</i>	Mardon skipper	Jackson Co., OR	1(1)	C (Federal), E (CA)
	<i>Euphyes dukesi</i>	Duke's Skipper	Wayne Co., MI	1(1)*	T (MI)
	<i>Atrytonopsis hianna</i>	Dusted Skipper	Newaygo Co., MI	1(1)	SC (MI, WI)
	<i>Atrytonopsis n. sp.1</i>	Crystal Skipper	Fort Macon, NC	5(5)	SC (Federal)
NOCTUIDAE	<i>Papaipema beeriana</i>	Blazing Star Borer	Washtenaw Co., MI	1(1)	SC (MI)
	<i>Papaipema sciata</i>	Culver's Root Borer	Jackson Co., MI	1(1)*	SC (MI)
	<i>Papaipema silphii</i>	Silphium Borer Moth	Washtenaw Co., MI	1(1)	T (MI)
	<i>Schinia lucens</i>	Leadplant Flower Moth	Ransom Co., ND	1(1)	E (MI)

^aCorresponding codes: T = threatened; E = endangered; SC = species of special concern; C = candidate for protection; * = sequenced

late with 100% pairwise identity (E value = 0) and all sequences were associated with *Wolbachia* strains from supergroup 2 (GenBank accession numbers KJ125429–KJ125436). Samples subjected to *MseI* restriction digest produced bands that matched the length predicted by restriction site mapping to known sequences for 16S and *fbpA*.

Discussion

The presence of *Wolbachia* was documented in 19 lepidopteran species of conservation concern, one of which (*N. m. mitchellii*) is the focus of captive rearing efforts (Tolson 2008). Multiple markers were utilized, a subset of individuals was sequenced, and restriction digests were conducted on the remaining individuals to minimize the possibility of false

positives. However, caution is urged when interpreting these results because a small sample size was used for many taxa and the data were not randomly sampled (though the conservation status of these taxa precludes random sampling). Thus, while these species tested positive for infection, the limited sampling reported here cannot be extended to the entire range of species. Furthermore, geographically comprehensive sampling would be required for any management plans for infected species.

Accurately determining the presence of infection and its induced phenotype requires the use of multiple methods. While necessary to detect the presence of infection, molecular genetic tools alone cannot identify the induced phenotype, because induced phenotypes are

not monophyletic and the same strain might induce different phenotypes in different species (Zhou et al. 1998; Baldo et al. 2006). Molecular methods must be used in conjunction with demographic observations to determine the effects of *Wolbachia*. Sequencing alone cannot allay concerns regarding the effects of *Wolbachia*. For example, *Wolbachia* from the *Culex pipiens* / *quinquefasciatus* complex are identical at multiple loci, yet crosses between some strains result in cytoplasmic incompatibility (Duron et al. 2006; Atyame et al. 2011). Indeed, even utilizing the MLST protocol, in which five variable *Wolbachia* loci are sequenced, may or may not distinguish strains (Baldo et al. 2006). The *Wolbachia* from the invasive vinegar fly *Drosophila suzukii* is identical to *wRi* (the strain found in *Drosophila simulans*) at all five MLST loci, and yet had polymorphisms at the *dnaA* locus and numerous inversions; these differences were only observed after genomic sequencing was conducted (Siozios et al. 2013). Confirming infection by a CI strain requires experimental crosses between infected and uninfected individuals. In addition to *Wolbachia*, there are a number of other reproductive manipulators associated with insects, such as *Cardinium*, *Rickettsia*, and *Spiroplasma*, that should also be screened for and considered (Duron et al. 2008).

Assuming deleterious effects of an infection, there are two additional concerns when managing *Wolbachia*: (1) the possibility that the induced phenotype is suppressed (Hornett et al. 2006), and (2) the possibility that there are multiple strains occurring sympatrically in the same population (Hiroki et al. 2004). In the case of suppressed phenotypic effects, some populations may appear uninfected because they have been able to ameliorate the phenotypic effects of infection, though they remain infected. These populations have been able to

suppress the infection phenotype, and while *Wolbachia* are still detectable using molecular methods, the infection phenotype is absent (Hornett et al. 2006). Transmission of *Wolbachia* from a suppressing population to an uninfected population may result in the expression of the induced phenotype and all of the subsequent consequences. While suppression might allow populations to escape the consequences of infection, the concealed presence of *Wolbachia* increases the likelihood of inadvertently introducing an infection through captive management or translocation. In the case of infection by multiple strains of *Wolbachia* (Reuter and Keller 2003; Hiroki et al. 2004), each strain might induce a different phenotype.

