Morphometrics of Eggs, Nymphs, and Adults of Bactericera cockerelli (Hemiptera: Triozidae), Grown on Two Varieties of Tomato Under Greenhouse Conditions

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MORPHOMETRICS OF EGGS, NYMPHS, AND ADULTS OF BACTERICERA COCKERELLI (HEMIPTERA: TRIOZIDAE), GROWN ON TWO VARIETIES OF TOMATO UNDER GREENHOUSE CONDITIONS

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ABSTRACT

The effects of 2 varieties of tomato, Solanum lycopersicum L., i.e., ‘Charanda F1’ and ‘Rafaello’, were evaluated on the morphometrics of Bactericera cockerelli (Sulc). Eggs, nymphs, and adults of B. cockerelli were collected from 2 varieties of tomato, ‘Charanda F1’ and ‘Rafaello’, under greenhouse conditions in the Colegio de Postgraduados, Campus Montecillo, Texcoco, the State of Mexico, during the periods Oct-Dec 2009 and Jan-Mar 2010. Since 2000-2001 the B. cockerelli cultures were maintained on tomato with no exposure to agrochemicals. Adult B. cockerelli were kept in individual growth chambers constructed of wooden frames covered with organza cloth, under 14:10 h L:D and temperatures ranging from 10 to 25 °C. For morphometric analysis of the eggs, the following variables were investigated: egg length (LH), egg width (AH), and pedicel length (PED). For nymphs, the variables were: body length (LC), body width (AC), and antennal length (ANT). For adults, the variables were: body length (LC), body width at thorax (ACT), wing length (LALA), and wing width (ANALA). The 2 tomato varieties were found to have differential morphometric effects on B. cockerelli as follows: no significant differences on egg length ($F_{1,41} = 0.57; P = 0.4551$), but egg width was differentially affected by the variety of tomato ($F_{1,41} = 11.92; P = 0.0013$). There were significant differential effects of tomato variety on nymphs: body length ($F_{4,324} = 1199.2; P < 0.0001$), body width ($F_{4,324} = 900.72; P = 0.0001$); and antennae length ($F_{4,324} = 883.9; P = 0.0001$). Body length of the adults ($F_{1,117} = 7.11; P = 0.0087$) was differentially affected by the 2 different tomato varieties. None of the plants showed any symptoms of infection by ‘Candidatus Liberibacter solanacearum’, which is known to cause effects on B. cockerelli fitness traits. Body width and antennal length of nymphs can be recommended to differentiate all 5 nymphal instars on this pest species, which has practical implications.

Key Words: Potato psyllid, tomato psyllid ‘Charanda F1’, ‘Rafaello’, Psylloidae, morphology

