

First Report of Hecatera dysodea (Denis and Schiffermüller) (Noctuidae) in the Pacific Northwest of the United States

Authors: Landolt, Peter J., Worth, Richard A., and Zack, Richard S.

Source: The Journal of the Lepidopterists' Society, 64(4): 192-196

Published By: The Lepidopterists' Society

URL: https://doi.org/10.18473/lepi.v64i4.a3

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Journal of the Lepidopterists' Society 64(4), 2010, 192–196

FIRST REPORT OF HECATERA DYSODEA (DENIS AND SCHIFFERMÜLLER) (NOCTUIDAE) IN THE PACIFIC NORTHWEST OF THE UNITED STATES

PETER J. LANDOLT

USDA, ARS Yakima Agricultural Research Laboratory, 5230 Konnowac Pass Road, Wapato, WA 98951; email: peter.landolt@ars.usda.gov

RICHARD A. WORTH

Oregon Department of Agriculture, 635 Capitol St. NE, Salem, OR 97301

AND

RICHARD S. ZACK

Department of Entomology, Washington State University, Pullman, WA 99164

ABSTRACT. New geographic records are reported for the noctuid moth *Hecatera dysodea* (Denis & Schiffermüller). It is a Palearctic species, but is now found in a broadly contiguous area of Oregon and Washington in the United States. This area is comprised of 7 counties across much of the north of Oregon and into 4 counties of southern Washington. Moths were captured in several types of survey traps baited with insect pheromones and feeding attractants, as well as blacklight traps, from 2003 to 2009. Larvae were collected on flower stalks of prickly lettuce, *Lactuca serriola* L. (Asteraceae), from June into September, suggesting multivoltinism. Collection records over time indicate a possibly rapidly expanding distribution of the species.

Additional key words: host plant, introduction, moth, trap, Oregon, Washington

Hecatera dysodea (Denis & Schiffermüller) has a widespread distribution in Europe and North Africa, east into Russia, western China and India (Wiltshire 1957; Poole 1989; Ivinskis & Miatleuski 1999; Hacker et al. 2002). The species was thought to have become extinct in England but has since reappeared (Skinner 1984; Pratt 1986; Agassiz & Spice 1998; Honey 2002), and may be expanding its distribution in other areas of Europe such as Denmark and Sweden (http://www.lepidoptera.dk/dysodea.htm (Hacker et al. 2002). Reasons for its prior decline in England are discussed by Pratt (1986).

The larvae feed on the flower buds, flowers, and seeds of species of *Lactuca* (lettuce), *Crepis* (hawksbeard) and *Sonchus* (sow thistle) (Asteraceae) (South 1920; Bretherton *et al.* 1979; Skinner 1984; Pratt 1986). Bretherton *et al.* (1979) report it as an occasional pest on domestic lettuce, *Lactuca sativa* L. However, Pratt (1986) logically suggests that such claims are in reference only to lettuce grown for seed production.

The larva is described by Beck (1999); the adult is illustrated by South (1920), Skinner (1984), and Hacker et al. (2002). The adult and male genitalia are also figured in this report (Fig. 1 & 2). We summarize first collections of *H. dysodea* in North America (e.g. Hodges et al. 1983), within Oregon and Washington of the United States.

METHODS AND RESULTS

Oregon. Collection information is summarized in Table 1 and Figure 3. First Oregon records for the species were in Wasco County, with the earliest moth capture in 2003 near Dufur, in a sticky wing type trap baited with a pheromone lure for trapping Helicoverpa armigera (Hübner), and placed as part of an insect survey by the Oregon Department of Agriculture. In 2005, 28 H. dysodea adults were captured in The Dalles area of Wasco County in Lindgren funnel traps (PheroTech Inc., Delta, BC, Canada) baited with ethanol lures and used in a survey for scolytid beetles. An additional 11 moths were captured between 5 June and 21 September 2006 in the same area with the same type of trap. In 2006, moths were also captured in the Dufur and The Dalles areas, in Universal Moth Traps (Great Lakes IPM, Vestaburg, MI) baited with a floral based chemical attractant, and in The Dalles in The floral lure was a blend of blacklight traps. phenylacetaldehyde, methyl salicylate, beta myrcene, and methyl-2-methoxy benzoate, developed for trapping flower-visiting Noctuidae such as the alfalfa looper Autographa californica (Speyer) (Landolt et al. 2007). The floral lure was tried as an attractant for H. dysodea because of the reported affinity of the moth for flowers (Bretherton et al. 1979). In 2006, seven larvae were collected from prickly lettuce, Lactuca serriola L.

