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Clonal plants' traits and responses following fire disturbance in *Pinus ponderosa* Lawson & C. Lawson ecosystems in the Colorado Front Range¹

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Abstract. Mountain ecosystems face increasing disturbance impacts (e.g., wildfire) due to climate change, leading to significant changes in vegetation composition and landscape dynamics. Clonal plants, characterized by their vegetative reproduction strategies, offer a range of ecologically important traits to cope with disturbances. However, clonality is often excluded in postfire vegetation studies in mountain ecosystems, and our knowledge of the ability of clonal plants to respond to differing disturbance regimes is limited. We studied the response of understory vegetation, based on composition and clonal traits, in burned and unburned *Pinus ponderosa* Lawson & C. Lawson (ponderosa pine) stands in the Colorado Front Range. Data from 40 total burned ($n = 20$) and unburned plots ($n = 20$) (10 m \times 10 m) included relative plant cover and five environmental variables. Burned plots had greater overall clonal plant cover and diversity compared to unburned plots. Although there were statistically distinct differences in species composition between burned and unburned plots, these differences may not be ecologically meaningful. A fourth-corner analysis showed relationships between elevation and slope and type of clonal growth organs, suggesting that elevation, slope, and fire history interact significantly in shaping vegetation community structure. Combined responses of lateral spread and ramet length of *Arctostaphylos uva-ursi* L. (kinnikinnick) differed significantly in areas impacted by wildfire to those unburned, whereas that of *Antennaria parvifolia* Nutt. (small-leaf pussytoes) showed no differences, suggesting a species-specific response of clonal traits following fire. Our findings indicate that clonal plants dominate the understory in burned *P. ponderosa* stands. We suggest managers and modelers account for plant clonal traits for a comprehensive understanding of postfire forest recovery.

Key words: community response, mountain ecosystems, plant clonality, vegetation dynamics, wildfire

Mountain ecosystems face various disturbances due to climate change, with wildfires being significant contributors (Hansen *et al.* 2001; IPCC 2023). Wildfires shape forest ecosystems, altering ecological processes, landscape dynamics, and vegetation structures (Johnston *et al.* 2016; Stevens-Rumann and Morgan 2019). Mixed-intensity fires can boost understory vegetation diversity by reducing fire-sensitive species and altering growing conditions (Keyser *et al.* 2008), and postfire tree mortality frees up resources and reduces competition for resilient understory vegetation, including clonal plants (Tubbesing *et al.* 2020). Although much research examines the response of vegetation to fire, few studies

focus on clonal plant response, even though we know clonal plants behave differently than nonclonal plants (Franklin *et al.* 2021). Knowing that clonal plants make up most species (Klimešová *et al.* 2021), we attempt to tease out the different response of clonal and nonclonal understory plants in Colorado's *Pinus ponderosa* Lawson & C. Lawson (ponderosa pine) forests.

Clonal plants, perennials that reproduce vegetatively through ramets, are initially dependent on the parent but subsequently develop their own root and shoot systems, forming genetically identical interconnected individuals (Franklin *et al.* 2020; Klimešová *et al.* 2021). These highly modular plants excel in rapid colonization, stress tolerance, and efficient resource sharing (Stuefer 1996), giving them an edge over nonclonal plants in some disturbed environments.

Clonal traits, such as clonal growth organs (CGOs) (e.g., epigeogenous and hypogeogenous rhizomes, bulbs, horizontal aboveground stems, and root splitters) support clonal plants' recovery postdisturbance. Clonal growth organs are structures that enable a plant to reproduce asexually and spread by producing new genetically identical individuals. Lateral spread and connection persistence

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are other clonal traits that also aid in recovery (Klimeš *et al.* 1997; Klimešová *et al.* 2019). After disturbances alter soil nutrients, rhizomes and stolons explore nutrient-rich patches, colonizing bare ground through horizontal growth (Klimeš *et al.* 1997). This lateral spread helps clonal plants compete for resources postdisturbance (Pausas *et al.* 2018). Herben *et al.* (2012) highlighted a trade-off between vegetative and seed reproduction, with varying reproduction methods benefiting clonal plants based on environmental conditions, but in general, clonal plants do well in heterogenous conditions (Louâpre *et al.* 2012; Liu *et al.* 2016) typical of postfire environments (Schoennagel *et al.* 2008; Das Gupta *et al.* 2019).

Wildfires shape vegetation patterns, making understanding plant responses vital, especially clonality. Exploring vegetation recovery trajectories helps predict resilience in disturbed mountain ecosystems (Viana-Soto *et al.* 2020; Seidl and Turner 2022), guiding effective forest management and conservation efforts (Hart and Chen 2006). Although studies have explored understory responses to wildfires in *P. ponderosa* forests, few focus on clonality's role in postfire recovery. Our goal was to fill this gap by studying postfire responses and clonal plant traits in burned and unburned *P. ponderosa* plots. Our objectives were to (a) compare clonal plant cover and diversity in burned *versus* unburned plots; (b) examine plant species composition changes in these plots, considering clonal *versus* nonclonal contributions; and (c) examine clonal traits (lateral spread, ramet length) of *Arctostaphylos uva-ursi* L. (kinnikinnick) and *Antennaria parvifolia* Nutt. (small-leaf pussytoes) across burned and unburned plots.

We hypothesized that burned plots would show greater clonal plant percent cover and diversity due to their adaptability in disturbed conditions and rapid recolonization ability. Thus, we anticipated significant community composition differences. Clonal plants' unique vegetative spread and recovery mechanisms should be more evident postfire. Clonal traits like lateral spread and ramet length in *A. uva-ursi* and *A. parvifolia* can indicate how these plants respond to disturbances (Klimešová *et al.* 2019). Lateral spread shows horizontal expansion and resource acquisition, whereas ramet length reflects vertical growth, highlighting clonal plant fitness and growth strategies (Klimešová and Klimeš 2008). We also hypothesized significant

differences in these clonal traits between burned and unburned plots, suggesting plastic responses to the postfire environment.

Methods. STUDY AREA. This study was conducted in *P. ponderosa* forests of the Arapaho and Roosevelt National Forests (ARNF) in the Colorado Front Range after the 2020 Cameron Peak Fire, which is considered one of the largest wildfires in the history of Colorado (NPS 2021) (Fig. 1). The ARNF covers about 1.5 million acres and is primarily coniferous. *Pinus ponderosa* dominates at elevations between 1,800 and 2,500 m, accompanied by understory species such as graminoids, forbs, and shrubs (Fryer 2018). Other conifers like *Pinus contorta* Douglas ex Loudon (lodgepole pine), *Pseudotsuga menziesii* (Mirb.) Franco (Douglas-fir), *Pinus flexilis* James (limber pine), and *Juniperus scopulorum* Sarg. (Rocky Mountain juniper) appear at mid-high elevations (2000–2500 m). Plots receive an average annual precipitation of 38.3 cm, with temperatures varying from 2 °C in January to 32 °C in June (data from 1991 to 2020; Western Regional Climate Center 2007).