RESUMEN

Se evaluó el efecto de dos variedades de jitomate (‘Charanda F1’ y ‘Rafaello’) en la geomorfometría de B. cockerelli (Sulc). Se colectaron huevos, ninñas y adultos criados en dos variedades de jitomate ‘Charanda F1’ y ‘Rafaello’, en condiciones de invernadero en el Colegio de Postgraduados, Campus Montecillo, Texcoco, México, en el periodo Octubre-Diciembre 2009-Enero-Marzo 2010. Las condiciones generales de la cría fueron: la colonia de insectos origen se integró desde el periodo 2000-2001 sobre plantas de tomate, sin aplicación de agroquímicos; los adultos fueron mantenidos en jaulas individuales de marco de madera cubiertos con tela de organza, bajo un periodo de 14 h de luz, con un rango de temperatura de 10 a 25 °C. En el análisis de morfometría de huevos se consideraron las variables: Longitud de Huevo (LH), Ancho de Huevo (AH) y Longitud de Pedicelo (PED). En ninñas, Largo de Cuerpo (LC), Ancho de Cuerpo (AC) y Longitud de Antenas (ANT). En adultos, Longitud de Cuerpo (LC), Ancho de Cuerpo en Tórax (ACT), Longitud de Ala (LALA), Ancho de Ala (ANALA). Se encontró que el efecto de las variedades de jitomate en la geomorfometría, no presenta diferencias significativas sobre la Longitud de huevos ($F_{1,41} = 0.57; P = 0.4551$); aunque el Ancho de hueso sí resultó afectado por la variedad de jitomate sobre la que se criaron los insectos ($F_{1,41} = 11.92; P = 0.0013$). Los estados ninflares, para las variables Largo de Cuerpo ($F_{4,324} = 1199.2; P < 0.0001$), Ancho de Cuerpo ($F_{4,324} = 900.72; P < 0.0001$) y Largo de An-
The evolution of insect populations is closely linked with that of the species they feed upon, and this seems especially the case of some genera and species of the Psylloidea (White & Hodkinson 1985; Burckhardt & Lauterer 1997; Hodkinson 2009). The insects of the Psylloidea feed on the sap of host plants by introducing their stylets and making use of the sugars and amino acids of the plant that they cannot synthesize themselves (Thao et al. 2000; Percy 2003). Bactericera cockerelli (Sulc) (Hemiptera: Triozidae), variously known as the tomato psyllid and potato psyllid, is an insect pest species that feeds mainly on plants of the Solanaceae family and some members from several other families of plants (Pletsch 1947; Burckhardt & Lauterer 1997; Crespo-Herrera et al. 2012). There is evidence relating to the effect of the host plants on the development and morphology of insects from diverse genera and taxonomic families, such as Bemisia tabaci (Berlinger 1986; Perring et al. 1993; Huang et al. 2008) and the pea aphid, Acyrthosiphon pisum Harris (Frantz et al. 2010). Some studies focused specifically on the effects of certain Solanaceae on the reproductive ability of B. cockerelli (Liu et al. 2004; Liu et al. 2006a; Yang & Liu 2009; Vargas-Madríz et al. 2011), or on its fitness (Nachappa et al. 2012).

The architecture and the content of secondary metabolites of cultivated plants are some characteristics that cause differences in the feeding and oviposition behaviors of phytophagous insects, as happens in the case of the psyllid, Boreioglycaspis melaleuca Moore, on Melaleuca quinquenervia (Cav.) S. T. Blake (Myrtales: Myrtaceae), whose leaf nitrogen and carotene contents are factors that modify the behavior of the mentioned psyllid. Also Hodkinson et al. (2001) and Casteel et al. (2006) proved that the Mi-1.2 gene in wild tomato (Solanum lycopersicum L.; Solanales: Solanaceae) affects the reproductive ability of B. cockerelli. This gene confers resistance in tomato against Meloidogyne spp., the potato aphid (Macrosiphum euphorbiarum Thomas) and the silverleaf whitefly (Bemisia argentifoli (Bellows and Perring). The main effect of the Mi-1.2 gene is to cause necrotic zones to develop around the areas attacked by nematodes and repels females of Meloidogyne spp., thus reducing oviposition. Some reports mentioned the possibility of the existence of at least 2 ‘biotypes’ of B. cockerelli in several geographical zones of the US, Mexico and Guatemala (Jackson et al. 2009; Nelson et al. 2011), captured from diverse host plants such as tomato and chili pepper (Capsicum spp.; Solanaceae) (Liu et al. 2006b; Liu et al. 2007), potato (Abdullah 2008), and eggplant (Solanum melongena, L.; Solanales: Solanaceae) (Yang & Liu 2009). Majority of the work done focuses on studying variation in the reproductive ability of B. cockerelli feeding on different host plants, but the effects of host plants the morphology of B. cockerelli have not been investigated. Such effects of host plants on insect morphology are fundamental in understanding morphological variation within insect species, and they are considered as evidence to determine ‘biotypes’ (Claridge & Den Hollander 1983). Thus, the present study is intended to give evidence on the possible effect of 2 varieties of tomato on the morphology of eggs, nymphs, and adults of B. cockerelli under greenhouse conditions.