Volume 64, Number 4



Fig. 1. Adult male moth of *Hecatera dysodea* reared from larva collected on *Lactuca serriola* in Dufur, Oregon.

near Dufur and were reared to the adult stage following protocols similar to that described below for studies in Washington. However, these larvae were maintained in plastic bags instead of paper cups.

After 2006, *H. dysodea* were found over a wider area in Oregon. Moths were captured in survey traps baited with ethanol lures in Baker, Malheur, Umatilla, and Union Counties in 2007. In 2009, larvae were found on prickly lettuce flower stalks in Gilliam County, and two adults were collected from the side of a residence in the city of Portland, Multnomah County.

Washington. Collection information is summarized in Table 2 and Figure 3. In the state of

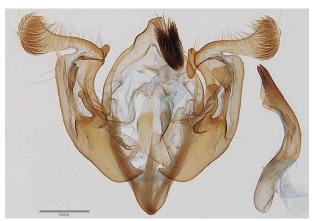


FIG. 2. Male genitalia with aedeagus removed and figured laterally. Note the slightly asymmetrical morphology of the valvae.

Washington, the first collections of *H. dysodea* moths were two adult specimens at porch lights of a residence in Stevenson, Skamania County. In the summer of 2009, prickly lettuce flower stalks along several roadways in south central and southeastern Washington were searched for Lepidoptera larvae. This included: 1) State Route 14, which parallels the Columbia River, from Paterson west to Vancouver, 2) Interstate Highway 5 from Vancouver north to Tacoma, 3) U.S. Route 97 from the Columbia River north to Yakima, 4) Interstate Highway 82 from Yakima to Richland, 5) the vicinity of Yakima, 6) the vicinity of Pullman in Whitman County, and 7) State Route 24 from Colfax in Whitman County to Othello

Table 1. Collection information for *Hecatera dysodea* in the state of Oregon. Old world bollworm (OWB) trap was baited with the sex pheromone of *Helicoverpa armigera*.

County	Location	Date	Number	Stage	Collection Note
Baker	Baker City	27 VII 2007	1	adult	ethanol trap
Gilliam	Rts I-84 X SR 74	12 VIII 2009	several	larvae	on L. serriola
Malheur	Ontario	V to IX 2007	5	adult	ethanol trap
Multnomah	Portland	7 VIII 2009	2	adult	on wall
Umatilla	Hermiston	29 VI 2007	1	adult	ethanol trap
	Hermiston	15 X 2007	1	adult	ethanol trap
Union	La Grande	VII to IX-2007	9	adult	ethanol trap
Wasco	Dufur	24 IX 2003	1	adult	OWB trap
	The Dalles	V to VI 2005	28	adult	ethanol trap
	The Dalles	VI to IX 2006	11	adult	ethanol trap
	Dufur area	VI to IX 2006	10	adult	floral lure trap
	Dufur	2 VII 2006	7	larvae	on L. serriola
	The Dalles	VI to IX 2006	6	adult	floral lure trap
	The Dalles	VI toVII 2006	4	adult	blacklight trap

in Adams County. Generally, when a patch of prickly lettuce was sampled, 30 plants were searched. Prickly lettuce plants are visually distinctive and are readily identified when flower stalks are developed (see Whitson *et al.* 1996). When captured, caterpillars were placed in 280 ml wax coated paper cups with plastic lids, along with cuttings of the host plant with flower buds and flowers. Cups with larvae were placed in an insulated chest for transport to the laboratory.

Larvae were reared to the adult stage in the laboratory, and were kept in the same wax-coated paper cups. Fresh cut sections of prickly lettuce flower stalks were placed in each cup with larvae daily, and any dead larvae or moldy frass and plant material were removed at that time. When larvae were ca 2 cm in length, 2 cm of potting soil (a one to one mixture of sand and peat moss) was placed in the bottom of the cup as a pupation medium. After larvae burrowed into and remained with the soil, the cups were transferred to screened cages for adult emergence. An emergence cage was maintained for each field site that yielded larvae. Emergence cages were checked daily for moths, which were killed and prepared as voucher specimens.

