

Tidal Adaptation of a Circadian Clock Controlling a Crustacean Swimming Behavior

Author: Akiyama, Tadashi

Source: Zoological Science, 14(6): 901-906

Published By: Zoological Society of Japan

URL: https://doi.org/10.2108/zsj.14.901

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <u>www.bioone.org/terms-of-use</u>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Tidal Adaptation of a Circadian Clock Controlling a Crustacean Swimming Behavior

Tadashi Akiyama*

Ushimado Marine Laboratory, Okayama University, Ushimado 701-43, Japan

ABSTRACT—Cumacean *Dimorphostylis asiatica* exhibits an endogenous swimming rhythm, free-running period of which spontaneously changes from circa 12-hr to circa 24-hr in the laboratory. Administration of a 4-hr pulse of hydrostatic pressure stimulus and a 4-hr light pulse confirmed that the circa 12-hr rhythm is circatidal and the circa 24-hr rhythm circadian, with respect to the phase setting characteristics. The activity records provide evidence of a "splitting" phenomenon, which suggests that these two types of rhythms are governed by (an) identical pacemaker(s). This species appears to have acquired tidal synchrony by making use of flexibly coupled circadian pacemakers, accompanied by expansion of their subtidal habitat to shallow, tide-affected area.

INTRODUCTION

Diurnal vertical migration (DVM) of marine zooplanktonic crustaceans has been reported for several taxonomic groups such as copepods (Enright, 1977), a shrimp (Pearcy, 1970) and cumaceans (Corey, 1970). This behavior is predominantly based on nocturnal swimming, which is largely governed by circadian rhythm (Enright and Hamner, 1967).

In the Seto Inland Sea of Japan, a cumacean, Dimorphostylis asiatica, exhibits a temporal pattern of DVM modified by tidal cycles: the animals swimm up to the water surface at nocturnal high tides (Akiyama and Yoshida, 1990). The endogenous swimming rhythm of freshly collected animals is circatidal but not circadian (Akiyama, 1995). Compared with the generally noisy, less persistent circatidal activity pattern of intertidal organisms (Neumann, 1981), the free-running rhythm of D. asiatica is highly precise and persistent, which is rather a common feature of circadian rhythms. In addition, unaccountable change in the free-running period from circa 12-hr to circa 24-hr occurred within several days of recording in most of the specimens (Akiyama, 1995). One plausible explanation is that the "circatidal" rhythm of D. asiatica spontaneously changes into a circadian rhythm in the laboratory.

An effective method to discriminate circatidal rhythms from circadian rhythms is the administration of rhythm-specific agents: circatidal rhythms are phased by some tidally caused stimulus such as mechanical agitation (Enright, 1965; Hastings, 1981), temperature (Williams and Naylor, 1969), salinity (Bolt and Naylor, 1985), or hydrostatic pressure (Morgan, 1965; Northcott *et al.*, 1991a, b), but circadian rhythms

* Corresponding author: Tel. +81-86934-5210; FAX. +81-86934-5211. are predominantly entrained by light. In this study, phase responsiveness of the circa 12-hr and the circa 24-hr rhythms of *D. asiatica* to a 4-hr pulse of hydrostatic pressure stimulus and a 4-hr light pulse were compared.

MATERIALS AND METHODS

Adult males of *Dimorphostylis asiatica* were collected at night along the coast of the Seto Inland Sea at Ushimado, Okayama Prefecture, using electric torch lamp (100 V, 200 W) and a hand net. Each freshly collected specimen was transferred to an air tight cylindrical recording aquaria (3 cm in diameter, 7.5 cm in height) containing 20 ml sea water and 1 mm layer of boiled mud substratum. The aquaria were kept under constant darkness in light tight incubators. The activity of the specimens was monitored using photoelectric switches and personal computers (NEC PC-8800). The double plotted actogram for each specimen was illustrated using another personal computer (NEC PC-9800).

A 4-hr hydrostatic pressure stimulus equivalent to 3 m of water was made using an electric air pump (100 V, 100 W; IWAKI Co.), which was connected to the recording aquaria. The intensity of the stimulus was controlled by a hand made air bulb and tightly packed wool which also eliminated high frequency air vibration. A 4-hr light pulse (approximately 1000 lx) was given using cool fluorescent lamps (100 V, 20 W). Water temperature was kept at 15°C and 20°C for the circa 12-hr rhythms, and 10°C and 15°C for the circa 24-hr rhythms, depending on seasonal changes of ambient sea water at the collection site.

