Development and Characterization of 15 Microsatellite Markers for Cephalotaxus fortunei (Cephalotaxaceae)

Authors: Wang, Chunbo, Guo, Zhiyou, Huang, Xilian, and Huang, Lu

Source: Applications in Plant Sciences, 4(5)

Published By: Botanical Society of America

URL: https://doi.org/10.3732/apps.1500129
**Development and Characterization of 15 Microsatellite Markers for Cephalotaxus fortunei (Cephalotaxaceae)**

CHUNBO WANG, ZHIYOU GUO, XILIANG HUANG, and LU HUANG

2Department of Life Sciences, Qiannan Normal College for Nationalities, Duyun 558000, People’s Republic of China; and 3State Key Laboratory of Biocontrol, School of Life Sciences, Sun Yat-sen University, Guangzhou 510275, People’s Republic of China

- **Premise of the study:** To survey population variation and the adaptive evolution of *Cephalotaxus fortunei* (Cephalotaxaceae), an endemic and endangered conifer in China, microsatellite markers were developed and characterized for this species.
- **Methods and Results:** Based on the Fast Isolation by AFLP of Sequences Containing repeats (FIASCO) protocol, 15 microsatellite markers were developed for *C. fortunei*, 13 of which were polymorphic within a sample of 75 individuals representing five natural populations. The number of alleles per locus ranged from one to seven. The expected and observed heterozygosities were 0.108–0.738 and 0.000–1.000, respectively. Ten polymorphic loci were also successfully amplified in *C. oliveri*.
- **Conclusions:** These polymorphic loci provide a valuable tool for population genetic analysis of *C. fortunei*, which will contribute to its management and conservation.

**Key words:** Cephalotaxaceae; *Cephalotaxus fortunei*; cross-amplification; FIASCO; genetic analysis; microsatellite primers.

_Cephalotaxus fortunei_ Hook. is a perennial, coniferous shrub or small tree belonging to the family Cephalotaxaceae. Endemic to China, _C. fortunei_ is mainly distributed from the subtropical regions up to the northernmost Qinling Mountains and Huai River in central China, occurring in locations with an elevation between 200 and 3700 m (Zhou et al., 1997). Because it contains the anticancer alkaloid harringtonine, _C. fortunei_ is important for medicinal use in treating leucocytoma (Shi et al., 2010). Its natural populations in China face threats of deforestation, other human-induced disturbances, and overexploitation. At present, _C. fortunei_ is listed as a Category V threatened plant by the international Conifer Specialist Group (He et al., 1996). Thus, a deeper understanding of genetic variation and population structure of this species using polymorphic DNA markers will provide valuable information for developing conservation strategies.

In this study, we developed 15 microsatellite loci for _C. fortunei_ using the Fast Isolation by AFLP of Sequences Containing repeats (FIASCO) approach (Zane et al., 2002), and we also examined their ability to be cross-amplified in _C. oliveri_ Mast.

**METHODS AND RESULTS**

Seventy-five individuals of _C. fortunei_ from five populations were collected in its natural distribution area from 2014 to 2015, and voucher specimens were deposited at the herbarium of Qiannan Normal College for Nationalities (Appendix 1). Young and healthy leaves were preserved in silica gel. All samples were stored at −20°C until processed. Total genomic DNA was extracted using a modified cetyltrimethylammonium bromide (CTAB) protocol with −20°C propanol pretreatment to eliminate polysaccharides (Su et al., 1998).

The FIASCO method was used to develop microsatellite loci using one individual of _C. fortunei_ from the Guizhou population (voucher: CB Wang 201406, JPG [QNCNI]). Approximately 3 μg of DNA was digested with _MseI_ (New England Biolabs, Ipswich, Massachusetts, USA). The DNA digestion fragments were linked to an _MseI_ adapter pair (F: 5’-TACTGAGACT-CAT-3’, R: 5’-GAGGATGAGTCCGTGAG-3’) with T4 ligase at 4°C overnight (Pan et al., 2011). A diluted digestion-ligation mixture (1:10) was directly amplified using the following program: 95°C for 30 s, 60°C for 20 min, and 72°C for 1.5 min for 23 cycles with _MseI_ primers (5’-GAGGATGAGTCCGTGAGTAA-3’). Then, we used 5’-biotinylated (AC)15 and streptavidin-coated magnetic beads (Promega Corporation, Madison, Wisconsin, USA) to hybridize and capture the PCR product (Miao et al., 2012). Enriched fragments were recovered with PCR amplification as described above, using _MseI_-N as the primers. Purifying with a multifunctional DNA Extraction Kit (OMEGA Bio-Tek, Norcross, Georgia, USA), the PCR products were then ligated into pTA2 vector (Toyobo, Osaka, Japan) and transformed into _E. coli_ DH5α competent cells. A total of 80 clones were selected by blue-white screening and tested by PCR using M13+/-M13 as primers. Seventy-six positive clones were chosen to be sequenced on an ABI Prism 3730 automated DNA sequencer (Invitrogen, Guangzhou, China). Out of the 76 clones, 27 clones contained simple sequence repeats, of which 12 were discarded because they were unsuitable for designing primers. Primers for the remaining 15 sequences were designed using Primer Premier 5.0 (Clark and Gorley, 2001).

