BioOne COMPLETE

# Brucella suis Infections in Collared Peccaries in Venezuela

Authors: Lord, Veronica R., and Lord, Rexford D.

Source: Journal of Wildlife Diseases, 27(3) : 477-481

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-27.3.477

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

*Journal of Wildlife Diseases*, 27(3), 1991, pp. 477–481
© Wildlife Disease Association 1991

# *Brucella suis* Infections in Collared Peccaries in Venezuela

**Veronica R. Lord**[1] **and Rexford D. Lord,**[2] [1] Laboratory of Brucellosis, Institute of Veterinary Investigations, Box 70, Maracay 2102, Venezuela; [2] Biology Department, Indiana University of Pennsylvania, Indiana, Pennsylvania 15705, USA

ABSTRACT: A bacteriologic and serologic study was conducted on two ranches in the states of Apure and Guarico, Venezuela for brucellosis in collared peccaries (*Tayassu tajacu*). One hundred thirty-nine peccaries were necropsied and tissues were cultured. Forty-three isolations of *Brucella suis* biovar 1, were made from lymph nodes and spleens of 25 males and 18 females. Antibody to *Brucella* sp. was detected in sera from 122 animals by the rapid plate agglutination, standard tube agglutination, 2-mercaptoethanol, rivanol, complement fixation and card tests. Young animals had infection and reactor rates nearly as high as the older animals indicating most were infected at a relative early age. Results suggest that this species may transmit brucellosis when living with domestic animals. This is the first report of *B. suis* biovar 1 from collared peccaries in Venezuela.

*Key words:* Brucellosis, collared peccary, *Brucella suis*, prevalence, sero survey, *Tayassu tajacu*.

Brucellosis, caused by several species of *Brucella*, has been detected in numerous wild animals (Moore and Schnurrenberger, 1981; Witter, 1982; Garin-Bastuji et al., 1990). Some of these species might become reservoirs of infection for susceptible domestic animals (Corey et al., 1964; Hayes, 1977).

In several countries serum antibody has been detected and *Brucella* spp. isolated from wild animals but it remains uncertain whether they serve as reservoirs of the disease (Szyfres and Tome, 1966; Davis et al., 1979; De La Vega et al., 1979; Boer et al., 1980). Collared peccaries (*Tayassu tajacu*) are well distributed in Venezuela and are found on ranches with cattle which are of considerable economic importance to the country.

The principal objectives of this study were to sample tissues and sera from peccaries from the Llanos of Venezuela, for the purpose of isolating *Brucella* sp. and surveying for serum antibody against *Brucella* sp.

One hundred thirty-nine peccaries were collected by shooting with a rifle on two ranches, one in the state of Guarico (07°45′ to 07°55′N, 69°18′ to 69°23′E) and the other in the state of Apure (09°16′ to 09°21′N, 67°15′ to 67°22′E). Spleen, liver, mesenteric and retropharyngeal lymph nodes, sera and eyes were taken from all animals.

Spleens were washed in a solution of 0.85% NaCl, immersed in 95% alcohol, flamed, seared with a hot spatula and a 1 $cm^3$ piece removed for inoculation of solid culture media (Albimi *Brucella* sp. agar, trypiticase soy agar, and Kuzdas Morse) to which 5% fetal bovine serum was added (Alton et al., 1975). Lymph nodes were ground with mortar and pestal with a solution of 0.85% NaCl (pH 6.8) and inoculated on the same culture media.

Plates were incubated ≤10 days at 37 C in 10% $CO_2$, and examined daily after 48 hr. Colonies were observed with a steroscopic microscope according to the method of Henry (Alton et al., 1975). Possible *Brucella* sp. isolates were stained by the Gram and Koster technique (Alton et al., 1975). Several colonies from each sample, with typical characteristics of *Brucella* sp., were harvested and inoculated on agar slants (potato agar and trypticase soy agar) and on Petri plates. Isolates were incubated at 37 C to determine $CO_2$ dependency and growth on media containing 5% fetal bovine serum. They were examined further by the following tests: acriflavine (1:1,000), immersion in crystal violet (1:40), motility, urease activity, production of catalase, production of oxidase, production of $H_2S$, fermentation of lactose (MacConkey agar), production of hemolysis (blood agar), and reduction of nitrates



Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

TABLE 1. Characteristics of the 43 isolates of *Brucella* sp. from collared peccaries in Venezuela.

