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Source: Zoological Science, 20(4) : 449-459

Published By: Zoological Society of Japan

URL: https://doi.org/10.2108/zsj.20.449
Cloning of the Genes for the Pituitary Glycoprotein Hormone α and Follicle-Stimulating Hormone β Subunits in the Japanese Crested Ibis, *Nipponia nippon*

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ABSTRACT—We have isolated a part of the gene for the pituitary glycoprotein hormone common α subunit (PGHα) and the whole gene for the follicle-stimulating hormone β subunit (FSHβ) in the Japanese crested ibis (*Nipponia nippon*), a critically endangered bird species in East Asia. The nucleotide sequence of a part of the PGHα gene (5026 bp) contained three exons holding the whole coding and 3' untranslated regions, but lacked a 5' untranslated region. Its exon-intron structure was similar to that in mammals, but different from that in teleosts in the location of the second intron. For the FSHβ gene, the nucleotide sequence of 7633 bp was assembled from two phage clones. The exon-intron structure of three exons and two introns was similar to that observed in mammals and teleosts. In the putative promoter region of the ibis FSHβ gene, a progesterone responsive element (PRE)-like sequence and two AP-1 responsive element-like sequences reported in the ovine FSHβ gene were not conserved in complete form. The increased number of ATTAA motifs in the putative 3' untranslated region in comparison with those in Japanese quail and chicken FSHβ cDNA suggested that more rapid degradation of FSHβ mRNA occurs in this species. Deduced amino acid sequences of the ibis PGHα and FSHβ showed high similarities with those of the corresponding subunits of other avian species. This is the first report on the genomic sequences of the PGHα and FSHβ in an avian species.

Key words: gonadotropin, PGHα, FSHβ, gene, the Japanese crested ibis

INTRODUCTION

Follicle-stimulating hormone (FSH) and luteinizing hormone (LH) are the main members of gonadotropins and they also belong to the pituitary glycoprotein hormone family which also includes thyroid-stimulating hormone (TSH). All these hormones are heterodimeric molecules composed of a common α subunit and a hormone-specific β subunit. The α and β subunits are encoded by different genes and synthesized as separate peptides. So far, the genes encoding the gonadotropin subunits have been characterized in mammals and teleosts. As for gonadotropin subunits in avian species, cDNA sequences have been reported in Japanese quail (Ando and Ishii, 1994; Kikuchi *et al.*, 1998), chicken (Noce *et al.*, 1989; Foster *et al.*, 1992; Shen and Yu, 2002), turkey (Foster and Foster, 1991; You *et al.*, 1995) and two species of duck (Hsieh *et al.*, 2001), and amino acid sequences have been chemically determined in ostrich (Koide *et al.*, 1996). However, genomic information such as exon-intron structure and regulatory sequences in the 5' flanking region has not been available in avian species. Accordingly, cloning of avian gonadotropin subunit genes will provide us important information on the evolution of gonadotropin genes and their regulation.

The Japanese crested ibis *Nipponia nippon* (order Ciconiiformes, family Threskiornithidae) is a critically endangered species in East Asia. This species was formerly widespread throughout Japan, Korea, China and an adjacent part in southeastern Russia. At present, the only wild population of approximately 170 individuals survives in Yang Xian, Shaanxi Province, China. In addition, captive populations are kept in China and Japan. Since 1980s, Ishii and his collaborators (Ishii, 1999; Wingfield *et al.*, 2000) have developed endocrinological methods for the artificial breeding of
endangered birds. They have shown that hormone therapy using gonadotropins from closely related species in reproductive quiescent female Japanese quails resulted in gonadal maturation, egg-laying and production of reproductive active offspring (Wakabayashi et al., 1992; Ishii, 1999). Thus, homologous gonadotropin administration is considered to be effective in stimulating reproductive activity in the ibis. However, chemical isolation of gonadotropins from the ibis or related species for the hormone therapy program is almost impossible. Rather, it seems practical to clone the genes for the gonadotropin subunits of the ibis and to generate large quantities of gonadotropins using the recombinant technique.