In 2009, from 9 July to 20 August, 99 Lepidoptera larvae were collected from flower stalks of prickly lettuce plants at eight sites in four counties in the state of Washington (Table 2). Seventy-two *H. dysodea* adult moths emerged from these samples from 28 July to 9 October 2009. Although some larvae died while



FIG. 3. Counties of the states of Oregon and Washington positive for collection of *Hecatera dysodea*.

held in the laboratory, we did not see the emergence of any parasitoids.

Anecdotal observations were made of larvae during field sampling. Larvae varied in coloration and size, with smaller larvae green with a pale lateral stripe, and larger larvae tan to light brown. At several sites where multiple larvae per plant were encountered (Drano Lake, Mary Hill State Park, Roosevelt, Wishram), larvae collected on a single plant were of multiple sizes, indicating that oviposition on that plant occurred

Table 2. Collection information for *Hecatera dysodea* in the state of Washington.

County	Location	Date	Number	Stage	Collection Note
Benton	5 mi S Prosser	8 VII 2009	7	larvae	on L . $serriola$
Klickitat	Dallesport	18 VII 2009	7	larvae	on L. serriola
	Goldendale	8 VII 2009	1	larva	on L. serriola
	Goldendale	18 VII 2009	5	larvae	on L. serriola
	Mary Hill State Park	3 VII 2009	20	larvae	on L. serriola
	Mary Hill State Park	8 VII 2009	15	larvae	on L. serriola
	Mary Hill State Park	15 VIII 2009	6	larvae	on L. serriola
	Roosevelt	8 VII 2009	10	larvae	on L. serriola
	Wishram	18 VII 2009	13	larvae	on L. serriola
Skamania	Drano Lake	18 VII 2009	10	larvae	on L. serriola
	Stevenson	8 VIII 2008	1	adult	porch light
	Stevenson	9 IX 2008	1	adult	porch light
Yakima	Union Gap	12 VII 2009	1	larva	on L . $serriola$
	Union Gap	20 VIII 2009	2	larvae	on L . $serriola$

Volume 64, Number 4

over a period of at least several days (multiple oviposition bouts). All larvae burrowed into the potting soil and pupated within a cell made of soil. These cells were particles of soil loosely bound together to make an oblong sphere-shaped structure encompassing an air space holding the pupa. The pupae remained within this cell after eclosion and adult emergence.

DISCUSSION

The known distribution of H. dysodea in North America now includes a broad area within the Pacific Northwest of the United States, comprised of a contiguous area of seven Oregon and four Washington counties (Figure 3). Temporal and geographic aspects of the collection records suggest the distribution of the insect has expanded since 2006. However, the absence of positive collection records does not demonstrate its absence in an area. The host plant utilized, prickly lettuce, is widespread and can be extremely abundant. Collection sites near Dufur and in The Dalles in Wasco County of Oregon, as well as near Prosser in Benton County and in Union Gap of Yakima County, Washington are among abundant patches of prickly lettuce in areas of physical disturbance and irrigated agriculture. The population of this moth could very rapidly increase in those counties as well as in similar habitat of the Columbia River Basin to the north and the Palouse Prairie to the east. The host plant is also very abundant at disturbed sites in low elevation areas west of the Cascade Mountains, such as the Williamette Valley of Oregon and Puget lowlands of Washington.

At present, the introduction, range, and spread of this Eurasian species in the Pacific Northwest are not of particular concern. Prickly lettuce is itself an introduced Palearctic weed (Whitson et al. 1996). Two other introduced weeds that are potential hosts of H. dysodea in the Pacific Northwest are Sonchus asper (L.) (prickly sow thistle) and Crepis setosa (Haller) (rough hawksbeard). Two native species of wild lettuce, Lactuca pulchella (Pursch) (blue lettuce) and Lactuca biennis (Moench) Fernald are also potential hosts. There is commercial agricultural production of lettuce (Lactuca sativa L.) seed in the Pacific Northwest that may be at risk if the moth continues to expand its distribution. For example, commercial lettuce seed production occurs in Skagit County, Washington well to the north and west of the present known distribution of *H. dysodea*.

In Britain, the moth is reported to have one generation per year (Skinner 1984). The moth flight occurs there in late June and July, larvae are present

on plants in July and August, and overwintering is in the pupal stage. However, in our Oregon and Washington records, the moth was collected in every month from May into October. Also, larvae were present on host plants at least from late June into late August. These observations suggest more than one generation of this insect per year in the Pacific Northwest of the United States.

