

Sex-specific Cortisol and Sex Steroids Responses in Stressed Sockeye Salmon during Spawning Period

Authors: Kubokawa, Kaoru, Yoshioka, Motoi, and Iwata, Munehiko

Source: Zoological Science, 18(7): 947-954

Published By: Zoological Society of Japan

URL: https://doi.org/10.2108/zsj.18.947

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Sex-specific Cortisol and Sex Steroids Responses in Stressed Sockeye Salmon during Spawning Period

Kaoru Kubokawa^{1*}, Motoi Yoshioka² and Munehiko Iwata³

¹Department of Marine Bioscience, Ocean Research Institute, University of Tokyo, Nakano, Tokyo, 164-8639 Japan.

²Faculty of Bioresources, Mie University, Tsu, Mie, 514-8507 Japan.

³School of Fisheries Sciences, Kitasato University, Sanriku, Iwate, 022-0101 Japan.

ABSTRACT—Response to stress during the spawning period of sockeye salmon (*Oncorhynchus nerka*) was investigated by measuring cortisol, selected steroids and glucose levels in the plasma of cultured fish under acute confinement conditions. Fish were individually placed in nets and sacrificed to obtain blood samples after each confinement period: 0, 3, 5, 15 and 30 min, respectively. In males, acute stress increased cortisol levels in the plasma at 15 min and testosterone, 11-ketotestosterone and glucose levels in the plasma within 3 min. In females, the cortisol level was higher than that in males and did not significantly change at 15 and 30 min. Testosterone levels in females decreased at 15 min in confinement. Progesterone, 17-hydroxyprogesterone and 17,20 β -dihydroxy-4-pregnen-3-one levels did not change with confinement time in females. To know the reproductive stage of the fish used in the stress experiment we collected blood and measured sex steroid hormones in different fish from the same pond one week and one day before the experiment. In these fish, the sex steroids in males and females changed with the pattern of breeding. Cortisol levels were higher in females than in males for one week. This study demonstrates that the cortisol level in females is higher than in males during their spawning period, and that females might be less sensitive to changes in the level of cortisol caused by acute stress than are the males.

Key Words: Stress, Cortisol, Sex steroid, Breeding season, Salmon

INTRODUCTION

Collection of blood samples from wild salmon is necessary to study their physiological conditions by measuring hormones and other chemicals in the blood. However, capture, handling, and bleeding of fish cause stress, and the stress induces changes in the circulating hormone levels (Greenberg and Wingfield, 1987; Carragher and Sumpter, 1990). In teleosts, cortisol is a major stress hormone and its plasma level increases are positively correlated with stress levels (Billard and Gillet, 1981; Pickering et al., 1982; Sumpter et al., 1986). Many studies of adult salmon also reported a correlation between plasma cortisol concentration and stress (Billard and Gillet, 1981; Pickering et al., 1982, Barry et al., 1995; Pickering and Pottinger, 1987a,b; Carragher et al., 1989; Carragher and Sumpter, 1990; Foo and Lam, 1993; Chopin et al., 1996; Jardine et al., 1996; Waring, et al., 1997, Carruth et al., 2000). Acute handling and confinement (stressors) of juvenile and immature Atlantic salmon for 3 hours is accompanied by an increase concentration of plasma cortisol (Carey and McCormick, 1998). During physiological changes such as a smoltification and sexual maturation, cortisol levels increase but androgen levels in plasma inversely decrease due to stress (Barry et al., 1995; Pickering et al., 1987). In mature salmon, stress caused by the administration of exogenous cortisol induced suppression of reproductive and gamete development (Carragher et al., 1989; Carragher and Sumpter, 1990; Foo and Lam, 1993). In mammals, the effect of stress on reduction of gonadal maturation and reproductive activity is induced by a decline in plasma concentrations of reproductive hormones. Starvation is one of the inducers of stress and strongly induces the gonadal degradation. However, migrating salmon maintain their reproductive activity even when they are exposed to the physiological stressors of osmotic regulation and starvation during the spawning period (Robertson et al., 1961; Fagerlund, 1967). Reproductive hormones for controlling reproduction in migrating salmon were produced and stimulated the reproductive phenomena of spawning during starvation (So et al., 1985; Dye et al., 1987; Truscott et al., 1986; Ikuta, 1996). The physiological mechanisms responsible for the differences in the reproductive response to stress between mammal and migrating salmon have not been determined yet.

FAX. +81-3-5351-6488.