### MATERIALS AND METHODS

The experiments were conducted under greenhouse and laboratory conditions at the Colegio de Postgraduados, Campus Montecillo, Texcoco, the State of Mexico. Temperature and RH were recorded every 30 min, using electronic sensors and data logger, HOBO, model H-08-032-08 #333699, placed on the tomato plants used in the experiments. The tomato varieties used in this study were the 2 hybrids: ‘Charanda F1’ and ‘Rafaello’, which are grown frequently by tomato farmers in Mexico because of their good agronomic behavior, i.e., very long fruiting periods (indeterminate development) and long postharvest shelf life (Tighelaar 1986; Pérez & Castro 1999; Mendoza-De Jesús et al. 2010; Del Val et al. 2010).

**Bactericera cockerelli Colonies**

The 2 tomato varieties were seeded in planting trays; 20 plants from each of the 2 varieties were selected for the study. Seedlings were transplanted into polyethylene bags containing 2 kg of 50% Canadian Growing Mix 1VM and 50% Tezontle (porous volcanic rock) when the plants were 31 d old. Plants were moved to growth chambers (62 cm long × 95 cm wide × 95 cm high) framed with wooden boards and covered with plain weave mesh to exclude insects, especially Bemisia tabaci.
was used to identify the species. Genitalia of nymphal growth were registered daily until they were later adjusted to 100 eggs. Hatching and the number of eggs laid was recorded. This colony, 65 adults in their reproductive stage were taken randomly to guarantee a cohort of 100 eggs per chamber, and 65 adults were introduced per chamber. From each colony, 65 adults in their reproductive stage were taken randomly to guarantee a cohort of 100 eggs on 1 d in each growth chamber. Three chambers were set up for each tomato variety, with 5 plants per chamber, and 65 adults were introduced per chamber. After 24 h, the adults were taken out and the number of eggs laid was recorded. This was later adjusted to 100 eggs. Hatching and nymphal growth were registered daily until they transformed into adults. Genitalia of B. cockerelli was used to identify the species.

To identify B. cockerelli abdomens and genitalia were removed from 5 adult males following protocols of Crawford (1910) and Crawford (1914). The abdomens were placed for 1 to 2 min in 10% potassium hydroxide to eliminate abdominal fat. The abdomens were cleaned by placing them in hot (80 °C) distilled water for 15 min, and 1% acetic acid was used to rinse off the potassium hydroxide from each specimen. The abdomens were then placed in 70% alcohol, and then in 100% potassium hydroxide to eliminate abdominal fat. The abdomens were placed for 1 to 2 min in 10% potassium hydroxide to eliminate abdominal fat. The abdomens were cleaned by placing them in hot (80 °C) distilled water for 15 min, and 1% acetic acid was used to rinse off the potassium hydroxide from each specimen. The abdomens were then placed in 70% alcohol, and then in 100% alcohol for 1 min to finish cleaning the cuticle. During the cleaning process alcohol was changed with an insulin syringe 3 times to eliminate any plant debris. After removing the abdomens from alcohol, they were placed in xylene to dealcoholize and clear them; xylene was changed twice. Then each abdominal specimen was carefully placed on a slide, using small cardboard pieces to avoid crushing the abdomens. Canadian balsam was used as the mounting medium because it has the appropriate refraction index for viewing by stereo microscope. Genitalia were later observed in the stereo microscope and specimens under study was confirmed as B. cockerelli based on the keys described by Crawford (1910 & 1914) and (Tuthill 1945). Photographs of the taxonomic characters of each developmental stages were taken based on the descriptions by Lehman (1930), Tuthill (1945), Marín-Jarillo et al. (1995), Burckhardt & Lauterer (1997), and Abdullah (2008).