Specimens of *H. dysodea* from Oregon were deposited in the Canadian National Collection of Insects, Ottawa, Ontario, and the United States National Museum of Natural History, Smithsonian Institution, Washington, DC (USNM). Specimens of *H. dysodea* from Washington were deposited in the USNM, the collection of the Washington State Department of Agriculture, and the M. T. James Entomological Collection, Department of Entomology, Washington State University.

ACKNOWLEDGEMENTS

We thank J. Donald Lafontaine, Steve Passoa, Mike Pogue, and Lars Crabo for identification of specimens, J. Donald Lafontaine, Steve Passoa, Martin Honey, and Colin Plant for help with references, Steve Valley for digital images, Shirley Reed, William Rivers, and Todd Adams for field support, and Jim Hansen for preparing the distribution map. John Davis of Stevenson, Washington kindly contributed collection information for Portland, Oregon and Stevenson, Washington. Helpful suggestions to improve the manuscript were made by James Hansen, Michael Pogue and Wee Yee. This study was supported in part by funding to the Oregon Department of Agriculture from USDA-CAPS grants #03-8584-0698 CA, #05-8584-0260 CA, #07-8584-0260 CA, and #07-8584-1095, U.S. Forest Service grant 2007CA-11062754-220 and funding to Washington State University from the Washington State Potato Commission.

LITERATURE CITED

AGASSIZ, D. J. L. & W. M. SPICE. 1998. The return of the small Ranunculus. Entomol. Rec. and J. of Var. 110:229–232.

BECK, H. 1999. Die Larven der Europäischen Noctuidae: Revision der Systematik der Noctuidae, Herbipoliana 5, Vol. I, 864 pp.

BRETHERTON, R. F., B. GOATER, & R. I. LORIMER. 1979. Noctuidae, pp. 120–278. In J. Heath & A. M. Emmet (eds.), The Moths and Butterflies of Great Britain and Ireland, Volume 9, Sphingidae–Noctuidae, Noctuinae and Hadeninae. Curwen Press, London.

HACKER, J., L. RONKAY, & M. HREBLAY. 2002. Noctuidae Europaiae, Volume 4, Hadeninae I. Entomological Press, Sorr, Denmark. 419 pp.

Hodges, R. W., T. Dominick, D. R. Davis, D. C. Ferguson, J. G. Franclemont, E. G. Munroe, & J. A. Powell (eds). 1983. Checklist of the Lepidoptera of America North of Mexico. The Wedge Entomological Research Foundation. 284 pp.

HONEY, M. R. 2002. The small Ranunculus, *Hecatera dysodea* ([Denis & Schiffermüller], 1775) (Lepidoptera: Noctuidae), and its return to Surrey (V.C. 17). Entomol. Gazette 53:92.

IVINSKIS, P. & J. MIATLEUSKI. 1999. Data on Noctuidae (Lepidoptera) of Turkmenistan. Acta Zoologica Lituanica 9: 201–208.

LANDOLT, P. J., A. PANTOJA, A. HAGERTY, L. CRABO, & D. GREEN. 2007. Moths trapped in Alaska with feeding attractant lures and the seasonal flight patterns of potential agricultural pests. Can. Entomol. 139:278–291.

- Poole, R. W. 1989. Lepidopterum Catalogus (New Series). Fascicle 118, Part 1. Noctuidae. E. J. Brill/Flora and Fauna Publications, New York, 500 pp.
- Pratt, C. 1986. A modern review of the demise of *Hecatera dysodea* D. & S.: the small ranunculus. Entomol. Rec. and J. of Var. 98:70–78, 114–118, 154–158.
- SKINNER, B. 1984. Color identification guide to moths of the British Isles. William Clowes Ltd. Publisher, London. 276 pp.
- SOUTH, R. 1920. The moths of the British Isles. Frederick Warne and Co., Ltd., London. 355 pp.
- WHITSON, T. D., L. C. BURRILL, S. A. DEWEY, D. W. CUDNEY, B. E. NELSON, R. D. LEE, & R. PARKER. 1996. Weeds of the West. (Fifth edition). Published by The Western Society of Weed Science. 630 pp.
- ence. 630 pp.
 WILTSHIRE, E. P. 1957. The Lepidoptera of Iraq. Bartholomew Press, Dorking, UK. 160 pp.

Received for publication 6 November 2009; revised and accepted $23\,\mathrm{March}\ 2010.$