

# Nest design and parental care of Striped Woodhaunter Automolus subulatus

Authors: Conejo-Barboza, Karla, Sánchez, César, Sandoval, Luis, and Greeney, Harold F.

Source: Bulletin of the British Ornithologists' Club, 140(4): 468-476

Published By: British Ornithologists' Club

URL: https://doi.org/10.25226/bboc.v140i4.2020.a10

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <a href="https://www.bioone.org/terms-of-use">www.bioone.org/terms-of-use</a>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

## Nest design and parental care of Striped Woodhaunter Automolus subulatus

by Karla Conejo-Barboza, César Sánchez, Luis Sandoval & Harold F. Greeney

Received 12 August 2020; revised 19 November 2020; published 9 December 2020 http://zoobank.org/urn:lsid:zoobank.org:pub:2E85A1D4-A262-48A7-B03E-F379AC3E5410

Summary.—The western and trans-Andean populations of Striped Woodhaunter Automolus subulatus are sometimes considered separate species. We discuss previously published data on the nesting of Striped Woodhaunter and present novel information concerning the nest, eggs, nestlings and parental care of western A. s virgatus and trans-Andean A. s. subulatus. Nest placement and architecture of the two populations are similar to each other and to other Automolus species. However, Striped Woodhaunter build shorter nest tunnels than other related species and genera. All similarities in nest design, nestbuilding behaviour and parental care presented herein support the genetic clade including Automolus, Thripadectes and Clibanornis, but do not differentiate between the subspecies of Striped Woodhaunter. More studies are required about adult attendance and nest design within this clade, taking into account more samples across the species' range.

The systematics of the non-monophyletic genus Automolus (Furnariidae: Philydorini) are still under scrutiny (Claramunt et al. 2013, Schultz et al. 2017). The most recent changes that have gained acceptance are the subsuming of Hyloctistes within Automolus (Claramunt et al. 2013) and the splits of Pará Foliage-gleaner A. paraensis from Olive-blacked Foliage-gleaner A. infuscatus (Claramunt et al. 2013, Clements et al. 2019) and Chiriquí Foliage-gleaner A. exsertus from Buff-throated Foliage-gleaner A ochrolaemus (Freeman & Montgomery 2017, Chesser et al. 2018). Like other furnariids (Irestedt et al. 2006), nesting behaviour and nest architecture of Automolus spp. may prove useful for testing DNA-based taxonomic arrangements. However, the nesting biology of Automolus species is well known only for Chiriquí Foliage-gleaner (Skutch 1952, 1969) and White-eyed Foliage-gleaner A. leucophthalmus (Euler 1900, J. C. R. Magalhães in Remsen 2003a, Marini et al. 2007, Cockle & Bodrati 2017) but poorly known or unpublished for the other seven species (Remsen 2003a).

Striped Woodhaunter A. subulatus is distributed from eastern Nicaragua south to western Ecuador and, east of the Andes, from southern Venezuela and south-east Colombia to northern Bolivia and western Amazonian Brazil (Stiles & Skutch 1995, Remsen 2003a). Some authors treat western and trans-Andean populations as separate species, Western Woodhaunter A. virgatus and Amazonian Woodhaunter A. subulatus (Ridgely & Greenfield 2001, Hilty 2003, del Hoyo & Collar 2016) based largely on vocal differences (Ridgely & Tudor 1994, Freeman & Montgomery 2017). This split is considered premature by other authorities (see Remsen 2003b) and herein we follow Clements et al. (2019). Here we discuss and clarify published data on the nesting of Striped Woodhaunter and present novel information concerning the nests, eggs, nestlings and parental care for two subspecies, A. s. virgatus and A. s. subulatus.

Historical data for A. s. subulatus and A. s. assimilis. - The first published nest description attributed to Striped Woodhaunter was presented by Sclater & Salvin (1873). They quoted the notes of E. Bartlett, which accompanied a specimen of A. subulatus collected at Chamicuros, Loreto, eastern Peru (05°30'S, 75°30'W, sensu Stephens & Traylor



TABLE 1 Monitoring dates, localities, nest contents and main observations of Striped Woodhaunter Automolus subulatus nests found in western Colombia (nest 1; Zyskowski & Prum 1999, Hilty 2003; K. J. Zyskowski in litt. 2018), in central-west Costa Rica (nest 2) and in eastern Ecuador (nests 3–7).