Thus, cloning of the genes for the gonadotropin subunits in the Japanese crested ibis is invaluable because it provides not only the first information on the genomic structure of these subunits in avian species, but also the molecular information which is necessary for recombinant gonadotropins. Here, we report the isolation and characterization of a nucleotide sequence of a part of the gene for the pituitary glycoprotein hormone common $\alpha$ subunit (PGH$\alpha$) and the sequence of the whole gene for the follicle-stimulating hormone $\beta$ subunit (FSH$\beta$) in the Japanese crested ibis.

**MATERIALS AND METHODS**

**Isolation of the ibis PGH$\alpha$ gene**

When the last male Japanese crested ibis of the Japanese origin (named Midori) died in 1995, most of his organs were preserved in liquid nitrogen (Ishii, 1999). Genomic DNA was extracted from approximately 15 mg of the preserved kidney using a GenomicPrep Cells and Tissue DNA Isolation Kit (Amersham Biosciences, NJ). The extracted DNA was digested with EcoRI and separated in agarose gel electrophoresis. DNA fragments of 4–10 kb long were recovered from the gel and ligated with the Lambda ZAPII vector (Stratagene, CA). An original library of 2.5 x 10$^6$ clones was generated and amplified once to 1.3 x 10$^{11}$ clones. Approximately 2.5 x 10$^5$ clones in the amplified library were screened by plaque hybridization. To prepare a hybridization probe, the PGH$\alpha$ cDNA fragment of Japanese quail (Ando and Ishii, 1994) was randomly labeled with $\alpha$-32PdCTP (Amersham Biosciences, UK) and used as a probe. Prehybridization was performed at 42°C for more than 2 hr with denatured salmon sperm DNA (0.2 mg/ml) in a hybridization buffer containing 6 x SSC (1 x SSC: 150 mM NaCl, 15 mM sodium citrate, pH 7.0), Denhardt's solution (0.02% Ficoll, 0.02% bovine serum albumin, and 0.02% polyvinylpyrrolidone), and 0.1% SDS. Hybridization was carried out at 50°C overnight, and membranes were washed once with 3 x SSC containing 0.1% SDS at 50°C for 20 min. Following these steps, three positive clones were isolated. Based on identification by restriction enzyme digestion, a clone (IF4) was selected and a region of this insert was subcloned into the pBluescript phagemid vector (Stratagene, CA) for sequencing.

Because IF4 was found to cover only a 5' part of the FSH$\beta$ gene, screening was repeated to obtain the remaining 3' part. The plFPCR insert was digested with BamHI and then a fragment of approximately 180 bp was recovered to exclude a part overlapping with IF1. This fragment was randomly labeled with $\alpha$-32PdCTP and used as a probe. Prehybridization was performed as described above, and hybridization was carried out at 60°C overnight. Membranes were washed once with 1 x SSC containing 0.1% SDS at 40°C for 20 min and once with 0.1 x SSC containing 0.1% SDS at 60°C for 20 min. After third round of screening, one positive clone (IF1) was isolated. The insert fragment was digested with EcoRI and subcloned into the pBluescript phagemid vector (Stratagene, CA) for sequencing.

Because IF1 was found to cover only a 5' part of the FSH$\beta$ gene, screening was repeated to obtain the remaining 3' part. The plFPCR insert was digested with BamHI and then a fragment of approximately 180 bp was recovered to exclude a part overlapping with IF1. This fragment was randomly labeled with $\alpha$-32PdCTP and used as a probe. Since the genomic library did not give any positive signal, we constructed other genomic library with the Lambda EMBL3 vector and BamHI-digested genomic DNA. The resultant library was 1.6 x 10$^5$ clones and was screened without amplification. Hybridization was carried out at 50°C overnight, and membranes were washed once with 3 x SSC containing 0.1% SDS at 50°C for 20 min. Consequently, three positive clones were isolated. Based on identification by restriction enzyme digestion, a clone (IF4) was selected and a region of this insert was subcloned into the pBluescript phagemid vector. Nucleotide sequence was determined with a BigDye Terminator v3.0 Ready Reaction Cycle Sequencing Kit and an ABI PRISM 377 DNA Sequencing System (Applied Biosystems, CA).