FIELD AND UNDERSTORY VEGETATION MEASUREMENTS. Sampling area was delineated and informed based upon the combination of historical fire data in the Colorado Front Range and on-site inspection. Prior to onfield visit, we reviewed historical data on the Cameron Peak wildfire to provide guidance on sampling locations and understand the spatial extent and severity of the fire (ArcGIS 2023). We then systematically drove through *P. ponderosa* stands within the potential sampling area to inspect potential burned and unburned plots. Based upon onfield inspection, the 20 burned plots were sampled by assessing *P. ponderosa* stands that had experienced complete canopy fire consumption during the wildfire event. Similarly, the 20 unburned plots used as controls were sampled plots that had not experienced fire damage in at least the last two decades (Fig. 2). Although sampling techniques relied on visual assessment during field visits, the locations of plots were recorded using GPS devices (Garmin eTrex 32x, Garmin Ltd., Olathe, KS) to ensure accurate locations of plots. Plot sampling was conducted in a systematic manner to minimize bias and ensure representation across the study area. Each of the 40 total burned and unburned plots was 10 m × 10 m, with plots at least 50 m apart. In every

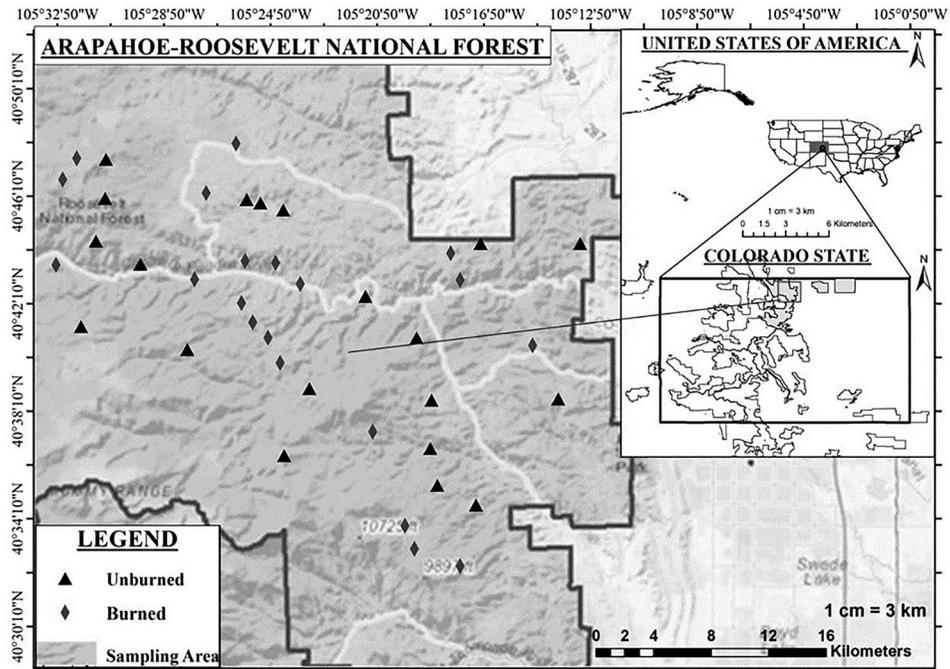


FIG. 1. Map of the 40 plots sampled from understory vegetation distributed within burned ($n = 20$) and unburned ($n = 20$) *Pinus ponderosa* stands in the Colorado Front Range.

sampled plot, we took the following environmental data from the plot center: elevation, slope, aspect, longitude, and latitude (Appendix 1).

Past studies assert that using plant cover data effectively characterizes plant communities, describes plant-environment interactions, and monitors plant

ecosystems through time (Mueller-Dombois and Ellenberg 1974; Anderson 1986). Visual estimates of relative percent cover were done using the ocular method of estimating percent cover for all plant species in each plot, both clonal and nonclonal (Helm and Mead 2004). We estimated percent

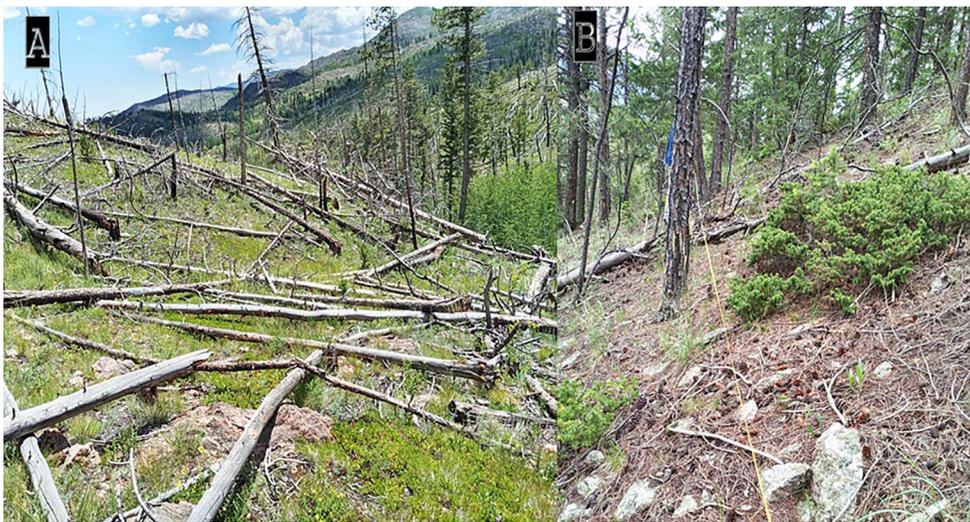


FIG. 2. Photograph of the study area showing burned (A) and unburned (B) *Pinus ponderosa* stands in the Colorado Front Range.

cover of each plant species separately within each plot by pooling plant species abundances before calculating relative percent cover. The choice of percent cover over an ordinal cover scale is supported by a recent study by Dengler and Dembicz (2023). In their study, the authors found that direct percent cover estimation produces lower mean absolute and relative errors compared to the use of ordinal scales, particularly for cover values above 1% (Dengler and Dembicz 2023). If a plant could not be identified, we followed the Assessment Inventory and Monitoring methods (Toews *et al.* 2011) and initially named that plant as an unknown species. The plant was collected, with detailed descriptions and photos, brought to the University of Northern Colorado herbarium to be keyed and identified, and once identified, reclassified to species in the data set. Taxonomic identification followed Ackerfield (2015). CLO-PLA 3 (Klimeš and Klimešová 1999; Klimešová and Klimeš 2008), and TRY plant trait databases (Kattge *et al.* 2020) were used to label species as clonal or nonclonal based on their dominant CGO such as epigeogenous rhizome, hypogeogenous rhizome, bulb, root splitter, and horizontal aboveground stem. In addition, the USDA PLANTS Database (USDA 2024) was used to provide other ecological traits of species, including life form (forb, shrub or graminoid) and the photosynthetic pathway that each species utilizes (C3, C4, or CAM).

