Solar UV Upregulates Photoprotection but Slows Photosynthesis in Subalpine Australian Plants

Authors: William T. Salter, Tarryn L. Turnbull, Heinz Rennenberg, and Mark A. Adams
Source: Arctic, Antarctic, and Alpine Research, 49(4) : 673-685
Published By: Institute of Arctic and Alpine Research (INSTAAR), University of Colorado
URL: https://doi.org/10.1657/AAAR0017-024
Solar UV upregulates photoprotection but slows photosynthesis in subalpine Australian plants

William T. Salter¹,*¹, Tarryn L. Turnbull¹, Heinz Rennenberg²³, and Mark A. Adams⁴

¹Centre for Carbon, Water and Food, University of Sydney, 380 Werombi Road, Brownlow Hill, NSW 2570, Australia
²Institute of Forest Botany and Tree Physiology, Chair of Tree Physiology, University of Freiburg, Georges-Koehler-Allee 53/54, 79110, Freiburg, Germany
³College of Sciences, King Saud University, P.O. Box 2455, Riyadh 11451, Saudi Arabia
⁴Department of Chemistry and Biotechnology, Faculty of Science, Engineering and Technology, Swinburne University, PO Box 218, Hawthorn, Victoria 3122, Australia

*Corresponding author’s email: william.salter@sydney.edu.au

ABSTRACT
Recent literature suggests a shifting paradigm in relation to photobiology associated with ultraviolet (UV) radiation. UV has been repeatedly shown to be less detrimental to plant performance than previously thought. Nonetheless, relatively few plant species have been studied; too few to make definitive statements about effects of UV on plants at the ecosystem scale. We present findings of a field-based study using natural solar radiation, coupled with UV screening films, to determine physiological costs and benefits of exposure to solar UV for three species representative of subalpine Australian flora: a tree (Eucalyptus pauciflora), a forb (Geranium antrorsum), and a grass (Poa hiemata). Photochemical and photosynthetic responses to UV exclusion varied among species; exposure to UV was of no consequence to the structure, chemistry, or incidence of photoinhibition for E. pauciflora. UV was effectively screened at the leaf surface of P. hiemata. The response of G. anstrorsum to UV exclusion suggests greater susceptibility to photodamage; less successful in screening against UV and exposure reduced rates of photosynthesis, despite increased capacity to scavenge reactive oxygen species (via accumulating ascorbate). This study clarifies that responses to UV are highly species-specific, and that the endemic native flora is seemingly well-adapted to mitigate negative effects.

INTRODUCTION
Identifying positive relationships between solar ultraviolet (UV) light and plant performance remains poorly studied, particularly in natural ecosystems. For example, the literature overwhelmingly contains examples of UV causing plant damage, or negatively affecting photobiology and growth, resulting in the general perception that UV radiation is detrimental to plants. Wargent and Jordan’s (2013) Tansley Review attributes the balance of findings to methodology, with the dominance of ‘negative’ studies attributable to unrealistic wavebands or intensity, or targeting exposed mesophyll cells (rather than cells embedded within, and protected by leaf structures, screening compounds, and biochemical processes). Plants reduce their exposure to UV by reflecting around 5% of incident UV radiation at the leaf surface. Of more significance are screening chemicals in the outer leaf structures that together can absorb up to a further 90% of incident UV. Cuticular waxes, particularly of alpine plants, have distinct UV-A and UV-B absorption peaks (Jacobs et al., 2007; Bruhn et al., 2014); the growth of trichomes and accumulation of UV-absorbing hydroxycinnamic acids in these cells is promoted by exposure to UV (Karabourniotis et al., 1992; Tattini et al., 2000), and the accumulation of flavonoids and related phenolic compounds in epidermal (and to a lesser extent mesophyll) cells can attenuate UV-B and UV-A wave-
Eucalyptus pauciflora was Poa hiemata ′ Geranium antrorsum 674 / William T. Salter et al. / Arctic, Antarctic, and Alpine Research