E-mail: kubokawa@ori.u-tokyo.ac.jp

^{*} Corresponding author: Tel. +81-3-5351-6487;

In the present study, sockeye salmon were investigated to study the effects of acute stress on circulating levels of cortisol, sex steroids and glucose, which increase in relation to stress during the spawning period.

MATERIALS AND METHODS

Fish

Twenty-five mature male and 25 mature female sockeye salmon ($Oncorhynchus\ nerka$), reared in one of the ponds of the Nikko Branch of the National Research Institute of Aquaculture, Chugushi, Nikko City, were used for a stress experiment on 9 September 1993. The females were bearing ovulated eggs in the body cavity, and males contained milt, which could be stripped. Their body weights (mean \pm S.E.M.) were 725.0 \pm 106.4 g and 613.6 \pm 81.3 g in males and females, respectively. All fish were bled without anesthesia immediately after the confinement time. These fish had ovulated or spermiated but had not spawned. In addition, 10 female and 10 male cultured sockeye salmon were similarly sacrificed to monitor levels of cortisol, sex steroids and glucose in plasma on 2 and 8 September 1993 before the stress experiment. Their body weights (mean \pm S.E.M.) were 775.2 \pm 96.9 and 761.5 \pm 49.6 in males and females, respectively.

Stress experiment

A stress experiment was performed on cultured sockeye salmon

on 9 September 1993. The experimental design is detailed in a previous paper (Kubokawa *et al.*, 1999). Briefly it is described below. A small pool, $2.0 \text{ m} \times 0.5 \text{ m}$, was built near the pond. This size was large

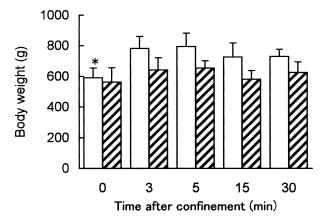


Fig. 1. Mean of body weights of sockeye salmon that were used in stress experiments at each confinement time. Males (open column) and females (shaded column) were used in groups of five. *Significantly different at P≦0.01 when compared between initial time and 3 min of confinement in males.

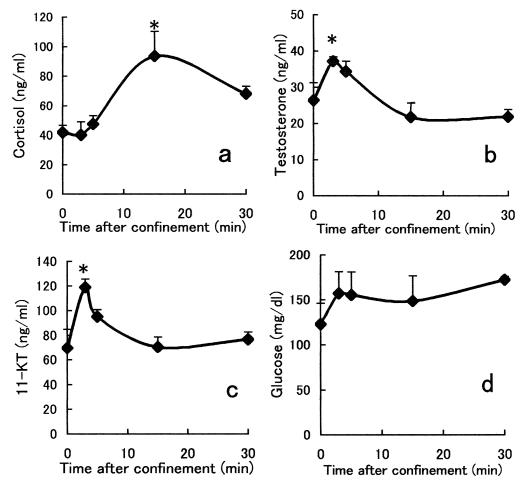


Fig. 2. Changes in plasma cortisol (a), testosterone (b), 11-ketotestosterone (11-KT) (c) and glucose (d) levels in male sockeye salmon exposed to the acute stress during 3, 5, 15 and 30 min. *Significantly different at P≦0.01 when compared with the hormone concentration at the initial time. Five fish were used for each confinement time.

enough to contain ten confinement nets used to hold the experimental fish. Pond water was circulated continuously through the pool. Bleeding of fish was performed at the edge of the pond. Soon after capture from the culture pond, each fish was confined in a small bag made of fishing net and put into the small pool. Blood was collected through the net within 0.5 min after the bag was removed from the pool. Five males and five females out of 50 fish were bled and then sacrificed to check for gonadal maturation after each confinement time, 0, 3, 5 15 and 30 min. One ml of blood was collected from the caudal vein of each fish with a 1 ml sterilized syringe. Blood samples

were immediately centrifuged and the resulting plasmas were stored at -80°C until radioimmunoassay of hormones and glucose measurement could be made. Unhandled fish, controls for the stress experiment, were corresponded to fish that was sacrificed at time zero without the confinement stress.

