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New Zealand *Pneumolaelaps* Berlese (Acari: Laelapidae): description of a new species, key to species and notes on biology

QING-HAI FAN¹, ZHI-QIANG ZHANG², ROBERT BROWN³, SANTHA FRANCE¹ & SHAUN BENNETT¹

¹ Plant Health & Environment Laboratory, Ministry for Primary Industries, 231 Morrin Road, Auckland, New Zealand. E-mail: Qinghai.Fan@mpi.govt.nz
² Landcare Research, Private Bag 92170, Auckland, New Zealand & School of Biological Sciences, The University of Auckland, Auckland, New Zealand. E-mail: zhangz@landcareresearch.co.nz
³ Landcare Research, PO Box 69040, Lincoln 7640, New Zealand.

Abstract

Mites of the genus *Pneumolaelaps* Berlese are often found in association with bumble bees (Hymenoptera: Apidae). Approximately sixty species of the genus have been described worldwide but only two of them have been recorded in New Zealand, viz. *Pneumolaelaps bombicolens* (Canestrini, 1885) and *P. breviseta* (Evans & Till, 1966). A new species, *Pneumolaelaps niutirani*, collected from hives of honeybee *Apis mellifera* and nests of the German wasp *Vespula germanica* (Vespidae) brings the number of species in New Zealand to three. We herein review *P. bombicolens* and *P. breviseta*, describe the new species, *Pneumolaelaps niutirani*, with notes on its biology, and provide a key to the species known in New Zealand. This is the first report of *Pneumolaelaps* in association with a vespid wasp.

Key words: Mesostigmata, *Apis mellifera*, *Bombus* spp., *Vespula germanica*

Introduction

The genus *Pneumolaelaps* Berlese, 1920 has had a history of unsettled classification for many years until the last two decades. The name *Pneumolaelaps* was originally proposed for a subgenus of the genus *Hypoaspis* by Berlese (1920), who designated *Iphis bombicolens* Canestrini, 1885 as its type species. This classification of *Pneumolaelaps* as a subgenus of *Hypoaspis* was followed for over five decades (e.g. Bregetova 1977). However, some authors considered it a separate genus (Willmann 1953; Hunter 1966). It is now classified in the subfamily Melittiphidinae of the family Laelapidae along with other genera that contain species closely associated with Hymenoptera (Casanueva 1993; Lindquist et al. 2009).

Mites of the genus *Pneumolaelaps* are common inhabitants of nests of bumblebees (Apidae) (Costa 1966; Hunter 1966; Hunter & Husband 1973; Macfarlane 2005; OConnor & Klimov 2012), honeybees (Apidae) (OConnor & Klimov 2012), the solitary bee *Megachile torrida* (Megachilidae) (Elsen 1973), and wasps (Vespidae) (present study). There are about 60 species described in the genus. Two species were previously recorded from New Zealand, *Pneumolaelaps bombicolens* (Canestrini, 1885) and *P. breviseta* (Evans & Till, 1966) (Macfarlane 2005). The main purpose of this paper is to describe a new species with notes on its biology and provide additional information on the two other species in New Zealand. We also give a key to species found in New Zealand to facilitate identification of these mites.
Material and methods

Taxonomy

Specimens were illustrated using a drawing tube attached to a Nikon interference-phase contrast microscope, and then re-examined, measured and imaged with a Zeiss interference-phase contrast microscope. Images were edited with Helicon Focus and Photoshop CS4. Lengths of legs were measured from the bases of trochanters to the tips of tarsal claws. All measurements are given in micrometers (μm). The chaetotaxy of the idiosoma and legs follows Evans & Till (1965) and Lindquist & Evans (1965).

Acronyms. BMNH: British Museum (Natural History), now The Natural History Museum (NHM), London, UK: NZAC: New Zealand Arthropod Collection: PANZ: Plant Health & Environment Laboratory, Auckland, New Zealand.