For these reasons, management programs should screen a representative subset of individuals propagated in captivity to verify if infection is present. This study may serve as a foundation for examining other at-risk arthropods, many of which have captive propagation programs. Screening of these species is necessary to determine the extent of *Wolbachia* infection within populations and across species. Future studies should also seek to determine the nature of the *Wolbachia*-induced phenotypes in these species. Information on the prevalence, geographic extent, and phenotypic effects of *Wolbachia* might prove critical for effective management of threatened and endangered arthropods.

Acknowledgements

The authors wish to thank David Cuthrell from the Michigan Natural Features Inventory, Nick Haddad from North Carolina State University, and Travis Longcore from the Urban Wildlands Group and Jana Johnson and Moorpark College, for providing specimens. We would also like to thank

Christopher Hoving from the Michigan Department of Natural Resources and Environment, Tameka Dandridge from the United States Fish and Wildlife Service, and Barry Williams from Michigan State University for providing their expertise and comments on this project. This research was supported in part by a fellowship from the Scriber Scholars in Butterfly Conservation, a Plant Sciences Research Fellowship at Michigan State University, and a USFWS Preventing Extinction grant supporting C. A. Hamm, and funding from U.S. Fish and Wildlife Service, Carlsbad Field Office. The Texas State Research Enhancement Program provided additional support. This paper was generated from the symposium “Conservation of Lepidoptera” at the 2009 Annual Meeting of the Entomological Society of America. We wish to thank Clinton D. Pilcher, Carol L. Pilcher, and Marlin Rice of the Entomological Society of America for facilitating this symposium. The authors wish to thank Drew Davis for his helpful comments.

References

- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *Journal of Molecular Biology* 215: 403–410.
- Atyame C, Delsuc F, Pasteur N, Weil M, Duron O. 2011. *Molecular Biology and Evolution* 28: 2761–2772.
- Baldo L, Desjardins CA, Russell JA, Stahlhut JK, Werren JH. 2010. Accelerated microevolution in an outer membrane protein (OMP) of the intracellular bacteria *Wolbachia*. *BMC Evolutionary Biology* 10: 48.
- Baldo L, Hotopp JCD, Jolley KA, Bordenstein SR, Biber SA, Choudhury RR, Hayashi C, Maiden MCJ, Tettelin H, Werren JH. 2006. Multilocus sequence typing system for the endosymbiont *Wolbachia pipientis*. *Applied and Environmental Microbiology* 72: 7098–7110.
- Brookes M, Graneau Y., King P., Rose O. Thomas C and Mallet J.B. 1997. Genetic analysis of founder bottlenecks in the rare British butterfly *Plebejus argus*. *Conservation Biology* 11: 648–661.
- Brownlie JC, Cass BN, Riegler M, Witsenburg JJ, Iturbe-Ormaetxe I, McGraw EA, O'Neill SL. 2009. Evidence for metabolic provisioning by a common invertebrate endosymbiont, *Wolbachia pipientis*, during periods of nutritional stress. *PLOS Pathogens* 5: e1000368.
- Crone E, Pickering D, Schultz C. 2007. Can captive rearing promote recovery of endangered butterflies? An assessment in the face of uncertainty. *Biological Conservation* 139: 103–112.
- Duron O, Bernard C, Unal S, Berthomieu A, Berticat C, Weill M. 2006. Tracking factors modulating cytoplasmic incompatibilities in the mosquito *Culex pipiens*. *Molecular Ecology* 15: 2061–3071.
- Duron O, Bouchon D, Boutin S, Bellamy L, Zhou L, Engelstädter J, Hurst GD. 2008. The diversity of reproductive parasites among arthropods: *Wolbachia* do not walk alone. *BMC Biology* 6:27.
- Hilgenboecker K, Hammerstein P, Schlattmann P, Telschow A, Werren JH. 2008. How many species are infected with *Wolbachia*?--A statistical analysis of current

data. *FEMS Microbiology Letters* 281: 215–220.