| | | |
|---|---|---|
| Bacteriological morphology: | | Cocabacilli |
| Motility: | | 37 C—negative, 22 C—negative |
| Requirement for serum: | | none |
| Dependency on $CO_2$: | | none |
| Sensitivity to thionin: | | yes |
| Sensitivity to fuchsin: | | no |
| Sensitivity to safranin: | | no |
| Development in 1 mg/ml i-erythritol: | | positive (100% growth) |
| Development in 5 IU[a]/ml penicillin: | | positive (50% growth) |
| Production of $H_2S$ (days): | | positive for 5 days |
| Activity of urease (minutes): | | positive (after 15 min) |
| Activity of catalase (seconds): | | positive (immediate) |
| Fermentation of lactose (MacConkey agar): | | negative |
| Reduction of nitrates: | | positive |
| Utilization of citrate: | | negative |
| Monospecific sera—A: | | positive agglutination |
| —M: | | negative agglutination |
| —R: | | negative agglutination |
| Sensitivity to Tbilisi phage: | 1 RTD | no lysis |
| | $\times 10^4 \times$ RTD | lysis |

[a] International Units.

and citrate (Alton et al., 1975; Cowan and Steele, 1979; MacFaddin, 1980). The dye sensitivity of *Brucella* sp. isolates was determined by adding basic 0.1% fuchsin (1:25,000, 1:50,000, 1:100,000), 0.1% thionin (same dilutions), and 1% safranin (1:5,000) to Albimi *Brucella* sp. agar. Growth on media containing i-erythritol (1 mg/ml) and penicillin (5 International Units/ml) was also studied (Huddleson, 1931). The media were inoculated with bacterial suspensions prepared in 0.85% sterile NaCl solution at a similar density with reference strains (*B. abortus* 544, *B. melitensis* 16 M, *B. suis* 1330). Plates were divided into four quarters for inoculation with a calibrated platinum loop and incubated at 37 C for 72 hr. Monospecific antisera, anti-A, anti-M and anti-R, were used to determine which of the agglutinins predominated in the isolates. Two concentrations of the Tbilisi phage (Routine Test Dilution (RTD) and 10,000 × RTD) were used (Alton et al., 1975).

For the metabolic tests, the following substrates were used in sequential groups:

TABLE 2. Oxidative rates $QO_2(N)$ of 43 isolates of *Brucella* sp. from peccaries in Venezuela with four amino acids and four carbohydrate substrates.

| | Group I | | Group II | | Group III | | | |
|---|---|---|---|---|---|---|---|---|
| Isolates or reference strain | L-alanine | L-glutamic acid | D,L-ornithine | L-lysine | L-arabinose | D-galactose | D-ribose | D-glucose |
| *B. suis* biovar 1 observed range . . . | 60–68 | 61–69 | 160–166 | 141–149 | 120–127 | 280–287 | 304–308 | 372–378 |
| 544[a] | 140 | 322 | 41 | 18 | 160 | 289 | 108 | 86 |
| 16M[b] | 82 | 98 | 37 | 36 | 25 | 38 | 42 | 57 |
| 1,330[c] | 65 | 68 | 159 | 148 | 124 | 283 | 304 | 375 |

[a] *B. abortus*, reference strain.
[b] *B. melitensis*, reference strain.
[c] *B. suis*, reference strain.

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

TABLE 3. Antibody levels to *Brucella* sp. in peccaries from Venezuela.

| Number of animals[a] | RPA test[b] | STA test[c] | Card test | Rivanol | 2-mercapto-ethanol test[d] | Complement fixation | Group |
|---|---|---|---|---|---|---|---|
| 12 | 100 IU[e] | 200 IU | positive | 200 IU | 200 IU | 104 IU | 1 |
| 39 | 200 | 100 | positive | 200 | 200 | 52 | |
| 18 | 100 | 100 | positive | 50 | 50 | 26 | |
| 11 | 50 | 100 | positive | — | 25 | 13 | 2 |
| 19 | 25 | 50 | positive | — | 25 | 13 | |
| 10 | 50 | 50 | positive | — | 50 | 13 | |
| 4 | 50 | 50 | positive | — | 25 | 13 | |
| 2 | 25 | 50 | — | — | — | — | 3 |
| 3 | 50 | 25 | — | — | — | — | |
| 4 | 25 | 25 | — | — | — | — | |

[a] 17 animals were negative for all serological tests.
[b] Rapid plate agglutination test.
[c] Standard tube agglutination test.
[d] 2-mercaptoethanol test.
[e] International Units.

Group I used L-alanine and L-glutamic acid; Group II used amino acids of the urea cycle, D,L-ornithine and L-lysine; and Group III used the carbohydrates, L-arabinose, D-galactose, D-ribose and D-glucose (Meyer and Morgan, 1962).

