

## Serum Antibody Prevalence for Herpesvirus sylvilagus, Bacillus piliformis and California Serogroup Arboviruses in Cottontail Rabbits from Pennsylvania

Authors: Dressler, R. L., Ganaway, J. R., Storm, G. L., and Tzilkowski, W. M.

Source: Journal of Wildlife Diseases, 24(2): 352-355

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-24.2.352

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <u>www.bioone.org/terms-of-use</u>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

## Serum Antibody Prevalence for *Herpesvirus sylvilagus, Bacillus piliformis* and California Serogroup Arboviruses in Cottontail Rabbits from Pennsylvania

**R. L. Dressler**,<sup>1</sup> J. R. Ganaway,<sup>2,3</sup> G. L. Storm,<sup>4</sup> and W. M. Tzilkowski,<sup>5</sup> <sup>1</sup> Maine Department of Inland Fisheries and Wildlife, P.O. Box 1298, Bangor, Maine 04401, USA; <sup>2</sup> National Institutes of Health, Bethesda, Maryland 20205, USA; <sup>3</sup> Present address: Microbiological Associates, Inc., Rockville, Maryland 20850, USA; <sup>4</sup> Pennsylvania Cooperative Fish and Wildlife Research Unit, University Park, Pennsylvania 16802, USA; and <sup>5</sup> School of Forest Resources, The Pennsylvania State University, University Park, Pennsylvania 16802, USA

ABSTRACT: A serologic survey of 60 eastern cottontail rabbits (*Sylvilagus floridanus*) from three counties in Pennsylvania was conducted in March 1983. Serum antibody prevalences for *Herpesvirus sylvilagus* and La Crosse virus (California serogroup) were <4%. There was no evidence of previous exposure to either Jamestown Canyon or snowshoe hare viruses (California serogroup). Antibody to trivittatus virus (California serogroup) was found in 60% of the 20 cottontails from York County. No cottontails had antibodies to *Bacillus piliformis*, the etiologic agent of Tyzzer's disease.

Key words: Cottontail, Sylvilagus floridanus, Herpesvirus sylvilagus, Bacillus piliformis, Tyzzer's disease, California group arboviruses, serology, serological survey.

Populations of the eastern cottontail rabbit (*Sylvilagus floridanus*) have been declining over portions of its original range (Edwards et al., 1981). Land-use changes have been recognized as a principal cause of these reductions at the regional level (Chapman et al., 1982). Many other factors such as disease, predation, adverse weather, dispersal, competition and nutritional inadequacies can decrease local populations (Chapman et al., 1982).

Herpesvirus sylvilagus, Bacillus piliformis (the etiologic agent of Tyzzer's disease), and California serogroup arboviruses have been reported in populations of cottontails (Ganaway et al., 1976; Lewis and Hinze, 1976; Seymour and Yuill, 1981). The epizootiology of *H. sylvilagus* in Wisconsin, the causative agent of a lymphoproliferative disease of the cottontail rabbit, was described by Lewis and Hinze (1976) and Spieker and Yuill (1976, 1977a, b) described various aspects of transmission of the virus. Tyzzer's disease in freeliving cottontails from Maryland (Ganaway et al., 1976) is a contagious, fatal, enteric and hepatic disease caused by B. piliformis. La Crosse (LAC) and trivittatus (TVT) of the California serogroup arboviruses infect cottontails (Moulton and Thompson, 1971; Pinger et al., 1975). Little is known about the occurrence of these pathogens in cottontails from Pennsylvania. Thus, the objective of the present study was to determine the serum antibody prevalence to H. sylvilagus, B. piliformis, and California serogroup arboviruses in geographically separate populations of eastern cottontail rabbits from Pennsylvania.

In March 1983, 20 cottontails (10 adult males, 10 adult females) were live-trapped from several sites in each of three counties in Pennsylvania: Somerset (southcentral Pennsylvania, 39°57'N, 78°55'W), Westmoreland (southwestern, 40°20'N, 79°22' W), and York (southeastern, 40°5'N, 77°2'W). Cottontails from Somerset County were trapped from reclaimed stripmined sites near Roxbury, Pennsylvania. Cottontails from Westmoreland County were trapped on farmland areas near Latrobe, Pennsylvania. Cottontails from York County were trapped on farmland and State Game Lands near Dillsburg, Pennsylvania.

Blood collection was by heart puncture with Vacutainer vacuum tubes (Becton, Dickinson and Company, Rutherford, New Jersey 07070, USA). Blood samples were allowed to clot for 30 min after collection, and then centrifuged at 1,000 g for 10 min. At least 0.5 ml of sera was immediately frozen at -10 C for shipment to the School of Veterinary Medicine, University of Wisconsin–Madison (Madison, Wisconsin 53706, USA) for *H. sylvilagus* and California serogroup arbovirus serological testing. An additional 0.5 ml of frozen sera was shipped to the National Institutes of Health (Bethesda, Maryland 20205, USA) for *B. piliformis* testing.