Nest	Date found	Days monitored	Location/elevation	Coordinates	Nest contents	Observations
1	15 Feb 1976		San Isidro, Buenaventura, dpto. Valle del Cauca, Colombia	03°27′0″N 77°10′0″W	2 nestlings	Nestlings more than half-grown
2	19 Dec 2002		Finca Rafiki Safari Lounge, Santo Domingo, Perez Zeledón, prov. San José, Costa Rica; 130 m	09°27′41″N 83°59′39″W	1 egg	Egg: 28.1 × 21.0 mm
3	15 May 2004	15–20 May 2004 and 25 Jun 2004	Near La Selva Jungle Lodge, c.75 km north-east of Coca, adjacent to Lake Garzacocha, prov. Sucumbíos, Ecuador; 250 m	00°29′53″S 76°22′23″W	2 fresh eggs	Eggs: 24.2 × 17.2 and 22.8 × 17.1 mm. Adult behaviour documented on video.
4	17 Feb 2012	17, 20, 25 Feb 2012	Cabañas Yankuam, south of río Pastaza, south (right) bank of río Nangaritza, prov. Zamora- Chinchipe, Ecuador; 1,100 m	04°15′0″S 78°39′30″W	empty	Burrow excavation.
5	26 Sep 2012	26 Sep 2012	Boanamo, near the prov. Pastaza/ Orellana border, Ecuador; 230 m	01°15′45″S 76°22′54″W	2 eggs (1 inviable*)	Eggs: $24.2 \times 17.8$ and $23.7 \times 17.9$ * mm; mass: $3.8$ and $3.5$ g.
6	5 Mar 2013	5, 7, 8, 10 Mar 2013	Gareno Lodge, south of río Napo, prov. Napo, Ecuador; 400 m	01°01′59″S 77°23′42″W	empty	Nest cup construction. Adult behaviour documented on video.
7	6 Mar 2013	7, 10 Mar 2013	Gareno Lodge, south of río Napo, prov. Napo, Ecuador; 400 m	01°02′01″S 77°23′15″W	2 eggs, 1 hatched	Eggs: $25.4 \times 18.5$ and $25.4 \times 18.5$ mm; mass: $4.2$ and $4.3$ g. Nestling mass: $4.7$ g. Adult behaviour documented on video.

1983). However, the description, of a cup nest built 2–3 m above ground among dead palm fronds and holding two spotted eggs, is clearly in error, as first noted by Zyskowski & Prum (1999). The last-named authors provided the only other published information on the nest architecture of Striped Woodhaunter, including a photograph, based on a nest collected by N. Wheelwright in western Colombia (nest 1; Table 1). This nest was described as a platform-like cup of loosely interlaced leaf petioles placed at the end of an earth tunnel, and the photograph revealed two nestlings probably less than half-grown at the time of discovery (Zyskowski & Prum 1999). K. Zyskowski (in litt. 2018) kindly provided additional details (Table 1) on this nest which, based on its locality, is attributable to A. s. assimilis.

Nest and egg of A. s. virgatus.—We examined a nest of A. s. virgatus collected by J. E. Sánchez & E. M. Carman at Finca Rafiki Safari Lounge, Costa Rica (nest 2; Table 1), and deposited at the Museo Nacional de Costa Rica, San José (MNCR-ONH772). Sánchez et al. (2004) provided a habitat description for the locality. Nest 2 was collected from a burrow excavated in a dirt bank adjacent to a forest trail (cavity with tunnel, sensu Simon & Pacheco 2005), with an expanded inner chamber at the end of a tunnel. The nest itself was a shallow, platform-like cup composed entirely of loosely interwoven leaf rachises (Fig. 1A). We detected both leaflet scars and extra-floral nectaries on most of the rachises, suggesting that they were probably from a plant in the family Fabaceae. No additional details concerning the burrow are provided on the specimen label, but we were able to measure the nest platform (Table 2; on Fig. 2, see measurements 10-15). A single, unmarked white egg

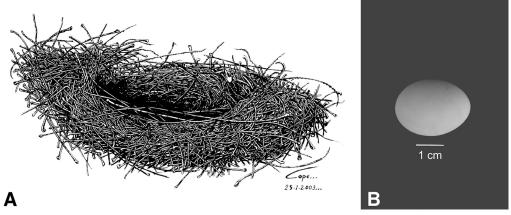


Figure 1. (A) Nest of Striped Woodhaunter Automolus subulatus virgatus, collected at Finca Rafiki, Santo Domingo, Perez Zeledón, San José Province, Costa Rica (nest 2) (© Alberto Pérez). (B) Immaculate white egg found in the nest (Karla Conejo-Barboza)

accompanied the nest (MZUCR-H205; Table 1; Fig. 1B) but the specimen label provides no details regarding clutch size or egg development. The size of the hole opened in the egg, however, suggested that it may have contained a well-developed embryo when collected.