lengths, respectively (Caldwell et al., 1983; Agati and Tattini, 2010). The synthesis of flavonoid compounds is induced by exposure to UV radiation in some plant species; however, many do not act solely to screen UV and have multiple photoprotective roles, including as antioxidants (Agatti et al., 2012; Landi et al., 2015). UV that reaches the mesophyll can affect photosynthetic apparatus in similar fashion to excess radiation of longer wavelengths, in particular via accumulation of reactive oxygen species (ROS) in the thylakoid membrane (Foyer et al., 1994) and/or by inhibiting repair of PSI (Nishiyama et al., 2006). At the cellular level, UV wavelengths can exacerbate oxidative stress (Barta et al., 2004; Snyrchova et al., 2007) via the formation of oxygen radicals from peroxidases and oxidases, chlorophyll molecules and metabolic reactions in peroxisomes, as well as in the electron transport chain (for reviews see Kataria et al., 2014; and Czégény et al., 2016). UV radiation also promotes the production of hydrogen peroxide (H$_2$O$_2$), the stable form of oxidized oxygen, and its subsequent photosensitive breakdown to highly oxidizing hydroxyl radical (Czégény et al., 2014). Free radicals can be neutralized by the ascorbate (Asc) and glutathione (GSH) antioxidative system (Foyer and Noctor, 2011; Noctor et al., 2012; Toth et al., 2013), as well as by other enzymatic (including superoxide dismutase, catalases) and non-enzymatic antioxidants (including a-tocopherols, carotenoids, phenolic compounds) (for review see Ahmad et al., 2010).

There is strengthening recognition of positive UV-plant relationships. For example, Day et al. (1992) highlighted that plants do not necessarily have to protect their mesophyll from UV, with herbaceous dicots endemic to mountain ecosystems transmitting 20–40% of UV to the mesophyll (c.f. 4% in adjacent woody dicots and grasses). Similarly, exposure to solar UV is far from universally damaging to plant photosynthetic physiology. UV may directly (via chlorophyll a absorption: McCree, 1972; Inada, 1976; Turnbull et al., 2013) and, to a lesser extent, indirectly (via reabsorption of UV-induced blue-green fluorescence: 1% contribution to photosynthesis in Johnson and Day, 2002; 7.8–9.8% in Mantha et al., 2001) drive oxidative photosynthesis in a range of plants. Most recently, Wargent et al. (2015) recorded that the photosynthetic performance of Lactuca sativa was enhanced when exposed to UV during early development, and suggested that UV acted as a priming agent for primary productivity.

Australian plants are arguably exposed to some of the brightest (Salby, 2012) and most UV-enriched light environments on Earth (Roy et al., 1995), far in excess of requirements for photosynthesis (Amthor, 2010). In subalpine ecosystems in southeast Australia, plants are exposed to intense visible and UV radiation because of latitude (Gies et al., 2004), altitude (Blunthaler et al., 1997), and albedo and snow cover (Lee et al., 2015). Theory thus suggests that native (well-adapted) plants from these systems should exhibit mechanisms and structures (for example, promotion of antioxidant biosyntheses, alteration of morphology/phenology) that mitigate and limit photobiological stress as, in addition to an inherently energetic light environment, they are also subject to extreme variations in temperature, and availability of water and nutrients, that limit growth and diminish photosynthetic requirements for light (Bornman et al., 2015). We recently provided evidence that Pimelea ligustrina, a woody shrub common to Australia’s subalpine region, employs a suite of photobiological adaptations in order to utilize UV-A for photosynthesis (Turnbull et al., 2013). P. ligustrina has no effective cuticle and limited concentrations of phenols in the epidermal layer. Solar UV-A passes relatively unimpepted to the mesophyll layer, which is in turn enhanced in the UV-A absorbing photo-pigments chlorophyll a and lutein (a carotenoid pigment). Consequently, P. ligustrina uses UV-A to raise photosynthetic rates ~12% above that driven by the visible component of solar radiation. Furthermore, our data suggest the capacity to use UV-A for photosynthesis features strongly in subalpine flora, with 26 of the 55 most prominent species (mostly herbaceous dicots) also demonstrating that UV-A can directly be absorbed by chlorophyll a in intact leaves (Turnbull et al., 2013).

We sought to test if UV-A driven photosynthesis might feature among other major plant genera in the Australian subalpine region. We chose a canopy dominant tree (Eucalyptus pauciflora), an herbaceous dicot (Geranium antrorsum), and the common snow grass (Poa hiemata) as study species. In addition, we sought to test if exposure to UV during leaf development influenced the capacity to utilize UV-A and visible wavelengths together for photosynthesis. We deployed screens above plants growing in situ for the entire growth season, and combined field measurements of gas exchange with laboratory analysis of leaf fluorescence, redox chemistry, and leaf anatomy.