Antibody to *H. sylvilagus* was evaluated by 80% plaque reduction neutralization in tissue culture using methods described by Hinze (1971). All samples were tested at a dilution of 1:10.

*B. piliformis* originally was isolated from a laboratory rabbit (Ganaway et al., 1971). To prepare an enzyme-linked immunosorbent assay (ELISA) antigen, *B. piliformis* was grown in primary chicken embryonic liver cell cultures (Ganaway et al., 1985b). Details of the ELISA techniques were as published previously (Ganaway et al., 1985a).

Antibodies to LAC, TVT, snowshoe hare (SSH), and Jamestown Canyon (JC) California serogroup viruses were measured by microneutralization tests described by Pantuwatana et al. (1972) as modified by Ksiazek and Yuill (1977). Calculation of end points for virus titrations and log neutralization indices followed Reed and Muench (1938). Reference antibodies were hyperimmune mouse ascitic fluids prepared by the method of Brandt et al. (1967). Prepared virus stocks were LAC in the fifth suckling mouse brain (SMB) passage, TVT in the 17th SMB passage, SSH virus in the 23rd SMB passage, and JC in the sixth SMB passage (Ksiazek and Yuill, 1977). Titers <10 for each disease agent were considered negative.

Two rabbits, one adult female from Somerset County and one adult male from York County were positive for *H. sylvilagus*. Lewis and Hinze (1976) indicated that antibody prevalence was higher in adult males than in other sex and age groups. The low prevalence found in our study (<4%) suggests that *H. sylvilagus* was not a major source of mortality for cottontails in Pennsylvania during 1983.

Serologic evidence of SSH or JC was not found in any of the rabbits tested. Two adult male cottontails (10%) from Westmoreland County tested positive for LAC virus. La Crosse virus produces disease in humans and asymptomatic infection occurs in several small mammals, although not cottontails (Seymour and Yuill, 1981). Moulton and Thompson (1971) reported a 15% antibody prevalence for LAC virus in cottontails in Wisconsin. An earlier study in Pennsylvania reported LAC antibody prevalence in 254 cottontails to be <1%(Kradel et al., 1978). Our results fall between the values reported in these two studies.

One cottontail (5%) from Somerset County and twelve cottontails (60%) from York County tested positive for TVT virus. Titers ranged from 20 to 640. No sex specificity was evident for prevalence of antibodies to TVT virus. Neither LAC nor TVT cause clinical disease in cottontails but are maintained in cottontail populations and transmitted to other hosts by arthropods (Seymour and Yuill, 1981). Kradel et al. (1978) reported the prevalence of antibody to TVT was <1% of cottontails (n = 254) tested in central Pennsylvania. Pinger et al. (1975) reported 46% of cottontails (n = 22) from Iowa tested positive for TVT virus. Furthermore, they indicated a close association between the TVT virus vector, a mosquito (Aedes trivittatus), and the eastern cottontail. We are unable to explain the wide disparity in serum antibody prevalence for TVT virus between York County and the other collection areas. An attempt was not made to determine vector prevalence in the study area at the time blood samples were taken.

Serologic evidence of exposure to *B. pil-iformis* was not found in any of the 60 rabbits. Prevalence of Tyzzer's disease in free-ranging rabbit populations is not well known. Ganaway et al. (1976) hypothe-

sized that this disease may be an important population controlling factor for cottontails because most exposed animals eventually succumb to the disease. Because we found no serologic evidence of *B. piliformis*, we concluded that Tyzzer's disease apparently has not occurred in these populations, antibody decay was rapid, or infected animals died.

Our results provide no evidence to support a contention that the selected diseases were widespread in the rabbit population of Pennsylvania during 1983.

This paper is a contribution of the School of Forest Resources, The Pennsylvania State University, University Park, authorized as Journal Series Number 7583 of the Pennsylvania Agriculture Experimentation Station, and Pennsylvania Cooperative Fish and Wildlife Research Unit Paper Number 281. This research was supported by funds provided by the Pennsylvania Game Commission, National Rifle Association, and The Pennsylvania State University (School of Forest Resources and Institute for Research on Land and Water Resources). References to trade names used herein does not imply endorsement by the authors. The authors gratefully acknowledge the assistance provided by T. M. Yuill during serological testing. His comments on the methodology section of the manuscript also were appreciated. We thank A. Fairbrother, E. Burgess, and T. H. Spencer for assistance in serological analyses, many landowners for access to their properties for collection of cottontail sera, and J. E. Hudgins for assistance in the field.