Measurement of cortisol, sex steroids and glucose

Cortisol and steroid levels were measured at the University of Tokyo by using antibodies established in previously published papers. For males, concentrations of cortisol, testosterone and 11-

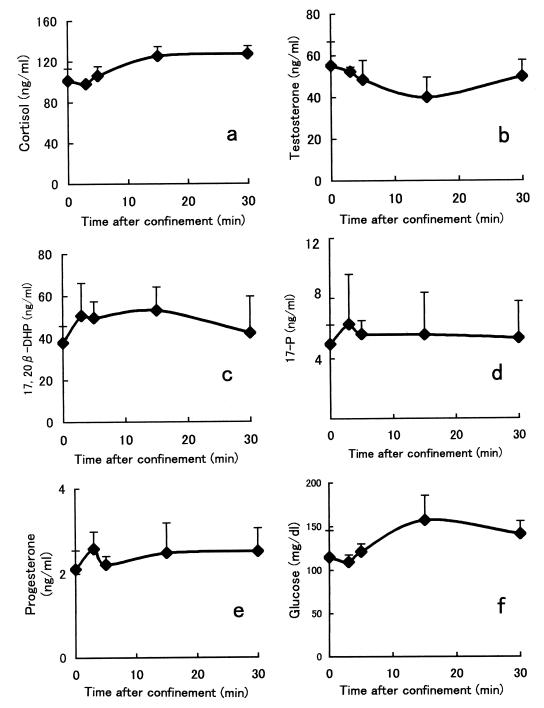


Fig. 3. Changes in plasma cortisol (a), testosterone (b), 17,20 β-dihydroxy-4 pregnen-3-one (17,20 β-DHP) (c), 17-hydroxyprogesterone (17-P) (d), progesterone (e) and glucose (f) levels in female sockeye salmon exposed to the acute stress during 3, 5, 15 and 30 min. *Significantly different at P≦0.01 when compared with the hormone concentration at the initial time. Five fish were used for each confinement time.

ketotestosterone (11-KT) were determined by radioimmunoassays. For females, concentrations of cortisol, estradiol-17 β (E₂), testosterone, 17-hydroxyprogesterone (17-P), and 17,20 β -dihydroxy-4-pregnen-3-one (17, 20 β -DHP) were determined by radioimmunoassays. The procedures of radioimmunoassays followed those in Takahashi *et al.* (1985) for cortisol, Lou *et al.* (1985) and Shimizu *et al.* (1985) for

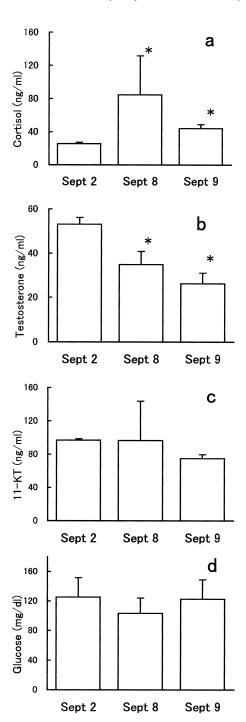


Fig. 4. Changes in mean plasma cortisol (a); testosterone (b), 11-ketotestosterone (11-KT) (c) and glucose (d) levels in unhandled male sockeye salmons on one day and one week before stress experiment. Results on 9 September correspond to values of time zero at the experiment. *Significantly different at P≦0.01 when compared with hormone levels on 2 September. The number of fish used was 10, 10 and 5 on September 2, 8 and 9, respectively.

testosterone, E $_2$, progesterone, 17-P and 17,20 β -DHP, and Lou *et al.* (1986) for 11-KT. Plasma E $_2$ levels in females were less than the minimal detectable value, 30 pg/ml. Intraassay and interassay coefficients of sex steroid hormones were described previously (Zairin *et al.*, 1992). For both males and females, the glucose concentration in plasma was determined by using a commercial kit, Glucose CII-test Wako (Wako Pure Chemical Inc. Ltd., Osaka). The linear range of standard glucose was 0.5 mg/ml to 4 mg/ml.

Statistics

The analysis of variance with one-way layout followed by Duncan's multiple range tests was employed to compare body weight, hormone and glucose levels between animal groups in the confined-stress experiment, and for comparison between unhandled salmon captured on different days.