### Materials and Methods

#### Site Description and Treatments

We studied snow gum (Eucalyptus pauciflora Sieber x Spreng), rosetted cranesbill (Geranium antrorsum Carolin.), and soft snow grass (Poa hiemata Vickery) growing in a subalpine grassy woodland in the Snowy Mountains region of New South Wales (NSW) (36°05′36″S,
148°31′42″E; 1550 m a.s.l.). Photosynthetic measurements were made on clear, sunny days in March 2013 (midday visible photon flux density [PFD] = 1700 μmol photon m⁻² s⁻¹; midday UVA PFD = 50 μmol photon m⁻² s⁻¹). Samples for fluorescence, reflectance, microscopy, and chemical analyses were collected upon completion of photosynthetic measurements. The youngest fully expanded leaves (all formed under treatment conditions) were used for all analyses.

To control the spectral quality reaching our chosen plant species, we deployed two filters of contrasting spectral transmissivities (Fig. 1); one that transmits visible light but excludes UV (UV exclusion; #3114; Rosco Australia PTY Ltd., Artarmon, NSW, Australia) and a control filter that transmits visible light and UV (Control; B6191-50; Churchill & Coombes Pty Ltd., Lidcombe, NSW, Australia). We positioned 15 control and 15 UV-exclusion filters along an 800 m transect in locations where the study species grew within a 1 m² quadrat. The filters were in place for five months from October 2012 until March 2013.

Leaf Gas Exchange

An infrared gas analyzer (IRGA) (LI-6400; Li-Cor Inc., Lincoln, Nebraska, U.S.A.) was used to measure photosynthetic rates in situ under fully saturating light conditions (PFD > 1500 μmol photon m⁻² s⁻¹, as determined by a light response curve, data not shown). Chamber temperature and moisture matched ambient, whereas reference CO₂ concentration was 400 ppm and flow rate retained at 350 μmol s⁻¹. The IRGA was fitted with a clear-topped 2 × 3 cm leaf chamber and glass filters to control the spectral quality. To quantify the contribution of UVA to photosynthesis, photosynthetic rates were measured when the leaf was in receipt of wavelengths exceeding 320 nm (PAR + UVA) (N-WG320; Schott Australia Pty. Ltd., French’s Forest, NSW, Australia) and at wavelengths greater than 395 nm (PAR – UVA) (GC395; Schott Australia Pty. Ltd., French’s Forest, NSW, Australia). Leaves were allowed to stabilize in the chamber for 3 min prior to measurement. Light saturated rates of photosynthesis (Aₚ) and stomatal conductance (gₛ) were logged for 4 min at 30 s intervals.

**Fluorescence Spectroscopy**

Fluorescence profiles of fresh leaves were captured with a Cary Eclipse fluorescence spectrophotometer (Agilent Technologies, Mulgrave, Victoria, Australia) and a Cary Eclipse fiber optics accessory using a scan rate of 120 nm min⁻¹. Fluorescence emission spectra were collected from samples using excitation λ 380 nm (emission λ scan = 400–700 nm). Epidermal capacity to screen UV was estimated from the ratio of fluorescence emission at λ 440 (epidermal hydroxycinnamic acids) to emission at λ 685 nm (chlorophyll in the mesophyll), and chlorophyll content was estimated from the fluorescence emission at λ 685 to emission at λ 720 nm (Lenk and Buschmann, 2006). To determine chlorophyll a fluorescence, excitation spectra were collected for samples at emission λ 685 nm (excitation λ scan = 340–475 nm) and λ 730 nm (excitation λ scan = 370–475 nm). Data points between λ 380 nm and 405 nm were modeled using a sigmoidal curve fitted to the remaining 30 points at an interval of 1 nm between 370 nm and 425 nm (Prism 6; Graphpad Software, La Jolla, California, U.S.A.). All fitted curves had an R-squared value greater than 0.99.