Observations on mite-host association

Two Vespa germanica (Fab.) colonies were obtained in April 2012 by excavating wild nests found in Lincoln, Canterbury, South Island, New Zealand. For each nest, adult specimens were separated by caste and held for up to 3–4 weeks in 3 L polycarbonate storage containers (H: 12 cm, L: 23, W: 15 cm) (Sistema, New Zealand) modified with a 5 x 12 cm ventilation hole on the short sides covered with stainless steel mesh (2-mm mesh). Remaining comb from nest excavations containing gyne and male pupae was kept with several workers to aid the adult emergence. As new gynes and males emerged, they were separated into new holding containers. The wasps were allowed to feed ad libitum from 20 mL of 30% sucrose water solution until they were used for experiments. Live and freshly-dead individuals were inspected under a dissecting microscope (Leica, Germany, model# M80) and mite numbers and location on the wasps were recorded. Living wasps were anesthetized with CO₂ before inspection. Thirty-five gynes, 17 workers, and 22 males were inspected.

Results

Pneumolaelaps Berlese, 1920

Type species: Iphis bombicolens Canestrini, 1885; by original designation.

Pneumolaelaps bombicolens (Canestrini, 1885)

(Figures 1, 9–14)

Iphis bombicolens Canestrini, 1885: 96.

Diagnosis. FEMALE: dorsal idiosomal shield (Fig. 1A) reticulate throughout, most cells as wide as long; bearing 42–46 pairs of setae, all smooth and tapered; setae j2–j3, z2–z4, s1–s4 and r2–r5 approximately twice as long as j1 and other setae. Presternal and sternal shields (Figs 1B, 9A) clearly separate; presternal shield with 3 transverse cells at each side; posterior margin of sternal shield obviously concave. Epigynal shield (Fig. 1B, 11A) medially reticulated with 6–8 large cells posterior to st5, margins of cells straight or curved but not sinuous; without a pair of longitudinal internal sclerites between st4. Peritrematal shield (Fig. 10A) broadly enlarged around stigmatic opening, where about as wide as length of poststigmatal plate; poststigmatal plate broadly triangular. Anal...
shield (Fig. 12A) approximately 1.5× as long as wide. Deutosternal groove (Fig. 1B, 14A) with 6 rows of 1–4 denticles per row. Setation on segments of legs same as that in *P. niutirani* sp. nov.

**FIGURE 1.** *Pneumolaelaps bombicolens* (Canestrini, 1884). Adult female (differential interference contrast microscope images). A, dorsum; B, venter.


**New Zealand: Auckland (AK):** 1 female, honey bee (*Apis mellifera*), 16.ix.2002, by A. Yearsley (PANZ, 03/2002/2545); 1 female, ex sticky board in honeybee (*A. mellifera*) hive, 14.vi.2005, by A. Wei (PANZ, 03/2005/1806); Birkenhead, 1 female, ex sticky board in honeybee...
(A. mellifera) hive, 1.vi.2005, A. Wei (PANZ, 03/2005/1749); Drury, 1 female, ex sticky board in honeybee (A. mellifera) hive, 14.vi.2005, by A. Wei (PANZ, 03/2005/1805); Howick, 5 females, ex sticky board in honeybee (A. mellifera) hive, 1.vi.2005, by A. Wei (PANZ, 03/2005/1739); Lynfield, 1 female, ex sticky board in honeybee (A. mellifera) hive, 14.vi.2005, by G. Matthews and A. Wei (PANZ, 03/2005/1829); Swanson, 2 females, ex sticky board in honeybee (A. mellifera) hive, 2005, by A. Wei (03/2005/1752); Waikowhai, 1 female, ex sticky board in honeybee (A. mellifera) hive, 14.vi.2005, by A. Wei (PANZ, 03/2005/1828); Waitakere, 1 female, ex sticky board in honeybee (A. mellifera) hive, 16.vi.2005, by A. Wei (PANZ 03/2005/1847); Whenuapai, 3 females on two slides, ex sticky board in honeybee (A. mellifera) hive, 14.vi.2005, by J. Green and A. Wei (PANZ, 03/2005/1830).

Bay of Plenty (BP): Bethlehem Road, 1 female, ex sticky board in honeybee (A. mellifera) hive, 2.v.2011, by H. Fergusson (PANZ, T11_02119).