Linear regression lines of consecutive activity onsets (illustrated as diagonal lines in Figs. 1, 3, 5 and 7) were calculated to determine the phase of the rhythm and the phase shifts. Phase of treatments of the hydrostatic and light pulses was determined as offset time of treatments to activity onsets (circadian time 12), which was calibrated by the free-running periods. For the circa 12-hr rhythms, phase shifts of two conjugated activities were calculated separately (so one or two phase shifts are plotted for each treatment).

RESULTS

Phase responses of the circa 12-hr rhythms

Figure 1 illustrates the circa 12-hr swimming rhythms of *Dimorphostylis asiatica* exposed to a 4-hr pulse of hydrostatic pressure. As well as phase shifts of the rhythms, the treatment resulted in two types of responses: persistence of the circa 12-hr rhythm after the treatment (Fig. 1A-D) and change of the activity rhythm to a circa 24-hr cycle (Fig. 1E, F). The phase responses to the stimulus are summarized as phase response curves (PRCs, Fig. 2). Although it is unknown whether these PRCs are comparable with the PRCs of circatidal rhythms previously reported (Enright, 1976; Naylor and Williams, 1984; Harris and Morgan, 1984; Northcott *et*

al., 1991b), including the unaccountable change of the activity pattern, the several hours of phase shifts (especially in Fig. 2B) strongly suggests that the circa 12-hr rhythm perceive hydrostatic pressure as an entraining agent.

The circa 12-hr swimming rhythms exposed to a 4-hr light pulse are shown in Fig. 3. Most of the specimens retained the circa 12-hr rhythms after the treatments. The treatments did not cause large phase shifts. The PRC to the light pulses is illustrated in Fig. 4. Phase advances and delays, most of which were within 2 hr, did not relate to the circadian time at which the pulses were given. Thus the circa 12-hr rhythm of *D. asiatica* is a "circatidal" rhythm with respect to the phase setting characteristics.

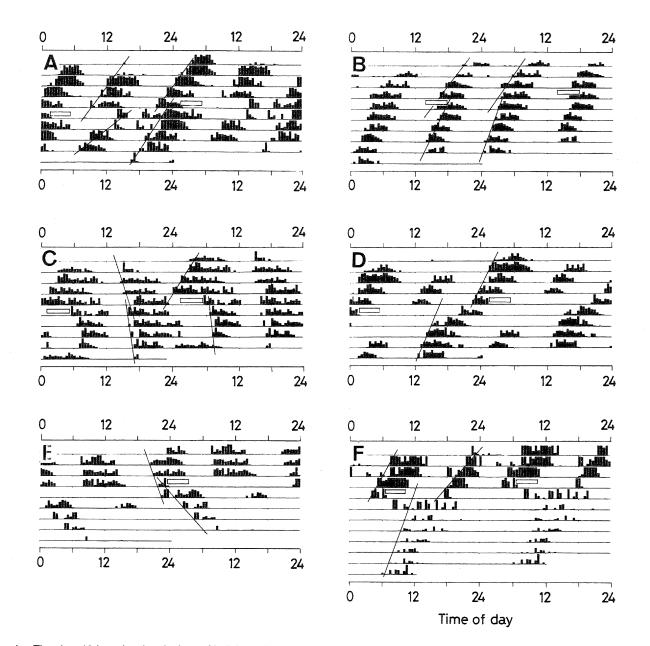


Fig. 1. The circa 12-hr swimming rhythms of individual *Dimorphostylis asiatica* exposed to a 4-hr pulse of hydrostatic pressure. Rectangles indicate the times of treatment. Diagonal lines indicate the linear regression lines of consecutive activity onsets.

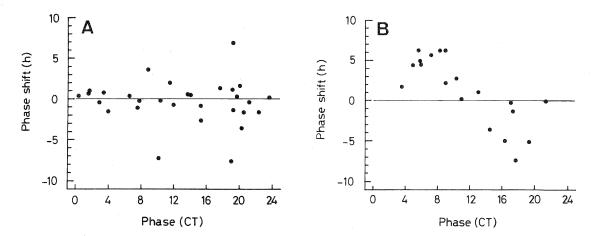


Fig. 2. Phase response curves (PRCs) of the circa 12-hr swimming rhythms exposed to the hydrostatic pressure pulses. (A) PRC of the specimens which retained circa 12-hr rhythms after the treatments (20 specimens). (B) PRC of the specimens whose activity rhythms changed to the circa 24-hr rhythms after the treatments (19 specimens). Classification of the phase responses into these two types summarized in (A) and (B) depended on the activity pattern on fourth and fifth days after the treatment. For example, the case shown in Fig. 1D was classified into (A) and the case in Fig. 1E into (B).