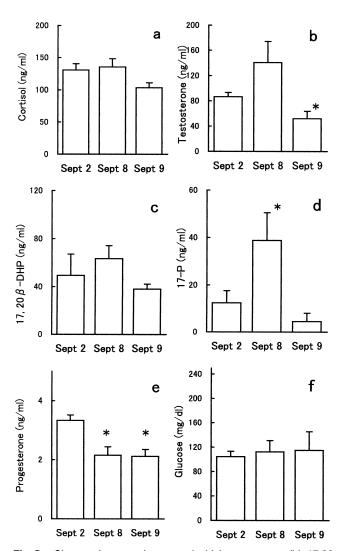


Fig. 5. Changes in mean plasma cortisol (a), testosterone (b), 17,20 β-dihydroxy-4 pregnen-3-one (17,20 β-DHP) (c), 17-hydroxyprogesterone (17-P) (d), progesterone (e) and glucose(f) levels in unhandled female sockeye salmons on one day and one week before stress experiment. Results on 9 September correspond to values of time zero at the experiment. *Significantly different at P≤0.01 when compared with hormone levels on 2 September. The number of fish was 10, 10 and 5 on 2,8 and 9 September, respectively.

RESULTS

Body weight

The body weight of males at initial time was significantly lower than for other groups of males (P≦0.01, Fig.1). It is suspected that the smaller size fish were easier to catch than the larger fish using the net method. A similar tendency was shown for females although it was not significant (Fig.1).

Cortisol, sex steroids and glucose in stressed fish

Fish were collected from the culture pond and immediately measured to determine the cortisol, sex steroids and glucose levels in plasma for each confinement time. The mean initial cortisol level in males (41.9±10.6 ng/ml) was significantly lower than that in females (101.5±18.3 ng/ml) (Fig.2a and 3a) (P≦0.01). The differences in cortisol levels between males and females appeared throughout the range of confinement times. Under the stress of confinement, cortisol levels increased at 15 min in males (increment at 5 min and 15 min being significant at P≤0.01) (Fig.2a), while the level slightly decreased at 30 min. In females, cortisol levels slightly increased after 15 and 30 min. (Fig.3a). The testosterone levels in males were significantly lower than those in females throughout the confinement time (Fig.2b and 3b) (P≤0.01). In males, the initial values of testosterone and 11-KT were 26.4 ±11.7 ng/ml and 69.7±35.1 ng/ml, respectively. Testosterone and 11-KT levels significantly increased at 3 min (P≤0.01) and gradually decreased with time and were retained at 30 min after confinement (Fig.2b and c).

In females, initial values of sex steroids were 55.3 \pm 27.2 ng/ml in testosterone, 37.9 \pm 7.8 ng/ml in 17,20 β -DHP, 4.5 \pm 3.6 ng/ml in 17-P, and 2.1 \pm 0.4 ng/ml in progesterone (Fig.3b, c, d and e). The increase of testosterone levels at 3 min in males was not observed in females, but the level gradually decreased with time as it did in the males. The mean levels of 17,20 β -DHP, 17-P, and progesterone did not show a significant increase nor decrease with time of the confinement stress (Fig. 3c, d and e). E $_2$ levels were lower than the detection limit of 30 pg/ml (data not shown). The plasma glucose levels increased at 3 min in males (Fig.2d), and gradually with time in females (Fig.3f).

Cortisol, sex steroids and glucose in unhandled fish

In blood samples were taken from unconfined fish, stress, cortisol, steroids and glucose levels were measured one week before and one day before in the acute stress experiment, in order to confirm the reproductive stage of these fish. The mean cortisol levels in males increased on 8 and 9 September (Fig. 4a). In females, no significant difference in the mean cortisol level was observed over three days. (Fig.5a). The mean cortisol level in females was higher than in males even on 8 September.

In males, the mean testosterone level was significantly decreased on 8 and 9 September (P≤0.01)(Fig.4b). The mean 11- KT and glucose levels were almost constant over three days (Fig.4c and d). In females, testosterone increased on 8

September and decreased on 9 September (Fig. 5b). There was no significant difference in increase of testosterone. The mean level of 17-P was significantly increased on 8 September and decreased on 9 September (P \leq 0.01)(Fig.5d). The mean levels of 17,20 β -DHP and glucose did not change throughout the three days (Fig.5c and f). The mean levels of progesterone significantly decreased on 8 and 9 September (P \leq 0.01) (Fig.5e).

Correlation Analysis

To confirm the generality of a relationship between cortisol and sex steroid levels exhibited in the confinement experiment, simple correlation coefficients between hormone levels of all fish used in the experiment were calculated and the significance of these correlations was tested. In males, the cortisol level was significantly and negatively correlated with testosterone level, and weekly and negatively with 11-KT level (P \leq 0.01). Testosterone and 11-KT levels were positively correlated well (P \leq 0.01) (Table 1). However, in females, no significant correlation was found between levels of cortisol and any of the sex steroids measured. Positive and week correlation was obtained between levels of testosterone and progesterone, testosterone and 17-P, and progesterone and 17,20 β -DHP (Table 2).