Host: Apis mellifera, Bombus hortorum (?), Bombus ruderatus (?), Bombus terrestris.

Biology: Donovan (2007) found P. bombicolens (as Pneumonyssus bombicolens) from queens of two bumble bee species, Bombus terrestris and B. hortorum, and noted that one queen, one worker and two males of B. hortorum each carried one large mite, probably P. bombicolens. Large mites, most likely P. bombicolens, were found with collapsed eggs of bumble bee in the laboratory (Donovan 2007).

Remarks. It should be noted that bumble bee queens of at least three species were imported from England to New Zealand during 1883–1885 (Macfarlane & Griffin 1990). An adult female P. bombicolens collected from Nelson on 11 November 1949 indicates this species has been in New Zealand for at least sixty-six years.
**Pneumolaelaps breviseta** (Evans & Till, 1966)  
(Figures 2, 9–14)

*Hypoaspis breviseta* Evans & Till, 1966: 193

**Diagnosis.** FEMALE: dorsal idiosomal shield (Fig. 2A) marginally reticulate, ornamentation in medial area very faint, most cells as wide as long; bearing 42–46 pairs of setae (*px1–3* present), *j2–j3, z2–z4, s1–s4* and *r2–r5* about as long as *j1* and slightly longer than other setae. Presternal and sternal shields (Figs 2B, 9B) clearly separate; presternal shield with 3 faint transverse cells at each
side; posterior margin of sternal shield obviously concave. Epigynal shield (Fig. 2B, 11B) with 6–8 faint cells posterior to st5, margins of cells straight or curved but not sinuous; internal sclerites between st4 about the size of iv2. Peritrematal shield (Fig. 10B) broadly enlarged around stigmatic opening, where about as wide as length of poststigmatic plate; poststigmatic plate broadly triangular. Anal shield (Fig. 2B, 12B) 1.2×–1.5× as long as wide. Deutosternal groove (Fig. 14B) with 6 rows of 1–4 denticles per row. Setation on segments of legs same as that in P. niutirani sp. nov.


**Host:** Apis mellifera, Bombus muscorum, B. terrestris, B. hortorum.

**Distribution:** Britain (Evans & Till 1966), New Zealand (Macfarlane 2005; present study).

**Remarks.** Three adult females of this species were collected from Nelson on 11 November 1949. This indicates that this species has been in New Zealand for at least sixty-six years.

**Pneumolaelaps niutirani Fan & Zhang, sp. nov.**

(Figures 3–14)

**Diagnosis.** FEMALE: dorsal idiosomal shield (Figs 3A, 5A) strongly reticulate throughout, most cells wider than long; setae j2–3, z2–z4, s1–4 and r2–5 as long as or slightly longer and stouter than j1 and others. Presternal and sternal shields (Figs 4A, 5B, 9D) fused; posterior margin of sternal shield slightly convex. Epignyal shield (Figs 4A, 5B, 11D) medially reticulated with 6–8 large cells posterior to st5, margins of cells sinuous; a pair of longitudinal internal sclerites (Figs 4A, 5B) present between st4. Peritrematal shield (Figs 4A, 10D) broadly enlarged around stigmatic opening, where about twice width of poststigmatic plate; poststigmatic plate narrow and faint. Peritreme running anteriorly to posterior margin of coxa I. Anal shield (Figs 4A, 5B, 12D) about as long as wide. Deutosternal groove (Figs 4C, 14D) with 6 transverse rows of denticles, each row bearing 1–5 denticles. Peritrematal shield (Figs 4A, 10D) broadly enlarged around stigmatic opening, where about as wide as length of poststigmatic plate; poststigmatic plate narrow and faint. Peritreme running anteriorly to posterior margin of coxa I. Anal shield (Figs 4A, 5B, 12D) about as long as wide. Deutosternal groove (Figs 4C, 14D) with 6 transverse rows of denticles, each row bearing 1–4 teeth.