Nests of A. s. subulatus.—HFG studied five nests of A. s. subulatus found between 2004 and 2013 at four localities in eastern Ecuador (Table 1): near La Selva Jungle Lodge (nest 3; Fig. 3A), Cabañas Yankuam (nest 4), Boanamo (nest 5; Fig. 3B-C) and Gareno Lodge (nests 6-7; Fig. 3D). Habitat was similar at all four localities, all representing mosaics typical of relatively undisturbed western Amazonian forest (see Greeney 2017, Greeney et al. 2018 for detailed descriptions). HFG visited the nests periodically to ascertain their status and contents. He checked nest contents either directly or using a small lighted mirror, and made direct observations of adult behaviours. When possible to document nest attendance by adults, he filmed nest activity at nests 3, 6 and 7 (Table 1) by placing a video camera on a tripod 1.5 m tall, 3 m from the nest entrance. Due to its position the video camera could not film activity within the inner chamber. Behaviour of the adults appeared to be unaffected by the presence of the camera.

All Ecuadorian nests were sited in earth burrows (cavity with tunnel, sensu Simon & Pacheco 2005) as described for assimilis and virgatus with entrances at a mean height of 124 cm (range = 60–230 cm; SD = 63.5 cm; Table 2) above ground (Fig. 2, measure 1). Nest 3 was in the root mass of an overturned Cecropia tree (Urticaceae), nest 4 was in a 1.5 m-tall bank with a 60 cm overhang along a road-cut, nest 5 was in streamside bank below an overhang of dirt and roots, and the other two nests were in the large (c.3 m tall) root masses of trees felled by wind action. The burrows' entrances led to tunnels that varied in slope from downward at a c.30° angle (nest 3) to sloping slightly upward, and opened into enlarged chambers containing the nest (nests 6 and 7). Nest cups of A. s. subulatus were platform-like structures of loosely arranged, stiff, unbranched leaf rachises that were barely sufficiently cohesive to remain intact when removed from the burrow. In the case of nest 5 (Fig. 3B), all rachises appeared to be from the same species of plant, but the taxonomic affinities of the nest materials were not examined closely at the other nests.

Measurements of A. s. subulatus burrows (Table 2; Fig. 2, measurements 2–9) were: entrance height = 7.1 cm (6.0–8.5 cm; SD = 1.1 cm; n = 4) and entrance max. diameter = 9.1 cm (8–11 cm; SD = 1.3 cm; n = 4); min. tunnel height = 5 cm (n = 1); tunnel width = 8.5 cm (7-10 cm; SD = 2.1 cm; n = 2); tunnel length (from entrance lip to start of inner chamber) =