 Morphometry of Eggs, Nymphs, and Adults of the B. cockerelli

Eggs, nymphs, and adults of B. cockerelli grown on 2 varieties of tomato, ‘Rafaello’ and ‘Charanda’ were collected in 2009-2010. The eggs and nymphs were collected from the leaves of the host plants using entomological forceps and placed in jars with 70% ethanol. The egg, nymph and adults samples were dried and placed on slides in order to photograph them with a Carl Zeiss Tessovar photomicroscope connected to a Pixera digital camera. Care was taken to standardize the age of the insects used. Eggs were used regardless of whether they were fertile or infertile. The metric ruler was also photographed to calibrate the Image Tool (Wilcox et al. 2002) measuring software. The photographs were digitized in JPEG format.

The taxonomic differentiation of B. cockerelli adults was done using taxonomic keys described by (Crawford 1910a, 1910b, 1911a, 1911b, 1914). In the morphometric analysis of the eggs, the following variables were considered: egg length (LH), egg width (AH), and pedicel length (PED).

The measured variables in the case of the nymphs were: body length (LC), body width (AC), and antennae length (ANT). The nymphs were classified according to instars, from instar 1 to 5, to immediately carry out the measurements of the mentioned variables.

In the case of morphometry of adults, the following variables were considered: body length (LC), body width at the thorax (ACT), wing length (LALA), and wing length (ANALA). Measurements of 30 adult specimens were made. Data on measurements of various life stages were analyzed using one-way analysis of variance (ANOVA, SAS Institute Inc. 2005). The effect of the factors: ‘variety’ and ‘sex’, and their possible interaction on the morphometric measured variables were assessed. The significance of the considered factors was tested through the application of the GLM procedure in SAS (SAS 2005) with alpha = 0.05. The Tukey test was used (P = 0.05) to separate means of measurements involving the different instars; and the F test was used to assess the interaction between sex and tomato variety. The possible significance of the differences in the means of the morphological measurements due to the ‘variety’ and ‘sex’ factors were tested by Student’s normalized t test, at the same significance level.

RESULTS

In 2009, the incubation period for the egg stage was 7 days on ‘Charanda F1’ and 8 days on ‘Rafa-
Table 1. Duration of the Nymph and Adult Stages of Bactericera cockerelli Reared on 2 Tomato Varieties Under Greenhouse Conditions in Montecllo State of Mexico, Oct-Dec 2009 and Jan-Mar 2010

<table>
<thead>
<tr>
<th>STAGES</th>
<th>CHARANDA F1’ (days)</th>
<th>RAFAELO’ (days)</th>
<th>CHARANDA F1’ (days)</th>
<th>RAFAELO’ (days)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Egg</td>
<td>7</td>
<td>8</td>
<td>Egg</td>
<td>13</td>
</tr>
<tr>
<td>Nymph 1</td>
<td>11</td>
<td>9</td>
<td>Nymph 1</td>
<td>19</td>
</tr>
<tr>
<td>Nymph 2</td>
<td>13</td>
<td>11</td>
<td>Nymph 2</td>
<td>16</td>
</tr>
<tr>
<td>Nymph 3</td>
<td>16</td>
<td>16</td>
<td>Nymph 3</td>
<td>16</td>
</tr>
<tr>
<td>Nymph 4</td>
<td>17</td>
<td>15</td>
<td>Nymph 4</td>
<td>17</td>
</tr>
<tr>
<td>Nymph 5</td>
<td>19</td>
<td>20</td>
<td>Nymph 5</td>
<td>17</td>
</tr>
<tr>
<td>Adult</td>
<td>44</td>
<td>45</td>
<td>Adult</td>
<td>41</td>
</tr>
<tr>
<td>Egg-Adult</td>
<td>63</td>
<td>68</td>
<td>Egg-Adult</td>
<td>69</td>
</tr>
</tbody>
</table>

Note: The time of development from the egg to adult is not the sum of the different stages because do not necessarily at the end of one stage indicate the star of the next stage.
variables obtained on tomato varieties showed differential effects of tomato varieties and sexual gender (Table 7). Sexual gender differentially affected anterior wing length and anterior wing width variables on the tomato varieties.