**Description.** FEMALE: Dorsal idiosomal shield (Figs 3A, 5A) oval, 505 (487–518) long and 328 (315–338) wide; reticulate throughout, most cells wider than long, no more than 4 cells between j6–j6 or J1–J1; idiosomal shield usually bearing 37 pairs of setae including 22 pairs of podonotals (j1–6, z1–6, s1–6 and r2–5) and 15 pairs of opisthonotals (J1–5, Z1–5 and S1–5); a single or a pair of setae (Jx1) may be present (9% chance) between J1 and J2, and/or a single or a pair of setae (Jx4) may be present (22% chance) behind or around J4 in some individuals, rarely with Jx3 and/or Zx5; all setae smooth and not reaching bases of setae in next row. Measurements (n=5): lengths: j1 18 (17–20), j2 26 (22–28), j3 23 (20–25), j4 18 (17–22), j5 17 (16–22), j6 17 (16–21); z1 15 (14–19), z2 23 (20–25), z3 24 (22–25), z4 22 (20–24), z5 18 (17–21), z6 20 (18–23); s1 22 (18–24), s2 25 (20–27), s3 24 (23–27), s4 22 (21–24), s5 22 (20–24), s6 23 (21–25); r2 21 (20–24), r3 24 (22–26), r4 25 (23–28), r5 22 (20–24); J1 17 (16–20), J2 17 (16–20), J3 16 (14–19), J4 15 (14–17), Jx4 15 (13–18), J5 19 (18–22); Z1 19 (17–22), Z2 20 (18–23), Z3 16 (14–19), Z4 18 (15–19), Z5 19 (16–22); S1 22 (17–23), S2 20 (18–22), S3 14 (13–18), S4 15 (13–18), S5 17 (15–20); distances: j1–j1 20 (18–23), j2–j2 22 (19–24), j3–j3 49 (44–51), j4–j4 53 (45–55), j5–j5 48 (45–51), j6–j6 63 (54–68); z5–z5 130 (120–
Dorsal shield with 15 pairs of discernible pore-like structures.

FIGURE 3. *Pneumolaelaps niutirani* Fan & Zhang, sp. nov. Adult female (line drawings). A, dorsum; B, chelicera; C, epistome; D, prodorsal setae; E, palp; F, palpal apotele.

Ventral idiosoma (Figs 4A, 5B). Tritosternum with plumose laciniae; tritosternal base (Fig. 4A) trapezoid-shaped, anterior base 12 (9–13), posterior base 22 (19–23), altitude 17 (15–19); laciniae (69–77) each with 8–11 pectinates. Sternal and presternal shields (Figs 4A, 5B, 9D) fused together; presternal area reticulated with transversally elongate cells; sternal shield with posterior margin medially convex, its anterior and lateral areas reticulated, postero-median region forming a triangular smooth area; reticulated cells elongate and compressed in anterior area; shield bearing 3 pairs of long setae st1–3 and 2 pairs of lyrifissures (iv1-2). Measurements (n=5): lengths: st1 51 (50–55), st2 58 (55–60), st3 58 (55–62); distances: st1–st1 58 (55–60), st1–st2 35 (33–39), st2–st2 92 (88–94), st2–st3 46 (44–48), st3–st3 119 (117–123); lyrifissures iv1 posteromedial to st1, iv2 posterolateral to st2. Setae st4 and lyrifissure iv3 on soft cuticle; endopodal plate medial of coxae III–IV separate from
sternal shield. Epigynal shield (Figs 4A, 5B, 11D) 223 (219–228) long and 94 (90–96) wide at level of st5, with hyaline anterior margin broadly rounded and posterior margin arched, reticulate throughout, with 6–8 large cells posterior to st5; margins of cells sinuous; bearing a pair of setae st5 on margins, and flanked by a pair of lyrifissures iv5 on soft cuticle posterolateral to st5; a pair of longitudinal internal sclerites (Figs 4A, 5B), 21 (18–22) long, 6 (5–7) wide, present between st4. Measurements (n=5): lengths: st4 42 (42–50), st5 49 (44–52). Spermathecal apparatus with tubulus annulatus widened near ramus sacculus (Fig. 4B), ramus sacculus slightly sclerotized. External metapodal platelets elongate, 37 (35–40) long and 13 (10–15) wide; inner metapodal platelets very small, 13 (12–14) long and 4 (3–5) wide; paragenital platelet tiny, beside seta ZV1. Peritrematal shield (Figs 4A, 10D) broadly enlarged, 29 (28–32) wide at level of stigmatic opening, about twice the width of peritreme; peritreme broad and free from exopodal platelets and dorsal shield, running