Table 1. Correlation coefficiencies of changes among cortisol, sex steroids, glucose and body weight in male

	Т	11-KT	Glucose	BW
Cortisol	-0.65	-0.54	0.2	0.006
T		0.84	-0.2	0.35
11-KT			-0.07	0.42
Glucose				0.45

T: testosterone, 11-KT:11-Ketotestosterone BW: body weight

Table 2. Correlation coefficiencies of changes among cortisol, sex steroids, glucose and body weight in female

	Т	DHP	Р	17-P	Glucose	BW
Cortisol	-0.26	0.11	0.1	-0.06	0.3	-0.18
T		0	0.58	0.41	0.19	0.3
DHP			0.42	0.38	-0.05	0.05
Р				0.34	0.35	0.13
17-P					-0.04	0.35
Glucose						0.08

T: testosterone, DHP:17, 20 β -dihydroxy-4-pregnen-3-one, P: progesterone, 17-P: 17-hydroxyprogesterone, BW: body weight

DISCUSSION

In the present study, we demonstrated the sex-specific response to acute stress in migrating salmon during the spawning period. The plasma cortisol level in mature female land-locked sockeye salmon had already reached the high level when challenged by confinement stress, and did not increase for 30 min into the experiment. However mature male sockeye salmon responded well to the acute stress with increas-

ing cortisol concentrations. In addition, we measured plasma cortisol concentrations in fish from the culture pond and the cortisol level in males was lower than in females before spawning. It was considered that the plasma cortisol concentration in females increased faster than in males, and that the sensitivity to stress in females is lower than in males before spawning. The high levels of cortisol in females suggested that the mechanism for synthesis of cortisol might cause damage on a part of the hypothalamus – pituitary – interrenal (HPI) axis at the end of a spawning period.

Carruth *et al.* (2000) reported that the plasma cortisol concentration in a landlocked subspecies of sockeye salmon increased in association with upstream migration, and that there were no statistically significant differences between the sexes at any stage of gonadal development. His result is different from our result with respect to the sex-specific response to stress. Two possible explanations are considered in for the differing results. First, the fish used in his study were acclimated for one week after they were caught in the river. Second, the time period in which blood samples were collected in his study was longer than that in ours, which was 0.5 min. These differences in experimental design may have influenced the cortisol level in either males or females.

In order to measure the cortisol concentration without stress, a cannulation system and a large culture pond would be needed to collect blood samples over time. However, it is very difficult to keep fish that have been cannulated in a large pond and sample their blood using this system. Accordingly, the base value used to measure hormone level changes in the acute stress experiment corresponded to the value at time zero, because a control fish is only stressed by the collection of blood at the beginning of the experiment.

When migrating salmon die after spawning, the cortisol level in both sexes had increased to extraordinary high levels. However, there is no reports that female cortisol levels are higher than males before spawning except for those in our previous paper (Kubokawa, et al., 1999). Many investigators have studied on the elevation of stress-induced cortisol in male and female teleost under various physiological and reproductive conditions (Bonga, 1997). The increase of plasma cortisol levels during a spawning migration is also considered to be cause by some sort of stress, for example, the stressors such as long-distance migration and change from sea water to fresh in ocean-run salmonid. However, Carruth et al. (2000) reported that landlocked sockeye salmon, which migrate in short-distance from lake to natal spawning river, showed the elevation of plasma cortisol levels during an upstream migration without the stressors described above. It is suggested that the elevation of cortisol in an upstream migration might be influenced greatly by an endogenous system. Our result, that is the different endocrine response to stress between females and males of landlocked sockeye salmon, also indicates the important role of an endogenous system. Females may have been stressed prior to the spawning by ovarian maturation and ovary maintenance. Another possible explanation may be that females are more severely stressed by the large volume of ovaries than that of testis. In the future, a study design to determine the effect of an artificial increase of gonadal volume on cortisol levels might be a productive approach.

It was reported that, when immature all-female salmon produced by triploid treatment and mature all-female salmon produced by diploid treatment were challenged with 2.5 hr of confinement stress, and cortisol and glucose levels in the immature salmon increased approximately twice as much as in the mature salmon (Sadler *et al.*, 2000). This result along with ours suggests that the bio-defense system against stress becomes insensitive in mature female salmon, and that an excess amount of stress stops the synthesis or secretion of additional cortisol internally.