Comparison of the mean values of the morphometric variables of *B. cockerelli* adults indicated that the ‘Charanda’ variety induced greater body length, antennae length, anterior wing length and width, while the Rafaello variety resulted in greater body width at the thorax and anterior wing width (Table 7).

**DISCUSSION**

The durations of the egg and the first instar in 2009 were similar to those reported by Yang et al. (2009), who observed a value of 6 days for the same variable on eggplant and bell pepper, however the durations of the other instars were very different. The same tendency occurred in 2010. A possible explanation for this phenomenon could be the different range of temperature used in this work (21 °) in contrast with the range used by Yang et al. (2009) (26.7 °) and the different plant hosts used in both studies. Great differences in durations within the life cycle of the *B. cockerelli* reported by different authors indicates the need for more careful research on this species under the specific conditions of different tomato producing areas. In Mexico tomato production occurs in many areas with different climatic conditions; and precise knowledge of the life cycle in producing areas is needed to make decisions about controlling this insect pest. Our results of the egg incubation period coincide with those reported by Lehman (1930), Pack (1930), Knowlton & Janes (1931), List (1939), Pletsch (1947) and Abdullah (2008), but they do not coincide with respect to the duration of the nymph, adult, and the total biological cycle. The differences could be due to different plant varieties and environmental conditions used in the different studies. Nachappa et al. (2012) recently reported some effects of *Candidatus Liberibacter solanacearum* (a.k.a. ‘Ca. L. psyllaurous’) on the fitness of *Bactericera cockerelli*, but not on the morphology. In this study we did not consider the possible effect of this bacterium on the morphometry of the insect. Yet, as an anonymous reviewer suggested, the presence of this bacterium has the potential to cause what appear to be cultivar effects resulting from pathogen-induced changes in phloem pressure, nutritional status, etc. However in our study none of the plants showed symptoms of *Candidatus Liberibacter solanacearum* infection. Therefore we do not think that any of the observed effects were caused by the influence of bacteria.

**Egg Morphometry**

There was no significant difference in lengths of the eggs (*F*1, 41 = 0.57; *P* = 0.4551) obtained from the 2 different tomato varieties (Table 2), but egg width was affected by the tomato variety on which the insects were cultured (*F*1, 41 = 11.92; *P* = 0.0013). In this regard Morales & Cermeli (2001) point out that the preference in oviposition of an insect on a host plant is influenced by its age and the environmental conditions where the plant developed. Future studies should investigate the reason for differences in eggs morphometrics in-

<table>
<thead>
<tr>
<th>Variable</th>
<th>‘CHARANDA F1’</th>
<th>‘RAFAELLO’</th>
<th>t</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Egg Length</td>
<td>285.97 ± 14.92</td>
<td>296.87 ± 38.04</td>
<td>-1.25</td>
<td>0.22NS</td>
</tr>
<tr>
<td>Egg Width</td>
<td>129.18 ± 6.65</td>
<td>140.18 ± 15.53</td>
<td>-3.04</td>
<td>0.005**</td>
</tr>
<tr>
<td>Pedicel Length</td>
<td>184.03 ± 23.36</td>
<td>175.24 ± 21.09</td>
<td>1.29</td>
<td>0.20NS</td>
</tr>
</tbody>
</table>

NS = Not significant (*P* ≤ 0.05); ** significant (*P* ≤ 0.01).

Table 3. Mean values ( ± 1SD) of morphometric variables of *Bactericera cockerelli* nymphs (instars 1 to 5) grown on 2 tomato varieties in MonteCillo State of Mexico, 2009-2010. Measurement data from all 5 instars were pooled.