Hiroki M, Tagami Y, Miura K, Kato Y. 2004. Multiple infection with *Wolbachia* inducing different reproductive manipulations in the butterfly *Eurema hecabe*. *Proceedings Of The Royal Society B-Biological Sciences* 271: 1751–1755.

Hornett EA, Charlat S, Duploux AMR, Davies N, Roderick GK, Wedell N, Hurst GDD. 2006. Evolution of male-killer suppression in a natural population. *PLOS Biology* 4: e283.

Landis DA, Fiedler AK, Hamm CA, Cuthrell DL, Schools EH, Pearsall DR, Herbert ME, Doran PJ. 2011. Insect conservation in Michigan prairie fen: addressing the challenge of global change. *Journal of Insect Conservation* 16: 131–142.

Nice CC, Gompert Z, Forister ML, Fordyce JA. 2009. An unseen foe in arthropod conservation efforts: The case of *Wolbachia* infections in the Karner blue butterfly. *Biological Conservation* 142: 3137–3146.

Reuter M, Keller L. 2003. High levels of multiple *Wolbachia* infection and recombination in the ant *Formica exsecta*. *Molecular Biology And Evolution* 20: 748–753.

Richardson DM, Hellmann JJ, McLachlan JS, Sax DF, Schwartz MW, Gonzalez P, Brennan EJ, Camacho A, Root TL, Sala OE, Schneider SH, Ashe DM, Clark JR, Early R, Etterson JR, Fielder ED, Gill JL, Minter BA, Polasky S, Safford HD, Thompson AR, Vellend M. 2009. Multidimensional evaluation of managed relocation. *Proceedings Of The National Academy Of Sciences USA* 106: 9721–9724.

Siozios S, Cestaro A, Kaur R, Pertot I, Rota-Stabelli O, Anfora G. 2013. Draft genome of the *Wolbachia* endosymbiont of *Drosophila suzukii*. *Genome Announcements* 1: e00032-13.

Simões P, Mialdea G, Reiss D, Sagot MF, Charlat S. 2011. *Wolbachia* detection: an assessment of standard PCR Protocols. *Molecular Ecology Resources* 11: 567–572.

Snyder NFR, Derrickson SR, Beissinger SR, Wiley JW, Smith TB, Toone WD, Miller B. 1996. Limitations of Captive Breeding in Endangered Species Recovery. *Conservation Biology* 10: 338–348.

Stahlhut JK, Desjardins CA, Clark ME, Baldo L, Russell JA, Werren JH, Jaenike J. 2010. The mushroom habitat as an ecological arena for global exchange of *Wolbachia*. *Molecular Ecology* 19: 1940–1952.

Tolson P. 2008. Rearing Mitchell's satyr at the Toldeo Zoo - a first step towards eventual re-introduction in secure habitats. *News of the Lepidopterists' Society* 50: 42–43.

U.S. Fish & Wildlife Service. *Environmental Conservation Online System*. Available online: http://ecos.fws.gov/tess_public/

Van Oosterhout C, Smith AM, Hänfling B, Ramnarine IW, Mohammed RS, Cable J. 2007. The guppy as a conservation model: implications of parasitism and inbreeding for reintroduction success. *Conservation Biology* 21: 1573–1583.

Weeks AR, Turelli M, Harcombe WR, Reynolds KT, Hoffmann AA. 2007. From parasite to mutualist: rapid evolution of *Wolbachia* in natural populations of *Drosophila*. *PLOS Biology* 5: e114.

Werren JH, Baldo L, Clark ME. 2008.
Wolbachia: master manipulators of
invertebrate biology. *Nature Reviews
Microbiology* 6: 741–751.

Zhou W, Rousset F, O'Neill S. 1998.
Phylogeny and PCR-based classification of
Wolbachia strains using wsp gene sequences.
*Proceedings Of The Royal Society Of London
Series B-Biological Sciences* 265: 509–515.