FIGURE 4. Pneumolaelaps niutirani Fan & Zhang, sp. nov. Adult female (line drawings). A, venter; B, tubulus annulatus and ramus sacculus; C, subcapitulum.
anteriorly to posterior margin of coxa I; poststigmatal plate reduced, 20 (19–22) long), shorter than width of peritrematal plate around stigmatic opening, bearing a pore. Anal shield (Figs 4A, 5B, 12D) inversely subtriangular, about as long as wide, length 83 (82–93) and width 85 (80–92); reticulated throughout; post-anal seta (21 (19–22)) as long as para-anal setae (22 (21–24)); cribrum with a transverse band of dense spicules. Soft cuticle with simple ventral setae, subequal in length. Setal lengths: JVI 34 (31–35), JV2 30 (29–33), JV3 27 (26–28), ZVI 33 (30–33), ZV2 27 (26–29), ZV5 17 (15–19).

FIGURE 5. Pneumolaelaps niutirani Fan & Zhang, sp. nov. Adult female (differential interference contrast microscope images). A, dorsum; B, venter.
Gnathosoma. Epistome (Figs 3C, 13D) subtriangular with anterior margin smooth and nearly rounded. Deutosternal groove (Figs 4C, 14D) with 6 transverse rows of denticles, preceded anteriorly and followed posteriorly by a smooth ridge devoid of teeth; typically first row bearing a large denticle, rows 2–5 each with 1–3 denticles; 6th row with 4 small denticles; corniculi horn-like, convergent; internal malae projecting medially, fringed laterally, labrum extending beyond tip of corniculi; hypostomal setae simple, inner posterior setae \( h_3 \) > palpcoxal setae \( pc \) > anterior setae \( h_1 \); lengths: \( h_1 \) 26 (24–27), \( h_2 \) 16 (14–16), \( h_3 \) 43 (40–45), \( pc \) 33 (31–34); distances: \( h_1 \)–\( h_1 \) 19 (18–21), \( h_3 \)–\( h_3 \) 27 (25–28), \( pc \)–\( pc \) 53 (50–55). Cheliceral fixed digit (Fig. 3B) bearing 2 medial teeth, followed by setiform pilus dentilis, then two large blunt teeth; dorsal seta nearly twice as long as pilus dentilis, posteriad of dorsal lyrifissure; movable digit with 2 medial teeth. Palpal setae (Fig. 3E) simple, \( al_1 \) and \( al_2 \) of genu slightly thickened, \( al_2 \) with hyaline sheath; setation of palp segments from trochanter to tarsus: 2—5—6— 12+2 solenidia—6+9 solenidia; palp tarsal claw 2-tined (Fig. 3F), with ventral tine (8–9 long) tapered and dorsal tine finger-shaped (12–14 long).

Legs (Figs 6A, 6B, 7A, 7B). Each pretarsus bearing a pair of claws with large membranous pulvillae. Legs I with tarsus (excluding pretarsus) 1.5× as long as tibia, about 2× for legs II–IV. Coxae without prominent serrations or spur-like processes on distal margins. Tarsus I without prominently elongated setae apically. Tarsi II–IV with apical set processes \( ad \) and \( pd \) long, as long as pretarsi (to the base of claws). All setae simple. Setation on segments of legs (I–IV): coxae 2—2—2—1; trochanters 6 (1 1/3 1)—5 (1 0/3 1)—5 (1 0/3 1); femora 13 (2 2/3 2)—11 (2 2/3 2/1)—6 (1 2/1 1/0)—6 (1 2/1 1/0); genua 13 (2 3/2 3/1)—11 (2 3/2 3/1)—9 (2 2/1 1/2)—10 (2 2/1 1/2)—10 (2 2/1 1/2)—8 (2 1/1 2/1)—10 (2 1/1 3/1 2); tarsi 30° setae and 9° solenidia—18—18—18.