**Analysis of the second intron in the ibis FSH$\beta$ gene in two individuals**

Nucleotide sequences in the second intron of the FSH$\beta$ gene were determined in two individuals. One was Midori of Japanese origin, and the other was Long-Long which had been lent from China to Japan and died in Japan in 1994. The genomic DNA of Long-Long was extracted from the cryo-preserved kidney (Ishii, 1999). Genomic DNA extraction, PCR using the primers FSH-F and FSH-R, cloning and sequencing were performed as described above. After sequencing, the PCR products were digested with EcoRI and subjected to electrophoresis.

**RESULTS**

**Isolation and nucleotide sequence of the ibis PGH$\alpha$ gene**

Ten positive clones were isolated from the genomic library and nucleotide sequence of a clone (pIA1) was determined (Fig. 1). The determined sequence of 5026 bp was compared with PGH$\alpha$ cDNA sequences of turkey (Foster and Foster, 1991), chicken (Foster et al., 1992), Japanese quail (Ando and Ishii, 1994) and two species of duck (Hsieh et al., 2001). Comparison results showed remarkable
sequence similarities which enabled us to infer the exon-intron structure (Fig. 2). We could recognize three putative exons containing the whole coding region and 3' untranslated region within this sequence, but we were unable to find a sequence corresponding to most of 5' untranslated region. Exon-intron junctions were determined according to the GT-AG rule (Breathnach and Chambon, 1981). Then, amino acid sequence was deduced and aligned with those of PGHα of turkey (Foster and Foster, 1991), chicken (Foster et al., 1992), Japanese quail (Ando and Ishii, 1994), ostrich (Koide et al., 1996), and two species of duck (Hsieh et al., 2001) (Fig. 3). Three amino acid residues in the putative ibis PGHα precursor were different from those in each of the other birds over a signal peptide of 24 residues, while 96 residues in a mature protein were completely identical to those of the other birds except for ostrich (ostrich mature protein differed in three amino acid residues from those of the other birds). Ten cysteine residues forming disulfide bonds and two putative N-linked glycosylation sites were conserved among all these birds. These findings assured that the acquired nucleotide sequence encoded the ibis PGHα precursor molecule.

All the PGHα genes previously reported in mammals and teleosts consist of four exons and three introns (Fiddes and Goodman, 1981; Goodwin et al., 1983; Burnside et al., 1988; Gordon et al., 1988; Kato et al., 1990; Golos et al., 1991; Huang et al., 1992; Suzuki et al., 1995). The exon-intron structure in the ibis PGHα gene was compared with that of carp (Huang et al., 1992), mouse (Gordon et al., 1988), bovine (Goodwin et al., 1983) and human (Fiddes and Goodman, 1981) (Fig. 4). The result indicated that the clone we obtained lacked a 5' part corresponding to the first exon of other vertebrates. By aligning amino acid sequences, locations of the introns in the coding region were compared. The location of the first intron in the part of the ibis PGHα gene was identical with that of the corresponding intron (the second intron) in mammals, but different from that in teleosts with an amino acid residue (Fig. 5), while the location of the second intron (the third in mammals and teleosts) was conserved through the ibis, mammals and teleosts.