CLONAL TRAITS MEASUREMENTS. Ten plots each from burned and unburned areas were selected to study clonal traits' responses in *A. uva-ursi* and *A. parvifolia*. These plots were chosen based on their distribution. We aimed to compare lateral spread and ramet length between burned and unburned plots, as these traits indicate clonal reproduction and recovery postdisturbance (Klimeš and Klimešová 2008; Pausas *et al.* 2018). Measurements followed Klimešová *et al.* (2019) protocols, with lateral spread measured between offspring and rooting units, and ramet length from base to tip. Five measurements per plant were used to calculate an average, and five individuals were averaged for each plot.

DATA ANALYSES. We were primarily interested in assessing the relative percent cover (i.e., cover of all species in each plot summed to 100%) and diversity of clonal plants in burned and unburned plots of *P. ponderosa* stands. Diversity variables (i.e., richness and evenness) were calculated using

relative cover of full species matrix for 40 total plots with 27 clonal taxa/species in PC-ORD. Since percent cover and diversity data did not meet assumptions of normality and homoscedasticity of variances, unpaired nonparametric Mann-Whitney U-tests were conducted to assess whether each of our response variables—relative percent cover, richness, and evenness—differed between burned *versus* unburned plots for clonal plants. For clonal plant trait measurements, a MANOVA (Multivariate Analysis of Variance) was used to detect the effects of burned *versus* unburned plots on lateral spread and ramet length for each clonal plant *A. uva-ursi* and *A. parvifolia* in a combined model. Although lateral spread and ramet length may respond differently, analyzing them together allowed for a comprehensive assessment of trait responses. All analyses were conducted using R Statistical Software (version 4.3.1; R Core Team 2021, Vienna, Austria).

PC-ORD 7.10 (McCune and Mefford 1999) was used to conduct multivariate analyses to study species composition differences in burned and unburned plots and the roles of clonal and nonclonal plants. We used four multivariate analyses: NMS (nonmetric multidimensional scaling), MRPP (multi-response permutation procedures), ISA (indicator species analysis), and FCA (fourth-corner analysis) to explore trait-environment relationships.

Nonmetric multidimensional scaling ordination points (burned and unburned plots in our case) in a reduced-dimensional space, preserving the pairwise dissimilarity among them. This helps to visualize patterns and relationships in ecological communities (McCune and Grace 2002). Nonmetric multidimensional scaling was used because the data did not follow linear, parametric assumptions for normality and homogeneity of variances, which is common with ecological community data (Clarke 1993; McCune and Grace 2002). Autopilot with the “Slow and Thorough” setting using “Sørensen distance” measure was used to run 50 iterations to create a configuration for the ordination including 104 clonal and nonclonal taxa/species in the primary species matrix and 40 total burned and unburned plots (Appendix 2). The second matrix had five environmental variables collected at each plot from the center (elevation, slope, aspect, latitude, and longitude). Multi-response permutation procedures and ISA were performed to complement and help interpret the NMS ordination.

Multi-response permutation procedures is a non-parametric statistical test that determines whether there are significant differences between groups based on multivariate response variables (Pillar 2013). In this case, we used MRPP to assess the dissimilarity in species composition between burned and unburned plots using the “Sørensen distance” measure. Based on the weighting technique, the analysis can generate the *T* statistic, which measures the difference between groups, with more negative values indicating stronger separation; the *A* statistic, which reflects within-group homogeneity, ranging from -1 (no agreement) to 1 (perfect agreement); and *P*, which is the probability of smaller or equal delta (McCune and Mefford 1999).

Indicator species analysis was used to determine which plants were significantly associated with burned and unburned plots. The analysis helps to determine which species (clonal or non-clonal) are indicators of these specific environmental conditions (Dufrière and Legendre 1997; McCune and Grace 2002).

The fourth-corner analysis explores species’ trait-environment relationships. It is called the “fourth-corner” because it combines three corners of species ecological traits, environment variables, and species cover data, and then leaves the fourth corner to examine the relationship among species, environmental variables, and plant ecological traits (Dray et al. 2014). Specifically in this study, FCA was used to determine whether environmental variables had any relationship with species ecological traits and with differences in species composition observed in the NMS ordination.

Results. RELATIVE PERCENT COVER AND DIVERSITY AREA OF CLONAL PLANTS. We identified 104 understory species (27 clonal and 77 nonclonal) in all 40 burned and unburned plots. Invasive species were rare in all plots. Clonal plants (Table 1) and nonclonal plants were mainly forbs, with a few shrubs and graminoids. Both plot types had an overstory dominated by *P. ponderosa* but included *P. flexilis* and *P. menziesii* to a lesser extent. *Achillea millefolium* L. (common yarrow), *Campanula rotundifolia* L. (bluebell bellflower), and *A. parvifolia* showed the greatest cover among forbs. *Arctostaphylos uva-ursi*, *Artemisia ludoviciana* Nutt. (white sagebrush), *Artemisia frigida* Willd. (prairie sagewort), *Cercocarpus montanus* Raf. (alder-leaf mountain mahogany), and *Purshia tridentata* (Pursh.) D.C. (antelope bitterbrush) were the

shrubs with the most cover. In terms of graminoids, *Carex rossii* Boott (Ross’ sedge), *Bromus tectorum* L. (cheatgrass), and *Poa bulbosa* L. (bulbous bluegrass) were the species with the greatest cover in our plots.

Clonal plant percent cover differed significantly between burned and unburned plots ($U = 332$; $p < 0.001$), being greater in burned plots (Fig. 3). Burned plots averaged 12 species, whereas unburned had 6, with standard deviations of 1.883 and 1.071, respectively. Clonal plant richness was significantly higher in burned plots ($U = 397$; $p < 0.001$; Fig. 4), but evenness did not differ between plot types ($U = 203.5$; $p = 0.935$; Fig. 5).

COMMUNITY STRUCTURE. Species community composition in burned and unburned plots using NMS ordination yielded a three-dimensional solution explaining 64.3% of the variance ($p = 0.004$). The final stress level was 18.11 with a solution instability of < 0.0001 after 102 iterations. Non-metric multidimensional scaling axis 1 explained 25.2% of the variance, capturing a significant portion of species composition variation. The positive end of axis 1 correlated with higher *C. rossii*, *Pascopyrum* Å. Löve (wheatgrass), *Helianthus pumilus* Nutt. (little sunflower), *P. bulbosa*, and *Mertensia ciliata* E. James ex. Torr G. Don (tall fringed bluebells). Conversely, the negative end of axis 1 correlated with *Cirsium arvense* (L.) Scop. (Canada thistle), *Geranium* sp. (geranium), and *A. millefolium*. The only environmental variable that was positively correlated with NMS axis 1 was slope, whereas elevation and latitude were negatively correlated. Plots formed distinct burned and unburned groups in ordination space (Fig. 6).