<table>
<thead>
<tr>
<th>Variable</th>
<th>CHARANDA F1</th>
<th>RAFAELLO</th>
<th>t</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body Length</td>
<td>1273.75 ± 423.11</td>
<td>1294.00 ± 415.89</td>
<td>-0.44</td>
<td>0.66NS</td>
</tr>
<tr>
<td>Body Width</td>
<td>842.76 ± 313.53</td>
<td>836.41 ± 298.07</td>
<td>0.19</td>
<td>0.85NS</td>
</tr>
<tr>
<td>Antennae Length</td>
<td>209.38 ± 95.06</td>
<td>208.28 ± 80.87</td>
<td>0.11</td>
<td>0.91NS</td>
</tr>
</tbody>
</table>

NS = Not significant (*P* ≤ 0.05), *n* = 165 specimens.
Reduced by ‘Rafaello’ and ‘Charanda F1’. Possibly differences in phytochemicals produced by these varieties are responsible for this response of the insect. Orozco-Cardenas et al (2001) and Orozco-Cárdenas & Ryan (1999) reported high production of \( \text{H}_2\text{O}_2 \) in tomato plants after the attack of some aphids, and that this might provide an early defense in being able to cause an often drastic reduction in insect feeding (Harborne 1993).

**Nymphal Morphometry**

The nymphal instar of *B. cockerelli* is one of the least studied developmental stages in the American continent, whereas exhaustive studies on the nymphal stages of the Psylloidea superfamily have been conducted in Europe (White & Hodkinson 1985; Ossiannilsson 1992). Body length \((F_{4, 324} = 1199.2; P < 0.0001)\), body width \((F_{4, 324} = 900.72; P < 0.0001)\), and antennae length \((F_{4, 324} = 883.93; P < 0.0001)\) variables were significantly differentiated for each *B. cockerelli* nymphal instar. These distinctions have practical implications, because they can be used to differentiate the 5 nymphal instars of *B. cockerelli*. Instar classification can be adequately accomplished by body length, except in the cases of instars 2 and 3, which overlap in body length. However all 5 instars can be reliably differentiated by body width and antennal length. This is of a much practical value for any researcher who needs to sort the nymphal instars of this triozid insect.

**Adult Morphometry**

Mean body length of *B. cockerelli* adults reared on ‘Rafaello’ tomato differs from that of those reared on ‘Charanda’ tomato \((F_{1, 117} = 7.11; P = 0.0087)\). However sexual gender per se did not affect body length \((F_{1, 117} = 0.09; P = 0.7603)\). This seems strange given that it is known that the females have a greater body size than the males (Plesch 1947; Burckhardt & Lauterer 1997). Possible such a difference could be detected by analyzing a very large sample. Body width at the tho-

### Table 4. Variation factors and significance on the morphometry of *Bactericera cockerelli* nymphs grown on 2 tomato varieties in Montecillo State of Mexico, 2009-2010.

<table>
<thead>
<tr>
<th>Source</th>
<th>Degrees of freedom</th>
<th>Mean squares</th>
<th>Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body length</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Instar</td>
<td>4</td>
<td>1349749.38**</td>
<td></td>
</tr>
<tr>
<td>Variety</td>
<td>1</td>
<td>36808.62NS</td>
<td></td>
</tr>
<tr>
<td>Instar*variety</td>
<td>4</td>
<td>28028.38*</td>
<td></td>
</tr>
<tr>
<td>Body width</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Instar</td>
<td>4</td>
<td>7215007.08**</td>
<td></td>
</tr>
<tr>
<td>Variety</td>
<td>1</td>
<td>1651.16NS</td>
<td></td>
</tr>
<tr>
<td>Instar*variety</td>
<td>4</td>
<td>6797.31NS</td>
<td></td>
</tr>
<tr>
<td>Antennae length</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Instar</td>
<td>4</td>
<td>593524.51**</td>
<td></td>
</tr>
<tr>
<td>Variety</td>
<td>1</td>
<td>99.38NS</td>
<td></td>
</tr>
<tr>
<td>Instar*variety</td>
<td>4</td>
<td>4634.35**</td>
<td></td>
</tr>
</tbody>
</table>

NS = Not significant \((P \geq 0.05)\); *, ** Significant with \((P \leq 0.05)\) and highly significant with \((P \leq 0.01)\).