**Leaf Anatomy**

Slides for microscopy analysis were prepared as per Turnbull et al. (2013). Fresh leaf sections (1 cm × 2 mm) were fixed with 2.5% glutaraldehyde solution in 0.1 M phosphate buffer saline (PBS). These were refrigerated at 4 °C for four days to allow full penetration of the fixative. Leaf samples were then dehydrated in a graded series of ethanol, cleared with xylene, and infiltrated with paraffin using an automated processor (TissueTek VIP, Sakura, Tokyo, Japan). The samples were then embedded in blocks of paraffin and cut into 4 μm sections using a microtome (Leica RM2165; Leica Microsystems GmbH, Wetzlar, Germany). The leaf sections were picked up on glass slides and dried at 56 °C. Xylene

**FIGURE 1.** Spectral light environment of each treatment in relation to natural insolation; black solid line—sunlight, gray solid line—control, gray dotted line—UV exclusion. The total values shown represent the sum of total PFDs of the UV waveband (λ 280–400 nm) and the VIS waveband (λ 400–700 nm) in μmol m⁻² s⁻¹.
was used to deparaffinize the slides, which were then rehydrated through a graded series of ethanol. Slides were bathed in 0.1% Toluidine Blue stain, rinsed with distilled water, dehydrated, cleared, and mounted with Ultramount (Fronine, Sydney, NSW, Australia).

Slides were imaged using a light microscope (Olympus BX51; Olympus Imaging Australia, North Ryde, NSW, Australia) connected to a digital camera (Olympus DP70; Olympus Imaging Australia, North Ryde, NSW, Australia). Images were captured at 100× and 400× magnifications and were analyzed using PhotoShop software (Adobe Systems Inc., San Jose, California, U.S.A.). Measurements were made of abaxial and adaxial cuticle depth, abaxial and adaxial epidermal depth, and total leaf thickness.

**Ascorbate and Its Redox Status**

Ascorbate contents were analyzed using the method of Haberer et al. (2007). 20 mg of homogenized frozen leaf material was added to 500 μL of 5% metaphosphoric acid (MHO₄), vortexed, and centrifuged for 30 min (12,000 g, 4 °C). Supernatant (100 μL) was transferred to a new tube and neutralized with 20 μL of 1.5 M triethanolamine (TEA) and 100 μL of 150 mM Na-phosphate buffer. Reduced ascorbate was measured directly; total ascorbate was measured after complete reduction by DTT (50 μL, 10 mM, 15 min at room temperature). Excess DTT was removed with NEM (50 μL, 0.5%). Samples for reduced and total ascorbate analysis were then treated in the same way. We added 200 μL of 10% trichloroacetic acid (TCA), 200 μL of 44% ortho-phosphoric acid (HPO₄), and 200 μL of 4% 2,1-dipyridyl to tubes successively. Finally, 100 μL of 3% FeCl₃ was added and samples incubated in a water bath at 37 °C for 60 min. Of the final solution, 800 μL was transferred to a cuvette for analysis at 525 nm in a spectrophotometer (Beckman DU650, Beckman, Krefeld, Germany) as described by Schupp and Rennenberg (1988). Peaks were identified and quantified using a standard solution (0.2 mM cysteine, 0.1 mM g-glutamyl-cysteine and 1 mM glutathione in 0.01 M HCl) and concentrations normalized per unit fresh mass.

**Statistical Analyses**

Data were tested for normality and identification of outliers (SPSS Statistics Ver. 21; IBM Australia Ltd., St Leonards, NSW, Australia), and then two-way analyses of variance (ANOVA) were used to determine growth treatment effects and differences between species (Prism 6; Graphpad Software, La Jolla, California, U.S.A.). A paired t-test was used to determine the contribution of UV-A to photosynthesis (Prism 6).