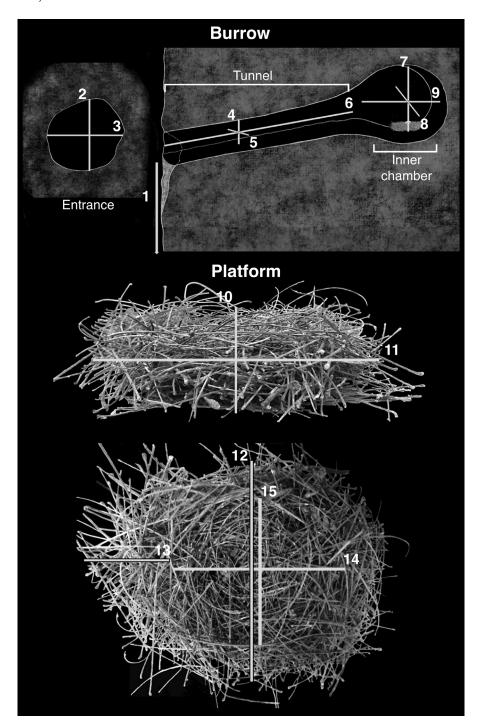


Figure 2. From top to bottom: schematic internal view of the burrow of a nest of Striped Woodhaunter Automolus s. subulatus, based on those found in eastern Ecuador (nests 3 and 5) and, a lateral and superior view of the platform nest of A. s. virgatus collected in central-west Costa Rica (nest 2). As in Table 2, numbers correspond to burrow height (1), entrance height (2), entrance max. diameter (3), tunnel height (4), tunnel max. diameter (5), tunnel length (6), inner chamber height (7), inner chamber max. diameter (8), inner chamber min. diameter (9), platform height (10), external max. diameter (11), external min. diameter (12), wall thickness (13), internal max. diameter (14), internal min. diameter (15) (Karla Conejo-Barboza)

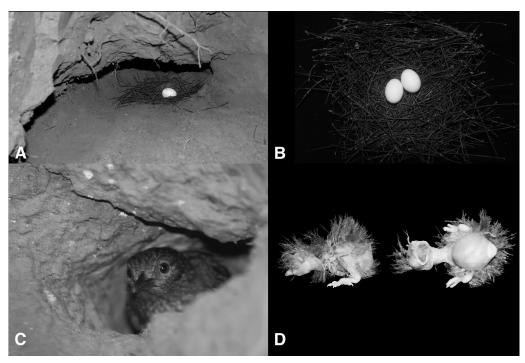


Figure 3. (A) Inner chamber and nest platform of Striped Woodhaunter Automolus s. subulatus, found near La Selva Jungle Lodge, c.75 km north-east of Coca, adjacent to Lake Garzacocha, Sucumbíos province, Ecuador (nest 3). (B) Nest platform, eggs and (C) an adult Striped Woodhaunter A. s. subulatus, found at Boanamo, near the Pastaza and Orellana province border, Ecuador (nest 5). (D) Chicks of Striped Woodhaunter A. s. subulatus, found in a nest at Gareno Lodge, south of the río Napo, Napo province, Ecuador (nest 7) (Harold F. Greeney)

26.3 cm (20–31 cm; SD = 5.2 cm; n = 4). Inner chamber height = 11 cm (10–12 cm; SD = 1.4 cm; n = 2); inner chamber max. diameter = 18 cm (13–23 cm; SD = 7.1 cm; n = 2); inner chamber min. diameter = 14 cm (12–16 cm; SD = 2.8 cm; n = 2). The inner chamber max. and min. diameter were measured perpendicular to each other on the horizontal plane (Fig. 2). HFG measured only the platform of nest 5 (Fig. 2).

Eggs and nestlings of A. s. subulatus.—The complete clutch at three of the five A. s. subulatus nests comprised two immaculate white eggs, although some were slightly stained pale brown, probably from the surrounding earth of the inner chamber (Fig. 3A-B). When the adults were not at the nests, HFG measured and photographed the eggs (nests 3, 5 and 7; Table 1; Fig. 3A-B) and one newly hatched nestling (nest 7; Table 1; Fig. 3D). Mean measurements of six eggs were 24.3 mm (22.8-25.4 mm, SD = 1.0 mm; Table 1) × 17.8 mm (17.1-18.5 mm, SD = 0.6 cm; Table 1). The masses of three eggs with advanced embryonic development were 4.1 g (3.8-4.3 g; SD = 0.3 g; Table 1). An undeveloped and slightly damaged egg weighed during the latter half of incubation had a mass of 3.5 g (nest 5). On HFG's final visit to nest 7 (Table 1), at 16.30 h, it contained a single nestling that probably hatched on the morning of the same day based on its physical appearance and mass. The second egg was lightly pipped, suggesting that the eggs' hatching would occur c.24 hours apart. The nestling weighed 4.7 g. It had long, densely plumose, grey natal down on its capital, spinal dorsal, spinal pelvic, alar, ventral sternal, femoral and crural regions (sensu Proctor & Lynch 1993). The skin was pinkish, including the tarsi and toes, with the cloaca and surrounding skin noticeably more whitish. Its nails were dusky white, as was the bill,

TABLE 2

Measurements (cm) of all Striped Woodhaunter *Automolus subulatus* nests that we found, in central-west Costa Rica (nest 2) and eastern Ecuador (nests 3–7). Ent. = entrance, max. = maximum, diam. = diameter, I. cham. = Inner chamber, min. = minimum, Ext. = external, Int. = internal. Numbers in parentheses correspond to the measurement's numbers in Fig. 2.