Nonmetric multidimensional scaling axis 2 contributed an additional 23.4% of the variance. Species strongly correlated with the positive end of axis 2 included *Taraxacum officinale* F. H. Wigg. (common dandelion), *Potentilla fissa* Nutt. (big-flower cinquefoil), and *C. rossii*. *Viburnum edule* (Michx.) Raf. (squashberry), *C. rotundifolia*, and *A. millefolium* showed weaker positive correlations with axis 2. Species negatively correlated with this axis included *Erigeron eximius* Greene (sprucefir fleabane), *Sedum lanceolatum* Torr. (spearleaf stonecrop), *P. bulbosa*, and to a lesser extent, *Draba graminea* Greene (Rocky Mountain draba). All environmental variables had weak

Table 1. Clonal plants with dominant clonal growth organs (CGOs) in sampled plots of the *Pinus ponderosa* stands in the Colorado Front Range (from CLO-PLA 3 & TRY databases).

| Species | CGO |
|---|---|
| <i>Achillea millefolium</i> L. | epigeogenous stem (rhizome) ¹ |
| <i>Antennaria parvifolia</i> Nutt. | epigeogenous stem (rhizome) |
| <i>Arctostaphylos uva-ursi</i> L. | epigeogenous stem (rhizome) |
| <i>Bromus inermis</i> Leyss. | hypogeogenous stem (rhizome) ² |
| <i>Campanula rotundifolia</i> L. | epigeogenous stem (rhizome) |
| <i>Cerastium arvense</i> L. | root-splitter ³ |
| <i>Carex rossii</i> Boott | epigeogenous stem (rhizome) |
| <i>Cirsium arvense</i> (L.) Scop. | roots with adventitious buds ⁴ |
| <i>Chenopodium album</i> L. | horizontal aboveground stem ⁵ |
| <i>Cystopteris fragilis</i> (L.) Bernh. | epigeogenous stem (rhizome) |
| <i>Draba graminea</i> Greene | root-splitter |
| <i>Koeleria macrantha</i> (Ledeb.) Schult. | root-splitter |
| <i>Oxytropis campestris</i> (L.) DC. | bulb ⁶ |
| <i>Phleum pratense</i> L. | epigeogenous stem (rhizome) |
| <i>Pseudocymopterus montanus</i> (A. Gray) J.M. Coult. & Rose | epigeogenous stem (rhizome) |
| <i>Poa bulbosa</i> L. | epigeogenous stem (rhizome) |
| <i>Poa pratensis</i> L. | hypogeogenous stem (rhizome) |
| <i>Pulsatilla patens</i> (L.) Mill. | epigeogenous stem (rhizome) |
| <i>Purshia tridentata</i> (Pursh) DC. | epigeogenous stem (rhizome) |
| <i>Ribes aureum</i> Pursh | roots with adventitious buds |
| <i>Rubus idaeus</i> L. | epigeogenous stem (rhizome) |
| <i>Salix</i> L. | horizontal above-ground stem |
| <i>Symphoricarpos orbiculatus</i> Moench | epigeogenous stem (rhizome) |
| <i>Taraxacum officinale</i> F.H. Wigg. | root-splitter |
| <i>Thalictrum alpinum</i> L. | epigeogenous stem (rhizome) |
| <i>Tragopogon dubius</i> Scop. | epigeogenous stem (rhizome) |
| <i>Vicia sativa</i> L. | epigeogenous stem (rhizome) |

¹ An organ originating from stems that grows aboveground.

² A clonal growth organ formed underground.

³ A plant whose primary root system splits into buds and ramets without the need for secondary roots.

⁴ A plant's main roots, including the hypocotyl and adventitious roots, which can develop on their own or in response to an injury.

⁵ A clonal growth organ rooting in the soil and providing connection between offspring plants or formed by a creeping axis of a plant.

⁶ A short underground stem surrounded by fleshy leaves or leaf bases, storing nutrients.

correlations with axis 2. However, the unburned plots tended to cluster at the negative end of axis 2, whereas burned plots clustered toward the positive end of this axis.

The third NMS axis accounted for 13.4% of the explained variance. The positive end of axis 3 had strong correlations with *Castilleja puberula* Rydb. (shortflower Indian paintbrush) and *V. edule*. The negative end was strongly correlated with *A. ludoviciana*, *P. tridentata*, *Heterotheca villosa* (Pursh) Shinners (hairy false goldenaster), *H. pumilus*, *Bromus inermis* Leyss. (smooth brome), and *C. montanus*. Slope was strongly negatively correlated, whereas elevation, aspect, longitude, and longitude all had negative correlations.

Multi-response permutation procedures ($T = -9.970$, $A = 0.021$, $p \leq 0.001$) analysis revealed

statistically significant differences in vegetation composition between burned versus unburned plots, albeit a low A value questions ecological significance. Indicator species analysis identified 16 indicator species for both burned and unburned plots out of the 104 taxa/species in the species matrix ($p \leq 0.05$) with 63% clonal indicating burned plots and one clonal species indicating unburned plots (Table 2).

Fourth-corner analysis revealed no significant relationships between elevation, slope, latitude, longitude, and life form. Environmental variables had weak correlations with clonality (all $p > 0.05$). However, CGOs were associated with elevation and slope. Specifically, elevation related to horizontal aboveground stems, epigeogenous rhizomes, and root splitters ($p = 0.025$, $p = 0.019$, and $p =$

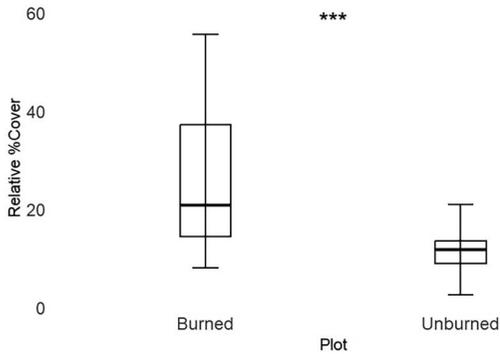


FIG. 3. Relative percent cover of clonal plants from burned versus unburned *Pinus ponderosa* stands in the Colorado Front Range.

0.032, respectively), whereas slope related to epigeogenous rhizomes ($p = 0.023$). Environmental variables did not relate with photosynthetic type (all $p > 0.05$). Overall, elevation, slope, and fire history significantly influenced vegetation structure, consistent with NMS ordination results (Table 3).

CLONAL TRAITS RESPONSE IN BURNED AND UNBURNED PLOTS. MANOVA results suggested that the combined responses of lateral spread (LS) and ramet length (RL) of *A. urva-ursi* were significantly higher in burned plots compared to unburned plots (Pillai Trace $F = 0.5624$, $p < 0.001$; Fig. 7). Although these traits in *A. urva-ursi* varied in plasticity after fire in burned plots, unburned plots showed less variability. In contrast, *A. parvifolia* showed no significant differences in LS or RL between burned and unburned plots (Pillai Trace $F = 0.1042$, $p = 0.393$; Fig. 8), with unburned plots also displaying less trait variability.