We used 75 individuals from five populations to test the polymorphism of the newly developed primer pairs (Table 1). The PCR amplifications were performed in a 20-μL reaction containing 1 μL of genomic DNA, 2 μL of PCR buffer, 0.5 μL 10 mM each primer, 1 μL 10 mM dNTP mixture, and 1 unit Taq DNA polymerase (TaKaRa Biotechnology Co., Dalian, China). PCR profiles were as follows: an initial denaturation at 94°C for 5 min; followed by 35 cycles at 94°C for 45 s, annealing temperature for 30 s, extension at 72°C for 1 min; and final extension at 72°C for 10 min (Table 1). PCR products were electrophoresed on 6% polyacrylamide denaturing gels by silver staining using a 50-bp ladder. Thirteen of the 15 loci were found to be polymorphic. The sizes of all amplification products matched the expected lengths.
The effective number of alleles ($A_e$), observed heterozygosity ($H_o$), expected heterozygosity ($H_e$), and departure from Hardy–Weinberg equilibrium (HWE) were estimated by GenAIEx version 6.4 (Peakall and Smouse, 2006). Linkage disequilibrium (LD) across all populations was tested using GENEPOP version 4.0.10 (Rousset, 2008). The occurrence of null alleles was investigated using MICRO-CHECKER version 2.2.3 (Van Oosterhout et al., 2004). The number of alleles per locus varied from one to seven, with a total of 247 alleles scored.

### CONCLUSIONS

In this study, we developed 15 microsatellite loci for *C. fortunei*, 13 of which were polymorphic. The genetic information based
Table 3. Genetic diversity in five Cephalotaxus oliveri populations using 10 polymorphic microsatellite loci originally developed in C. fortunei.a

<table>
<thead>
<tr>
<th>Locus</th>
<th>Changyang population (N=15)</th>
<th>Hupingshan population (N=15)</th>
<th>Fanjingshan population (N=15)</th>
<th>Anfu population (N=15)</th>
<th>Daweishan population (N=15)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>A</td>
<td>A_e</td>
<td>H_o</td>
<td>A</td>
<td>A_e</td>
</tr>
<tr>
<td>CF1</td>
<td>3</td>
<td>2.766</td>
<td>0.439</td>
<td>0.382</td>
<td>5</td>
</tr>
<tr>
<td>CF2</td>
<td>5</td>
<td>3.432</td>
<td>0.628</td>
<td>0.571</td>
<td>3</td>
</tr>
<tr>
<td>CF3</td>
<td>4</td>
<td>2.488</td>
<td>0.482</td>
<td>0.276</td>
<td>2</td>
</tr>
<tr>
<td>CF4</td>
<td>5</td>
<td>3.653</td>
<td>0.927</td>
<td>0.803</td>
<td>4</td>
</tr>
<tr>
<td>CF5</td>
<td>3</td>
<td>2.072</td>
<td>0.372</td>
<td>0.210</td>
<td>3</td>
</tr>
<tr>
<td>CF6</td>
<td>3</td>
<td>1.364</td>
<td>0.518</td>
<td>0.454</td>
<td>4</td>
</tr>
<tr>
<td>CF7</td>
<td>4</td>
<td>3.632</td>
<td>0.7398</td>
<td>0.454</td>
<td>4</td>
</tr>
<tr>
<td>CF8</td>
<td>4</td>
<td>3.281</td>
<td>0.835</td>
<td>0.706</td>
<td>1</td>
</tr>
<tr>
<td>CF9</td>
<td>3</td>
<td>2.737</td>
<td>0.243</td>
<td>0.217</td>
<td>5</td>
</tr>
<tr>
<td>CF10</td>
<td>3</td>
<td>2.455</td>
<td>0.306</td>
<td>0.258</td>
<td>2</td>
</tr>
</tbody>
</table>

Note: A = actual number of alleles; A_e = effective number of alleles; H_o = observed heterozygosity; N = sample size for each population.

aLocality and voucher information is available in Appendix 1.

on these newly developed microsatellite loci will contribute to the management and conservation of C. fortunei. In addition, the successful cross-species amplification of the loci in C. oliveri implies that they will provide an opportunity to further investigate the adaptive evolution of Cephalotaxus species.

LITERATURE CITED