Measurements	Nest 2 (cm)	Nest 3 (cm)	Nest 4 (cm)	Nest 5 (cm)	Nest 6 (cm)	Nest 7 (cm)
Burrow height (1)		110	120	230	60	100
Ent. height (2)		7.5		8.5	6	6.5
Ent. max. diam. (3)		11		9	8	8.5
Tunnel height (4)		5				
Tunnel max. diam. (5)		10		7		
Tunnel length (6)		31		20	24	30
I. cham. height (7)		12		10		
I. cham. max. diam. (8)		23		13		
I. cham. min. diam. (9)		16		12		
Platform height (10)	4.2					
Ext. max. diam. (11)	14.9			14		
Ext. min. diam. (12)	12.4					
Wall thickness (13)	3.4					
Int. max. diam. (14)	6.2			5.5		
Int. min. diam. (15)	6.0					
Depth (16)	1.9			1.5		

except the dusky-grey tip. The prominent egg tooth, tomia and inflated rictal flanges were bright white and the mouth lining was pale pink, similar to the skin colour (Fig. 3D).

Behaviour of A. s. subulatus adults. - At two nests (4 and 6) HFG observed nest construction over the course of 3-4 days. Nest 4 was visited three times during burrow excavation and, when first found, was c.10 cm deep. Three days later the tunnel was c.1.5 cm deeper, and eight days after discovery it was c.15 cm deep. A single adult was flushed from the nest on both the first (09.30 h) and final (14.30 h) visits. On both occasions the adult emerged with soil on its bill, flew directly into dense vegetation 3–5 m from the nest, and vocalised continually for the 4-5 minutes that HFG remained at the nest. The vocalisation, presumably an alarm call, was nearly identical to that recorded at a nearby locality by B. M. Whitney, given by an adult in response to playback of the same vocalisation (www.xenocanto.org/86344). HFG was unable to determine if both sexes participated in excavation. The burrow of nest 6, when discovered, contained an empty but apparently fully formed nest. HFG visited the nest six times over the course of five days, between 06.15 h and 17.00 h, without observing an adult. Three days after discovery, during one hour of video observation (08.30-09.30 h), HFG recorded a single adult visit. The adult, of unknown sex, arrived with a single leaf petiole in its bill and remained in the burrow for c.3 minutes before flying away.

When first encountered, nest 3 contained a single egg showing no signs of development and a second with a tiny (>1 mm) embryo. Based on the experience of HFG with the embryonic development of numerous tropical suboscine passerines, we estimate that the clutch was completed 2–4 days prior and suspect that the undeveloped egg was inviable. HFG recorded adult incubation rhythms (on/off-bouts), between 06.00 h and 18.00 h (sunrise to sunset) on the five consecutive days following discovery of nest 3. As he was able to record only entrances and exits at the nest burrow, he inferred that the eggs were

covered during the entire period an adult was inside. Both adults incubated the eggs, as evidenced by the observation of adults replacing each other at the nest, but HFG could not distinguish the sexes. During the entire observation period, adults spent 63.5% of daylight hours warming the eggs. Daily percentage attendance for the five days was 64.5, 43.4, 37.2, 79.2 and 80.0%, respectively. On HFG's final visit to nest 7 (Table 1), direct observations of adults at the nest revealed that both brooded the nestling and delivered single, very small (1–3 mm) prey items.

## Discussion

The nest placement and architecture of the three Striped Woodhaunter subspecies reported here are similar to those reported for other Automolus (A. leucophthalmus: Euler 1900, J. C. R. Magalhães in Remsen 2003a, Marini et al. 2007, Cockle & Bodrati 2017; A. ochrolaemus: Van Tyne 1926; A. paraensis: Snethlage 1935, Pinto 1953; A. exsertus: Skutch 1952, 1969). In particular, the exclusive use of leaf rachises in nest construction appears to be ubiquitous in Automolus, but their nests are otherwise similar in form and placement to the nests of related genera (Thripadectes, Clibanornis and Buff-fronted Foliage-gleaner Philydor rufum: Derryberry et al. 2011) being platforms of loosely woven material placed at the end of upward-angled earth burrows (Skutch 1969, Kiff et al. 1989, Strewe 2001, Remsen 2003a, Maillard et al. 2006, Faria et al. 2008, Botero-Delgadillo & Guayara 2009, Zyskowski & Greeney 2010, Miller et al. 2012, Smith & Londoño 2013, Cockle & Bodrati 2017). The nest descriptions reported here demonstrate that general nest placement and design support the strong relationship within genera of the Automolus-Thripadectes-Clibanornis clade (see Cockle & Bodrati 2017), and do not appear to vary between currently recognised subspecies or populations of Striped Woodhaunter (Remsen 2003a, Clements et al. 2019).