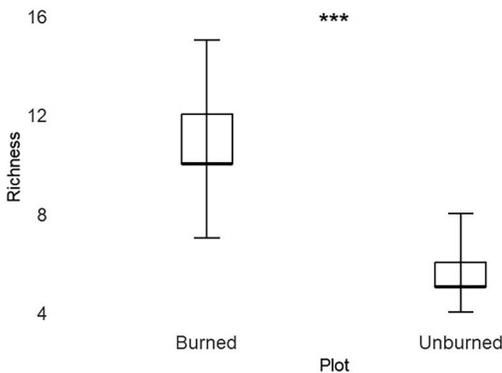


FIG. 4. Richness of clonal plants from burned versus unburned *Pinus ponderosa* stands in the Colorado Front Range.

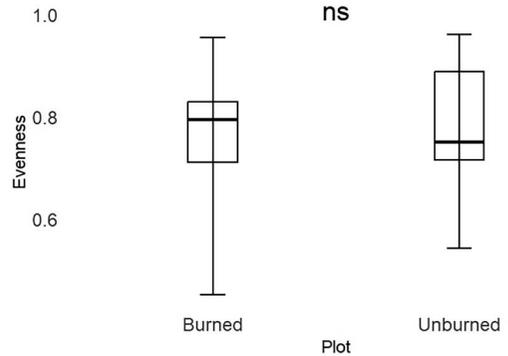


FIG. 5. Evenness of clonal plants from burned versus unburned *Pinus ponderosa* stands in the Colorado Front Range.

Discussion. RELATIVE PERCENT COVER AND DIVERSITY OF CLONAL PLANTS. Our study focused on the response of understory clonal plants in *P. ponderosa* stands in the Colorado Front Range following fire. We expected burned plots to show greater clonal plant cover and diversity than unburned plots. We also predicted that the different responses of clonal and nonclonal plants would result in significant changes in overall species composition between burned and unburned plots. In addition, we hypothesized that clonal traits of *A. urva-ursi* and *A. parvifolia* would differ

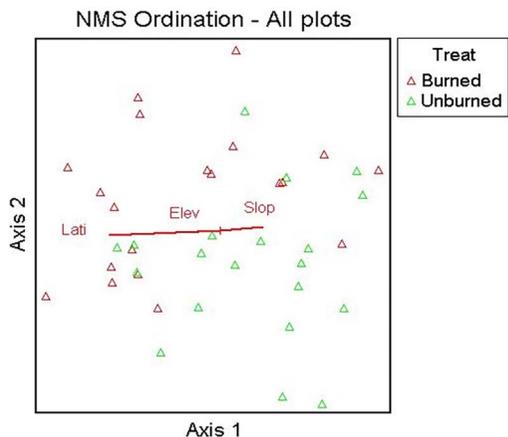


FIG. 6. NMS ordination of the understory species composition for all 40 burned and unburned plots of *Pinus ponderosa* stands in the Colorado Front Range. A three-dimensional solution was recommended, but only two dimensions are shown here. Burned plots are marked by red triangles, whereas unburned plots are marked by light green triangles with five environmental variables (elevation, slope, aspect, latitude, and longitude).

Table 2. Indicator species analysis (ISA) results for species across study plots from *Pinus ponderosa* stands in the Colorado Front Range.

| Species | Plant type | Plot indicated | Observed indicator value | P-value |
|--|------------|----------------|--------------------------|---------|
| <i>Achillea millefolium</i> L. | Clonal | Burned | 80.0 | 0.0002 |
| <i>Campanula rotundifolia</i> L. | Clonal | Burned | 67.0 | 0.0004 |
| <i>Purshia tridentata</i> (Pursh) DC | Clonal | Burned | 55.0 | < 0.001 |
| <i>Oxytropis campestris</i> (L.) DC | Clonal | Burned | 45.0 | 0.002 |
| <i>Taraxacum officinale</i> F.H. Wigg. | Clonal | Burned | 37.3 | 0.041 |
| <i>Cirsium arvense</i> (L.) Scop. | Clonal | Burned | 35.0 | 0.009 |
| <i>Poa pratensis</i> L. | Clonal | Burned | 35.0 | 0.007 |
| <i>Rubus idaeus</i> L. | Clonal | Burned | 25.0 | 0.023 |
| <i>Thalictrum alpinum</i> L. | Clonal | Burned | 25.0 | 0.045 |
| <i>Mertensia ciliata</i> (James ex Torr.) G. Don | Nonclonal | Burned | 25.0 | 0.043 |
| <i>Potentilla fissa</i> Nutt. | Nonclonal | Burned | 66.8 | 0.001 |
| <i>Artemisia ludoviciana</i> Nutt. | Nonclonal | Burned | 48.6 | 0.046 |
| <i>Festuca saximontana</i> Rydb. | Nonclonal | Burned | 40.0 | 0.002 |
| <i>Bromus madritensis</i> L. | Nonclonal | Burned | 30.0 | 0.020 |
| <i>Verbascum thapsus</i> L. | Nonclonal | Burned | 45.0 | 0.001 |
| <i>Cystopteris fragilis</i> (L.) Bernh. | Clonal | Unburned | 25.0 | 0.048 |

in their clonal traits' responses between burned and unburned plots. Overall, the findings partially supported our hypotheses. Although burned plots did show increased clonal plant cover and diversity as predicted, the changes in species composition may not be ecologically significant. Additionally, the clonal traits responses of the two studied plants did not differ much between burned and unburned plots.

Several studies indicate that plant clonality plays a key role in ecosystem recovery and community

restoration after disturbances (e.g., Fahrig *et al.* 1994; Oborny *et al.* 2000; Van Mantgem *et al.* 2020). Here, we found greater clonal plant cover in burned plots, attributing this to their resilience, colonization ability, and vegetative reproduction from strong bud banks and CGOs like rhizomes and stolons (Klimešová and Martínková 2022). In addition, postfire environments often exhibit increased heterogeneity, affecting soil, microclimate, and vegetation at broader scales (Schoennagel *et al.* 2008). Although our study focused on community-level effects, clonal plants thrive in these varied postfire landscapes, benefiting from quick regrowth and

Table 3. Results of fourth-corner analysis (FCA) examining the relationship of environmental variables with species ecological traits from *Pinus ponderosa* stands in the Front Range of Colorado.

| Environmental variable | Species ecological trait | F value | P value |
|------------------------|--------------------------|---------|---------|
| Elevation | clonality | 0.745 | 0.837 |
| | life form | 7.734 | 0.632 |
| | CGOs | 35.401 | 0.002 |
| Slope | photosynthetic type | 5.812 | 0.319 |
| | clonality | 0.71 | 0.822 |
| | life form | 5.174 | 0.698 |
| Aspect | CGOs | 23.21 | 0.028 |
| | photosynthetic type | 2.633 | 0.433 |
| | clonality | 4.994 | 0.380 |
| Latitude | life form | 4.499 | 0.436 |
| | CGOs | 5.384 | 0.601 |
| | photosynthetic type | 2.182 | 0.437 |
| Longitude | clonality | 1.544 | 0.753 |
| | life form | 9.643 | 0.372 |
| | CGOs | 11.218 | 0.272 |
| Longitude | photosynthetic type | 3.712 | 0.465 |
| | clonality | 4.676 | 0.395 |
| | life form | 8.676 | 0.127 |
| Longitude | CGOs | 4.281 | 0.724 |
| | photosynthetic type | 3.647 | 0.330 |