Changes in plasma sex steroids from the beginning of sexual maturation to spawning were reported in upstream migrating sockeye salmon (Ikuta, 1996; Truscott et al, 1986). Moreover, Ikuta measured annual changes of steroid hormone levels in adult sockeye salmon at Nikko Branch of the National Research Institute of Aquaculture (unpublished data). In males, the testosterone level increased during the migration and the highest level was approximately 50 ng/ml when they gathered at the mouth of the stream. At the final stage of migration and before spawning, 11-KT elevated to approximately 80 ng/ml and testosterone level decreased at that time. In our study, the decrease of testosterone and the high and constant level of 11-KT consistent with his data. In females, maximum levels of testosterone, 17-P and progesterone were approximately 200 ng/ml, 25 ng/ml and 4 ng/ml, respectively before spawning, and then these steroids decreased in the spawning. In the spawning, 17,20 β-DHP was slightly increased to approximately 20 ng/ml. In our study, the significant increase of 17-P on 8 September may show the spawning condition, but no spent females were found on 8 and 9 September. Mean levels of the other steroids in females approximately consistent with his data. In comparison, males used in this study might be spermiated and females ovulated, but they were not spawned.

The inhibitory effects of stressors on reproductive performance have been established for many different teleost species (Bonga, 1997). However, the mechanism by which stress inhibits reproduction and levels in the endocrine cascade at which stress acts are less clear. In sexually mature males of brown and rainbow trout, acute confinement stress for 1 hr elevated plasma cortisol and ACTH levels and reduced plasma testosterone and 11-KT levels (Pickering et al., 1987; Pankhurst and Van Der Kraak, 2000). In sexually mature females of brown and rainbow trout, plasma testosterone levels were depressed by acute stress or treatment with exogenous cortisol (Pottinger et al., 1996; Pankhurst and Van Der Kraak, 2000). These androgen level findings are the same as those in other species. Furthermore, plasma levels of gonadotropin (GTH or GTH-II) were unaffected by stress, but the gonadotropin contents in the pituitary were decreased (Pickering et al., 1987; Carragher et al., 1989; Pankhurst and Van Der Kraak, 2000). On the other hands, implantation of testosterone or

11-KT reduced the elevation of the stress-induced plasma cortisol levels in both mature rainbow and brown trout (Pottinger, *et al.*, 1996; Young *et al.*, 1996). It is suggested that in mature males and females of salmonid the HPI axis regulates gonadal steroidogenesis, and gonadal steroids might regulate stress-induced activity of the HPI axis (Bonga, 1997; Pottinger *et al.*, 1996; Pankhurst and Van Der Kraak, 2000). However, our study in females indicated that the cortisol does not directly regulate gonadal testosterone secretion. Further experiment needs to reveal the direct effects of gonadal steroids on interrenal and indirect effects on interrenal function via the hypothalamus-pituitary axis.

Under stress, catecholamines act directly on the liver to stimulate glycogenolysis and result in mobilization of glucose (Axelrod and Reisine, 1984). Furthermore, glucose level is a correlate of corticosteroid level under an acute stress (Thomas and Neff, 1985; Vijayan et al., 1994; Rotllant and Tort, 1997; Vijayan et al., 1997). In our study, in females, testosterone and glucose levels increased without an increase in cortisol levels. It is more plausible that another stress hormone such as adrenaline is responsible for the glycogenolysis (Wingfield et al. 1992). About a manner of testosterone, we could discuss on the possibility of the existence of interrenal steroid, although there is no published data on it in teleost. In mammals, over 45 steroids including androgens have been isolated and chemically characterized from adrenal gland extracts, and studied on the function (Norman and Litwack, 1997). Recently, Fearon et al. (1998) reported that the differential control of adrenal androgens and cortisol in human. Thus, the existence of interrenal androgen may be needed to consider on the relationship between the function of HIP axis and stress in teleost.

ACKNOWLEDGEMENTS

We thank Mr. T. Watanabe, Hokkaido Prefectural Fish Hatchery, who was a student in Kitasato University for assisting with the radio-immunoassays performed at the University of Tokyo. We also thank Dr. J. Sullivan for helping with this manuscript. Mr. K. Muto and Mr. S. Oda, Nikko Branch of the National Institute of Aquaculture kindly cooperated and helped us by their technical assistance with the fieldwork. This study was supported by a grant from the Ministry of Education, Science, Sports, Culture and Technology of Japan and the Mishima Kaiun Foundation to K.K.