Perhaps of significance, we found that tunnel length of Striped Woodhaunter burrows is generally shorter (26.3 cm; 20-31 cm; SD = 5.2 cm) than reported for related genera (81.4 cm; 38–200 cm; SD = 47.5 cm; Van Tyne 1926, Remsen 2003a, Marini et al. 2007, Faria et al. 2008, Botero-Delgadillo & Guayara 2009, Zyskowski & Greeney 2010, Miller et al. 2012, Cockle & Bodrati 2017). We know from other burrow nesters that habitat and nest design (e. g., entrance size and orientation, and tunnel length) may be important for the regulation of appropriate nest microclimates (Ellis 1982, Haggerty 1995, Ke & Lu 2009). The single nest of A. s subulatus that we observed with a downward-sloping entrance tunnel (nest 3), may reflect regional variation in architecture based on local microclimate or, alternatively, may have been an error by the adults or one forced by roots, rocks, or other obstructions within the substrate.

At present, we are unable to confirm that both sexes of Striped Woodhaunter participate in burrow excavation and nest construction, as is known for some species in the Automolus-Thripadectes-Clibanornis clade (see Cockle & Bodrati 2017). We can confirm, however, that both parents participate in incubation and chick provisioning. This behaviour is shared among most furnariids including all members of the Automolus-Thripadectes-Clibanornis clade studied to date (Remsen 2003a, Cockle & Bodrati 2017) but differs vs. other relatives in the Philydorini (sensu Derryberry et al. 2011), e.g., Ochre-breasted Foliagegleaner Anabacerthia lichtensteini and Sharp-billed Treehunter Heliobletus contaminatus, which have uniparental care (Cockle & Bodrati 2017). Although based on relatively small sample sizes, it appears that parental attendance during incubation may be higher in Striped Woodhaunter (63.5%) than has been reported for Chiriquí Foliage-gleaner (58.0%: Skutch 1952). A preliminary interpretation of these data might be that the shorter entrance tunnels of Striped Woodhaunter burrows, which may promote more rapid loss of heat within the nest (Ke & Lu 2009), may promote improved attendance. We suggest that further



information on adult attendance and nest design within this group may uncover interesting correlations. Furthermore, the seemingly rare occurrence of downward-inclined burrows in Striped Woodhaunter merits further investigation.

### Acknowledgements

We thank M. Corrales-Ugalde, J. Azofeifa-Solano, G. M Kirwan and K. Cockle for their comments that improved the manuscript; and A. Pérez 'Cope' for the illustration of the Costa Rican nest. For permitting access to their collections, we also thank the Departamento de Historia Natural, Museo Nacional de Costa Rica, and Museo de Zoología, Universidad de Costa Rica, especially C. Pineda, G. Alvarado, S. Bolaños and M. Springer. LS acknowledges the support provided by Vicerrectoría de Investigación, Universidad de Costa Rica, under project B9123 'Biología reproductiva de las aves tropicales: plasticidad fenotípica e historia natural'. HFG is grateful to Rudolphe A. Gelis, Eliot T. Miller and José Simbaña for their help with field work, to John V. Moore, Matt Kaplan, and the Population Biology Foundation for supporting his field work, and Pollywog Productions and Ryan Killackey for enabling his trip to Boanamo as part of the documentary feature film Yasuni Man.