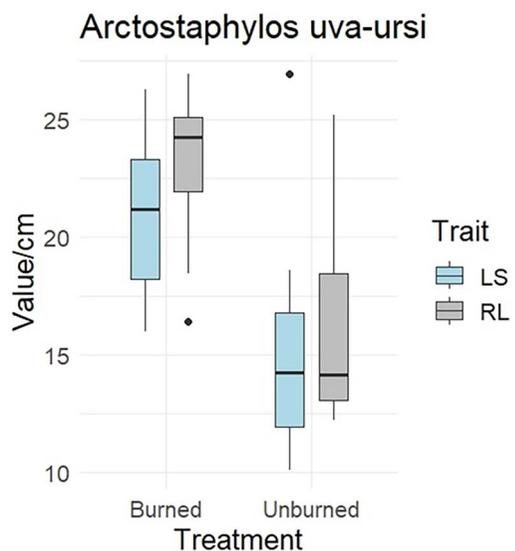


FIG. 7. Response of lateral spread and ramet size of *Arctostaphylos uva-ursi* for burned versus unburned *Pinus ponderosa* stands in the Colorado Front Range.

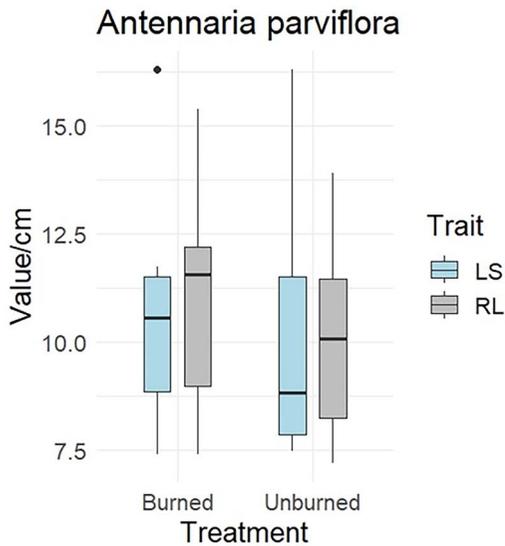


FIG. 8. Response of lateral spread and ramet size of *Antennaria parvifolia* for burned versus unburned *Pinus ponderosa* stands in the Colorado Front Range.

resource exploitation (Svensson *et al.* 2013). This increased cover is also linked to traits like physiological integration and resource allocation, crucial for clonal plant dominance in disturbed ecosystems (Liu *et al.* 2016).

Our data suggest that clonal plant species richness increases postfire. Clonal plants had more species in burned plots compared to unburned ones, consistent with other studies (e.g., McMahon *et al.* 2017; Pausas *et al.* 2018). However, clonal plant evenness did not differ between burned and unburned plots. Similar evenness suggests fire does not change the relative abundance of clonal species. This evenness could influence community stability and resilience postfire, affecting recovery ability (Sasaki and Lauenroth 2011; Battisti *et al.* 2016).

COMMUNITY STRUCTURE. Nonmetric multidimensional scaling ordination axis 1 primarily represents elevation gradient in vegetation community structure, likely influenced by topography, especially in unburned ponderosa pine plots. On the positive end of axis 1, species such as *C. rossii*, *Pascopyrum* sp., *H. pumilus*, *P. bulbosa*, and *M. ciliata* were associated at higher elevations, indicating cooler conditions. These species likely thrive in cooler conditions typical of higher elevations in the unburned ponderosa communities. The negative end of axis 1 was correlated with *C. arvense*, *Geranium* sp., and

A. millefolium linked to lower elevations, possibly due to the dense *P. ponderosa* canopy cover. Although elevation emerges as the main driver of species along axis 1, particularly in unburned plots, there might be an interaction between elevation and fire history in shaping vegetation community structure.

Nonmetric multidimensional scaling axis 2 represents a postfire disturbance and species-specific response gradient. Burned plots mainly cluster at the positive end, including species such as *T. officinale*, *C. rossii*, and *P. fissa*, which can thrive postfire due to clonality or wind-dispersed seeds (Toth 1991). These species benefit from postfire microhabitats and establish quickly. On the negative end of axis 2, species like *E. eximius*, *S. lanceolatum*, *P. bulbosa*, and *D. graminea* apparently preferred less disturbed habitats. Further, unburned plots mostly clustered at the negative end of this axis and we believe fire regimes in the Colorado Front Range likely influenced these patterns. It should be noted that “unburned” really implies burned less recently, as all plots have burned at some time in this *P. ponderosa* ecosystem.

Nonmetric multidimensional scaling axis 3 corresponds to variation in species cover related to postfire vegetation structure. The positive end had only *C. puberula* and *V. edule*, which are not indicative of postfire environments in the literature. However, at the negative end, we saw species with greater cover commonly found in postfire environments. These species included *A. ludoviciana*, *P. tridentata*, *H. villosa*, *B. inermis*, and *C. montanus*. Species cover along this axis also correlated negatively with slope, indicating slope’s role in shaping species distribution. Steeper slopes might experience different fire behavior, affecting postfire community structure (Bassett *et al.* 2015).

The data suggests that there are slight compositional differences between burned and unburned plots, but we interpret these cautiously. Indicator species analysis identified 16 significant indicator species, but only three had values above 60. This low indicator value is likely due to many species being widespread across plots. Clonal plants like *A. millefolium*, *C. rotundifolia*, *C. arvense*, *Oxytropis campestris* (L.) DC. (field locoweed), *P. tridentata*, *T. officinale*, and *Poa pratensis* L. (Kentucky bluegrass) were significant indicators in burned plots, aligning with our expectation that clonal plants respond more directly to fire disturbance. This supports the idea that clonal plants enhance postfire ecosystem resilience (Falk *et al.* 2019).

We found no relationships between clonality, life form, photosynthetic type, and environmental variables like elevation, slope, aspect, longitude, and latitude. This aligns with our prediction that these traits are relatively stable across species regardless of environmental variations. However, responses within these traits can vary; for example, clonal plants may exhibit diverse strategies postdisturbance.

Significantly, elevation and slope were related to certain CGOs, like horizontal aboveground stems and rhizomes. This suggests clonal plants allocate resources to these CGOs more at higher elevations or steeper slopes, possibly enhancing their ability to spread and access resources postdisturbance (Štöastná *et al.* 2012). Elevation also strongly correlated with NMS axis 1 and was related to CGOs in the FCA. This indicates that elevation and fire history interact to shape vegetation in these Colorado Front Range ecosystems, suggesting a nuanced response to environmental conditions (Klimešová *et al.* 2012).