### Table 5. Differentiation of mean morphometric variables \((M)\) of nymphal instars of *Bactericera cockerelli* grown on 2 tomato varieties in Montecillo State of Mexico, 2009-2010.

<table>
<thead>
<tr>
<th>Instar</th>
<th>Body length</th>
<th>Body width</th>
<th>Antennae length</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>652.22 d</td>
<td>390.05 e</td>
<td>92.71 e</td>
</tr>
<tr>
<td>2</td>
<td>1135.92 c</td>
<td>709.47 d</td>
<td>159.71 d</td>
</tr>
<tr>
<td>3</td>
<td>1184.34 c</td>
<td>769.74 c</td>
<td>181.18 e</td>
</tr>
<tr>
<td>4</td>
<td>1681.47 b</td>
<td>1145.66 b</td>
<td>287.04 b</td>
</tr>
<tr>
<td>5</td>
<td>1763.17 a</td>
<td>1187.58 a</td>
<td>323.61 a</td>
</tr>
</tbody>
</table>

Means with the same letter in the same column are not statistically different, Tukey \((P < 0.05)\).

### Table 6. Factors of variation and significance on the morphometry of *Bactericera cockerelli* adults reared on 2 tomato varieties in Montecillo State of Mexico, 2009-2010.

<table>
<thead>
<tr>
<th>Source</th>
<th>Degrees of freedom</th>
<th>Mean squares and significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body length</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Variety</td>
<td>1</td>
<td>859536.13**</td>
</tr>
<tr>
<td>Sex</td>
<td>1</td>
<td>20358.07NS</td>
</tr>
<tr>
<td>Variety*sex</td>
<td>1</td>
<td>8595.36NS</td>
</tr>
<tr>
<td>Body width</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Variety</td>
<td>1</td>
<td>318.60NS</td>
</tr>
<tr>
<td>Sex</td>
<td>1</td>
<td>2787.54NS</td>
</tr>
<tr>
<td>Variety*sex</td>
<td>1</td>
<td>387.95NS</td>
</tr>
<tr>
<td>Antennae length</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Variety</td>
<td>1</td>
<td>121260.12**</td>
</tr>
<tr>
<td>Sex</td>
<td>1</td>
<td>74570.69</td>
</tr>
<tr>
<td>Variety*sex</td>
<td>1</td>
<td>121789.77**</td>
</tr>
<tr>
<td>Forewing length</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Variety</td>
<td>1</td>
<td>5627.59NS</td>
</tr>
<tr>
<td>Sex</td>
<td>1</td>
<td>301567.74**</td>
</tr>
<tr>
<td>Variety*sex</td>
<td>1</td>
<td>22741.19NS</td>
</tr>
<tr>
<td>Forewing width</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Variety</td>
<td>1</td>
<td>35009.00**</td>
</tr>
<tr>
<td>Sex</td>
<td>1</td>
<td>64688.65**</td>
</tr>
<tr>
<td>Variety*sex</td>
<td>1</td>
<td>15816.40**</td>
</tr>
</tbody>
</table>

*, ** = \(F\) test significance \((P < 0.05 \text{ and } P < 0.01)\); NS = not significant.
TABLE 7. EFFECT OF TOMATO VARIETIES AND SEXUAL GENDER OF *Bactericera cockerelli* ADULTS ON THEIR MORPHOMETRY IN MONTECILLO STATE OF MEXICO, 2009-2010.

