Perpetuation of Avian Influenza in the Americas: Examining the Role of Shorebirds in Patagonia

Authors: Graciela Escudero, Vincent J. Munster, Marcelo Bertellotti, and Pim Edelaar
Source: The Auk, 125(2) : 494-495
Published By: American Ornithological Society
URL: https://doi.org/10.1525/auk.2008.2408.2
Perpetuation of avian influenza in the Americas: Examining the role of shorebirds in Patagonia.—Aquatic birds are considered the natural reservoir of all influenza viruses (Webster et al. 1992). Low-pathogenic avian influenza (AI) viruses have been isolated from Anseriformes (at least 36 species of ducks and 8 species of geese), Charadriiformes (10 species of shorebirds, 9 species of terns), and a few additional waterfowl species (Olsen et al. 2006).

Prevalence of AI virus is higher in fall than in spring in all studies of ducks (Anatidae) and gulls (Laridae) from Europe and North America (Hanson et al. 2003, 2005; Olsen et al. 2006; Munster et al. 2007; Wallensten et al. 2007). This temporal pattern of prevalence was attributed to the higher proportion of immunologically naive juveniles during fall migration. By contrast, the seasonal prevalence of AI viruses in shorebirds of North America showed a different picture. Krauss et al. (2004) found that prevalence of AI in shorebirds in the Delaware Bay area was 14.2% during northward spring migration but only 0.9% (15× lower) during southward fall migration. To explain this pattern, it has been proposed that an undetermined reservoir of AI viruses exists in South America where shorebirds may be exposed during the nonbreeding season and, subsequently, carry virus back north during spring migration (Krauss et al. 2004, 2007; Olsen et al. 2006; Wallensten et al. 2007).

Different hypotheses for the perpetuation of AI viruses have been proposed. Viruses could survive in frozen lakes, with reinfection of birds the following spring (Ito et al. 1995), or the AI-virus gene pool in aquatic birds may be perpetuated by low-level transmission among waterfowl throughout the year (Kawaoka et al. 1988, Krauss et al. 2004). Cold-temperate coastal and inland wetlands in southern South America that support large numbers of breeding and nonbreeding birds are, therefore, prime candidates to contribute to the South American virus reservoir hypothesized by Kraus et al. (2004, 2007).

We present results from 165 shorebirds sampled in freshwater and coastal wetlands in Chubut and Santa Cruz provinces in southern Argentinean Patagonia that were tested for all AI virus subtypes (see Fig. 1 for sampling locations). In this region, breeding and postbreeding Neotropical shorebirds and waterfowl are found in mixed flocks with nonbreeding Nearctic shorebirds. Five Neotropical and six Nearctic shorebird species with different migratory strategies were sampled between August 2004 and December 2005 (Table 1). Birds were captured using cannon nets, mist nets, and nest traps, or shot for other projects. Cloacal samples were collected using small sterile cotton swabs with a metal stem. The swabs were stored in transport media (Munster et al. 2007) and transported from the field in liquid

**Table 1.** Overview of the species sampled for AI virus, their migratory status, the environment in which the samples were taken, sample size per species, and 95% confidence interval (CI) for virus prevalence.

<table>
<thead>
<tr>
<th>Species</th>
<th>Migratory status</th>
<th>Environment</th>
<th>Sample size</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Magellanic Oystercatcher (H. leucopodus)</td>
<td>Neotropical migrant</td>
<td>Coast</td>
<td>20</td>
<td>0.00–11.7</td>
</tr>
<tr>
<td>American Oystercatcher (H. palliatus)</td>
<td>Neotropical resident</td>
<td>Coast</td>
<td>1</td>
<td>0.00–95.0</td>
</tr>
<tr>
<td>Least Seedsnipe (Thinocorus rumicivorus)</td>
<td>Neotropical migrant</td>
<td>Freshwater</td>
<td>2</td>
<td>0.00–77.6</td>
</tr>
<tr>
<td>Southern Lapwing (Vanellus chilensis)</td>
<td>Neotropical partial migrant</td>
<td>Freshwater</td>
<td>4</td>
<td>0.00–52.7</td>
</tr>
<tr>
<td>Two-banded Plover (Charadrius falklandicus)</td>
<td>Neotropical migrant</td>
<td>Coast</td>
<td>54</td>
<td>0.00–5.39</td>
</tr>
<tr>
<td>White-rumped Sandpiper (Calidris fuscicollis)</td>
<td>Nearctic migrant</td>
<td>Coast–freshwater</td>
<td>48</td>
<td>0.00–6.05</td>
</tr>
<tr>
<td>Pectoral Sandpiper (C. melanotos)</td>
<td>Nearctic migrant</td>
<td>Freshwater</td>
<td>2</td>
<td>0.00–77.6</td>
</tr>
<tr>
<td>Red Knot (C. canutus)</td>
<td>Nearctic migrant</td>
<td>Coast</td>
<td>1</td>
<td>0.00–95.0</td>
</tr>
<tr>
<td>Baird’s Sandpiper (C. bairdii)</td>
<td>Nearctic migrant</td>
<td>Freshwater–coast</td>
<td>24</td>
<td>0.00–11.7</td>
</tr>
<tr>
<td>Sanderling (C. alba)</td>
<td>Nearctic migrant</td>
<td>Coast</td>
<td>3</td>
<td>0.00–63.1</td>
</tr>
<tr>
<td>Lesser Yellowlegs (Tringa flavipes)</td>
<td>Nearctic migrant</td>
<td>Freshwater</td>
<td>6</td>
<td>0.00–39.3</td>
</tr>
</tbody>
</table>