CLONAL TRAITS RESPONSE IN BURNED AND UNBURNED PLOTS. The responses of clonal plant species *A. uva-ursi* and *A. parvifolia* to fire varied, highlighting the importance of understanding each species' unique postdisturbance behavior (Franklin *et al.* 2020; Bell *et al.* 2023). Specifically, *A. uva-ursi* exhibited increased lateral spread and ramet length in burned plots, potentially aiding its recovery and growth postfire. Although few studies focus on these species' responses to fire, Hutchings and de Kroon (1994) found that clonal traits like lateral spread can vary based on environmental conditions and genetics. This trait plasticity in response to disturbance is vital for resource competition and foraging (Dong and de Kroon 1994; Hutchings and de Kroon 1994; Dietz and Steinlein 2001). Such variability has practical implications for ecosystem management, especially in fire-prone areas. It underscores the need for tailored management plans that account for each species' specific needs postfire.

In summary, this study highlights that elevation, slope, and fire history collectively influence the understory vegetation of *P. ponderosa* stands in the Colorado Front Range. Three years after the fire, burned plots showed increased relative cover and diversity of clonal plants, displaying their resilience and adaptability to disturbances. Our findings are important for foresters, managers, and modelers to understand clonal plant performance and response

to changing fire regimes. Given the importance of clonal plants in postdisturbance recovery, future research should delve into the mechanisms behind their success. Long-term monitoring of postfire ecosystems will further elucidate the sustainability of clonal plants and their traits.

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Appendix 1

Elevation, slope, aspect, and GPS coordinates of plot locations in study area.

| Plot number | Plot type | Elevation | Slope | Aspect | Latitude | Longitude |
|-------------|-----------|-----------|-------|--------|----------|-----------|
| 1 | Burned | 1927 | 30 | 50 | 40.7927 | -105.54 |
| 2 | Burned | 1918 | 30 | 49 | 40.7797 | -105.54 |
| 3 | Burned | 1910 | 33 | 49.5 | 40.7713 | -105.45 |
| 4 | Burned | 1901 | 34 | 52 | 40.7339 | -105.30 |
| 5 | Burned | 1908 | 36 | 40 | 40.7267 | -105.55 |
| 6* | Burned | 2458 | 32 | 190 | 40.7178 | -105.46 |
| 7* | Burned | 2416 | 35 | 180 | 40.6660 | -105.41 |
| 8* | Burned | 2428 | 30 | 230 | 40.7530 | -105.39 |
| 9* | Burned | 2425 | 15 | 290 | 40.5650 | -105.33 |
| 10* | Burned | 2416 | 25 | 270 | 40.6769 | -105.25 |
| 11 | Burned | 2420 | 25 | 50 | 40.7173 | -105.29 |
| 12 | Burned | 2434 | 20 | 170 | 40.7280 | -105.41 |
| 13 | Burned | 2409 | 35 | 242 | 40.6233 | -105.35 |
| 14 | Burned | 2391 | 25 | 190 | 40.5401 | -105.29 |
| 15* | Burned | 2487 | 10 | 110 | 40.5508 | -105.32 |
| 16* | Burned | 2490 | 25 | 110 | 40.6816 | -105.42 |
| 17 | Burned | 2484 | 20 | 50 | 40.7030 | -105.43 |
| 18* | Burned | 2488 | 17 | 60 | 40.7149 | -105.39 |
| 19* | Burned | 2472 | 8 | 110 | 40.6912 | -105.43 |
| 20* | Burned | 2477 | 28 | 286 | 40.7292 | -105.43 |
| 21* | Unburned | 2103 | 30 | 326 | 40.7922 | -105.52 |
| 22* | Unburned | 2187 | 29 | 290 | 40.7679 | -105.52 |
| 23 | Unburned | 2230 | 31 | 310 | 40.7672 | -105.43 |
| 24 | Unburned | 2313 | 33 | 299 | 40.7651 | -105.42 |
| 25* | Unburned | 2352 | 36 | 302 | 40.7608 | -105.41 |
| 26 | Unburned | 2012 | 10 | 76 | 40.7415 | -105.52 |
| 27 | Unburned | 1995 | 25 | 25 | 40.7272 | -105.50 |
| 28 | Unburned | 1986 | 30 | 90 | 40.6501 | -105.39 |
| 29 | Unburned | 1953 | 34 | 84 | 40.7401 | -105.28 |
| 30 | Unburned | 1881 | 35 | 10 | 40.7073 | -105.36 |
| 31* | Unburned | 2486 | 15 | 28 | 40.5780 | -105.29 |
| 32 | Unburned | 2475 | 4 | 72 | 40.5902 | -105.31 |
| 33* | Unburned | 2473 | 10 | 254 | 40.6087 | -105.40 |
| 34* | Unburned | 2471 | 12 | 340 | 40.6887 | -105.53 |
| 35* | Unburned | 2477 | 10 | 104 | 40.6744 | -105.47 |
| 36 | Unburned | 2473 | 5 | 210 | 40.7401 | -105.22 |
| 37 | Unburned | 2471 | 4 | 300 | 40.6437 | -105.23 |
| 38* | Unburned | 2466 | 2 | 280 | 40.6816 | -105.32 |
| 39* | Unburned | 2464 | 2 | 128 | 40.6430 | -105.31 |
| 40* | Unburned | 2461 | 8 | 170 | 40.6130 | -105.31 |

* Plots chosen for clonal traits responses.

Appendix 2

Species list indicated by plot type (burned, unburned or both) and plant type (clonal or nonclonal) in study area.