Isolation and nucleotide sequence of the ibis FSHβ gene

Strategy to obtain the ibis FSHβ gene is illustrated in Fig. 6. The PCR gave a single band of approximately 800 bp and the determined 780 bp sequence was compared with FSHβ cDNA sequences of Japanese quail (Kikuchi et al., 1998) and chicken (Shen and Yu, 2002). This sequence was found to contain an intron of 582 bp flanked by partial exons of 23 and 175 bp of the putative FSHβ gene. A clone (IF1), which was isolated from the genomic library, contained an insert of approximately 10 kb, of which the nucleotide sequence of 4059 bp around the 3' part was determined. Comparison of the sequence with the FSHβ cDNA sequences of the two bird species revealed that IF1 contained a 5' part of the FSHβ gene, but lacked its 3' part. Then, three clones were isolated by repeated screening of the other genomic library. A partial nucleotide sequence of a clone (IF4), 3580 bp in size, was compared with the FSHβ cDNA sequences of the two bird species. IF4 was shown to contain a 3' part of the FSHβ gene which was missing in IF1.

By collecting the results from the two phage clones (IF1 and IF4) and the PCR fragment (pIFPCR), a sequence of 7633 bp was acquired (Fig. 7). Exon-intron structure was inferred by sequence similarities with the FSHβ cDNA in Japanese quail and chicken and exon-intron junctions were determined based on the GT-AG rule (Breathnach and Chambon, 1981). Particularly, we inferred the putative transcriptional start site and the polyadenylation site referring to the 5' and 3' end of the chicken FSHβ cDNA determined by rapid amplification of cDNA end (RACE) method. Finally, the assembled nucleotide sequence was found to contain 2820 bp of the 5' flanking region followed by three exons of 34, 160 and 2549 bp, separated by two introns of 458 and 582 bp. An amino acid sequence was deduced and aligned with those of Japanese quail (Kikuchi et al., 1998), chicken (Shen and Yu, 2002) and ostrich (Koide et al., 1996) (Fig. 8). The ibis amino acid sequence showed remarkable similarities to those of FSHβ of Japanese quail (94.7%) and chicken (95.4%). High similarity (93.4%) was also observed with the ostrich FSHβ except for a deletion of five residues in the C-terminus. Twelve cysteine residues and two putative N-linked glycosylation sites were conserved through these four bird species. These results enabled us to conclude that the nucleotide sequence we obtained encoded the FSHβ gene of the Japanese crested ibis.

The ibis FSHβ gene showed a structure with three exons and two introns. The similar exon-intron structure has been previously reported in mammals and teleosts (Jansen et al., 1988; Kim et al., 1988; Gharib et al., 1989; Hirai et al., 1990; Guzman et al., 1991; Kumar et al., 1995; Sohn et al., 1998; Rosenfeld et al., 2001). Locations of the first intron in the 5' untranslated region and the second intron between amino acid residue 33 and 34 of mature protein in the ibis FSHβ gene were also similar to locations reported for mammalian and teleostean counterparts. Comparison of the nucleotide sequence in the 5' flanking region between the ibis and non-avian species (teleosts and mammals)
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Met Gly Cys Tyr Gly Lys Tyr Ala Ala Val Thr Leu Thr Ile Ser Val Phe Leu His Leu
-20 -10

CTT CAT GCT TCC CCA GAT GGA GAG TTT CTC ATG CAG GGA acagtctcatgtgtctgcatgtagtcacgtctcattctctttgattttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttttt
Fig. 2. Nucleotide sequence of the part of the PGHα gene in the Japanese crested ibis. Exons are shown in capital letters, and introns and flanking regions are shown in lowercase letters. The polyadenylation signal, AATAAA, is underlined. Deduced amino acid residues are represented with three letter codes below the nucleotide sequence and numbered sequentially from the N-terminus. The nucleotide sequence data in this figure is available in the EMBL/GenBank/DDBJ Data Bank with Accession No. AB089503.

(A)

ibis  NGCGYKAVATLGVFLHILLHA  
duck  .D....S  
chicken  .D....T  
quail  .D....T  
turkey  .D....T  

(B)

ibis  FPFPFLMQGCEPLGENRFSSKPGAPIYQC  
duck  . . . .  
chicken  . . . .  
quail  . . . .  
turkey  . . . .  