**RESULTS**

**Leaf Gas Exchange**

Plants grown without exposure to UV exhibited faster rates of light-saturated photosynthesis and stomatal conductance than those exposed to UV ($F_{1,76} = 4.5; P < 0.05$, Fig. 2). Photosynthetic rates of *G. antrorsum* plants grown under UV exclusion were 19.7% faster than those grown in full sunlight ($P < 0.05$), corresponding with a 30% faster rate of stomatal conductance ($P < 0.05$, Fig. 2); *E. pauciflora* and *P. hiemata* showed similar trends.
Net photosynthetic rates of *P. hiemata* were ~60% slower than the other two species (*F*$_{1,76}$ = 99.2, *P* < 0.001). Intrinsic water use efficiency (WUE) varied significantly among species (*F*$_{2,76}$ = 5.9, *P* < 0.01) (Fig. 2). Responses of WUE to UV exposure were species-specific (interaction between species and UV treatment, *F*$_{2,76}$ = 99.2, *P* < 0.001); WUE was reduced in *P. hiemata* exposed to solar UV (*P* < 0.001), whereas *E. pauciflora* and *G. antrorsum* were unaffected (*P* > 0.05). No species was able to utilize UV-A to significantly increase photosynthetic rates (Fig. 3), with the maximum gain in photosynthetic rate from including UV-A in the measurement spectrum being 1.5–2% for *G. antrorsum* (*P* < 0.05).

**Fluorescence**

Fluorescence emission spectra of fresh leaves varied significantly among species (*F*$_{2,48}$ = 28.9, *P* < 0.001) (Fig. 4), indicating a potential disparity among species in the capacity of leaf surface chemistry to absorb UV-A. The spectra of *E. pauciflora* and *G. antrorsum* had identical peak profiles (λ = 423 nm, 445 nm, 460 nm, 486 nm, and 519 nm), indicating comparable chemical profiles, in contrast to *P. hiemata* that maintained a broad peak between λ = 410 nm to λ = 480 nm and sharp peaks at λ = 486 nm and λ = 519 nm. Total fluorescence emission, calculated as the area under the emission spectra, and capacity of epidermal cells to screen UV-A were least for *G. antrorsum* (both *P* < 0.001), with *E. pauciflora* leaves emitting the most fluorescence (70% greater than *G. antrorsum*) and *P. hiemata* best able to screen against UV-A (2.8-fold greater ability than *G. antrorsum*). Exposure to UV increased the capacity for epidermal screening against UV (*F*$_{1,48}$ = 6.2, *P* < 0.05, Table 1), with differences most clear in the grass *P. hiemata*, which displayed a 22% greater capacity to screen against UV-A when exposed to UV in the growth environment (*P* < 0.01).

Exposure to UV did not affect the chlorophyll content of leaves (*P* > 0.05; Table 1), although chlorophyll content did vary among species. *E. pauciflora* leaves had,
on average, 22% greater concentrations of chlorophyll a than G. antrorsum and 58% greater concentrations than P. hiemata (F = 27.0, P < 0.001). Fluorescence of Chl a (emission peak λ 685 nm and 730 nm) after excitation with UV-A wavelengths showed UV-A reached the mesophyll in all three species (Fig. 5), albeit the particular wavelengths able to excite Chl a varied among species (E. pauciflora: λ = 494.7, P < 0.001). Chl a was excited by wavelengths as low as 378 nm for G. antrorsum, whereas the capacity for UV-A to excite Chl a in E. pauciflora was negligible (λ > 395 nm). Exposure to UV during growth had no effect to excite Chl a in E. pauciflora.

### Table 1

Fluorescence emission peaks and ratios. Fluorescence emission peaks and peak ratios, using an excitation λ of 380 nm. Values represent means ± SEM for n = 9. Asterisks indicate significantly different means using a two-way ANOVA (⁎ P < 0.05, *** P < 0.001). Different lowercase letters represent significantly different means between species by Fisher’s LSD multiple comparison test (α = 0.05). Dot (•) indicates significantly different means between light treatments within species by Fisher’s LSD multiple comparisons test (α = 0.05).