A 1% Sorensen solution buffered with phosphates to pH 7.0 was prepared for each of the substrates (Meyer and Cameron, 1959). Packed bacteria cells were resuspended in Sorensen solution and adjusted to a dilution of 1:40 similar to a normal suspension. The density was determined in a spectrophotometer at a wave length of 420 nm. The normal suspension contained approximately 0.8 mg of nitrogen per ml. Manometric determinations were made using the Warburg apparatus (Clark, 1969). A substrate was considered to have been oxidated when the value of $QO_2(N)$ ($\mu$l oxygen uptake/mg nitrogen/60 min) was equal to or more than 50 $\mu$l (Meyer and Cameron, 1961a, b; Lord and Flores, 1983).

TABLE 4. Relation between age of peccaries, isolation of *Brucella* sp. and presence of *Brucella* sp. antibodies.

| Estimated age in years | Isolations/animals cultured (%) | Reactors/sera tested (%) |
|---|---|---|
| 1 | 4/18 (22) | 14/18 (78) |
| 2 | 7/23 (30) | 23/23 (100) |
| 3 | 18/51 (35) | 43/51 (84) |
| 4 | 14/47 (30) | 42/47 (89) |
| Total | 43/139 (31) | 122/139 (87.8) |

Blood samples were obtained from the jugular vein. The samples were allowed to clot, and were centrifuged, separated, and stored in a refrigerator until transport to the laboratory on wet ice. Peccary sera were tested in the rapid plate agglutination (RPA) test, standard tube agglutination (STA) test, 2-mercaptoethanol, rivanol, card test and complement fixation (CF) test, utilizing an antigen of *B. abortus* (strain 1119-3) (Alton et al., 1975).

Ages of peccaries were determined by the lens technique (Lord, 1959; Lord and Lord, 1988). All work was conducted in the Laboratory of Brucellosis (Institute of Veterinary Investigations, Maracay, Venezuela).

One hundred thirty-nine peccaries (70 males and 69 females) were taken from two ranches, and 556 tissue samples were cultured. *Brucella suis* was isolated from 43 (31%) of the peccaries (Tables 1, 2). On the basis of oxidative rates, all 43 isolates were determined to be biovar 1.

Serologic tests were considered positive when agglutination was observed in the card test, when there were 13 IU in the

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

CF test, and/or when titers of 100 IU were seen in the other tests. Reactors were animals whose sera reacted in any of the tests. High titers (26 IU in CF tests, 100 IU in other tests, or greater) of antibody were found in 69 (50%) of the 139 sera tested; 44 (32%) had low titers of antibody in all tests except rivanol; 9 (7%) of 26 had low levels of antibody in the RPA and STA tests, and 17 (12%) sera were negative to all tests.

Table 4 shows the relation between estimated age and the isolations and serologic reactors. The youngest animals had the lowest rates, though they were nearly as high as those of older animals. There was a slight tendency for higher isolation rates in animals with higher titers (32% of Groups 1 and 2; 22% of Group 3 of Table 3). Isolations were made from 25 males and 18 females, but serologic reactors were similar for both sexes.

Neither of the two ranches have feral or tame domestic pigs (*Sus scrofa*) running free on the open range. Nevertheless, there are other ranches in these states where domestic pigs range freely and *B. suis* has been detected in capybaras (*Hydrochaeris hydrochaeris*) from the Llanos (Lord and Flores, 1983). The peccaries could have been infected by transmission from capybaras or by a chain of transmission through peccaries, leading back to domestic pigs elsewhere. The high infection rate (31%) and a high rate of sero positively indicates an important segment of the peccary population was infected with *B. suis* biovar 1.

Although 1-yr-old animals had both the lowest isolation and reactor rates, these rates were nearly as high as those of the older animals, indicating that most became infected with *B. suis* at a relatively early age. This is the first report of *B. suis* biovar 1 in collared peccaries in Venezuela.

Peccaries are widely distributed in Venezuela, and likewise throughout many Latin American countries where bucellosis is a problem. These animals live on ranches with domestic animals such as swine, equines, bovines, and numerous wild animals, many of which may be adversely affected by *B. suis*. The epidemiologic significance of *B. suis* in peccaries is not known, but merits further investigation.

The authors express their thanks to Rafael Hoogestein, and Hernan Castellanos for their help in obtaining samples from peccaries.

## LITERATURE CITED

ALTON, G. G., L. M. JONES AND, D. F. PEITZ. 1975. Laboratory techniques in brucellosis. World Health Organization, Monograph Series 55, Geneva, Switzerland, 175 pp.