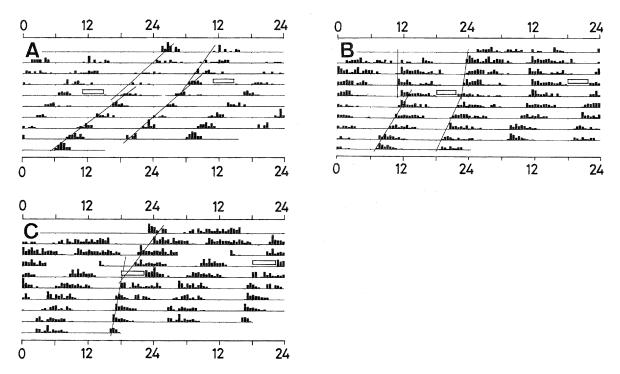


Fig. 3. The circa 12-hr swimming rhythms of *D. asiatica* exposed to a 4-hr light pulses. Rectangles indicate the times of treatment. Diagonal lines indicate the linear regression lines of consecutive activity onsets.

Phase responses of the circa 24-hr rhythms

Administration of the entraining agents to the circa 24-hr rhythm caused a change of phase setting accompanied by a change in free-running periods. Figure 5 shows the representative circa 24-hr swimming rhythm exposed to the hydrostatic pressure pulses, in which small phase shifts were caused by the treatments. The PRC (Fig. 6) indicates that the phase advances and delays, most of which are within 2 hr, did not relate to the circadian time at which the pulse was given. Large phase shifts more than 5 hr, caused by four specimens, appear to lie outside the normal distribution of the phase shifting response.

While the phase response to 4-hr hydrostatic pressure was obscure, the swimming rhythm appeared to be phased by 4-hr light pulses adjusting the activity onsets to the time of light off for each treatment (Fig. 7). The strong rephasing effect of the light pulses was also confirmed by the PRC (Fig. 8). Thus the circa 24-hr rhythm is definitely "circadian".

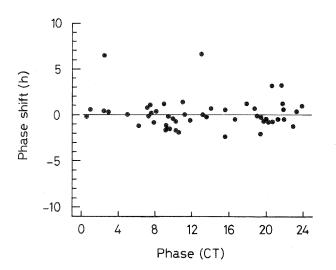


Fig. 4. Phase response curve of the circa 12-hr rhythms exposed to a 4-hr light pulse (32 specimens).

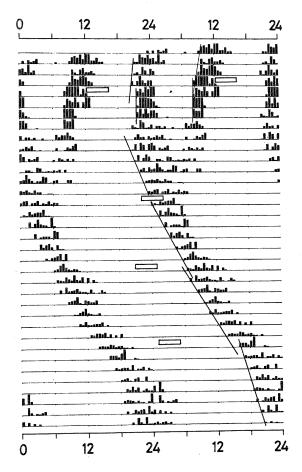


Fig. 5. The circa 24-hr swimming rhythm of *D. asiatica* exposed to 4-hr hydrostatic pressure pulses. Rectangles indicate the times of treatments. Diagonal lines indicate the linear regression lines of consecutive activity onsets.

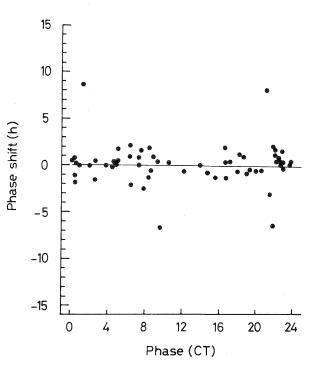


Fig. 6. A phase response curve of the circa 24-hr rhythms receiving a hydrostatic pressure pulses (38 specimens).