Fig. 3. Alignment of amino acid sequences of signal peptides (A) and mature proteins (B) of the PGHα precursor molecules of the Japanese crested ibis, duck (Hsieh et al., 2001), chicken (Foster et al., 1992), Japanese quail (Ando and Ishii, 1994), turkey (Foster and Foster, 1991) and ostrich (Koide et al., 1996). Residues identical to those in the Japanese crested ibis are indicated with dots. Ten conserved cysteine residues and two putative N-linked glycosylation sites are denoted by ˔ and ˝ respectively.

Fig. 4. Comparison of the exon-intron structure of the part of the PGHα gene in the Japanese crested ibis with those of PGHα gene of carp (Huang et al., 1992), mouse (Gordon et al., 1988), bovine (Goodwin et al., 1983) and human (Fiddes and Goodman, 1981). Solid and open boxes represent the coding and the untranslated regions of exons, respectively. Solid lines indicate introns and flanking regions. Thin broken lines between boxes link corresponding exons. A thick broken line in the Japanese crested ibis represents part whose nucleotide sequence is not available at present.

Fig. 5. Alignment of a part of amino acid sequences of PGHα for comparison of location of the second intron. Locations of the intron are shaded. Numbers at the top represent positions of amino acid residues in the PGHα in the Japanese crested ibis. References are as follows; carp (Huang et al., 1992), salmon (Suzuki et al., 1995), ibis (Fiddes and Goodman, 1981), rat (Burnside et al., 1988), mouse (Gordon et al., 1988), bovine (Goodwin et al., 1983), porcine (Kato et al., 1990), and human (Fiddes and Goodman, 1981).

Fig. 6. Structure and sequencing strategy of the FSHβ gene in the Japanese crested ibis. Insert fragments of pIFPCR, IF1 and IF4 are shown with arrows which indicate the direction and extent of sequencing. At the bottom, the structure of ibis FSHβ gene is shown with exons indicated as boxes. Solid and open boxes represent coding and untranslated regions, respectively. Restriction sites for BamHI are indicated with B.
revealed that the ibis sequence could be well aligned with the sequences in mammals within the promoter region (Fig. 9).

**Analysis of the second intron of the ibis FSHβ gene in two individuals**

Results of sequencing the PCR fragments of Midori and Long-Long showed that two types of sequences were present in the second intron of the FSHβ gene (Fig. 10). The difference between the two types was the 32 bp sequence in the 5' portion. Ten and five clones were sequenced for Midori and Long-Long, respectively. Only the shorter type was observed in Midori, whereas both the shorter (in two clones) and longer (in three clones) types were found in Long-Long. In addition to the length difference, three nucleotide substitutions were observed. A substitution, which was located in the most 3' position, changed the sequence "GAATTC" (the recognition site of EcoRI) in the shorter type into "CAATTC" in the longer type. Utilizing this sequence difference, we could differentiate the two types by EcoRI digestion of the PCR products. Namely, the PCR product containing only the shorter type (approximately 780 bp) was divided into two fragments of 550 and 230 bp, while the PCR product of the longer type (approximately 810 bp) was not cleaved. The digestion results showed that only two fragments of 550 and 230 bp were observed for Midori and three fragment of 810, 550 and 230 bp for Long-Long, indicating that Midori had only the shorter type and Long-Long had both types.