<table>
<thead>
<tr>
<th>TRAIT</th>
<th>Charanda F1</th>
<th>Rafaello</th>
<th>Female</th>
<th>Male</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>n</td>
<td>Mean</td>
<td>n</td>
<td>Mean</td>
</tr>
<tr>
<td>LC</td>
<td>60</td>
<td>1656.1</td>
<td>60</td>
<td>1486.8</td>
</tr>
<tr>
<td>ACT</td>
<td>57</td>
<td>542.8</td>
<td>58</td>
<td>546.1</td>
</tr>
<tr>
<td>LA</td>
<td>60</td>
<td>694.0</td>
<td>48</td>
<td>626.6</td>
</tr>
<tr>
<td>LALA</td>
<td>60</td>
<td>2118.0</td>
<td>58</td>
<td>2104.2</td>
</tr>
<tr>
<td>ANALA</td>
<td>60</td>
<td>780.9</td>
<td>59</td>
<td>815.2</td>
</tr>
</tbody>
</table>

**LC =** Body length; **ACT =** Body width at the thorax; **LA =** Antennae length; **LALA =** Anterior wing length; **ANALA =** Anterior wing width. ***,** significance of test (**P < 0.05** and **P < 0.01**); **NS** = not significant.

raxis is not affected by the tomato variety (**F** = 0.13; **P = 0.7145**), nor by the sexual gender of the insect (**F** = 1.18; **P = 0.2803**). Antennal length of the adults of *B. cockerelli* is affected by tomato variety (**F** = 7.29; **P = 0.0081**) and by sexual gender (**F** = 4.49; **P = 0.0365**). It is known that the males have larger antennae than females, perhaps because of mating behaviors. Related to this, an adult male-attractive pheromonal compound of *B. cockerelli* had been detected recently (Guedot et al. 2010). This is difference of antennal size between males and females is well documented for *Cacopsylla bidens* (Sule) (Soroker et al. 2004) and *Diaphorina citri* Kuwayama (Onagbola et al. 2008). Anterior wing length of *B. cockerelli* showed no variations induced by either of the 2 tomato varieties (**F** = 0.40; **P = 0.5301**), but there were highly significant differences due to the sexual gender of the insect (**F** = 21.25; **P < 0.0001**). This coincides with multiple results that indicate a greater wing size in females than in males (Knowlton & James 1931; Plesch 1947; Soroker et al. 2004; Liu et al. 2007). Anterior wing width was differentiated both by variety (**F** = 12.34; **P = 0.0006**) and the sex of the insect (**F** = 22.79; **P < 0.0001**). Yang & Liu (2009) presented developmental parameters of *B. cockerelli* on eggplant and chili pepper plants, and they obtained lower reproduction ratios in chili pepper, although the generation time was longer in this host plant. These researchers asserted worked that differences in its reproductive ability of *B. cockerelli* are a function of the host plant. This could cause the appearance of biotypes in *B. cockerelli*. It is important to point out that Yang & Liu (2009) did not provide a rigorous taxonomic identification of this insect. Moreover, their study lacked a morphometric analysis to relate the effect of the host plant to the insect’s morphology. The morphometric analysis proposed in the present paper enhances the data shown in the life tables for *B. cockerelli*, and it supplies precise information on the taxonomic identification of the studied insect. In many published scientific works the latter point is usually taken for granted, however specific conditions could cause changes in the table of life. The plants in our study did not show any symptoms ‘Candidatus Liberibacter solanacearum’ infection, and we do not think that any of the observed effects were caused by the influence of bacteria.

**CONCLUSIONS**

The tomato varieties, ‘Rafaello’ and ‘Charanda’, had a differential effect on *B. cockerelli* egg width, but not on egg length or egg pedicel length. Also, we were able to differentiate all 5 nymphal instars by body width and antennal length. Antennal length of nymphs has the potential to support the development of practical methodologies for identifying nymphal instars of *B. cockerelli* and of phylogenetically related insects.

Effects of the ‘Rafaello’ and ‘Charanda’ tomato varieties were identified on the morphometric variables of adults of *B. cockerelli*, i.e., body length, antennal length, and wing width. Sexual gender of insect influenced antennal length, wing length, and wing width. Additional research is needed to elucidate the precise way in which the host plants affect the morphology of the different development stages of *B. cockerelli*. It is necessary to evaluate the effects of number of additional solanaceous crop species on the population growth of *B. cockerelli*, and to relate life table parameters to the variation in morphometric dimensions.

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