DISCUSSION

The present results clearly show that the circatidal swimming rhythm of *Dimorphostylis asiatica* spontaneously changes to circadian in the laboratory. Now arises a question whether the underlying pacemakers of these rhythms are identical. The activity records of some specimens provide evidence of a "splitting" phenomenon which is informative about the underlying pacemaker(s): 1) two conjugated activities of the circa 12-hr rhythm were occasionally rephased by the pressure stimulus differently (Fig. 1C, D), which suggests that these activities are governed by two or more pacemakers and 2) these two conjugated activities fused gradually to become a single circa 24-hr rhythm (Fig. 7B, C), and rarely re-splitted to revert to the circa 12-hr rhythm (such a tendency is shown in Fig. 7C).

"Splitting" has been reported mainly in vertebrates, including mammals (Pittendrigh and Daan, 1976), birds (Gwinner, 1974), and lizards (Underwood, 1977). Recent studies have documented that splitting is also observed in circatidal rhythms of crab's locomotor activity (Palmer and Williams, 1986). To explain the splitting, one can postulate the existence of two circa 24-hr pacemakers whose phase angle difference determines whether the split (circa 12hr-period) or fused (circa 24hr-period) activity pattern is displayed (Pittendrigh and Daan, 1976). The circatidal and circadian rhythms of *D. asiatica* also appear to be governed by identical pacemakers but with different phase angle relationships.

Conversion of the activity rhythm in *D. asiatica* is possibly related to their low degree of habitat specialization, which generates various intensity of tidal impact according to the

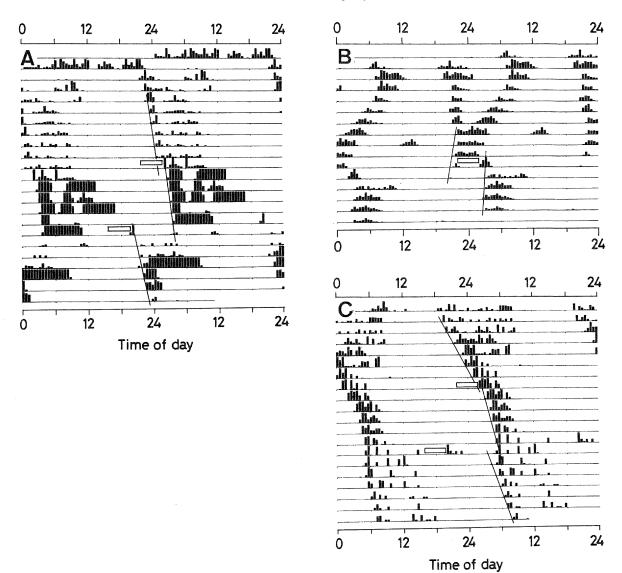


Fig. 7. The circa 24-hr swimming rhythms of *D. asiatica* exposed to a 4-hr light pulse. Rectangles indicate the times of treatment. Diagonal lines indicate the linear regression lines of consecutive activity onsets.

locality. For example, at Shijiki Bay (Nagasaki Prefecture, open to East China Sea), the animals live in muddy substratum in 13-50 m deep, where tidal impact would be small (Azuma, personal communication). In contrast, their habitat in the Seto Inland Sea of Japan (ca. 1.5-3.5 meter tidal amplitude) is situated in shallow water (within a few meters below mean low water spring tide), where there would be considerable tidal impact. It is concluded that, taking into consideration the general cumacean subtidal habitat with little tidal impact, this species would have developed tide-synchronized swimming on the bases of the circadian pacemaker(s) that it originally possessed.

Previous studies have hypothesized that "circatidal rhythms are tidally synchronized circadian rhythms" (Enright, 1976; Rodriguez and Naylor, 1972; Gibson, 1973). However, experimental supports are not sufficient, partly because the circatidal rhythms have been studied in a restricted number of species, most of which are decapod crustaceans such as shore crabs. More extensive studies using a variety of species would reveal the relationship between the circatidal and circadian rhythms.

ACKNOWLEDGMENTS

I thank late Prof. Dr. M. Yoshida, Prof. Dr. M. Yamamoto, Dr. H. Shirai and Dr. K. Ohtsu, Ushimado Marine Laboratory for encouragement of this work. I also thank Dr. I. G. Gleadall for revision of the early form of the manuscript. I was indebted to Dr. M. Saigusa, Okayama University for technical assistance.

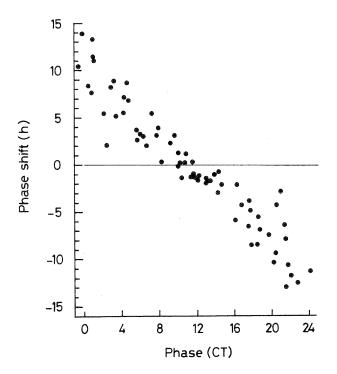


Fig. 8. A phase response curve of the circa 24-hr rhythms exposed to a 4-hr light pulses (51 specimens).