**DISCUSSION**

In the present study, we have isolated and characterized a nucleotide sequence of a part of the PGHα gene and FSHβ Genes in *Nipponia nippon* revealed that the ibis sequence could be well aligned with the sequences in mammals within the promoter region (Fig. 9).
the sequence of the entire FSHβ gene in the Japanese crested ibis. This is the first isolation and characterization of gonadotropin subunit genes in avian species. The exon-intron structure of the ibis PGHx and FSHβ genes was basically similar to that of the corresponding gene in mammals and teleosts. This result suggests that the same exon-intron structure exists generally through all avian species. However, there was a minor difference among animal groups. It was the location in the second intron (corresponding to the first intron in the partial ibis gene) in the PGHx was the location in the second intron (corresponding to the second exon-intron boundary). At present, the genomic structure of lungfish PGHx has not been clarified, it may provide some information to explain the location difference of this intron.

In the ibis FSHβ gene, we deduced the location of the putative transcriptional start site and the polyadenylation site referring to the 5' and 3' end of the chicken FSHβ cDNA (Shen and Yu, 2002). However, two TATAAA sequences in the putative promoter region and five potential polyadenylation signals in the putative 3' untranslated region were observed (Fig. 7). The proximal TATAAA sequence, which was 27 bp apart from the putative transcriptional start site, was conserved through the ibis and mammals, whereas the distal sequence was unique to the ibis and did not exist in mammals (Fig. 9). Then, the proximal sequence is considered to be more possible than the distal one, but the other possibility that the distal sequence is utilized cannot be excluded. Further studies are required to obtain a definite conclusion of the transcriptional start site and the polyadenylation site of the ibis FSHβ gene.

Kikuchi et al. (1988) and Shen and Yu (2002) have reported that five ATTAA motifs were found in the 3' untranslated region of the FSHβ cDNA in Japanese quail and chicken, respectively. This motif has been characterized as a signal for rapid degradation of mRNA coding for cytotkine or proto-oncogenes (Shaw and Kamen, 1986; Chen et al., 1995). We found eight ATTAA motifs in the putative 3' untranslated region of the ibis FSHβ gene. Akashi et al. (1994) have demonstrated that the number of this motif in the 3' untranslated region correlates with instability of mRNA using the chimeric rabbit β-globin gene. It is possible that the increased number of this motif in the ibis FSHβ gene induces more rapid degradation of the mRNA than the other birds.

The nucleotide sequence in the 5' flanking region of the ibis FSHβ gene showed no appreciable similarity to those in teleosts and mammals apart from those in the promoter region in mammals (Fig. 9). In the promoter region of the ovine FSHβ gene, a progesterone responsive element (PRE)-like sequence and two AP-1 responsive element-like sequences have been demonstrated to be responsible for progesterone and gonadotropin-releasing hormone (GnRH) regulation of the gene expression, respectively (Webster et al., 1995; Huang et al., 2001). In the bovine and porcine FSHβ genes, these sequences were completely identical, but several substitutions occurred in each of the sequences in the rat and human FSHβ genes. Function of these sequences in rat and human has not been clear. The corresponding sequences in the ibis had several substitutions from the mammalian sequences and the consensus sequence. These results suggest a possibility that the ibis and other vertebrates (teleosts and mammals) apply different regulation mechanisms to the FSHβ gene expression. Further analysis on the 5' flanking region will reveal the mechanism of ibis FSHβ gene expression.

Among avian species we compared, the amino acid sequences of the PGHx and FSHβ showed remarkable similarities. According to the taxonomy, extant birds are classified into two lineages, the Palaeognathae and the Neognathae. The Palaeognathae includes ostrich, and the Neognathae consists of two clades; the first clade contains two orders of Galliformes (chicken, Japanese quail and turkey) and Anseriformes (duck), and the second clade contains the rest orders (van Tuinen et al., 2000). Our study has

(A)

-20 MRTWCYVLLFCWKA2CCNG
ibis
chicken ...L.............Y.
quail ...L.............Y.

(B)

+1 CQLTMITAVEREECFC1TVNATWCSGYCFTRDPY
ibis
chicken .E.............L.
quail .E.............L.
ostrich .E.............L.

+38 N..................K.
ibis
chicken .E.............L.
quail .E.............L.
ostrich .E.............R.A.L...