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>E. pauciflora</th>
<th>G. antrorsum</th>
<th>P. hiemata</th>
</tr>
</thead>
<tbody>
<tr>
<td>Emission at λ 423 nm</td>
<td>6.4 ± 0.4</td>
<td>6.1 ± 0.5</td>
<td>a</td>
</tr>
<tr>
<td>Emission at λ 445 nm</td>
<td>5.0 ± 0.3</td>
<td>4.6 ± 0.3</td>
<td>a</td>
</tr>
<tr>
<td>Emission at λ 460 nm</td>
<td>4.1 ± 0.3</td>
<td>3.9 ± 0.3</td>
<td>a</td>
</tr>
<tr>
<td>Emission at λ 486 nm</td>
<td>4.0 ± 0.3</td>
<td>3.8 ± 0.2</td>
<td>a</td>
</tr>
<tr>
<td>Emission at λ 519 nm</td>
<td>6.4 ± 0.4</td>
<td>6.3 ± 0.3</td>
<td>a</td>
</tr>
<tr>
<td>Total fluorescence (area under spectra)</td>
<td>638 ± 39.5</td>
<td>605 ± 40.1</td>
<td>a</td>
</tr>
<tr>
<td>UV screening capacity (λ 440/685 nm)</td>
<td>5.8 ± 0.2</td>
<td>5.7 ± 0.2</td>
<td>a</td>
</tr>
<tr>
<td>Chlorophyll content (λ 685/720 nm)</td>
<td>1.8 ± 0.0</td>
<td>1.9 ± 0.1</td>
<td>a</td>
</tr>
</tbody>
</table>

Values represent means ± SEM (dashed lines) for n = 9.
fect on profiles of Chl $a$ excitation via UV-A ($P > 0.05$; Fig. 5).

**Leaf Anatomy**

Of the three study species, only the tree, *E. pauciflora*, had a waxy cuticle (Fig. 6; images shown are of leaves grown in full sunlight). Leaves of *E. pauciflora* were also more than twice as thick (463 µm) as those of the other two species (*G. antrorsum* 227 µm, *P. hiemata* 208 µm; $F_{2,73} = 155.3, P < 0.001$). The epidermis of *P. hiemata* was significantly thinner than that of the other two species ($F_{2,73} = 88.0, P < 0.001$), 13.9 ± 0.5 µm compared to 19.4 ± 0.6 µm in *E. pauciflora* and 21.6 ± 0.7 µm in *G. antrorsum*.

**FIGURE 5.** Spectral profiles of Chl $a$ fluorescence for fresh leaves of *E. pauciflora* (orange), *G. antrorsum* (orchid), and *P. hiemata* (green). Values represent excitation of the Chl $a$ emission peak at $\lambda$ 685 nm for plants grown under (a) full sunlight, and (b) UV exclusion; and of the Chl $a$ emission peak at $\lambda$ 730 nm for plants grown under (c) full sunlight, and (d) UV exclusion. Data shown are means ± SEM for $n = 9$. 
Adaxial epidermal cells were between 7% (P. hiemata) and 15% (G. antrosum) thicker than abaxial epidermal cells for all three species ($F_{2,73} = 75.4, P < 0.001$), consistent with a high light environment. Trichomes were present on the adaxial and abaxial surfaces of G. antrosum, but absent in the other two species. There were, however, no discernable anatomical differences between control and UV-exclusion plants in any species ($P > 0.05$).
Antioxidants and Their Redox Status

Foliar concentrations of ascorbate were, on average, 30% greater in plants exposed to UV ($F_{1,32} = 6.3, P < 0.05$; Fig. 7), albeit of unaltered redox state ($P > 0.05$). UV-induced ascorbate accumulation was greatest in *G. antrorsum*, with 50% greater concentrations in leaves grown in full sunlight than those grown without UV ($P < 0.05$). Concentrations of ascorbate differed among all three species ($F_{2,32} = 69.9, P < 0.001$), being greatest in *E. pauciflora* ($40.8 \pm 3.8 \mu mol \text{ g}^{-1} \text{ FW}$) and the least in *P. hiemata* ($6.5 \pm 1.7 \mu mol \text{ g}^{-1} \text{ FW}$). The ascorbate redox state of *P. hiemata* leaves was significantly greater than the other two species ($F_{2,32} = 31.5, P < 0.05$).

Leaf concentrations of glutathione (GSH), glutathione disulphide (GSSG) (Fig. 7), and the glutathione precursors cysteine (Cys) and γ-glutamylcysteine (γ-EC) (Table 2) were unaffected by exposure to solar UV during growth ($P > 0.05$) but varied considerably among species (GSH, $F_{2,42} = 30.2, P < 0.001$; GSSG, $F_{2,42} = 126.0, P < 0.001$; Cys, $F_{2,42} = 55.2, P < 0.001$; γ-EC, $F_{2,42} = 19.0, P < 0.001$; Fig. 7 and Table 2), with greatest levels of these compounds found in *G. antrorsum*. Glutathione redox state was also unaffected by solar UV ($P < 0.05$), but was much lower in *E. pauciflora* than the other two species ($F_{2,42} = 118.4, P < 0.001$).