REFERENCES

- Akiyama T (1995) Circatidal swimming activity rhythm in a subtidal cumacean *Dimorphostylis asiatica* (Crustacea). Mar Biol 123: 251–255
- Akiyama T, Yoshida M (1990) The nocturnal emergence activity rhythm in the cumacean *Dimorphostylis asiatica* (Crustacea). Biol Bull 179: 178–182
- Bolt SRL, Naylor E (1985) Interaction of endogenous and exogenous factors controlling locomotor activity rhythms in *Carcinus* exposed to tidal salinity cycles. J Exp Mar Biol Ecol 85: 47–56
- Corey S (1970) The diurnal vertical migration of some Cumacea (Crustacea, Pericardia) in Kames Bay, Isle of Cumbrae, Scotland. Can J Zool 48: 1385–1388
- Enright JT (1965) Entrainment of a tidal rhythm. Science 147: 864-867

Enright JT (1976) Plasticity in an isopod's clockworks: Shaking shapes

form and affects phase and frequency. J Comp Physiol 107: 13– 37

- Enright JT (1977) Copepods in a hurry: Sustained high-speed upward migration. Limnol Oceanogr 22: 118–125
- Enright JT, Hamner WM (1967) Vertical diurnal migration and endogenous rhythmicity. Science 157: 937–941
- Gibson RN (1973) Tidal and circadian activity rhythms in juvenile plaice, *Pleuronectes platessa*. Mar Biol 22: 379–386
- Gwinner E (1974) Testosterone induces "splitting" of circadian locomotor activity in birds. Science 185: 72–74
- Harris GJ, Morgan E (1984) The effects of salinity changes on the endogenous circatidal rhythm of the amphipod *Corophium voltator* (Pallas). Mar Behav Physiol 10: 199–217
- Hastings MH (1981) The entraining effect of turbulence on the circatidal activity rhythm and its semilunar modulation in *Eurydice pulchra.* J Mar Biol Assoc UK 61: 151–160
- Morgan E (1965) The activity rhythm of the amphipod *Corophium voltator* (Pallas) and its possible relationship to changes in hydrostatic pressure associated with tides. J Anim Ecol 34: 731– 746
- Naylor E, Williams BG (1984) Phase-responsiveness of the circatidal locomotor activity rhythm of *Hemigrapsus edwardsi* (Hilgendorf) to simulated high tide. J Mar Biol Assoc UK 64: 81–90
- Neumann D (1981) Tidal and lunar rhythms. In "Biological Rhythms (Handbook of Behavioral Neurobiology Vol 4)" Ed by Aschoff J, Plenum, New York, pp 351–380
- Northcott SJ, Gibson RN, Morgan E (1991a) The effect of tidal cycles of hydrostatic pressure on the activity rhythm of *Lipophrys pholis* (L.) (Teleostei). J Exp Mar Biol Ecol 148: 35–45
- Northcott SJ, Gibson RN, Morgan E (1991b) Phase responsiveness of the activity rhythm of *Lipophrys pholis* (L.) (Teleostei) to a hydrostatic pressure pulses. J Exp Mar Biol Ecol 148: 47–57
- Palmer JD, Williams BG (1986) Comparative studies of tidal rhythms.
 II. The dual clock control of the locomotor rhythms of two decapod crustaceans. Mar Behav Physiol 12: 269–278
- Pearcy WG (1970) Vertical migration of the ocean shrimp *Pandalus jordani*: A feeding and dispersal mechanism. Calif Fish Game 56: 77–140
- Pittendrigh CS, Daan S (1976) A functional analysis of circadian pacemakers in nocturnal rodents. V. Pacemaker structure: A clock for all seasons. J Comp Physiol 106: 333–355
- Rodriguez G, Naylor, E (1972) Behavioural rhythms in littoral prawns. J Mar Biol Assoc UK 52: 81–95
- Underwood H (1977) Circadian organization in lizards: the role of pineal organ. Science 195: 587–589
- Williams BG, Naylor E (1969) Synchronization of the locomotor rhythm of *Carcinus*. J Exp Biol 51: 715–725

(Received August 18, 1997 / Accepted September 13, 1997)