| Species | Plot Type | Plant Type |
|--|-----------|------------|
| <i>Achillea millefolium</i> L. | Both | Clonal |
| <i>Allium</i> sp. L. | Burned | Nonclonal |
| <i>Antennaria parvifolia</i> Nutt. | Both | Clonal |
| <i>Arctostaphylos uva-ursi</i> L. | Both | Clonal |
| <i>Arnica cordifolia</i> Hook. | Unburned | Nonclonal |
| <i>Artemisia frigida</i> Willd. | Both | Nonclonal |
| <i>Artemisia ludoviciana</i> Nutt. | Both | Nonclonal |
| <i>Artemisia tridentata</i> Nutt. | Both | Nonclonal |
| <i>Bouteloua gracilis</i> (Willd. ex Kunth) Lag. ex Griffiths | Burned | Nonclonal |
| <i>Bromus ciliatus</i> L. | Unburned | Nonclonal |
| <i>Bromus inermis</i> Leyss. | Burned | Clonal |
| <i>Bromus madritensis</i> L. | Burned | Nonclonal |
| <i>Bromus marginatus</i> Nees ex Steud. | Unburned | Nonclonal |
| <i>Bromus tectorum</i> L. | Both | Nonclonal |
| Cactaceae Juss. | Unburned | Nonclonal |
| <i>Calamagrostis rubescens</i> Buckley | Unburned | Nonclonal |
| <i>Campanula rotundifolia</i> L. | Both | Clonal |
| <i>Carex rossii</i> Boott | Both | Clonal |
| <i>Carex scopulorum</i> T. Holm. | Both | Nonclonal |
| <i>Carex</i> sp. L. | Unburned | Nonclonal |
| <i>Castilleja puberula</i> Rydb. | Both | Nonclonal |
| <i>Centaurea montana</i> L. | Burned | Nonclonal |
| <i>Cerastium arvense</i> L. | Both | Clonal |
| <i>Cercocarpus montanus</i> Raf. | Both | Nonclonal |
| <i>Chamaenerion angustifolium</i> (L.) Holub ssp. <i>angustifolium</i> | Burned | Nonclonal |
| <i>Chenopodium album</i> L. | Both | Clonal |
| <i>Chrysothamnus</i> sp. Nutt. | Both | Nonclonal |
| <i>Cirsium arvense</i> (L.) Scop. | Both | Clonal |
| <i>Cystopteris fragilis</i> (L.) Bernh. | Unburned | Clonal |
| <i>Draba graminea</i> Greene | Unburned | Clonal |
| <i>Echinocereus</i> sp. Engelm. | Unburned | Nonclonal |
| <i>Ericameria nauseosa</i> (Pall. ex Pursh) G.L. Nesom & Baird | Burned | Nonclonal |
| <i>Erigeron annuus</i> (L.) Pers. | Both | Nonclonal |
| <i>Erigeron eximius</i> Greene | Both | Nonclonal |
| <i>Eriogonum umbellatum</i> Torr. | Both | Nonclonal |
| <i>Erysimum capitatum</i> (Douglas ex Hook.) Greene | Both | Nonclonal |
| <i>Euonymus</i> sp. L. | Both | Nonclonal |
| <i>Euphorbia brachycera</i> Engelm. | Burned | Nonclonal |
| <i>Festuca arizonica</i> Vasey | Burned | Nonclonal |
| <i>Festuca saximontana</i> Rydb. | Burned | Nonclonal |
| <i>Gaillardia aristata</i> Pursh | Burned | Nonclonal |
| <i>Galium boreale</i> L. | Unburned | Nonclonal |
| <i>Galium triflorum</i> Michx. | Unburned | Nonclonal |
| <i>Geranium</i> sp. L. | Both | Nonclonal |
| <i>Helianthus pumilus</i> Nutt. | Both | Nonclonal |
| <i>Heracleum maximum</i> W. Bartram | Unburned | Nonclonal |
| <i>Hesperostipa comata</i> (Trin. & Rupr.) Barkworth | Both | Nonclonal |
| <i>Heterotheca villosa</i> (Pursh) Shinnery | Both | Nonclonal |
| <i>Heuchera bracteata</i> (Torr.) Ser. | Both | Nonclonal |
| <i>Heuchera parviflora</i> Bartlett | Both | Nonclonal |
| <i>Juniperus communis</i> L. | Both | Nonclonal |
| <i>Juniperus scopulorum</i> Sarg. | Unburned | Nonclonal |
| <i>Koeleria macrantha</i> (Ledeb.) Schult. | Both | Clonal |

Continued.

| Species | Plot Type | Plant Type |
|--|-----------|------------|
| <i>Lupinus angustifolius</i> L. | Both | Nonclonal |
| <i>Lupinus argenteus</i> Pursh | Both | Nonclonal |
| <i>Luzula parviflora</i> (Ehrh.) Desv. | Burned | Nonclonal |
| <i>Mertensia ciliata</i> E. (James ex. Torr) G. Don | Unburned | Nonclonal |
| <i>Oxalis albicans</i> Kunth | Unburned | Nonclonal |
| <i>Oxytropis campestris</i> (L.) DC. | Both | Clonal |
| <i>Packera multilobata</i> (Torr. & A. Gray ex A. Gray) W.A. Weber & Á. Löve | Both | Nonclonal |
| <i>Pascopyrum smithii</i> (Rydb.) Á. Löve | Unburned | Nonclonal |
| <i>Pascopyrum</i> sp. Á. Löve | Both | Nonclonal |
| <i>Penstemon</i> sp. Schmidel | Both | Nonclonal |
| <i>Penstemon strictus</i> Benth. | Both | Nonclonal |
| <i>Penstemon whippleanus</i> A. Gray | Burned | Nonclonal |
| <i>Phleum pratense</i> L. | Both | Clonal |
| <i>Physaria acutifolia</i> Rydb. | Burned | Nonclonal |
| <i>Pinus contorta</i> Douglas ex Loudon | Unburned | Nonclonal |
| <i>Pinus flexilis</i> James | Both | Nonclonal |
| <i>Pinus ponderosa</i> Lawson & C. Lawson | Both | Nonclonal |
| <i>Poa bulbosa</i> L. | Both | Clonal |
| <i>Poa nervosa</i> (Hook.) Vasey | Unburned | Nonclonal |
| <i>Poa pratensis</i> L. | Burned | Clonal |
| <i>Polemonium</i> sp. L. | Burned | Nonclonal |
| <i>Populus tremuloides</i> Michx. | Burned | Clonal |
| <i>Pseudocymopterus montanus</i> (A. Gray) J.M. Coult & Rose | Both | Clonal |
| <i>Pseudotsuga menziesii</i> (Mirb.) Franco | Both | Nonclonal |
| <i>Pulsatilla patens</i> (L.) Mill. | Both | Clonal |
| <i>Purshia tridentata</i> (Pursh) DC. | Both | Clonal |
| <i>Ranunculus adoneus</i> A. Gray | Unburned | Nonclonal |
| <i>Ribes aureum</i> Pursh | Both | Clonal |
| <i>Ribes</i> sp. L. | Both | Nonclonal |
| <i>Rosa</i> sp. L. | Burned | Nonclonal |
| <i>Rosa woodsii</i> Lindl. | Burned | Nonclonal |
| <i>Rubus idaeus</i> L. | Both | Clonal |
| <i>Salix</i> sp. L. | Burned | Clonal |
| <i>Sedum lanceolatum</i> Torr. | Both | Nonclonal |
| <i>Solidago multiradiata</i> Aiton | Both | Nonclonal |
| <i>Solidago simplex</i> Kunth | Both | Nonclonal |
| <i>Sporobolus cryptandrus</i> (Torr.) A. Gray | Burned | Nonclonal |
| <i>Symphoricarpos albus</i> (L.) S.F. Blake | Burned | Nonclonal |
| <i>Symphoricarpos orbiculatus</i> Moench | Burned | Clonal |
| <i>Taraxacum officinale</i> F.H. Wigg. | Both | Clonal |
| <i>Thalictrum alpinum</i> L. | Burned | Clonal |
| <i>Tradescantia occidentalis</i> (Britton) Smyth | Burned | Nonclonal |
| <i>Tradescantia</i> sp. L. | Burned | Nonclonal |
| <i>Tragopogon dubius</i> Scop. | Both | Clonal |
| <i>Trifolium pratense</i> L. | Both | Nonclonal |
| <i>Trisetum spicatum</i> (L.) K. Richt. | Burned | Nonclonal |
| <i>Verbascum thapsus</i> L. | Both | Nonclonal |
| <i>Viburnum edule</i> (Michx.) Raf. | Unburned | Nonclonal |
| <i>Vicia sativa</i> L. | Burned | Clonal |
| <i>Vulpia octoflora</i> (Walter) Rydb. var. <i>octoflora</i> | Burned | Nonclonal |