**Discussion**

Our results show clearly that whereas the reported capacity of *P. ligustrina* to utilize UV-A for photosynthesis (Turnbull et al., 2013) is not necessarily shared by other subalpine Australian species, neither can UV wavelengths be regarded as universally detrimental to the functioning of these plants. These results add to the emerging body of literature showing natural levels of UV radiation are not necessarily damaging to plants.

Despite their contrasting anatomy and chemistry, all three of the study species demonstrated capacity to use
UV-A for photosynthesis as indicated by Chl a fluorescing at wavelengths less than 400 nm (Fig. 3). As for P. ligustrina (Turnbull et al., 2013), G. antrorsum displayed negligible blue-green fluorescence when leaf surfaces were exposed to UV-A (Fig. 2) and had no cuticle (Fig. 4). Both E. pauciflora and P. hiemata strongly fluoresced under UV-A, a feature associated with the ability to use UV-A for photosynthesis (Turnbull et al., 2013). Whereas the excitation fluorescence profiles (Fig. 3) suggest the grass P. hiemata should have the greatest capacity to utilize UV-A wavelengths for photosynthesis, both the emission profile (Fig. 2) and capacity for UV-A absorption (Table 1) suggest that Chl a was stimulated to fluoresce upon receipt of blue-green fluorescence rather than directly by UV-A photons (Johnson and Day, 2002; Mantha et al., 2001). Regardless of whether UV-A was directly or indirectly (via blue green fluorescence) used for photosynthesis, including UV-A in the photosynthetically excited spectrum increased photosynthetic gain but by less than 2%. Work with lower plants (red algae) has highlighted that contributions of UV-A to photosynthesis are more important close to dawn and dusk, when visible light is limited (Gao and Xu, 2008). In higher plants, contributions of UV-A under light-limiting conditions are less clear. Of some relevance are diurnal fluctuations in UV screening compounds (Barnes et al., 2008) that offer increased protection during the middle of the day but allow additional UV to penetrate to the mesophyll closer to nighttime.

Plants mostly adapt to UV exposure via modified structures and chemistry such that evaluating impacts of UV wavelengths on photosynthesis and carbon gain is complex for terrestrial ecosystems (Jansen et al., 1998; Jordan, 2002). Removing exposure to UV did not instigate any structural change in our three study species over the course of our study period. Although this contrasts with some reports (Grammatikopoulos et al., 1998), it is consistent with strong co-regulation of leaf anatomy by light intensity (Krauss et al., 1997; Liakopoulos et al., 2006), which is also intense in subalpine areas. Leaves of E. pauciflora have thick cuticles and thick epidermal layers and exhibit strong structural screening against UV (Day et al., 1992). These are traits typical of woody perennials (Jordan et al., 2005) that have evolved in response to more direct radiation than understory grasses and herbs (Brodersen and Vogelmann, 2007). The literature contains many records of exposure to UV altering the array of leaf photoprotective compounds by increasing “screens” such as flavonols and hydroxycinnamic acids (Wargent et al., 2015), which dissipate high-energy UV either as heat (Demmig-Adams and Adams, 2006) or by re-fluorescing it as lower energy photons (i.e., visible photons, Johnson et al., 2000). Even so there is wide variation among life forms (Day et al., 1992) and diurnally (Barnes et al., 2008).

One of our study species, the grass P. hiemata, emitted more blue-red fluorescence (an analogue for UV screening capacity, Lenk and Buschmann, 2006) when exposed to solar UV. Similar responses to non-UV wavelengths have been recorded in dicot species (Wargent et al., 2015). Our data diverges from those of Day et al. (1992), who found grasses of subalpine areas in North America less able than the dominant woody plants to screen against UV. P. hiemata is widely distributed in subalpine areas of SE Australia and is often the dominant species on lower, treeless plains within inverted treeline landscapes. Our data show that P. hiemata has more than twice the capacity to screen against UV than the tree E. pauciflora and more than three-fold the capacity of the herb G. antrorsum.