Male and immature stages. Unknown.


**FIGURE 6.** *Pneumolaelaps niutirani* Fan & Zhang, sp. nov. Adult female (line drawings). A, leg I (trochanter-tibia); B, leg II (trochanter-tibia).

FIGURE 7. Pneumolaelaps niutirani Fan & Zhang, sp. nov. Adult female (line drawings). A, leg III (trochanter-tibia); B, leg IV (trochanter-tibia).
Holotype and a paratype will be deposited in NZAC. All other paratypes are deposited in PANZ and NZAC.

**Etymology.** The specific name, *niutirani*, is derived from the Maori “Niu Tirani”, meaning “New Zealand”, referring the locality from where the species were found. It is used here as a noun in apposition.

**Other specimen examined for comparison: Britain:** Cumberland: Gosforth, holotype female of *Pneumolaelaps minutissima*, ex *Bombus terrestris* queen, 3.v.1960, by D.C. Lee (NHM 1965:12:29:5).

**Remarks.** The new species is most similar to *P. minutissima* (absent from New Zealand) in the general appearance of the dorsal idiosomal shield. It can be easily distinguished from the latter and two New Zealand species, viz. *P. bombicolens* and *P. breviseta* by the characters listed in Table 1.

**FIGURE 8.** *Pneumolaelaps niutirani* Fan & Zhang, sp. nov. A, habitus; B, adult female dorsum (scanning electron microscope, SEM); C, ventral propodosoma (SEM).
TABLE 1. Distinguishing characters of Pneumolaelaps bombicolens (Pbo), P. breviseta (Pbr), P. minutissima (Pm) and P. niutirani sp. nov. (Pn).

<table>
<thead>
<tr>
<th>Character</th>
<th>Pbo</th>
<th>Pbr</th>
<th>Pm</th>
<th>Pn</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cells of reticulation of dorsal shield</td>
<td>j2, j3, z2–z4, s1–s4 and r2–r3</td>
<td>≈ 2x j1</td>
<td>≈ as long as j1</td>
<td>≈ as long as j1</td>
</tr>
<tr>
<td>Presternal and sternal shields (Fig. 9)</td>
<td>Separate</td>
<td>Strongly concave</td>
<td>Prominent, broadly triangular</td>
<td>Curved and plain</td>
</tr>
<tr>
<td>Posterior margin of sternal shield (Fig. 9)</td>
<td>Prominent, broadly triangular</td>
<td>Curved and plain</td>
<td>Absent</td>
<td>Distinctively longer than wide</td>
</tr>
<tr>
<td>Poststigmatic plate (Fig. 10)</td>
<td>Medially convex</td>
<td>Reduced, narrow and obscure</td>
<td>Sinuous</td>
<td>Present</td>
</tr>
<tr>
<td>Margins of cells on epigynal shield behind s3 (Fig. 11)</td>
<td>Curved</td>
<td>Sinuous</td>
<td>Present</td>
<td>Nearly as long as wide</td>
</tr>
<tr>
<td>Internal sclerites (Fig. 12)</td>
<td>Absent</td>
<td>Distinctively longer than wide</td>
<td>Nearly as long as wide</td>
<td>6 rows of 2–4 denticles</td>
</tr>
<tr>
<td>Deutostral groove (Fig. 14)</td>
<td>6 rows of 2–4 denticles</td>
<td>6 rows of 2–5 denticles</td>
<td>6 rows of 9–12 denticles</td>
<td>6 rows of 2–4 denticles</td>
</tr>
</tbody>
</table>

Biology. Pneumolaelaps niutirani sp. nov. is commonly found in association with the honeybee Apis mellifera and the German wasp Vespula germanica. In the sample collected from Canterbury in 2013, there was a high variation in the number of mites found among the individual adult wasps. Sixteen of the 35 gynes examined (46%) were found to have at least one mite on their body. Four of the male wasps (18%), had at least one mite on them at the time of inspection. There were no mites found on the 17 workers examined. The majority of the mites were found on the thorax and abdomen of V. germanica, and with few occurring on the head (Table 2).