BOEER, W. J., R. P. CRAWFORD, R. J. HIDALGO, AND N. ROBINSON. 1980. Small mammals and whitetailed deer as possible reservoir hosts of *Brucella abortus* in Texas. Journal of Wildlife Diseases 16: 19–24.

CLARK, J. M. 1969. Manometry: Calibration of Warburg flasks and manometers. Experimental Biochemistry. W. H. Freeman and Company, San Francisco, California, 228 pp.

COREY, R. R., L. J. PAULISSEN, AND D. SWARTZ. 1964. Prevalence of brucellae in the wildlife of Arkansas. Journal of Wildlife Diseases 36: 8.

COWAN, S.T., AND K. J. STEELE. 1979. Manual para la identificacion de bacterias de importancia medica. Editorial Continental, S.A., Mexico City, Mexico, 320 pp.

DAVIS, D. S., W. J. BOEER, J. P. MIMS, F. C. HECK, AND L. G. ADAMS. 1979. *Brucella abortus* in coyotes. I. A serological and bacteriological survey in eastern Texas. Journal of Wildlife Diseases 15: 367–372.

DE LA VEGA, E. C., C. GARCIA-CARRILLO, AND C. ARCE. 1979. Infeccion natural por *Brucella* en comadrejas (*Didelphis marsupialis*) en la Republica Argentina. Revista Medicina Veterinaria Argentina 60: 283–286.

GARIN-BASTUJI, B., J. OUDAR, Y. RICHARD, AND J. GASTELLU. 1990. Isolation of *Brucella melitensis* Biovar 3 from a chamios (*Rupicapra rupicapra*) in the southern French Alps. Journal of Wildlife Diseases 26: 116–118.

HAYES, F. A. 1977. Wildlife reservoirs of brucellosis. *In* Bovine brucellosis. An international symposium, R. P. Crawford and R. J. Hidalgo (eds.). Texas Agricultural and Mechanical University Press, College Station, Texas, pp. 269–276.

HUDDLESON, I. F. 1931. Differentiation of the species in the genus *Brucella*. American Journal of Public Health 21: 491–498.

LORD, R. D. 1959. The lens as an indicator of age in cottontail rabbits. The Journal of Wildlife Management 23: 358–360.

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use

———, AND V. R. LORD. 1988. Cross checking censuses and a model of the annual cycle of mortality and reproduction in capybaras (*Hydrochaeris hydrochaeris*). Studies on Neotropical Fauna and Environment 23: 213–244.

LORD, V. R., AND R. FLORES-C. 1983. *Brucella* spp. from the capybara (*Hydrochaeris hydrochaeris*) in Venezuela: Serologic studies and metabolic characterization of isolates. Journal of Wildlife Diseases 19: 308–314.

MACFADDIN, J. F. 1980. Pruebas bioquimicas para la identificacion de bacterias de importancia clinica. Editorial Medica Panamericana, S.A., Buenos Aires, Argentina, 301 pp.

MEYER, M. E., AND H. S. CAMERON. 1959. Comparative metabolism of species and types of organism within the genus *Brucella*. Journal of Bacteriology 78: 130–136.

———, AND ———. 1961a. Metabolic characterization of the genus *Brucella*. I. Statistical evaluation of the oxidative rates by which type I of each species can be identified. Journal of Bacteriology 82: 387–395.

——— AND ———. 1961b. metabolic characterization of the genus *Brucella*. II. Oxidative metabolic patterns of the described biotypes. Journal of Bacteriology 82: 396–400.

———, AND W. J. B. MORGAN. 1962. Metabolic characterization of *Brucella* strains that show conflicting identity by biochemical and serological methods. Bulletin of World Health Organization 26: 823–827.

MOORE, C. G., AND P. R. SCHNURRENBERGER. 1981. A review of naturally occurring *Brucella abortus* infections in wild mammals. Journal of the American Veterinary Medical Association 179: 1105–1112.

SZYFRES, B., AND J. G. TOME. 1966. Natural *Brucella* infection in Argentina wild foxes. Bulletin of World Health Organization 34: 919–923.

WITTER, J. F. 1982. Brucellosis. *In* Infectious diseases of wild mammals, J. W. Davis, L. H. Karstad, and D. O. Trainer (eds.), Iowa State University Press, Ames, Iowa, pp. 280–287.

*Received for publication 18 April 1990.*

Downloaded From: https://bioone.org/journals/Journal-of-Wildlife-Diseases on 28 Mar 2024
Terms of Use: https://bioone.org/terms-of-use