+111 G........................N.
ibis
chicken .G........................N.
quail .G........................N.
ostrich .G........................N.

Fig. 8. Alignment of the amino acid sequences of signal peptides (A) and mature proteins (B) of FSHβ precursor molecules of the Japanese crested ibis, chicken (Shen and Yu, 2002), Japanese quail (Kikuchi et al., 1998), and ostrich (Koide et al., 1996). Residues identical to those in the Japanese crested ibis are indicated with dots. Twelve conserved cysteines and two putative N-linked glycosylation sites are denoted by △ and ○, respectively.
Fig. 9. Comparison of nucleotide sequences of the promoter region of FSHβ genes of the Japanese crested ibis, ovine (Guzman et al., 1991), bovine (Kim et al., 1988), porcine (Hirai et al., 1990), rat (Gharib et al., 1989) and human (Jameson et al., 1988). The nucleotide sequence is numbered from the putative transcriptional start site in the ibis FSHβ gene. A progesterone responsive element (PRE)-like sequence (Webster et al., 1995) and two AP-1 responsive element-like sequences (Huang et al., 2001) reported in ovine FSHβ gene are boxed with solid line with consensus sequence attached in parenthesis. Parts in which TATAAA sequences are observed in the ibis are shaded.

<table>
<thead>
<tr>
<th>Species</th>
<th>Nucleotide Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ibis</td>
<td>TTTTGT-CT-TAAACAGGCTTGACTCCATGCTGTCGCTAGAATTACCTCACATTCCACCA</td>
</tr>
<tr>
<td>Ovine</td>
<td>TAGACTACTTCCTTTGCTAAGGCTTGACTCCATGCTGTCGCTAGAATTACCTCACATTCCACCA</td>
</tr>
<tr>
<td>Bovine</td>
<td>TAGACTACTTCCTTTGCTAAGGCTTGACTCCATGCTGTCGCTAGAATTACCTCACATTCCACCA</td>
</tr>
<tr>
<td>Porcine</td>
<td>TAGACTACTTCCTTTGCTAAGGCTTGACTCCATGCTGTCGCTAGAATTACCTCACATTCCACCA</td>
</tr>
<tr>
<td>Rat</td>
<td>TAGACTACTTCCTTTGCTAAGGCTTGACTCCATGCTGTCGCTAGAATTACCTCACATTCCACCA</td>
</tr>
<tr>
<td>Human</td>
<td>TAGACTACTTCCTTTGCTAAGGCTTGACTCCATGCTGTCGCTAGAATTACCTCACATTCCACCA</td>
</tr>
</tbody>
</table>

Fig. 10. Nucleotide sequences of the second intron of the FSHβ gene in the Japanese crested ibis, (L) the longer type and (S) the shorter type. Dashes indicate a gap inserted to maximize identity. Three nucleotide mismatches are marked with a reverse triangle. Part recognized by EcoRI in the short type is boxed.
provided the first information on the amino acid sequences of the PGHx and FSHβ in a bird from the second clade of the Neognathae. High similarities of the PGHx and FSHβ among ibis, chicken, Japanese quail and ostrich indicate that FSH is generally highly conserved over wide range of avian species.

Lastly, we found two (the shorter and longer) types of the sequence in the second intron of the FSHβ gene in the Japanese crested ibis. The shorter type was observed in both of two individuals we used, Midori of the Japanese origin and Long-Long of the Chinese origin, whereas the longer type was found only in Long-Long. These results suggested that geographic divergence occur in this species. At present, we are extending the sequence analysis of this intron to other individuals of both the Japanese and Chinese origins. Sequence analysis of the mitochondrial DNA of this species is also proceeding.

ACKNOWLEDGMENTS

This work is partly supported by Research Fellowships of the Japan Society for the Promotion of Science for Young Scientists.

REFERENCES


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(Received December 17, 2002 / Accepted January 14, 2003)