REFERENCES

- Axelrod J, Reisine TD (1984) Stress hormones: their interaction and regulation. Science 224: 452–459
- Barry TP, Malison JA, Lapp AF, Procarione LS (1995) Effects of selected hormones and male cohorts on final oocyte maturation, ovulation, and steroid production in walleye (*Stizostedion vitreum*). Aquaculture, 138: 331–347
- Billard R, Gillet C (1981) Stress, environment and reproduction in teleost fish. In "Stress and Fish" Ed by AD Pickering, Academic Press, New York, pp 185–208
- Bonga SEW (1997) The stress response in fish. Physiological Reviews 77: 591–625
- Carey JB, McCormick, SD (1998) Atlantic salmon smolts are more responsive to an acute handling and confinement stress than

- parr. Aquaculture 168:237-253
- Carragher JF, Sumpter, JP (1990) The effect of cortisol on the secretion of sex steroids from cultured ovarian follicles of rainbow trout. Gen Comp Endocrinol 77: 403–407
- Carragher JF, Sumpter JP, Pottinger TG, Pickering AD (1989) The deleterious effects of cortisol implantation on reproductive function in two species of trout, *Salmo trutta* L. and *Salmo gairdneri* Richardson. Gen Comp Endocrinol 76: 310–321
- Carruth LL, Dores RM, Maldonado TA, Norris DO, Ruth T, Jones RE (2000) Elevation of plasma cortisol during the spawning migration of landlocked kokanee salmon (*Oncorhynchus nerka kennerlyi*). Comp Biochem Phys C 127:123–131
- Chopin FS, Arimoto T, Inoue Y (1996) A comparison of the stress response and mortality of sea bream *Pagrus major* captured by hook and line and trammel net. Fisheries Res 28: 277–289
- Dye HM, Sumpter JP, Fagerlund UHM, Donaldson EM (1987) Changes in reproductive parameters during the spawning migration of pink salmon, *Oncorhynchus gorbuscha*. J Fish Biol 29: 167–176
- Fagerlund UHM (1967) Plasma cortisol concentration in relation to stress in adult sockeye salmon during the freshwater stage of their life cycle. Gen Comp Endocrinol 8: 197–207
- Fearon U, Clarke D, McKenna TJ, Cunningham SK (1998) Intra-adrenal factors are not involved in the differential control of cortisol and adrenal androgens in human adrenals. Eur J Endocrinol 138: 567–573
- Foo JTW, Lam TJ (1993) Serum cortisol response to handling stress and the effect of cortisol implantation on testosterone level in the tilapia, *Oreochromis mossambicus*. Aquaculture 115: 145–158
- Greenberg N, Wingfield JC (1987) Stress and reproduction: Reciprocal relationships. In "Hormones and Reproduction in Fishes, Amphibians, and Reptiles" Ed by DO Norris, RE Jones. Plenum Press, New York, pp 461–503
- Ikuta K (1996) Effects of steroid hormones on migration of salmonid fishes. Bul Nat Res Inst Aquaculture 0(Suppl 2) 23–27
- Jardine JJ, Van Der Kraak GJ, Munkitrick KR (1996) Capture and confinement stress in white sucker exposed to bleached kraft pulp mill effluent. Ecotoxicol Environ Safety 33: 287–298
- Kubokawa K, Watanabe T, Yoshioka M, Iwata M (1999) Effects of acute stress on plasma cortisol, sex steroid hormone and glucose levels in male and female sockeye salmon during the breeding season. Aquaculture 172: 335–349
- Lou SW, Aida K, Hanyu I, Sakai K, Nomura M, Tanaka, M, Tazaki S (1984) Endocrine profiles in the females of a twice-annually spawning strain of rainbow trout. Aquaculture 43: 13–22
- Lou S.W, Aida, K, Hanyu I, Sakai K, Nomura M, Tanaka M, Tazaki S (1986) Endocrine profiles in the males of a twice-annually spawning strain of rainbow trout, *Salmo gairdneri*. Gen Comp Endocrinol 64: 212–219
- Norman AW, Litwack G (1997) Steroid hormones: Chemistry, biosynthesis, and metabolism. In "Hormones", Academic Press, San Diego, pp 49–86
- Pankhurst NW, Van Der Kraak G (2000) Evidence that acute stress inhibits ovarian steroidogenesis in rainbow trout *in vivo*, through the action of cortisol 117: 225–237
- Pickering AD, Pottinger TG (1987a) Lymphocytopenia and interrnal activity during sexual maturation in the brown trout, *Salmo trutta* L. J Fish Biol 30: 41–50
- Pickering AD, Pottinger TG (1987b) Poor water quality suppresses the cortisol response of salmonid fish to handling and confinement. J Fish Biol 30: 363–374
- Pickering AD, Pottinger TG, Carragher J, Sumpter JP (1987) The effects of acute and chronic stress on the levels of reproductive hormones in the plasma of mature male brown trout, *Salmo trutta* L. Gen Comp Endocrinol 68: 249–259
- Pickering AD, Pottinger TG, Christie P (1982) Recovery of the brown trout, Salmo trutta L., from acute handling stress: a time course