#### References:

- Botero-Delgadillo, E. & Guayara, M. 2009. Primera descripción del Trepamusgos flamulado (Thripadectes flammulatus) en Colombia. Hornero 24: 103-106.
- Chesser, R. T., Burns, K. J., Cicero, C., Dunn, J. L., Kratter, A. W., Lovette, I. J., Rasmussen, P. C., Remsen, J. V., Stotz D. F., Winger B. M. & Winker K. 2018. Fifty-ninth supplement to the American Ornithological Society's Checklist of North American birds. Auk 135: 798-813.
- Claramunt, S., Derryberry, E. P., Cadena, C. D., Cuervo, A. M., Sanín, C. & Brumfield, R. T. 2013. Phylogeny and classification of Automolus foliage-gleaners and allies (Furnariidae). Condor 115: 375-385.
- Clements, J. F., Schulenberg, T. S., Iliff, M. J., Billerman, S. M., Fredericks, T. A., Sullivan, B. L. & Wood, C. L. 2019. The eBird Clements checklist of birds of the world: v2019. http://www.birds.cornell.edu/ clementschecklist/download/ (accessed 12 November 2019).
- Cockle, K. L. & Bodrati A. 2017. Divergence in nest placement and parental care of Neotropical foliagegleaners and treehunters (Furnariidae: Philydorini). J. Field Orn. 88: 336-348.
- Derryberry, E. P., Claramunt, S., Derryberry, G., Chesser, R. T., Cracraft, J., Aleixo, A., Perez-Emán, J., Remsen, J. V. & Brumfield, R. T. 2011. Lineage diversification and morphological evolution in a largescale continental radiation: the Neotropical ovenbirds and woodcreepers (Aves: Furnariidae). Evolution 65: 2973-2986.
- Ellis, J. H. 1982. Thermal nest environment and parental behavior of a burrowing bird, the Bank Swallow. Condor 84: 441-443.
- Euler, C. 1900. Descripção de ninhos e ovos das aves do Brasil. Rev. Mus. Paulista 3: 9-148.
- Faria, L. C. P., Carrara, L. A. & Rodrigues M. 2008. Biologia reprodutiva do fura-barreira Hylocryptus rectirostris (Aves: Furnariidae). Rev. Bras. Zool. 25: 172-181.
- Freeman, B. G. & Montgomery, G. A. 2017. Using song playback experiments to measures species recognition between geographically isolated populations: a comparison with acoustic trait analyses. Auk 134: 857-870.
- Greeney, H. F. 2017. Observations on the reproductive biology of Thrush-like Antpitta Myrmothera campanisona. Cotinga 39: 84-87.
- Greeney, H. F., Gualingua, D., Read, M., Medina, D., Puertas, C., Evans, L., Baihua, O. & Killackey, R. P. 2018. Rapid inventory, preliminary annotated checklist, and breeding records of the birds (Aves) of the Boanamo indigenous community, Orellana Province, Ecuador. Neotrop. Biodiver. 4: 10-44.
- Haggerty, T. 1995. Nest-site selection, nest design and nest-entrance orientation in Bachman's Sparrow. Southwest Natur. 40: 62-67.
- Hilty, S. L. 2003. Birds of Venezuela. Princeton Univ. Press.
- del Hoyo, J. & Collar, N. J. 2016. HBW and BirdLife International illustrated checklist of the birds of the world, vol. 2. Lynx Edicions, Barcelona.
- Irestedt, M., Fjeldså, J. & Ericson, P. G. P. 2006. Evolution of the ovenbird-woodcreeper assemblage (Aves: Furnariidae) - major shifts in nest architecture and adaptive radiation. J. Avian Biol. 37: 260-272.
- Ke, D. & Lu, X. 2009. Burrow use by Tibetan Ground Tits Pseudopodoces humilis: coping with life at high altitudes. Ibis 151: 321-331.
- Kiff, L. F., Marín, M. A., Sibley, F. C., Matheus, J. C. & Schmitt, N. J. 1989. Notes on the nest and eggs of some Ecuadorian birds. Bull. Brit. Orn. Cl. 109: 25-31.
- Maillard Z., O., Rocabado R., D. & Aguanta A., F. 2006. El nido y los polluelos de Philydor rufum, desde el subandino central de Bolivia. Kempffiana 2: 99–101.
- Marini, M. Â., Aguilar, T. M., Andrade, R. D., Leite, L. O., Anciães, M., Carvalho, C. E. A., Duca, C., Maldonado-Coelho, M., Sebaio, F. & Gonçalves, J. 2007. Biologia da nidificação de aves do sudeste de Minas Gerais, Brasil. Rev. Bras. Orn. 15: 367-376.