## LITERATURE CITED

- BRANDT, W. E., E. L. BUESCHER, AND F. M. HET-RICK. 1967. Production and characterization of arbovirus antibody in mouse ascitic fluid. The American Journal of Tropical Medicine and Hygiene 16: 339-347.
- CHAPMAN, J. A., J. G. HOCKMAN, AND W. R. ED-WARDS. 1982. Cottontails. *In* Wild mammals of North America: Biology, management, and economics, J. A. Chapman and G. A. Feldhamer (eds.). Johns Hopkins University Press, Baltimore, Maryland, pp. 83-123.
- Edwards, W. R., S. P. Havera, R. F. Labisky, J. A.

ELLIS, AND R. E. WARNER. 1981. The abundance of cottontails (*Sylvilagus floridanus*) in relation to agricultural landuse in Illinois (USA), 1956–1978. *In* Proceedings World Lagomorph Conference, K. Myers and C. D. MacInnes (eds.). University of Guelph, Guelph, Ontario, Canada, pp. 761–789.

- GANAWAY, J. R., A. M. ALLEN, AND T. D. MOORE. 1971. Tyzzer's disease of rabbits: Isolation and propagation of *Bacillus piliformis* (Tyzzer) in embryonated eggs. Infection and Immunity 3: 429-437.
- ——, R. S. MCREYNOLDS, AND A. M. ALLEN. 1976. Tyzzer's disease in free-living cottontail rabbits (Sylvilagus floridanus) in Maryland. Journal of Wildlife Diseases 12: 545–549.
- —, T. H. SPENCER, T. D. MOORE, AND A. M. ALLEN. 1985a. Isolation, propagation, and characterization of a newly recognized pathogen, cilia-associated respiratory bacillus of rats, an etiological agent of chronic respiratory disease. Infection and Immunity 47: 472–479.
- -----, -----, AND K. W. WAGGIE. 1985b. Propagation of the etiologic agent of Tyzzer's disease (*Bacillus piliformis*) in cell culture. Proceedings of the 8th ICLAS Symposium, Gustav Fischer Verlag, Stuttgart, New York, pp. 59-70.
- HINZE, H. C. 1971. New member of the Herpesvirus group isolated from wild cottontail rabbits. Infection and Immunity 3: 350–354.
- KRADEL, D. C., R. G. MCLEAN, AND G. L. STORM. 1978. Arbovirus activity of rabbits prior to expansion of a wastewater treatment system. In Ecology and pathology of wildlife in response to spray irrigation of chlorinated sewage effluent Phase I. Institute for Research on Land and Water Resources, The Pennsylvania State University, University Park, pp. 65-69.
- KSIAZEK, T. G., AND T. M. YUILL. 1977. Viremia and antibody response to La Crosse virus in sentinel gray squirrels (*Sciuris carolinensis*) and chipmunks (*Tamias striatus*). The American Journal of Tropical Medicine and Hygiene 26: 815-821.
- LEWIS, H. S., AND H. C. HINZE. 1976. Epidemiology of *Herpesvirus sylvilagus* infection in cottontail rabbits. Journal of Wildlife Diseases 12: 482–485.
- MOULTON, D. W., AND W. H. THOMPSON. 1971. California group virus infections in small forestdwelling mammals of Wisconsin. The American Journal of Tropical Medicine and Hygiene 20: 474–482.
- PANTUWATANA, S., W. H. THOMPSON, D. M. WATTS, AND R. P. HANSON. 1972. Experimental infection of chipmunks and squirrels with La Crosse and trivittatus viruses and biological transmission of La Crosse virus by Aedes triseratus. The American Journal of Tropical Medicine and Hygiene 24: 999-1005.
- PINGER, R. R., W. A. ROWLEY, Y. W. WONG, AND

D. C. DORSEY. 1975. Trivittatus virus infections in wild mammals and sentinel rabbits in central Iowa. The American Journal of Tropical Medicine and Hygiene 24: 1006–1009.

- REED, L. J., AND H. M. MUENCH. 1938. A simple method of estimating fifty percent end points. American Journal of Hygiene 27: 493-497.
- SEYMOUR, C., AND T. M. YUILL. 1981. Arboviruses. In Infectious diseases of wild mammals, J. W. Davis, L. H. Karstad, and D. O. Trainer (eds.). Iowa State University Press, Ames, Iowa, pp. 54– 86.

SPIEKER, J. O., AND T. M. YUILL. 1976. Herpesvirus

*sylvilagus* in cottontail rabbits: Antibody prevalence and flea burden relationships. Journal of Wildlife Diseases 12: 310–314.

- \_\_\_\_, AND \_\_\_\_\_. 1977a. Herpesvirus sylvilagus in cottontail rabbits: Evidence of shedding but not transplacental transmission. Journal of Wildlife Diseases 13: 85-89.
- , AND ——, 1977b. Herpesvirus sylvilagus in cottontail rabbits: Attempted laboratory transmission by two insect species. Journal of Wildlife Diseases 13: 90–93.

Received for publication 31 March 1987.