Whereas E. pauciflora and P. hiemata rely heavily on structural and chemical screens against UV, G. antrorsum does not. Herbaceous species generally show weaker screening capacity than other life forms (Day et al., 1992). A likely consequence is that significant amounts of UV reach the underlying mesophyll, stimulating generation of reactive oxygen species (ROS) (Czégeény et al., 2016) and potentially damaging cellular organelles involved with photosynthesis (Ivanova et al., 2008). Fittingly, G. antrorsum maintained the greatest concentrations of glutathione precursors. Concentrations of the glutathione precursors cysteine and γ-glutamylcysteine (γ-EC). Values represent means ± SEM for n = 15. Asterisks indicate significantly different means using a two-way ANOVA (*** P < 0.001). Different lowercase letters represent significantly different means between species by Fisher’s LSD multiple comparison test (α = 0.05).

<table>
<thead>
<tr>
<th>Redox component</th>
<th>E. pauciflora</th>
<th>G. antrorsum</th>
<th>P. hiemata</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Full sunlight</td>
<td>UV exclusion</td>
<td>Full sunlight</td>
</tr>
<tr>
<td>Cysteine (nmol g⁻¹ FW)</td>
<td>8.4 ± 1.10</td>
<td>5.7 ± 0.66 a</td>
<td>22.2 ± 2.54</td>
</tr>
<tr>
<td>γ-EC (nmol g⁻¹ FW)</td>
<td>3.7 ± 0.48</td>
<td>3.3 ± 0.31 a</td>
<td>6.0 ± 0.39</td>
</tr>
</tbody>
</table>

LSD multiple comparison test (α = 0.05).
Synthesized ascorbate upon exposure to UV (Foyer et al., 1994; Gao and Zhang, 2008; Kataria et al., 2012). Ascorbate and glutathione scavenge and detoxify ROS to prevent cellular damage (Foyer et al., 1994), ascorbate being the most responsive to light intensity (Grace and Logan, 1996; Peltzer and Polle, 2001). Whereas the constant oxidative state upon exposure to solar UV suggests the varying photoprotective strategies employed by each species prevent oxidative stress, we still observed reduced rates of photosynthesis for G. anstrosum plants exposed to UV. We speculate that this could be because of direct damage to photosynthetic reaction centers (PSII being particularly sensitive to UV, Ivanova et al., 2008), as has been found in other alpine ecosystems (Albert et al., 2010; Shi et al., 2011), or to diffusional limitations arising from reduced stomatal conductance that accompanies exposure to UV (Nogues et al., 1999), as illustrated by both G. anstrosum and E. pauciflora. Increased expression of the UV resistance locus8 (UVR8), a UV-B specific photoreceptor, has been used to explain decreased stomatal conductance following UV-B exposure (Brown et al., 2009). As UVR8 evolved with unicellular algae (Parihar et al., 2016) it is likely also present in monocots as well as dicots. We observed no regulation of g, by UV for the grass P. hiemata. Regardless, it appears diffusional limitations are unlikely to explain our decreased photosynthetic rate in G. anstrosum exposed to UV as WUE remained constant among treatments.

Conclusions

Recent studies show that exposure to solar UV can upregulate photosynthetic capacity for agricultural crops in temperate regions (Wargent et al., 2011, 2015; Davey et al., 2012). Here we demonstrate highly species-specific photobiological responses of native plants adapted to high UV exposure. Across three study species there were varying strategies to dissipate incoming UV radiation. Mild photoinhibition was recorded for one species, G. anstrosum, the species least structurally or chemically able to screen against UV wavelengths. We also found that increased photosynthetic rates do not necessarily follow the inclusion of UV-A wavelengths in the photosynthetic spectrum for plants that transmit UV wavelengths to the mesophyll, and finally, that UV exposure during growth appears an important regulator of plant water loss in dicots.

Acknowledgments

We thank Barry Aitchison for site access; David Coleman, Helen Liang, Svetlana Ryazanova, and Robin Shrestha for assistance with fieldwork; and Laura Leone for microscopy preparation. We also thank the staff and students at the lab of Prof. Rennenberg at the University of Freiburg for their assistance with redox analyses, with particular thanks to Dr. Jürgen Kreuzwieser, Monika Eiblmeier, and Ursula Scheerer.

References Cited


*MS submitted 26 February 2017*  
*MS accepted 25 August 2017*