- study. J Fish Biol 20: 229-244
- Pottinger TG, Carrick TR, Hughes SE, Balm PHM (1996) Testosterone, 11-ketotestosterone, and estradiol-17 β modify baseline and stress-induced interrenal and corticotropic activity in trout. Gen Comp Endocrinol 104: 284–295
- Robertson OH, Krupp MA, Thomas SF, Favour CB, Hane S Wexler BC (1961) Hyperadrenocorticism in spawning migratory and nonmigratory rainbow trout (*Salmo gairdnerii*); comparison with pacific salmon (genus *Oncorhynchus*). Gen Comp Endocrinol 1: 473–484
- Rotllant J, Tort L (1997) Cortisol and glucose responses after acute stress by net handling in the sparid red porgy previously subjected to crowding stress. J Fish Biol, 51: 21–28
- Sadler J, Wells RMG, Pankhurst PM, Pankhurst NW (2000) Blood oxygen transport, rheology and haematological responses to confinement stress in diploid and triploid Atlantic salmon, *Salmo salar*. Aquaculture 184:349–361
- Shimizu A, Aida K, Hanyu I (1985) Endocrine profiles during the short-reproductive cycle of an autumn-spawning bittering, *Acheilognathus rhombea*. Gen Comp Endocrinol 60: 351–371
- So YP, Idler DR, Truscot, B. and Walsh, JM (1985) Progesterones, androgens and their glucuronides in the terminal stages of oocyte maturation in landlocked Atlantic salmon (*Salmo salar*). J Steroid Biochem 23: 583–592
- Sumpter JP, Dye HM, Benfey TJ (1986) The effect of stress on blood ACTH, α -MSH and cortisol levels in salmonid fishes. Gen Comp Endocrinol 62: 377–385
- Takahashi A, Kubota J, Kawauchi H, Hirano T (1985) Effects of Nterminal peptide of salmon proopiocortin on internal function of the rainbow trout. Gen Comp Endocrinol 58: 128–135

- Thomas P, Neff JM (1985) Plasma corticosteroid and glucose responses of pollutants in striped mullet: different effects of naphthalene, benzo(a)pyrene and cadmium exposure. In "Marine pollution and physiology: Recent advances" Ed by FP Thurberg, A Calabrese, FJ Vernberg, WB Vernberg Univ. of South Carolina Press, Colombia, pp 63–82
- Truscott B, Idler DR, So YP, Walsh JM (1986) Maturational steroids and gonadotropin in upstream migratory sockeye salmon. Gen Comp Endocrinol 62: 99–110
- Vijayan MM, Pereira C, Grau EG, Iwama GK (1997) Metabolic responses associated with confinement stress in tilapia: The role of cortisol. Comp Biochem Physiol C 116: 89–95
- Vijayn MM, Pereria C, Moon TW (1994) Hormonal stimulation of hepatocyte metabolism in rainbow trout following an acute handling stress. Comp Biochem Physiol C 108: 321–329
- Waring CP, Stagg RM, Poxton MG (1996) Physiological responses to handling in the turbot. J Fish Biol 48: 161–173
- Wingfield JC, Vleck CM, Moore MC (1992) Seasonal changes of the adrenocortical response to stress in birds of the Sonoran desert. J Exp Zool 264: 419–428
- Young G, Thorarensen H, Davie PS (1996) 11-Ketotestosterone suppresses interrenal activity in rainbow trout (*Oncorhynchus mykiss*). Gen Comp Endocrinol 103: 301–307
- Zairin MJr, Asahina K, Furukawa K, Aida K (1992) Plasma steroid hormone profiles during HCG induced ovulation in female walking catfish *Clarias batrachus*. Zool Sci 9: 607–617

(Received October 6, 2000 / Accepted June 15, 2001)