- Miller, E. T., Greeney, H. F., Lichter-Marck, I., Lichter-Marck, E. & Cabrera, F., L. E. 2012. The breeding of the Henna-hooded Foliage-gleaner (Hylocryptus erythrocephalus), with notes on conservation concerns. Orn. Neotrop. 23: 517-527.
- Pinto, O. M. O. 1953. Sobre a coleção Carlos Estevão de pieles, ninhos e ovos das aves de Belém (Pará). Pap. Avulsos Zool., São Paulo 11: 111-222.
- Proctor, N. & Lynch, P. 1993. Manual of ornithology. Yale Univ. Press, New Haven, CT.
- Remsen, J. V. 2003a. Family Furnariidae (ovenbirds). Pp. 162-357 in del Hoyo, J., Elliott, A. & Christie, D. A. (eds.) Handbook of the birds of the world, vol. 8. Lynx Edicions, Barcelona.
- Remsen, J. V. 2003b. Split Hyloctistes virgatus from H. subulatus. Proposal (#40) to South America Classification Committee. http://www.museum.lsu.edu/~Remsen/SACCprop40.htm.
- Ridgely, R. S. & Greenfield, P. J. 2001. The birds of Ecuador, vol. 1. Cornell Univ. Press, Ithaca, NY.
- Ridgely, R. S. & Tudor, G. 1994. The birds of South America, vol. 2. Univ. of Texas Press, Austin.
- Sánchez, J. E., Barrantes, G. & Durán, F. 2004. Distribución, ecología y conservación de la avifauna de la cuenca del río Savegre, Costa Rica. Brenesia 61: 63-93.
- Schultz, E. D., Burney, C. W., Brumfield, R. T., Polo, E. M., Cracraft, J. & Ribas C. C. 2017. Systematics and biogeography of the Automolus infuscatus complex (Aves; Furnariidae): cryptic diversity reveals western Amazonia as the origin of a transcontinental radiation. Mol. Phyl & Evol. 107: 503-515.
- Sclater, P. L. & Salvin, O. 1873. On the birds of eastern Peru. Proc. Zool. Soc. Lond. 1873: 252–311.
- Simon, J. E. & Pacheco, S. 2005. On standardization of nest descriptions of neotropical birds. Rev. Bras. Orn. 13: 143–154.
- Skutch, A. F. 1952. Life history of the chestnut-tailed Automolus. Condor 54: 93–100.
- Skutch, A. F. 1969. Life histories of Central American birds. III. Pacific Coast Avifauna 35: 1-580.
- Smith, C. & Londoño, G. A. 2013. Notes on the nests, eggs, incubation period, and nestling development of Thripadectes holostictus moderatus. Wilson J. Orn. 125: 222–226.
- Snethlage, E. 1935. Beiträge zur Fortpflanzungsbiologie brasilianischer Vögel. J. Orn. 83: 532–562.
- Stephens, L. & Traylor, M. A. 1983. An ornithological gazetteer of Peru. Mus. Comp. Zool., Cambridge, MA.
- Stiles, F. G. & Skutch, A. F. 1995. Guía de aves de Costa Rica. Instituto Nacional de Biodiversidad, Heredia.
- Strewe, R. 2001. Notes on nests and breeding activity of fourteen bird species from southwestern Colombia. Orn. Neotrop. 12: 265-269.
- Van Tyne, J. 1926. The nest of Automolus ochrolaemus pallidigularis Lawrence. Auk 43: 546.
- Zyskowski, K. J. & Greeney, H. F. 2010. Review of nest architecture in *Thripadectes* treehunters (Furnariidae) with descriptions of new nests from Ecuador. Condor 112: 176-182.
- Zyskowski, K. J. & Prum, O. 1999. Phylogenetic analysis of nest architecture of Neotropical ovenbirds (Furnariidae). Auk 116: 891-911.
- Addresses: Karla Conejo-Barboza, Vázquez de Coronado, Patalillo, San José, Costa Rica, 11104, e-mail: k.conejobarboza@gmail.com. César Sánchez, Asociación de Ornitólogos Unidos de Costa Rica, San José, Costa Rica, 11695-1000, and Dept. of Biological Sciences & Museum of Natural Science, Louisiana State University, Baton Rouge, LA 70803, USA. Luis Sandoval, Asociación de Ornitólogos Unidos de Costa Rica, San José, Costa Rica, 11695-1000, and Escuela de Biología, Universidad de Costa Rica, Montes de Oca, San José, Costa Rica, 11501-2090. Harold F. Greeney, Yanayacu Biological Station & Center for Creative Studies, Cosanga, Ecuador, e-mail: antpittanest@gmail.com