TABLE 2. Summary of counts and locations of Pneumolaelaps niutirani sp. nov. found on Vespula germanica reproductive castes.

<table>
<thead>
<tr>
<th>Wasp caste</th>
<th>Location on body</th>
<th>Head</th>
<th>Thorax</th>
<th>Abdomen</th>
<th>Combined</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gynae (n=16)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean ± SEM</td>
<td></td>
<td>0.063 ± 0.063</td>
<td>9.88 ± 2.32</td>
<td>2.38 ± 0.73</td>
<td>12.31 ± 2.89</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>1</td>
<td>158</td>
<td>38</td>
<td>197</td>
</tr>
<tr>
<td>Male (n=4)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean ± SEM</td>
<td></td>
<td>0.25 ± 0.25</td>
<td>1.75 ± 0.48</td>
<td>0.75 ± 0.75</td>
<td>2.75 ± 1.44</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>1</td>
<td>7</td>
<td>3</td>
<td>11</td>
</tr>
</tbody>
</table>

FIGURE 11. Epigynal shield (differential interference contrast microscope images). A, P. bombicolens; B, P. breviseta; C, P. minutissima; D, P. niutirani.


Key to females of Pneumolaelaps known in New Zealand

1. Presternal and sternal shields separate; posterior margin of sternal shield concave; margins of cells on epigynal shield behind st5 straight or cured but not sinuous; cells of reticulation on dorsal shield about as wide as long .................................................................................................................................................. 2
P. niutirani Fan & Zhang, sp. nov.

2. Podonotal setae j2–3, z2–z4, s1–s4 and r2–s5 about as long as j1 and slightly longer than other setae; ornamentation in region of dorsal hexagon (j5-z5-j6) faint. 
P. breviseta Evans & Till, 1966

- Podonotal setae j2–3, z2–z4, s1–s4 and r2–s5 about twice as long as j1 and 2–4× as long as other setae; ornamentation in region of dorsal hexagon (j5-z5-j6) clearly distinguishable. 
P. bombicolens (Canestrini, 1885)


Discussion

Previous papers on the feeding behaviour of Pneumolaelaps provide scant information. Members of the genus mainly have been considered to be nectar and pollen feeders (Hunter & Husband 1973; Royce & Krantz 1989). Pneumolaelaps longanalis Hunter & Husband 1973, an associate of bumblebees, was known to feed on nectar and surface compounds of pollen (Royce & Krantz 1989). It is also known to feed on liquids from wounds of bumblebees, viz. the haemolymph of an accidentally decapitated bumblebee and the exposed air sac of a bumblebee (Hunter & Husband 1973). Pneumolaelaps hyatti (Evans & Till, 1966) was observed to feed on an acarid mite Kuzinia laevis (Dujardin, 1849) (Acaridae) (Costa 1966; Macfarlane 2005). The feeding habits of P. niutirani sp. nov. are unknown at the present. In this study, we reported the first case of Pneumolaelaps-Vespula association, but the nature of association remains to be studied. Many laelapid mites are very well preadapted to parasitism (Radovsky 1985). If P. niutirani sp. nov. were a true parasite of its host, then its offspring would be also found in the large number of samples from the hosts accumulated over the years. Despite the large number of specimens collected from bees, bumblebees and wasps over many years, only adult females of P. niutirani sp. nov. were found on adult bees and wasps. Thus adult females of this species likely mate with males and produce offspring off their hosts. In other mites associated with bees that commonly feed on pollen and nectar, similar observations have been made, e.g. mites of the family Ameroseiidae usually disperse via hosts as adult female mites.
(Evans 1963). Kar et al. (2015) also collected only females of *Neocypholaelaps novaehollandiae* Evans, 1963 (Ameroseiidae) from its insect hosts, but males and offspring were collected from flowers of plants where they feed on pollen and reproduce.

To understand the feeding habits of *P. niutirani* sp. nov., it will be useful to conduct direct observations on the feeding behaviour of the mite or obtain evidence of feeding via gut analysis (DNA). It will also be very useful to collect mites from soil and litter associated with the nests of wasps for the presence of mites.

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