**Fig. 1.** Sample locations in the Argentinean provinces Chubut and Santa Cruz.
nitrogen to Centro Nacional Patagónico (Pto. Madryn, Argentina), where they were stored in liquid nitrogen or at −20°C. All samples were shipped (at −20°C) to The Netherlands, arriving within 24 h (World Courier). RNA isolation and reverse-transcriptase polymerase chain reaction (RT-PCR) were performed at the Department of Virology of the Erasmus Medical Centre, Rotterdam, as described in Munster et al. (2005).

In contrast to predictions by Krauss et al. (2004) and Ito et al. (1995), all samples were negative for AI types, even though the technique for virus isolation used in the present study is more sensitive than techniques previously used for isolation in embryonated hens’ eggs (Gallus gallus domesticus; Munster et al. 2005). The probability of obtaining zero positive virus samples among 165 samples is \(1.06 \times 10^{-11}\) at the population infection rate of 14.2% found by Krauss et al. (2004) in migrating Nearctic shorebirds in spring (two-tailed test probability taken from binomial distribution; Sokal and Rohlf 1995). Using the binomial distribution to calculate the probability of finding zero positive samples when assuming a particular prevalence, we simulated the 95% confidence interval (CI) for AI prevalence across all species as 0.00–1.79% (CIs for each species—necessarily wider because of smaller sample sizes—are presented in Table 1). In support of our findings, AI virus was also not detected in another study that sampled 156 Red Knots (Calidris canutus rufa) in southern Argentina (D’Amico et al. 2007).

Our results show that neither Neotropical nor Nearctic shorebirds in southern Patagonia were shedding virus at the time of sampling. Hence, although the sample size is relatively small, we found no support for the hypothesis that Nearctic shorebirds become infected with AI at these high-latitude South American nonbreeding grounds.

The absence of AI viruses in shorebirds of southern Patagonia is an important result because of the potential role of this group of birds as reservoirs of AI viruses. More extensive surveillance of wild South American aquatic birds is required before making definitive statements, but our results may help narrow the search for AI viruses to other shorebird species that reach the area or to other stopover sites farther north.

Acknowledgments.—We are grateful to M. Abril, A. Farmer, M. and J. Torres Dowdall, J. L. Figetti, A. Nillni, and many others for assistance in the field. We thank R. Fouchier for fruitful discussions and sample analyses. Permits were issued by Dirección de Fauna Silvestre Santa Cruz (no. 406/05), Dirección de Fauna y Flora Silvestres de Chubut (nos. 10/04, 22/04, 02/05, 10/06), and Dirección de Áreas Protegidas Provincia del Chubut (no. 24/04). The export permit was issued by SENASA Argentina (no. 57863). G.E. is a member of Agencia Nacional de Investigaciones Científicas y Técnicas (ANPCyT). M.B. is a member of Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET). This research was financially supported by the Dutch Schure-Beijerinck-Popping Fonds.—Graciela Escudero, Centro Nacional Patagónico, Boulevard Brown 2825, U9120 ACF Puerto Madryn, Chubut, Argentina (e-mail: gescudero@cenpat.edu.ar); Vincent J. Munster, Department of Virology, Erasmus MC, Rotterdam, The Netherlands; Marcelo Bertellotti, Centro Nacional Patagónico, Boulevard Brown 2825, U9120 ACF Puerto Madryn, Chubut, Argentina; and Pim Edelaar, Department of Animal Ecology, Uppsala University, Sweden.

Literature Cited


Received 18 June 2007, accepted 15 December 2007